



Article **Do New-Generation Recycled Phosphorus Fertilizers Increase the Content of Potentially Toxic Elements in Soil and Plants?**

Magdalena Jastrzębska^{1,*}, Marta K. Kostrzewska¹, Agnieszka Saeid² and Wiesław P. Jastrzębski³

- ¹ Department of Agroecosystems and Horticulture, Faculty of Agriculture and Forestry, University of Warmia and Mazury in Olsztyn, Plac Łódzki 3, 10-718 Olsztyn, Poland; marta.kostrzewska@uwm.edu.pl
- ² Department of Engineering and Technology of Chemical Processes, Faculty of Chemistry, Wroclaw University of Science and Technology, Wyspiańskiego 42, 50-376 Wrocław, Poland; agnieszka.saeid@pwr.edu.pl
- ³ Department of Botany and Nature Protection, Faculty of Biology and Biotechnology, University of Warmia and Mazury in Olsztyn, Plac Łódzki 1, 10-727 Olsztyn, Poland; w.jastrzebski@uwm.edu.pl
- Correspondence: jama@uwm.edu.pl

Abstract: Phosphorus (P)-rich secondary raw materials can provide a valuable base for modern mineral fertilizers, provided that the new formulations do not load the soil–plant system with potentially toxic elements. Fertilizers from sewage sludge ash (SSA) and/or animal bones, activated by phosphorus-solubilizing bacteria (*Bacillus megaterium* or *Acidithiobacillus ferrooxidans*), were tested in field experiments in north-eastern Poland. The reference provided treatments with superphosphate and treatment without phosphorus fertilization. In one experiment, all P-fertilizers were applied at a P dose of 21 kg·ha⁻¹, and in the other three experiments, three P doses were adopted: 17.6, 26.4, and 35.2 kg·ha⁻¹. The effect of recycled fertilizers on the content of arsenic (As), chromium (Cr), nickel (Ni), copper (Cu), and zinc (Zn) in the soil, in wheat grain and straw (test plant), weeds, and post-harvest residues was investigated. The application of recycled fertilizers in P amounts up to 35.2 kg·ha^{-1} did not change the As, Cr, Ni, Cu, or Zn contents in the soil and plant biomass. The contents of these elements in soil were below the permissible levels for arable land in Poland. Their concentrations in wheat grain and straw did not exceed the permissible or suggested limits for plant material to be used for food and feed, while in the weed and post-harvest residue biomass, they usually fell within the biological plant variability ranges.

Keywords: phosphorus recovery; secondary raw materials; waste management; microbial solubilization; biofertilizers; heavy metals

1. Introduction

Although agriculture is the major food supplier for the growing world population, it is also one of the economic sectors with the largest environmental impact [1]. Among others, different studies have revealed a higher trend of accumulation of toxicants in different spheres of the environment, posing harmful effects to aerial, aquatic, and terrestrial health due to the injudicious applications of agrochemicals [2]. Potentially toxic elements (PTE), including heavy metals and metalloids, are at the top of the list of environmental toxicants from agricultural activities [3].

PTE are known to be naturally occurring elements [4], but they become dangerous when their concentration in different environmental compartments exceeds the prescribed levels [3,5]. Certain PTE, such as Cu, Fe, Ni, Zn, and even Cr(III), are essential elements to organisms [6]; however, they cause toxic effects when present in excessive quantities [7]. Their functions and potential toxicity have been comprehensively discussed in the literature [8–10]. Several PTE, e.g., As, Pb, Cd, and Hg, are non-essential to metabolic and other biological functions [11,12]. These are hazardous to organisms even at low levels [7] and harmful in various respects [13]. Persistent toxicity and bioaccumulation in food chains make PTE one of the most problematic types of pollutants [14].



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Among the agricultural PTE sources, fertilizers are the most common, and of these, phosphorus (P) fertilizers have been identified as the main source of PTE contamination of soil [9], especially if applied for a long time [15]. PTE are usually found in P-fertilizers because of the high amounts of these contaminants contained in raw materials, i.e., phosphate rocks (PR) [16–18]. PR contamination with PTE is natural, yet the level of these elements is variable, depending on the deposit source [19,20]. For example, the Cd concentration in PR in the world tends to be higher in North Africa (with the content up to 60 mg·kg⁻¹), Pb is higher in PR of Brazil (44.5 mg·kg⁻¹), and the highest Zn concentration occurs in the PR of Tunis (515 mg·kg⁻¹) [20].

Mined PR are either applied directly to soils or manufactured to produce water-soluble P-fertilizers [16], which are, therefore, accompanied by PTE [21]. PTE content is not included in the commercial fertilizer label, but according to Nziguheba and Smolders [22], for example, the average PTE concentrations in phosphate fertilizers sold on the European market were: 7.6 mg·kg⁻¹ As, 7.4 mg·kg⁻¹ Cd, 89.5 mg·kg⁻¹ Cr, 14.8 mg·kg⁻¹ Ni, 2.9 mg·kg⁻¹ Pb, and 166 mg·kg⁻¹ Zn. Many countries have regulations regarding limits on the maximum concentrations of PTE allowed in P-fertilizers [23,24]. Since Cd is the most studied toxic metal due to its high mobility and transfer rate from soil to crops [21], most of the fertilizer regulations relate Cd limits to P concentrations [25]. By 2036, the European Parliament (EP) is planning to tighten Cd limits to 20 mg per 1 kg P₂O₅ [26].

Phosphorus (P) is one of the essential elements needed for global food security. However, phosphate rocks (PR)—a primary P source—are finite and non-renewable [27]. Their uneven distribution over the globe and location mainly in geopolitically sensitive regions aggravate the problem and make many countries, including EU countries, dependent on raw material imports [27,28]. Moreover, the phosphate rock life cycle is, currently, predominantly linear, and there are approximately 80% P losses along the value chain "from mine to fork" [28,29]. The lost P ends up in waste, plant-unavailable P reservoirs in soils (legacy P), or in aquatic ecosystems [28,30,31].

Solutions to these problems can be found in closing the P loop, i.e., in recovering and recycling P as well as using it more efficiently [27,32]. Different types of bio-waste (e.g., municipal, food, agricultural) were valorized [33] and, among others, sewage sludge ashes (SSA) and animal bones have been identified as good raw materials for P-fertilizers [34]. Apart from being rich in P, these wastes can also serve as carriers of other macro- and micronutrients [35]. However, there are some barriers that must be overcome in order to use these resources effectively and safely for people and the environment [36].

Unlike sewage sludge, SSA is free of organic pollutants, but without further processing, it is poorly plant-available [37–40] and may contain elevated amounts of PTE, which restricts its direct use for agricultural purposes [41,42]. Thus, post-treatment is usually necessary in order to increase plant availability and to remove PTE [36]. There are several modern technologies for P recovery from SSA, such as thermo-chemical, acidic wet-chemical leaching, thermo-electric, and acidic wet-chemical extraction, which remove PTE from SSA or reduce their content to an acceptable level [43]. Finally, the PTE content in SSA varies widely depending on the sludge origins and treatment options [43]. For example, the thermochemically treated SSA from various plants of municipal sewage sludge combustion in Poland, tested by Smol et al. [43], contained 4.8–22.7 mg·kg⁻¹ As, 0.9–2.8 mg·kg⁻¹ Cd, 99.0–547.4 mg·kg⁻¹ Cr, 32.8-49.6 mg·kg⁻¹ Cu, <0.05 mg·kg⁻¹ Hg, 48.9-491.4 mg·kg⁻¹ Ni, 10.2–73.1 mg·kg⁻¹ Pb, and 1072.8–4459.9 mg·kg⁻¹ Zn, and met the Polish norms for trace elements covered by the legislation. The production of fertilizers in the P extraction processes from SSA is recommended [43], followed by a holistic evaluation of their effects in field conditions [44].

Bone raw material is not thought to be hazardous with PTE [35,45]. However, the primary P mineral in bones, calcium-deficient hydroxy-apatite, is much less soluble than conventional P fertilizers, although it is more soluble than PR [46,47]. Soil amendment using bones has been known since biblical times [48]. In modern times, bone meal (BM) and meat and bone meal (MBM) have been approved (with some limitations) for use

in agriculture as soil improvers [49] and have been reported from different countries to be effective in yield enhancement [50–54]. Recently, new technologies have emerged to produce bone-based fertilizers through thermal and/or chemical transformations [55–57].

An innovative alternative technology of recycled P fertilizer production is the inclusion of phosphorus-solubilizing microbes (PSM) into waste-based preparations [34]. Due to the release of weak organic (e.g., by *Bacillus megaterium*, bacteria naturally occurring in soil) or inorganic acids (e.g., by *Acidithiobacillus ferrooxidans*, bacteria active in strongly acidic (pH 2.5) environments), PSM increase the solubility of the phosphorus raw material [58]. Several recycled fertilizers have been developed using this technology [34] and evaluated for their agronomic utility in field experiments against traditional P-fertilizers [59]. In addition to satisfactory yields, the new formulations are expected to provide safety for field crop consumers and for the environment. This safety will not be guaranteed if the soil-plant system is loaded with PTE through the recycled fertilizer application. The issue of Cd and Pb presence in soil and plant biomass after the application of recycled PSM-containing fertilizers has been addressed previously [60]. This paper focuses on the effect of fertilizers from SSA and/or animal bones, activated by PSM (*Bacillus megaterium* or *Acidithiobacillus ferrooxidans*) on the content of As, Cr, Ni, Cu, and Zn in the soil, in wheat grain and straw (test plant), wheat associated weeds, and post-harvest residues.

2. Materials and Methods

2.1. Fertilizers

In field experiments, six recycled phosphorus fertilizers were tested against a commercial fertilizer superphosphate (SP). The new products, in the form of suspension or granular, were produced from sewage sludge ash (ash from the incineration of sewage sludge biomass from wastewater treatment; SSA) and/or animal (poultry) bones. Five of them were activated by PSM of the *Bacillus megaterium* or *Acidithiobacillus ferrooxidans* strains (Table 1). The recycled fertilizers were manufactured at the Institute of New Chemical Syntheses in Puławy (Poland), following a concept elaborated at the Wrocław University of Science and Technology (Poland). Raw materials originated from the following sources: SSA from the municipal wastewater treatment plant 'Łyna' in Olsztyn and bones from households or from the Firma Handlowo-Produkcyjna 'Podolski' (registered office in Lutomek, Wielkopolskie Voivodeship, Poland). Bacterial strains were acquired from the Polish Collection of Microorganisms at the Institute of Immunology and Experimental Therapy of the Polish Academy of Sciences in Wrocław (Poland). Table 2 shows the elemental composition of the recycled P-fertilizers, while the process of their production was described in separate articles [61,62].

Table 1. Recycled fertilizers tested in the experiments.

Fertilizer Symbol	Raw Material	Bacteria	Form
A _s Bm	sewage sludge ash (SSA)	Bacillus megaterium	suspension
B_sBm	bones	Bacillus megaterium	suspension
A_gAf	SSA	Acidithiobacillus ferrooxidans	granules
AB_gAf	SSA + bones	Acidithiobacillus ferrooxidans	granules
AB_{g}	SSA + bones	no bacteria	granules
АВ _д Вт	SSA + bones	Bacillus megaterium	granules

Recycled fertilizers were compared with superphosphate (SP). According to the commercial information provided on the label, this universal, concentrated P-fertilizer contains $40\% P_2O_5$ (17.4% P), 10% CaO (7.15% Ca), 5% SO3 (2% S), and microelements (B, Co, Cu, Fe, Mn, Mo, and Zn). According to an inductively coupled plasma–optical emission spectrometer (ICP-OES) analysis (the Chemical Laboratory of Multielemental Analysis at Wrocław University of Science and Technology accredited by ILAC-MRA and the Polish Center for Accreditation according to PN-EN ISO/IEC 17025 [63],; n = 3), apart from P (17.09%), this All P-fertilizers used in the experiments met the requirements of Polish and EU regulations for mineral fertilizers (Table 3).

Element	Unit	A_sBm	B _s Bm	A _g Af	AB _g Af	ABg	AB _g Bm
Р	% mass	0.176	0.259	9.24	7.50	6.10	5.87
Ν		0.255	0.350	0.530	3.190	3.82	3.14
Κ		0.487	0.217	0.963	0.727	0.846	0.772
Ca		0.694	0.373	12.513	12.012	11.369	10.725
Mg		0.119	0.009	2.442	1.176	1.296	1.188
S		0.055	0.046	1.38	1.50	2.28	1.91
Na		0.049	0.037	0.338	0.569	0.382	0.366
С		0.590	1.650	6.71	23.6	17.6	16.1
Fe	$ m gkg^{-1}$	1.679	0.022	33.0	15.1	15.4	14.4
Al	0 0	1.774	0.008	24.4	13.9	12.8	11.3
Zn		0.117	0.007	2.00	1.18	1.75	1.56
As	$mg \cdot kg^{-1}$	< 0.5	< 0.5	4.35	2.55	26.5	< 0.5
Cd	0 0	0.274	0.01	0.43	0.24	0.38	0.84
Cr		5.94	0.218	135	60.0	114	94.7
Cu		55.0	0.433	880	398	470	444
Ni		2.45	0.212	57.7	25.1	52.3	44.3
Pb		10.4	1.04	21.6	11.2	34.0	14.5

Table 2. Elemental composition of recycled P-fertilizers ¹.

¹ Analyses performed at the Chemical Laboratory of Multielemental Analysis at Wrocław University of Science and Technology accredited by ILAC-MRA and the Polish Center for Accreditation according to PN-EN ISO/IEC 17025 [63]; n = 3.

Recycled fertilizers were compared with superphosphate (SP). According to the commercial information provided on the label, this universal, concentrated P-fertilizer contains 40% P_2O_5 (17.4% P), 10% CaO (7.15% Ca), 5% SO3 (2% S), and microelements (B, Co, Cu, Fe, Mn, Mo, and Zn). According to an inductively coupled plasma–optical emission spectrometer (ICP-OES) analysis (the Chemical Laboratory of Multielemental Analysis at Wrocław University of Science and Technology accredited by ILAC-MRA and the Polish Center for Accreditation according to PN-EN ISO/IEC 17025 [63]; n = 3), apart from P (17.09%), this conventional fertilizer contained 25.57 mg·kg⁻¹ As, 15.66 mg·kg⁻¹ Cd, 186.1 mg·kg⁻¹ Cr, 22.35 mg·kg⁻¹ Cu, 44.29 mg·kg⁻¹ Ni, 10.43 mg·kg⁻¹ Pb, and 230 mg·kg⁻¹ Zn.

All P-fertilizers used in the experiments met the requirements of Polish and EU regulations for mineral fertilizers (Table 3).

Table 3. PTE limit values in mineral fertilizers according to Polish and EU regulations, $mg \cdot kg^{-1}$ DM.

Regulation	As	Cd	Cr	Cu	Hg	Ni	Pb	Zn
Poland [64]	50	50	n.s.	n.s.	2.0	n.s.	140	n.s.
EU [24]	40	60 ¹	n.s.	n.s.	1.0	100	120	n.s.

¹ per 1 kg P₂O₅; n.s.—not standardized.

2.2. Experimental Design and Agronomic Management

Recycled fertilizers were tested in four field experiments conducted in the years 2014–2016 (Table 4). Common wheat (*Triticum aestivum* ssp. *vulgare* Mac Key), winter or spring, was adopted as a test plant. In each experiment, the reference treatments for the recycled fertilizers were treatments with superphosphate (SP) fertilization and treatment without phosphorus fertilization (No P).

Experiment	Year	Test Plant	Recycled Fertilizers	Reference Treatments	P Doses, kg∙ha ⁻¹	Plant Protection (PP)
Ι	2014	spring wheat	A_sBm, B_sBm	No P, SP	21	PP-
Π	2015	spring wheat	A_sBm	No P, SP	17.6, 26.4, 35.2	PP-, PP+
III	2015	winter wheat	A_gAf , AB_gAf	No P, SP	17.6, 26.4, 35.2	PP+
IV	2016	winter wheat	AB_g, AB_gBm	No P, SP	17.6, 26.4, 35.2	PP+

Table 4. Field experiments conducted.

In Experiment I, all P-fertilizers were applied at a P dose of 21 kg·ha⁻¹, and in Experiments II–IV, three different P levels were established: 17.6, 26.4, and 35.2 kg·ha⁻¹. Suspension fertilizers were applied by the large-drop sprinkling of the soil, and solid fertilizers were manually scattered on the soil surface. The fertilizers were mixed with the soil by harrowing. In Experiment I, plant protection against agrophages (weeds, fungal pathogens, and pests) was not used (PP–); in Experiment II, two levels of plant protection were adopted—without protection (PP–) and with full protection (PP+); and in Experiments III and IV, full plant protection (PP+) was practiced. Other agricultural data for the experiments are shown in Table 5. Fertilization and other agrotechnical treatments were adjusted to the species requirements of the test plant and soil conditions and were consistent with the standards of good agricultural practice. Additionally, pesticides were applied according to the recommendations of the Institute of Plant Protection—National Research Institute in Poznań (Poland). All wheat cultivars used were assigned to the region according to Research Centre for Cultivar Testing (COBORU) in Słupia Wielka (Poland) recommendations.

Table 5. Basic agricultural data for the experiments.

	Experiment						
Item	Ι	II	III	IV			
Wheat cultivar	Trappe	Monsun	Julius	Julius			
Previous crop	spring barley	cereal-legume mixture	winter rape	winter rape			
Soil tillage system	plough tillage	plough tillage	plough tillage	plough tillage			
Fertilization							
– K, kg∙ha ^{−1}	99.6	83	83	83			
-	potassium chloride	potassium chloride	potassium chloride	potassium chloride			
$-N$, kg \cdot ha $^{-1}$	100	110	130	120			
	ammonium sulphate	ammonium sulphate	ammonium sulphate	ammonium sulphate			
Plant protection	_	_	_	_			
– herbicides		MCPA ¹	2.4-D + florasulam	2.4-D + florasulam			
– fungicides		azoxystrobin ¹	fenpropimorph +	fenpropimorph +			
		+ propiconazole	epoxiconazole + metrafenon	epoxiconazole + metrafenon			
		cyproconazole ¹	fluksapyroksad +	fluksapyroksad +			
		• •	piraklostrobina +	piraklostrobina +			
			epoxiconazole	epoxiconazole			
 insecticides 		lambda-cyhalothrin ¹	deltamethrin	deltamethrin			
 growth regulators 			trinexapac ethyl	trinexapac ethyl			
Sowing date	25.04.2014	9.04.2015	2.10.2014	15.10.2015			
Harvest date	11.08.2014	11.08.2015	5.08.2015	9.08.2016			

¹ applied only on plots with full plant protection (PP+).

Experiments I, III, and IV were established in a randomized block design and Experiment II in a parallel strip design. In each experiment, individual experimental treatments were executed in four replications (on four experimental plots of 20 m² each).

2.3. Study Site, Soil and Meteorological Conditions

The experiments with P-fertilizers were conducted at the Production and Experimental Station 'Bałcyny' Sp. z o.o. in Bałcyny (north-eastern Poland, Warmińsko-Mazurskie Province, 53°35′49″ N, 19°51′20″ E, 136.9 m above sea level) (Figure 1). The site is characterized by a temperate climate and geomorphological features that were predominantly formed during the Vistulian glaciation. The lowest air temperatures here are usually recorded in January and the highest in July. The average air relative humidity in the region varies from 71% in June to 90% in December [65]. The growing season lasts, on average, 215 days and falls in the months April to October [66].

In each experiment, wheat was grown in soil suitable for the species requirements (Table 6). The PTE content of the soil before the start of each experiment was within the range, deemed natural, and did not exceed the limits for agricultural use in Poland (see data and reference in Table 7).



Figure 1. Study site location.

Table 6. Soil characteristic before the start of the experiment	ıts.
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		Experi	ment	
Properties	I (n = 16)	II (n = 56)	III (n = 40)	IV (n = 40)
Soil type ¹	Luvisols	Luvisols	Luvisols	Luvisols
Soil texture ²	sandy clay loam	sandy loam	sandy loam	sandy loam
pH in KCl ³	6.23	5.32	5.51	5.23

ľ	roperties	I (n = 16)	II (n = 56)	III (n = 40)	IV (n = 40)
	C, $g \cdot kg^{-1}$	8.31	8.90	8.87	7.15
	N, $g \cdot kg^{-1}$	1.30	1.35	1.36	1.09
	P_{r} mg·kg ⁻¹	574	566	433	548
	K, mg·kg ^{-1}	2979	2895	3210	3304
	Mg, mg·kg ^{-1}	2070	2007	2253	2195
Total 4	Cd , $mg \cdot kg^{-1}$	0.025 (0.365)	0.292 (0.827)	0.309 (1.083)	0.133 (0.553)
10tal,	Pb, mg·kg ^{-1}	4.855 (13.198)	7.959 (25.099)	7.059 (22.895)	2.297 (18.180)
	As, mg·kg ^{-1}	2.012 (8.388)	0.574 (3.013)	<0.5 (<0.5)	2.067 (8.390)
	Cu, mg·kg ^{-1}	10.66 (14.97)	10.28 (14.92)	11.90 (19.44)	9.113 (10.860)
	$Cr, mg \cdot kg^{-1}$	19.96 (23.72)	19.60 (23.95)	20.85 (32.07)	23.34 (28.37)
	Ni, mg·kg ^{-1}	7.16 (11.25)	8.53 (14.78)	8.05 (18.27)	10.69 (15.71)
	$Zn, mg \cdot kg^{-1}$	33.60 (43.52)	60.30 (78.80)	51.08 (64.98)	41.69 (56.22)

Table 6. Cont.

¹ According to World reference base for soil resources 2014 [67]; ² based on agricultural map (1:5000) held by the Production and Experimental Station 'Bałcyny' Sp. z o.o. in Bałcyny (Bałcyny 15, 14-100 Ostróda, Poland); ³ determined by the potentiometric method [68]; ⁴ determined as described in Section 2.5, average values, maximum values in brackets.

Table 7.	. Reference	values	for PTE	l of the soil	l and plants,	, mg·kg $^{-1}$	DM.
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Content	As	Cr	Cu	Ni	Zn			
In Soil								
Geochemical background for Poland [69,70]	2-13	2–64	0.4–23.5	0.5–28.5	5.0–59			
In surface level of mineral soils in Poland [71]	0.6 - 10	5 - 100	0.5-25	0.5-60	10-225			
Permissible in Poland in arable land [72]								
– I ¹	10	150	100	100	300			
– II	20	300	150	150	500			
– III	50	500	300	300	1000			
Monitoring of arable soils [73]								
– Poland	0.73-20.7	2.4-49.1	1.2-320.1	1.0-71.1	5.9-6668.3			
– – average	3.63	11.0	10.3	9.8	78.8			
– – median	2.77	8.7	6.2	6.1	32.0			
– region ²	1.52-3.87	4.5-30.6	3.3-17.1	2.6-29.8	19.4–61.9			
	In Pla	nt						
Sufficient or normal [4]	1 - 1.7	0.1–0.5	5–30	0.1–5	27-150			
Excessive or toxic [4]	5-20	5-30	20 - 100	10 - 100	100-400			
Average in cereal grain [71]	0.03-0.6	0.003 - 0.4	2.6-6	0.1 - 0.5	15-60			
Permissible in Poland								
– in cereal grain [74] ³	0.20	n.s.	n.s.	n.s.	n.s.			
– in feed materials of plant origin [75,76]	2	n.s.	n.s.	n.s.	n.s.			
Other standards for food grain								
– FAO/WHO [77]	0.35 (0.2) ⁴	n.s.	n.s.	n.s.	n.s.			
– EU [78]	0.1–0.25 ⁵	n.s.	n.s.	n.s.	n.s.			
– PR of China [79]	0.5 (0.2) ⁶	0.1	n.s.	n.s.	n.s.			
Suggested maximum values in plant material								
[80,81] for food answere	:		20	10	FO			
– for food purposes	n.1.	n.1.	20	10	50			
– for feed purposes	n.1.	20	25-50	50	100			

¹ Sub-groups of arable land depending on soil compaction and pH; ² range for 4 measurement points (villages) located geographically closest to the test site (Bałcyny); ³ regulation repealed, no new; ⁴ for inorganic As in rice husked or polished (value in brackets); ⁵ for inorganic As in rice depending on its preparation and intended use; ⁶ value for rice in brackets; n.s.—not standardized; n.i.—no information.

The precipitation and air temperatures during the growing seasons were not favorable to the wheat plants (Table 8). In Experiments I–III, the growing seasons for the test plants (spring and winter wheat) tended to be very dry (rainfall deficits in May and July 2014 and in May and June 2015), while in Experiment IV, winter wheat developed under wet season conditions with excessive rainfall in July 2016. The water content of the soil may alter the PTE

2015

2016

1981-2010

2014

2015

2016

1981-2010

11-20

21-31

Total

1 - 10

11-20

21 - 31

Total

Total

1 - 10

11 - 20

21-31

Average

1 - 10

11 - 20

21-31

Average

1 - 10

11 - 20

21–31

Average

Average

0.4

7.4

28.5

0.3

9.0

19.4

28.7

30.1

0.0

2.7

-0.9

0.6

-8.4

-4.3

0.7

-3.8

-2.4

2.0

1.1

8.8

11.8

23.7

15.0

50.5

23.1

-1.8

-0.1

3.3

0.3

4.1

2.0

2.1

2.7

-1.6

0.0

26.8

46.0

0.6

7.0

12.9

20.5

30.7

4.2

4.7

4.9

4.6

2.9

2.2

5.5

3.6

1.8

10.9

4.4

23.4

4.8

19.2

9.1

33.1

29.8

7.0

8.5

12.9

9.5

4.1

7.1

10.4

7.2

10.3

9.1

6.9

8.8

7.7

availability to plants [82]. Moreover, heavy rainfall may also result in the leaching of PTE
from the surface layers of the soil and in their accumulation in depressions in the ground [83]

Month Year Days I Π III IV V VI VII VIII XII IX х XI Precipitation, mm 1 - 1016.7 37.3 0.0 9.5 0.0 15.0 15.711.812.6 11 - 202.3 21.5 6.8 0.0 20.86.2 12.8 5.6 8.6 2014 21-31 3.8 17.6 35.0 0.0 15.118.2 0.5 5.5 43.8 26.134.9 72.2 59.2 Total 20.430.8 21.3 21.2 56.6 1 - 1020.7 5.7 19.2 8.1 7.2 0.1 13.6 2.0 44.0 0.0 30.7 9.3

17.7

0.5

25.4

7.5

53.7

9.6

70.8

62.3

8.9

13.3

17.1

13.3

11.7

11.6

13.0

12.1

14.1

11.8

18.5

14.9

13.2

11.5

31.4

43.0

12.1

28.1

26.1

66.3

72.9

16.5

14.2

13.8

14.8

15.7

15.9

15.5

15.7

16.4

16.3

21.3

18.0

15.8

22.0

35.4

71.0

39.6

34.0

65.0

138.6

81.2

20.5

19.6

22.8

21.0

19.8

16.9

17.4

18.0

17.5

18.1

19.9

18.5

18.3

Air temperature, °C

0.0

11.0

13.0

54.5

10.4

7.0

71.9

70.6

22.2

17.2

14.6

17.9

23.3

21.1

19.5

21.3

17.7

15.6

19.3

17.6

17.7

6.8

0.4

51.2

7.5

1.2

8.4

17.1

56.2

15.0

17.0

11.6

14.5

15.2

15.9

11.6

14.2

17.5

15.1

11.7

14.7

13.0

18.6

2.2

20.8

65.1

9.2

22.0

96.3

51.2

11.3

12.2

5.4

9.5

7.9

6.0

6.0

6.6

9.0

5.7

6.1

6.9

8.1

39.3

10.8

80.8

39.9

27.7

10.6

78.2

46.1

8.5

4.7

0.0

4.4

6.3

7.9

1.2

5.1

2.7

2.7

2.1

2.5

2.8

26.4

44.7

80.4

41.4

19.2

17.2

77.8

42.6

-1.7

3.5

-3.2

-0.6

4.7

3.1

3.5

3.8

1.9

-0.3

1.4

1.0

-1.0

Table 8. Precipitation and air temperature during the study period according to the Meteorological Station in Bałcyny [84].

2.4. Sampling

2.4.1. Soil

Soil from the 0–0.30 m layer was sampled before the start of each experiment and after wheat harvest, using a hand-held twisting probe (Egner's soil sampler). Sampling was performed at 30 evenly spaced points in each plot, and a total of approximately 1 kg of soil was taken from an individual plot. The collected field-fresh soil samples were air-dried at room temperature (~22 °C) for several (5–7) days, thoroughly mixed, and sieved. Portions of approximately 300 g were then delivered to the accredited chemical laboratory (the Chemical Laboratory of Multielemental Analysis at Wrocław University of Science and Technology, Poland, accredited by ILAC-MRA and the Polish Center for Accreditation according to PN-EN ISO/IEC 17025 [63], Accreditation Certificate No. AB 696) for elemental analysis.

2.4.2. Wheat Grain

Grain samples weighing approximately 1 kg were taken from the volume resulting after combine harvesting from each plot. Out of these samples, portions of approximately 200 g of grain were weighed, and after cleaning from impurities and weed seeds, sent for chemical analyses.

2.4.3. Wheat Straw

Directly before harvest, wheat plants were picked up manually from two 1 meter-long rows of each plot. The wheat roots were then cut off at a height matching the combine harvester mowing height, and the spikes were removed. Straw (i.e., wheat stems with leaves) was cut into pieces of approximately 10 cm in length. The prepared samples were dried at room temperature for several days, and a portion of 50 g was then weighed from each sample and sent for chemical analyses.

2.4.4. Weeds

Weed samples were collected from a randomly designated 0.25 m² (0.5 m \times 0.5 m frame) area from each plot prior to wheat harvest. Weed roots were cut off and the aboveground biomass of weeds was dried at room temperature for several days. Afterwards, the samples in their entirety (different mass) were passed for chemical analyses.

2.4.5. Post-Harvest Residues

Post-harvest residues, i.e., wheat roots and bottom stem segments (stubble), and weed remnants left in the field after combine harvesting were sampled from each plot after wheat harvest. Soil columns of 0.40 m² and 0.30 m depth were dug up and then washed on sieves to remove soil particles. The plant material samples prepared in this way were dried at room temperature for several days and sent in their entirety for chemical analysis.

2.5. Chemical Analyses

The total soil contents of carbon (C) and nitrogen (N) were determined by Vario Macro Cube Elementar (C, H, N) analyzer (Elementar Analysensysteme, Langenselbold, Germany). As a standard solution, D-phenylalanine (C = 65.44%; N = 8.48%) was used.

An inductively coupled plasma–optical emission spectrometer (ICP–OES with a pneumatic nebulizer with an axial view—iCAP Duo Thermo Scientific, USA) was used to determine the contents of other elements in the soil and plant material. The Teflon vessels (microwave oven Milestone MLS-1200) and 5 mL of concentrated 65 mg·kg⁻¹ HNO₃ SuprapurTM (Merck) in the case of plant samples and with 10 mL of aqua regia in the case of soil samples were used to digest an appropriate mass (0.5 g) of sample materials (plant or soil). Then, all samples were diluted to 50 mL. The obtained samples were subjected to multi-elemental analyses using ICP-OES. The determination of PTE content was carried out with all the principles of measurement traceability, and certified reference materials were used to check the quality and metrological traceability. The detection levels for P, K, Mg, As, Cd, Cr, Cu, Ni, Pb, and Zn for the soil material were 3.59, 2.55, 1.17, 0.5, 0.01, 0.035, 0.27, 0.25, 0.15, and 0.32 mg·kg⁻¹, respectively, and for the plant material, the detection levels for As, Cr, Cu, Ni, and Zn were 0.05, 0.005, 0.04, 0.002, and 0.013 mg·kg⁻¹, respectively [85].

2.6. Calculations and Statistical Analysis

Based on the elemental composition of fertilizers (As content below the detection level (DL) replaced by 0.5 mg·kg⁻¹) (Table 1), and assuming that a dry soil from the 0–30 cm-deep layer and 1 ha area weighs 4500 t, the PTE input per 1 ha with fertilizers applied at different P doses was calculated, as well as the potential increase in soil PTE content (per 1 kg of soil).

The data on PTE content in soil and plant biomass were processed using an analysis of variance ANOVA (when its assumptions were met) or the alternative Kruskal–Wallis test (when ANOVA assumptions were not met). The Shapiro–Wilk W test and Levene's test were used to verify the normality of distribution of variables and the homogeneity of variance, respectively. Duncan's test or a multiple comparisons test was then applied to assess differences between objects. In statistical calculations, the PTE contents below the detection level (DL) were replaced by values equal to the DL. For each experiment, the median (Me) and the maximum value (Max) were determined. The calculations were performed using Statistica 13.3 software [86].

3. Results and Discussion

3.1. PTE Input from Fertilizers to Soil

The PTE presence in the tested P-fertilizers (Table 1) suggests the possibility of their accumulation in soil due to fertilizer application, which should potentially become apparent in comparison to no P treatments. The amount of PTE introduced into the soil with P fertilizers per ha and the potential increase in soil PTE content are shown in Table 9. Considering the limit values for annual loads of certain PTE established by EC [87], these quantities should be qualified as very small, and the soil enrichment in PTE is negligible for statistical analysis (compare differences in [15]).

Table 9. PTE amounts introduced into soil with P -fertilizers applied—range between the values for the lowest (17.6 kg·ha⁻¹) and the highest (35.2 kg·ha⁻¹) P dose.

	As	Cr	Cu	Ni	Zn
Fertilizers			Input per 1 ha		
	g	kg	kg	kg	kg
SP ¹	2.633-5.265	0.019-0.038	0.002-0.005	0.005-0.009	0.025-0.049
A_sBm	$5.00 - 10.00^{2}$	0.059-0.119	0.550 - 1.100	0.025-0.049	1.170-2.340
$B_s Bm$	3.398–6.795 ²	0.001-0.003	0.003-0.006	0.001-0.003	0.048-0.095
AgAf	0.829 - 1.657	0.026-0.051	0.168-0.335	0.011-0.022	0.381-0.762
AB_gAf	0.598 - 1.197	0.014-0.028	0.093-0.187	0.006-0.012	0.277 - 0.554
AB_{g}	7.646 - 15.292	0.033-0.066	0.136-0.271	0.015-0.030	0.505 - 1.010
$AB_g Bm$	0.150-0.300	0.028-0.057	0.133-0.266	0.013-0.027	0.468-0.935
Limit values per year ³	n.s	n.s. (0.04–12.0) ⁴	12	3	30
		Potential Increase	in Soil Content, µg k	g ⁻¹ of Soil DM	
SP	0.59-1.17	4.26-8.52	0.51-1.02	1.01-2.03	5.47-10.94
A _s Bm	1.11–2.22 ²	13.20-26.40	122.2-244.4	5.44 - 10.89	260.0-520
$B_s Bm$	$0.76 - 1.51^{2}$	0.33-0.66	0.65 - 1.31	0.32-0.64	10.6-21.14
AgAf	0.18-0.37	5.71-11.43	37.2-74.5	2.44 - 4.88	84.7-169
$AB_{g}Af$	0.13-0.27	3.13-6.26	20.8-41.5	1.31-2.62	61.5 - 123
ABg	1.70-3.40	7.31-14.62	30.1-60.3	3.35-6.71	112-224
AB _g Bm	0.03-0.07	6.31-12.62	29.6-59.2	2.95-5.90	104-208

¹ SP—superphosphate, A_sBm —SSA-based suspension fertilizer with *B. megaterium*, B_sBm —bone-based suspension fertilizer with *B. megaterium*, A_gAf —SSA-based granular fertilizer with *A. ferrooxidans*, AB_gAf —SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_gBm —SSA-bone-based granular fertilizer with *B. megaterium*; ² potentially maximum values; ³ according to Final Implementation Report for Directive 86/278/EEC on Sewage Sludge: 2013–2015 [87]; ⁴ range of national limits set by 20 Member States, excluding Poland (no set limits) [87].

The recycled fertilizers tested tended to introduce more PTE than SP applied at the same dose. The exceptions were lower As inputs with A_gAf , AB_gAf , and AB_gBm fertilizers, lower Cr and Ni inputs with B_sBm and Cr with AB_gAf fertilizers. It is noteworthy that even the relatively elevated amounts of Cu and Zn added to the soil with the recycled fertilizers were substantially lower than the doses of these micronutrients recommended for wheat [88].

3.2. PTE in Soil

The soil As, Cr, Cu, Ni, and Zn contents in the conducted experiments (Table 10) met the ranges considered natural under Polish conditions and did not exceed permissible levels for arable lands (Table 7). The average and median values in the experiments were not very far from those determined for the whole country. Even the highest values fell within the geochemical background ranges and were at the levels typical for the region. For As, the median values in the experiments usually ranked below the detection level (an exception was Experiment IV).

Experiment	P-Fertilizer	P-Dose, kg∙ha ^{−1}	Plant Protection	As	Cr	Cu	Ni	Zn
	No P	0	PP-	1.854	18.1	7.07↓	5.31↓	26.1↓
	SP 1	21		2.096	19.6	8.20↓	5.06↓	28.2↓
т	A_sBm	21		1.512	19.3	8.12↓	6.03↓	27.6↓
1	B_sBm	21		0.996	19.0	9.32↓	6.38↓	27.2↓
	Me			<dl< td=""><td>18.6</td><td>8.51</td><td>5.43</td><td>27.2</td></dl<>	18.6	8.51	5.43	27.2
	Max			6.885	22.8	10.93	9.04	30.8
	No P	0	PP-	0.776	$15.8\downarrow^2$	$6.17\downarrow$	7.67↓	43.3↓
	SP	17.6		<dl< td=""><td>$16.5\downarrow$</td><td>$6.87\downarrow$</td><td>$7.10\downarrow$</td><td>43.2↓</td></dl<>	$16.5\downarrow$	$6.87\downarrow$	$7.10\downarrow$	43.2↓
		26.4		<dl< td=""><td>15.3↓</td><td>$6.54\downarrow$</td><td>7.71↓</td><td>42.3↓</td></dl<>	15.3↓	$6.54\downarrow$	7.71↓	42.3↓
		35.2		2.598	15.3↓	5.37↓	$7.94\downarrow$	42.7↓
	A_sBm	17.6		3.047	$15.8\downarrow$	$6.47\downarrow$	7.98↓	42.9↓
		26.4		0.814	$15.4\downarrow$	5.80↓	7.59↓	42.3↓
		35.2		<dl< td=""><td>15.8↓</td><td>5.48↓</td><td>7.94 ↓</td><td>42.9↓</td></dl<>	15.8↓	5.48↓	7.94 ↓	42.9↓
		average		1.248	15.7↓	6.10↓	7.71↓	42.8↓
11	No P	0	PP+	<dl< td=""><td>$15.8\downarrow$</td><td>$6.51\downarrow$</td><td>7.34↓</td><td>43.3↓</td></dl<>	$15.8\downarrow$	$6.51\downarrow$	7.34↓	43.3↓
	SP	17.6		1.925	$16.0\downarrow$	5.62↓	7.57↓	43.1↓
		26.4		<dl< td=""><td>$17.4\downarrow$</td><td>5.79↓</td><td>7.56↓</td><td>43.5↓</td></dl<>	$17.4\downarrow$	5.79↓	7.56↓	43.5↓
		35.2		3.280	16.3↓	7.37↓	7.38↓	42.0↓
	A_sBm	17.6		<dl< td=""><td>$14.7\downarrow$</td><td>$6.11\downarrow$</td><td>7.59↓</td><td>42.9↓</td></dl<>	$14.7\downarrow$	$6.11\downarrow$	7.59↓	42.9↓
		26.4		<dl< td=""><td>15.3↓</td><td>5.37↓</td><td>$7.65\downarrow$</td><td>$41.8\downarrow$</td></dl<>	15.3↓	5.37↓	$7.65\downarrow$	$41.8\downarrow$
		35.2		<dl< td=""><td>16.3↓</td><td>7.82↓</td><td>7.16↓</td><td>42.3↓</td></dl<>	16.3↓	7.82↓	7.16↓	42.3↓
		average		1.101	16.0↓	6.37↓	7.46↓	42.7↓
	Me			<dl< td=""><td>14.7</td><td>5.54</td><td>7.34</td><td>42.4</td></dl<>	14.7	5.54	7.34	42.4
	Max			8.841	21.5	11.0	14.8	51.5
	No P	0	PP+	1.257	$19.8\downarrow$	7.29↓	8.77	38.0↓
	SP	17.6		1.428	19.7↓	$8.67\downarrow$	5.53	$40.1\downarrow$
		26.4		<dl< td=""><td>19.3↓</td><td>8.62↓</td><td>8.43</td><td>$41.9\downarrow$</td></dl<>	19.3↓	8.62↓	8.43	$41.9\downarrow$
		35.2		<dl< td=""><td>19.0↓</td><td>$8.34\downarrow$</td><td>5.87</td><td>$40.4\downarrow$</td></dl<>	19.0↓	$8.34\downarrow$	5.87	$40.4\downarrow$
	AgAf	17.6		<dl< td=""><td>19.2↓</td><td>9.32↓</td><td>7.15</td><td>$40.5\downarrow$</td></dl<>	19.2↓	9.32↓	7.15	$40.5\downarrow$
ш		26.4		1.113	19.8↓	9.26↓	8.76	37.3↓
111		35.2		<dl< td=""><td>19.3↓</td><td>$8.16\downarrow$</td><td>6.04</td><td>39.1↓</td></dl<>	19.3↓	$8.16\downarrow$	6.04	39.1↓
	AB _g Af	17.6		0.711	19.3↓	8.23↓	5.67	39.2↓
		26.4		0.806	19.6↓	7.20↓	6.24	38.7↓
		35.2		0.705	20.0↓	8.26↓	8.35	38.2↓
	Me			<dl< td=""><td>19.9</td><td>7.84</td><td>7.37</td><td>38.6</td></dl<>	19.9	7.84	7.37	38.6
	Max			2.950	23.2	13.56	12.2	47.4
	No P	0	PP+	1.319	$18.7\downarrow$	7.57↓	9.91	37.6↓
	SP	17.6		1.242	20.9↓	8.13↓	9.95	38.9↓
		26.4		1.356	19.6↓	7.65↓	9.40	38.1↓
		35.2		1.286	$18.1\downarrow$	7.33↓	9.99	38.2↓
	AB_g	17.6		0.931	$18.8\downarrow$	7.96↓	9.67	36.5↓
IV		26.4		1.154	19.7↓	7.76↓	10.30	37.1↓
τv		35.2		1.573	$20.4\downarrow$	7.99↓	10.20	43.1↓
	AB _g Bm	17.6		0.819	$15.5\downarrow$	7.99↓	10.39	$40.5\downarrow$
		26.4		1.513	20.5↓	8.62↓	10.20	37.9↓
		35.2		1.423	19.6↓	7.75↓	10.12	36.2↓
	Me			0.758	19.2	7.97	10.0	37.8
	Max			4.082	23.8	10.6	15.1	55.0

Table 10. PTE content in soil, $mg \cdot kg^{-1} DM$ (n = 4).

¹ SP—superphosphate, A_sBm—SSA-based suspension fertilizer with *B. megaterium*, B_sBm—bone-based suspension fertilizer with *B. megaterium*, A_gAf—SSA-based granular fertilizer with *A. ferrooxidans*, AB_gAf—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_gM—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_gM—SSA-bone-based granular fertilizer with *B. megaterium*; ² arrows (\downarrow) indicate significant decrease in relation to the starting state; for P-fertilization treatments (Experiments I–IV) and plant protection treatments (Experiment II), there were no significant differences.

Recycled fertilizers, similar to SP, did not significantly change the soil content of tested PTE as compared to no P treatments in any of the experiments, regardless of P dose. The application of PP+ compared to PP- (Experiment II) also did not change the soil PTE content.

The enrichment of soil PTE under P fertilization was not observed in any of the experiments. Moreover, in comparison with the initial state (before the experiments and fertilization treatments), a significant decrease in soil Cr content was observed in Experiments II-IV, Ni in Experiments I and II, and Cu and Zn in all experiments, but with no relation to the P fertilization applied. Since Cu, Ni, and Zn are micronutrients for plants [6], their depletion in soil was probably caused by plant uptake (wheat and weeds). The higher yields of P-fertilized plants and the resulting greater uptake of elements explain the statistically even decrease in soil PTE under P-treatments and under no P treatments. In particular, Cu and Zn, being key micronutrients for cereals [89], can be readily taken up by plants if available [90]. Although any biological role of Cr in plant physiology is known to date [91], it may be taken up along essential elements, such as sulfate through sulfate transporters [92]. In soils with a pH below 6.5, as in the experiments presented here, the solubility of PTE and, thus, their mobility and plant availability are increased [93]. Therefore, leaching and runoff of PTE, even of Cu and Zn, which are considered hardly leached from the soil, also cannot be excluded [88,94,95]. In none of the experiments was a change in the soil As content, in relation to the initial state, noted. According to Jiao et al. [96], under normal cropping practices, As accumulation in soils is insignificant. Chen et al. [97] reported that more than 90% of As introduced through fertilizer applications is expected to leach below the root zone over time.

Earlier studies by other authors indicate that feeding plants with fertilizers containing PTE, both from primary or secondary resources, may lead to an increase in their content in soil as a result of a longer-term application [95,98,99], or even during one growing season [98]. In contrast, based on a permanent arable field investigation, Uprety et al. [100] concluded that common cropping practices with the application of organic and inorganic fertilizers do not substantially enrich levels of As, Cd, Cr, Cu, Ni, and Pb in soils, even after they have been applied for more than 50 years. Chen et al. [15] found no increase in soil Cu, Cr, and Ni content after a 10-year application of 0.016, 0.042, and 0.022 kg of these elements, respectively, along with 400 kg·ha⁻¹ P, and no increase in As and Zn content when applied at rates up to 0.042 kg (with 200 kg·ha⁻¹ P) and 0.027 kg (with 100 kg·ha⁻¹ P), respectively. It is worth noting that the amounts of non-essential As introduced into the soil, together with all recycled fertilizers tested in the present study, were substantially lower (compare Table 9). However, considering the role of those PTE as essential micronutrients, particularly of Cu and Zn, their moderately higher quantities added to the soil along with the recycled fertilizers may be beneficial rather than problematic [90].

3.3. PTE in Plant Biomass

3.3.1. Wheat Grain and Straw

Wheat grain constitutes the basis of many food products worldwide, so its contamination with PTE is of particular concern [12,101]. The PTE content of straw becomes important if it is used for animal feed [102]. In all the conducted experiments, recycled fertilizers applied at three P rates showed no effect on the As, Cr, Cu, Ni, and Zn content in wheat grain or straw (Table 11). Similarly, PTE contents in grain and straw were not differentiated by SP.

Experiment	P-Fertilizer	P-Dose, kg∙ha ⁻¹	Plant Protection	As	Cr	Cu	Ni	Zn
	No P	0	PP-	<dl< td=""><td>0.307</td><td>2.85</td><td>0.182</td><td>22.1</td></dl<>	0.307	2.85	0.182	22.1
	SP ¹	21		<dl< td=""><td>0.118</td><td>2.77</td><td>0.132</td><td>21.9</td></dl<>	0.118	2.77	0.132	21.9
	A _s Bm	21		<dl< td=""><td>0.125</td><td>3.00</td><td>0.079</td><td>23.2</td></dl<>	0.125	3.00	0.079	23.2
1	B_sBm	21		<dl< td=""><td>0.223</td><td>2.74</td><td>0.034</td><td>21.5</td></dl<>	0.223	2.74	0.034	21.5
	Me			<dl< td=""><td>0.141</td><td>2.79</td><td>0.096</td><td>22.1</td></dl<>	0.141	2.79	0.096	22.1
	Max			0.050	0.748	3.45	0.362	25.9
	No P	0	PP-	0.054	0.233	3.70	0.028	40.9
	SP	17.6		0.059	<dl< td=""><td>3.70</td><td>0.065</td><td>42.3</td></dl<>	3.70	0.065	42.3
		26.4		<dl< td=""><td>0.193</td><td>3.32</td><td>0.027</td><td>39.1</td></dl<>	0.193	3.32	0.027	39.1
		35.2		<dl< td=""><td>0.196</td><td>3.55</td><td>0.086</td><td>41.2</td></dl<>	0.196	3.55	0.086	41.2
	A_sBm	17.6		<dl< td=""><td>0.008</td><td>3.76</td><td>0.031</td><td>47.6</td></dl<>	0.008	3.76	0.031	47.6
		26.4		<dl< td=""><td>0.058</td><td>3.64</td><td>0.058</td><td>39.7</td></dl<>	0.058	3.64	0.058	39.7
		35.2		<dl< td=""><td>0.084</td><td>3.87</td><td>0.033</td><td>42.8</td></dl<>	0.084	3.87	0.033	42.8
		average		0.047	0.111	3.65 b ²	0.047	41.9
II	No P	0	PP+	0.074	0.072	4.32	0.008	39.9
	SP	17.6		<dl< td=""><td>0.083</td><td>3.91</td><td>0.008</td><td>37.2</td></dl<>	0.083	3.91	0.008	37.2
		26.4		<dl< td=""><td>0.063</td><td>4.07</td><td>0.032</td><td>40.1</td></dl<>	0.063	4.07	0.032	40.1
		35.2		0.052	<dl< td=""><td>4.01</td><td>0.068</td><td>38.7</td></dl<>	4.01	0.068	38.7
	A_sBm	17.6		<dl< td=""><td>0.131</td><td>3.92</td><td>0.034</td><td>41.2</td></dl<>	0.131	3.92	0.034	41.2
		26.4		<dl< td=""><td>0.157</td><td>3.63</td><td>0.050</td><td>37.6</td></dl<>	0.157	3.63	0.050	37.6
		35.2		<dl< td=""><td>0.018</td><td>3.58</td><td>0.016</td><td>41.3</td></dl<>	0.018	3.58	0.016	41.3
		average		0.051	0.076	3.92 a	0.031	39.4
	Me			<dl< td=""><td>0.011</td><td>3.79</td><td>0.022</td><td>41.0</td></dl<>	0.011	3.79	0.022	41.0
	Max			0.125	0.521	4.67	0.235	56.4
	No P	0	PP+	<dl< td=""><td>0.033</td><td>2.31</td><td>0.179</td><td>26.4</td></dl<>	0.033	2.31	0.179	26.4
	SP	17.6		0.062	0.015	2.09	0.215	25.5
		26.4		<dl< td=""><td><dl< td=""><td>2.17</td><td>0.180</td><td>24.0</td></dl<></td></dl<>	<dl< td=""><td>2.17</td><td>0.180</td><td>24.0</td></dl<>	2.17	0.180	24.0
		35.2		<dl< td=""><td>0.159</td><td>2.09</td><td>0.245</td><td>24.2</td></dl<>	0.159	2.09	0.245	24.2
	$A_{\sigma}Af$	17.6		0.057	0.132	2.50	0.149	24.7
	8 -	26.4		0.081	<dl< td=""><td>2.39</td><td>0.187</td><td>24.2</td></dl<>	2.39	0.187	24.2
111		35.2		<dl< td=""><td>0.062</td><td>2.17</td><td>0.122</td><td>27.1</td></dl<>	0.062	2.17	0.122	27.1
	AB_gAf	17.6		0.063	0.112	2.39	0.287	27.5
	0,	26.4		0.058	0.133	2.43	0.182	24.6
		35.2		<dl< td=""><td>0.165</td><td>2.36</td><td>0.230</td><td>27.2</td></dl<>	0.165	2.36	0.230	27.2
	Me			<dl< td=""><td><dl< td=""><td>2.30</td><td>0.164</td><td>25.4</td></dl<></td></dl<>	<dl< td=""><td>2.30</td><td>0.164</td><td>25.4</td></dl<>	2.30	0.164	25.4
	Max			0.173	0.630	2.93	0.691	33.2
	No P	0	PP+	0.056	0.228	3.85	0.189	26.0
	SP	17.6		<dl< td=""><td>0.068</td><td>3.74</td><td>0.113</td><td>24.1</td></dl<>	0.068	3.74	0.113	24.1
		26.4		<dl< td=""><td>0.176</td><td>3.57</td><td>0.125</td><td>24.1</td></dl<>	0.176	3.57	0.125	24.1
		35.2		<dl< td=""><td>0.405</td><td>3.36</td><td>0.087</td><td>22.7</td></dl<>	0.405	3.36	0.087	22.7
	ABg	17.6		<dl< td=""><td>0.591</td><td>4.04</td><td>0.167</td><td>24.9</td></dl<>	0.591	4.04	0.167	24.9
IV	-	26.4		0.065	0.314	3.74	0.144	24.3
1 V		35.2		<dl< td=""><td>0.180</td><td>3.92</td><td>0.138</td><td>25.7</td></dl<>	0.180	3.92	0.138	25.7
	AB _g Bm	17.6		0.052	0.143	4.05	0.149	24.2
		26.4		<dl< td=""><td>0.179</td><td>3.95</td><td>0.185</td><td>25.6</td></dl<>	0.179	3.95	0.185	25.6
		35.2		<dl< td=""><td>0.247</td><td>3.90</td><td>0.250</td><td>27.0</td></dl<>	0.247	3.90	0.250	27.0
	Me			<dl< td=""><td>0.168</td><td>3.81</td><td>0.126</td><td>24.8</td></dl<>	0.168	3.81	0.126	24.8
	Max			0.087	0.992	5.11	0.390	33.4

Table 11. PTE content in wheat grain, $mg \cdot kg^{-1}$ DM (n = 4).

¹ SP—superphosphate, A_sBm—SSA-based suspension fertilizer with *B. megaterium*, B_sBm—bone-based suspension fertilizer with *B. megaterium*, A_gAf—SSA-based granular fertilizer with *A. ferrooxidans*, AB_gAf—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_g—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_g—SSA-bone-based granular fertilizer with *B. megaterium*, ² different letters indicate significant differences at p < 0.05 for plant protection treatments in Experiment II; for P-fertilization treatments (Experiments I–IV), there were no significant differences.

In all experiments, As, Cr, Cu, Ni, and Zn were typically found in wheat grain in amounts considered normal or sufficient and did not reach levels indicative of excessive or toxic content (Table 7). The As content in wheat grain was usually far below the value of 0.2 mg·kg⁻¹. This level was previously defined in Poland as the maximum permissible value for cereal grains for food purposes (the relevant regulation was abolished). Today, FAO/WHO and EU standards limit only the inorganic As content of rice grains for human consumption. The highest As content detected in grain was $0.173 \text{ mg} \cdot \text{kg}^{-1}$ (in Experiment III), while median values in all experiments were below the detection level. No standards for Cr, Cu, Ni, and Zn contents in cereal grains have yet been developed in Poland. Moreover, FAO/WHO and EU have also not provided limits for these metals in food grain (Table 7). In China, the consumption of grain is considered safe when Cr accumulation is under 1 mg kg^{-1} [79]. Although Cr contents in grain in the present study did not exceed this level, the highest value (0.992 mg \cdot kg⁻¹) determined in wheat fertilized with AB_g (Experiment IV) was dangerously close to it. However, in Experiment I, the highest Cr content in grain (0.748 mg·kg⁻¹) was found under no P treatment. Similarly, Kulczycki and Sacała [103] found an average Cr content of $0.77 \text{ mg} \cdot \text{kg}^{-1}$ in wheat grain with no Cr treatment. The Cu, Ni, and Zn contents were within ranges typical for cereal grains and usually did not exceed the maximum values suggested by Kabata-Pendias et al. [80] for plant material to be used for food and feed purposes (Table 7); an exception was the maximum Zn content in grain in Experiment II (56.4 mg·kg⁻¹). The Cu content in grain in Experiments II and IV was within the range of 3–6 mg·kg⁻¹, defined by Korzeniowska and Kantek [104] as optimal for plants and for grain consumers (humans and animals), while in Experiments I and III, it usually did not exceed the level of 3 mg·kg⁻¹. The Zn content, in turn, rarely reached or exceeded 45 mg, i.e., the target value of wheat grain Zn biofortification, which is considered optimal to meet the Zn needs of the human body [105].

The contents of As, Cu, Ni, and Zn in wheat straw in all experiments (Table 12) were normal or sufficient (Table 7). The Cr content in wheat straw was elevated, especially in Experiments II and IV, and the maximum value obtained in Experiment II (5.97 mg·kg⁻¹) even fell within the range considered excessive or toxic for plants. However, none of the values exceeded the permissible or suggested limits for a feed use plant material and no symptoms of Cr toxicity [92] were observed in wheat plants.

In each experiment, the content of As, Cr, and Ni in straw was noticeably higher than in grain, which is consistent with the common opinion that the pattern of accumulation of these PTE in the various parts of the plant is: roots > stem > leaves > seed/grain [106,107]. The relatively higher As content in winter wheat straw (Experiments III and IV) than in spring wheat straw (Experiments I and II) is explained by differences across the cultivars in PTE accumulation and translocation to different plant parts [108]. The noticeably higher Cr content in wheat straw in Experiments II and IV was probably due to the increased Cr solubility and availability in soil pH < 5.5 [4].

The level of Cu in wheat straw and grain was similar in three of the four experiments conducted, and only in Experiment III was a noticeably greater amount of Cu accumulated in straw than in grain. In the available literature, different patterns for the Cu accumulation in wheat grain and straw were reported: grain Cu > straw Cu [109], grain Cu \approx straw Cu [110], grain Cu < straw Cu [107]. Based on existing knowledge, the Cu amount loaded into wheat grain depends on both the Cu amount taken up by the plant post-anthesis and the amount that is remobilized (quite readily) from vegetative organs as they senesce [111]. According to Hill et al. [112], the Cu retranslocation from vegetation to grain may be limited due to Cu retention by senesced vegetation at marginal Cu content, and large numbers of tillers produced by wheat may have competed with the developing grain for retranslocated Cu. It seems that the latter is the best explanation for the described result from Experiment III, since the tillering coefficients for wheat in this experiment were the highest [113].

Experiment	P-Fertilizer	P-Dose, kg∙ha ⁻¹	Plant Protection	As	Cr	Cu	Ni	Zn
	No P	0	PP-	0.277	0.327	2.55	0.321	5.18
	SP ¹	21		0.127	0.288	2.54	0.292	2.83
	A_sBm	21		0.250	0.311	2.61	0.245	3.27
1	B_sBm	21		0.381	0.486	2.71	0.344	3.75
	Me			0.249	0.358	2.63	0.299	1.41
	Max			0.578	0.988	3.07	0.496	10.83
	No P	0	PP-	0.171	2.99	3.36	0.421	18.52
	SP	17.6		0.050	1.95	3.51	0.458	14.01
		26.4		0.156	3.18	3.78	0.535	13.69
		35.2		0.073	3.46	3.53	0.576	14.51
	A _s Bm	17.6		0.081	2.49	3.43	0.454	13.30
	5	26.4		0.106	1.98	3.49	0.337	14.22
		35.2		0.076	2.06	3.66	0.352	13.73
		average		0.102	2.59	3.54	0.447	14.57 a ²
II	No P	0	PP+	0.084	3.08	3.63	0.339	10.44
	SP	17.6		0.085	3.66	4.68	0.606	9.20
		26.4		0.121	2.05	4.01	0.307	9.41
		35.2		0.069	2.16	3.76	0.263	8.30
	A_sBm	17.6		0.112	3.58	3.69	0.692	9.35
		26.4		0.067	1.57	3.10	0.133	8.78
		35.2		0.050	2.24	4.47	0.396	9.84
		average		0.084	2.62	3.90	0.391	9.33 b
	Me			0.050	1.98	3.67	0.409	11.12
	Max			0.474	5.97	5.76	1.420	24.95
	No P	0	PP+	0.516	0.415	6.43	0.530	6.16
	SP	17.6		0.393	0.493	6.13	0.710	6.78
	01	26.4		0.373	0.484	6.11	0.284	6.41
		35.2		0.556	0.385	6.06	0.440	5.98
	A _a Af	17.6		0.343	0.315	5.83	0.277	5.98
	5)	26.4		0.582	0.531	6.37	0.221	6.28
III		35.2		0.458	0.492	4.08	0.236	6.20
	$AB_{\alpha}Af$	17.6		0.515	0.430	7.02	0.248	5.84
	5)	26.4		0.465	0.690	6.20	0.384	5.75
		35.2		0.415	0.599	6.08	0.558	6.12
	Me			0.426	0.450	6.17	0.284	6.29
	Max			0.961	1.546	9.26	1.492	8.23
	No P	0	PP+	0.453	0.99	4.12	0.234	7.36
	SP	17.6		0.643	0.71	6.34	0.310	7.54
		26.4		0.738	1.11	5.00	0.235	6.79
		35.2		0.448	1.03	5.39	0.256	6.82
	AB_{g}	17.6		0.505	0.79	5.96	0.503	7.25
157	0	26.4		0.475	1.29	5.04	0.296	6.17
1V		35.2		0.611	0.91	3.85	0.230	7.81
	AB_gBm	17.6		0.609	0.40	5.76	0.304	7.50
	2	26.4		0.786	1.25	5.72	0.288	8.34
		35.2		0.786	0.94	4.54	0.402	8.64
	Me			0.530	0.861	4.66	0.221	7.01
	Max			1.561	2.651	8.97	0.896	12.48

Table 12. PTE content in wheat straw, $mg \cdot kg^{-1} DM$ (n = 4).

¹ SP—superphosphate, A_sBm—SSA-based suspension fertilizer with *B. megaterium*, B_sBm—bone-based suspension fertilizer with *B. megaterium*, A_gAf—SSA-based granular fertilizer with *A. ferrooxidans*, AB_gAf—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_g—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_g—SSA-bone-based granular fertilizer with *B. megaterium*, ² different letters indicate significant differences at *p* < 0.05 for plant protection treatments in Experiment II; no letters—no significant differences for plant protection treatments or for P-fertilization treatments.

The Zn content in wheat straw was considerably lower than in wheat grain in all experiments. A similar relationship between straw Zn content and grain Zn content was shown by Al-Othman et al. [107]. Although Zn translocation from root to shoot following Zn uptake by the root is affected by many factors [114], it seems that adequate N supply was the reason for the greatest Zn accumulation in wheat grain in all fields in the present study, as suggested by other authors [115,116]. The particularly low straw Zn content in Experiment I was probably related to the low Zn abundance in the soil (Table 6).

The use of fertilizers containing PTE, both from primary and secondary raw materials, has been associated with concerns about the accumulation of these elements in edible plant parts, including cereal grain [99,117,118]. In the present study, the main raw material for recycled fertilizers was SSA. Although direct use of unprocessed SSA in agriculture is legally limited [119,120], it is recommended to produce fertilizers based on new SSA processing technologies that result in concentrated and pure fertilizer products [43,121]. To date, however, there are still few studies on SSA and SSA-based fertilizers in the context of their effect on PTE accumulation in usable/consumable plant parts [118,122,123].

Reports from pot experiments provide evidence that SSA originating from the 'Łyna' wastewater treatment plant, Olsztyn, used as P-fertilizer did not lead to a significant increase in PTE (Fe, Mn, Zn, Cu, Cd, Pb, Ni, Cr) concentrations in green forage maize [123] and only very slightly modified the PTE content (Cd, Cr, Cu, Ni, Mn, Zn) in the Virginia fanpetals plants [122]. Field studies by Iżewska and Wołoszyk [118] demonstrated that the application of SSA (from Pomorzany Sewage Treatment Plant in Szczecin) at rising P doses caused an increase in the content of Cd, Ni, and Pb in maize grain, Cd and Ni in maize straw, Cd and Pb in spring rape seeds, and Cd in rape straw. In the authors' previous paper, it was reported that SSA-based biofertilizers applied at reasonable amounts in field conditions did not increase the Cd and Pb content in wheat grain [60].

Many other studies have been published in which the secondary sources of nutrients did not contribute to increasing the PTE content of the crop plant grown in soil amended in this manner [124–127]. There are also opinions that the levels of elements such as Cu and Zn normally found in waste-based soil amendments may be beneficial rather than toxic to crop plants [90]. However, considering the chemical heterogeneity of secondary nutrient sources [43] and the complexity of the fertilizer/nutrient source–soil–plant system functioning [128], using them with caution is advisable until long-term studies have verified their full utility and safety.

Plant protection against agrophages (Experiment II) increased Cu content in wheat grain and decreased Zn content in wheat straw. The former effect is explained by the reduction in weed competition for Cu [129], and the latter by a 'dilution effect' [130] of Zn in wheat straw with considerably higher yields under PP+ than under PP- treatments (unpublished data). However, in higher grain yield under PP+, the Zn dilution was not statistically confirmed. No effect of plant protection on As, Cr, Ni, and Zn content in grain or on As, Cr, Cu, and Ni content in straw was proven. In comparison, Wołejko et al. [131,132] found higher Cu and Cr contents in grain and plants when wheat fertilized with granular sludge was protected with herbicide and fungicides in relation to the control treatment, while no differences were found in Cd, Ni, Pb, and Zn contents.

3.3.2. Weeds

Weeds compete with crops for nutrients [132], but by depleting PTE content in the shared rhizosphere, they may protect crops from excessive PTE uptake as well [133]. However, in some cases, accumulator weeds can enhance PTE uptake by neighboring crop plants [134] and alter PTE allocation in the different organs of crop plants (i.e., phytoenrichment). This phenomenon may be a potentially serious problem if it leads to an increase in the PTE accumulation in the edible parts of crop plants [133].

The As, Cr, Cu, Ni, and Zn contents determined in the weed biomass in the present study (Table 13) fell within the biological plant variability ranges (Table 7). Although none of the experiments conducted demonstrated any significant effect of the applied P-

fertilizer treatments on these PTE content in the weed biomass (Table 13), plant protection differentiated the content of As, Cr, Ni, and Zn in Experiment II.

Experiment	P-Fertilizer	P-Dose, kg ha ^{−1}	Plant Protection	As	Cr	Cu	Ni	Zn
	No P	0	PP-	0.180	0.537	5.13	0.331	16.4
	SP^{1}	21		0.329	0.404	5.28	0.339	16.2
Ι	A_sBm	21		0.318	0.670	5.13	0.377	17.7
	B _s Bm	21		0.254	0.524	4.91	0.384	14.7
	Me			0.242	0.539	5.19	0.335	16.2
	Iviax	2	DD	0.371	0.900	5.95	0.020	20.5
	NO P SP	0 17.6	PP-	0.935	2.217	7.62 8.01	1.254	76.8 77 7
	01	26.4		0.704	2.247	8.72	1.211	68.8
		35.2		0.677	1.937	8.15	1.355	80.5
	A_sBm	17.6		0.822	1.717	7.94	1.395	79.5
		26.4		1.095	2.178	8.73	1.429	76.1
		35.2		0.916	2.652	8.16	1.776	68.7
п		average		0.870 a ²	2.133 a	8.19	1.411 a	75.4 a
11	No P	0	PP+	0.140	1.545	8.80	0.908	64.1
	SP	17.6		0.322	1.718	8.18	0.850	44.2
		20.4		0.443	1.550	7.65	1.094	40.3 49.8
	$A_{a}Bm$	17.6		0.216	1.323	7.92	1.059	52.1
	3	26.4		0.146	2.129	7.11	1.294	52.0
		35.2		0.113	1.861	9.99	1.189	59.8
		average		0.298 b	1.708 b	8.20	1.055 b	52.9 b
	Me			0.476	1.832	8.04	1.178	65.5
	Max			1.748	4.446	11.02	2.300	105.7
	No P		PP+	1.838	0.853	10.7	3.49	75.6
	SP	17.6		1.586	0.298	9.7 14.4	4.86	83.1
		35.2		1.311	0.822	14.4 10.3	2.34	82.9
	$A_{\alpha}Af$	17.6		1.532	0.628	11.1	4.25	82.2
TTT	8 9	26.4		1.354	1.254	13.8	3.27	89.8
111		35.2		2.204	0.493	12.6	4.10	115.1
	AB _g Af	17.6		1.656	0.215	15.9	3.62	74.3
		26.4		1.648	0.351	10.3	4.76 5.91	105.6
		55.2		1.049	1.002	12.7	2.02	102.5
	Me Max			1.634	1.093	10.6 26.4	3.92 9.39	102.5 140 5
	NoP	0	DD 1	0.873	0.547	7.60	1 270	110.0
IV	SP	17.6	11+	0.875	1 878	7.00	2 161	40.4 50 1
	01	26.4		0.672	0.240	9.05	1.265	51.3
		35.2		0.836	1.251	8.06	1.488	40.7
	ABg	17.6		0.981	1.586	8.06	1.832	49.6
		26.4		1.059	1.032	8.36	2.060	55.9
	AB Rm	55.2 17.6		0.836	0.578	7.0U 9./1	1.700	43.0 62.2
	Auguni	26.4		0.030	0.240 0.458	8.81	2.234	59 5
		35.2		0.774	0.736	8.92	1.838	56.7
	Me			0.894	0.687	8.13	1.735	51.7
	Max			1.799	3.792	12.45	4.442	76.2

Table 13. PTE content in weeds, $mg \cdot kg^{-1}$ DM (n = 4).

¹ SP—superphosphate, A_sBm —SSA-based suspension fertilizer with *B. megaterium*, B_sBm —bone-based suspension fertilizer with *B. megaterium*, A_gAf —SSA-based granular fertilizer with *A. ferrooxidans*, AB_gAf —SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_gAf —SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_gAf —SSA-bone-based granular fertilizer with *B. megaterium*, ² different letters indicate significant differences at *p* < 0.05 for plant protection treatments in Experiment II; no letters—no significant differences for plant protection treatments.

The variability of PTE content in weed biomass within a single experiment and between experiments resulted from the species composition and structure of weed communities, along with differentiation related to weed developmental stages, competitive ability, and susceptibility to the herbicide applied. Weed biomass from the wheat field under PP– conditions was dominated by *Chenopodium album*, *Fallopia convolvulus*, and *Raphanus raphanistrum* (unpublished data), which are known to accumulate large amounts of As, Cr, Ni, and Zn [135–139]. The reduction in the share of these species in the community due to the herbicide use (PP+) contributed to a decrease in the content of the aforementioned PTE in the overall biomass of weeds and was manifested by a significant difference in relation to weed biomass under PP–. Relatively high Cu, Ni, and Zn contents in weed biomass were found in Experiment III. The weed community in this experiment was dominated by *Viola arvensis*, which proved to be quite resistant to the applied herbicide. *Violaceae* are generally considered highly tolerant to heavy metals [140], and the considerable ability of *Viola arvensis* to accumulate Cu, Ni, and particularly Zn, was also reported by other authors [135,136,141].

In the present experiments, the PTE contents in weed biomass often exceeded their content in wheat grain and straw. This can have both positive (protection against excessive PTE uptake by the crop) [133] and negative aspects (depriving the crop of a valuable nutrient if the given toxic element is a micronutrient in deficiency) [142].

Although PTE accumulation by weeds and the role of weeds in the phytoremediation of PTE-contaminated habitats has been frequently emphasized in the literature [138,143–145]; however, to date, weeds have not been studied for uptake and accumulation of PTE from recycled fertilizers applied to field crops. Galal and Shehata [146] reported from their experiment with rice irrigated with water canals receiving wastewater discharges that weeds, apart from limiting the growth and production of rice crops by accumulating large amounts of nutrients, had captured heavy metals from the soil and thus improved the quality of rice grains. However, Zubkova et al. [139] claim that the contribution of weeds to the general elimination of trace elements and heavy metals by plants of agrophytocenoses is insignificant.

Since the ecological/ecosystemic functions of weeds, including their contribution to the balance of organic matter, nutrient, and toxic element cycling in the agroecosystem, have begun to be appreciated, the need for research in this area has not weakened [146,147].

3.3.3. Post-Harvest Residues

PTE captured and stored in crop roots as well as in the straw (stubble) and weed parts remaining in the field after harvesting are temporarily immobilized in this biomass, avoiding potential rapid leaching into groundwater and water ecosystems [5]. They can then be, however, slowly released and used by succeeding plants [148,149].

In the present study, the PTE contents in post-harvest residues in the individual experiments (Table 14) were determined by the proportion between the biomass of the roots, stubble, and weeds. In addition, weed species composition, weed relative height in the wheat canopy, and the elemental content of those weed parts (roots, stems, or leaves) that predominated in the biomass left in the field, also mattered [150].

In all the conducted experiments, the content of As, Cu, Ni, and Zn in crop residues can be considered natural (Table 7), except for a slightly elevated maximum Ni value in Experiment IV and a particularly low Zn level in Experiment I, which were most likely due to the soil background (Table 6). The Cr content of crop residue biomass in all experiments fell already within the range of values classified as excessive or toxic; however, this could not be related to the P-fertilizers used, including recycled ones, as the same Cr level was also recorded under no P treatments. The P-fertilizer treatments did not differentiate the content of the tested PTE in post-harvest residues in any of the experiments conducted (Table 14).

Experiment	P-Fertilizer	P-Dose, kg∙ha ⁻¹	Plant Protection	As	Cr	Cu	Ni	Zn
	No P	0	PP-	0.577	5.64	3.52	2.08	7.79
	SP ¹	21		0.627	5.75	3.90	2.26	9.61
	A _s Bm	21		0.598	6.28	4.04	2.49	9.82
Ι	B_sBm	21		0.408	5.96	4.27	2.53	8.91
	Me			0.541	5.92	3.87	2.31	9.17
	Max			0.778	7.06	5.10	2.92	11.71
	No P	0	PP-	0.438	7.97	4.17	2.24	26.0
	SP	17.6		0.369	6.46	4.79	1.94	19.5
		26.4		0.603	8.64	4.59	2.26	19.7
		35.2		0.385	8.50	4.95	2.29	20.6
	A_sBm	17.6		0.392	8.58	4.57	2.21	21.2
		26.4		0.487	8.88	4.11	2.49	25.2
		35.2		0.548	7.23	4.41	2.31	19.9
		average		0.460 a ²	8.04	4.51 b	2.25	21.7
II	No P	0	PP+	0.372	10.83	4.67	1.88	24.0
	SP	17.6		0.318	11.35	5.64	1.95	21.9
		26.4		0.245	7.92	6.01	1.87	20.9
		35.2		0.369	9.10	5.03	2.12	25.0
	A _s Bm	17.6		0.320	7.16	4.82	2.84	22.0
	3	26.4		0.309	9.05	5.90	1.74	19.0
		35.2		0.395	8.13	4.81	2.32	23.3
		average		0.332 b	9.08	5.27 a	2.10	22.3
	Me			0 375	8 32	4 84	2.04	21.9
	Max			0.873	16.78	8.04	4.07	34.2
	No P	0	PP+	0.634	8.92	5.33	2.73	23.5
	SP	17.6		0.355	9.13	5.41	2.31	22.8
	01	26.4		0.333	9.81	5.63	2.74	20.8
		35.2		0.232	9.78	5 59	2.60	24.3
	A. Af	17.6		0.452	10.35	6.10	3.12	29.5
	1 1 g2 1j	26.4		0.132	10.00	5 38	3.10	29.0
III		25.4		0.407	9.80	5.30	2.68	23.0
	AB Af	17.6		0.407	9.00 8.80	5.27	2.00	23.0
	Abgrij	17.0 26.4		0.080	10.05	5.63	2.17	22.0
		35.2		0.000	8.51	5.74	2.33	24.3
	Mo			0 227	9 75	5 32	2 51	24.3
	Max			1.074	13.40	8.76	4.24	34.3
	No P	0	PP+	0.425	6.98	4.86	4.48	20.5
	SP	17.6		0.734	8.78	4.85	7.26	21.9
	01	26.4		0.674	8 80	5 19	6.72	22.0
		35.2		0 379	7.81	4 48	5.28	20.0
	ΔB	17.6		0.685	9.25	4 56	636	20.0
	2 sDg	26.4		0.695	7 43	4 13	6.46	21.3
IV		25.9		0.398	6.81	1.10 1 10	5.10	20.6
	AB. Rm	17.6		0.590	7 3/	4 58	5.02	20.0 19.8
	Auguni	26.4		0.007	9.24	т.00 5 10	7.86	22 O
		35.2		0.779	8.02	5.56	6.56	24.2
	<u>}</u>			0.(52		4 4 4	(00	01.4
	Me			0.652	7.757	4.61	6.32	21.4
	Max			1.801	15.297	7.19	11.54	29.7

Table 14. PTE content in post-harvest residues, $mg \cdot kg^{-1}$ DM (n = 4).

¹ SP—superphosphate, A_sBm—SSA-based suspension fertilizer with *B. megaterium*, B_sBm—bone-based suspension fertilizer with *B. megaterium*, A_gAf—SSA-based granular fertilizer with *A. ferrooxidans*, AB_gAf—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_g—SSA-bone-based granular fertilizer with *A. ferrooxidans*, AB_g—SSA-bone-based granular fertilizer with *B. megaterium*, ² different letters indicate significant differences at *p* < 0.05 for plant protection treatments in Experiment II; no letters—no significant differences for plant protection treatments or for P-fertilization treatments.

Previous studies have shown that roots of crops cultivated in agricultural soil tend to accumulate greater amounts of PTE than the above-ground parts, acting as a barrier to their translocation and protecting edible parts from PTE contamination [107,151,152]. For Cr, it has been reported that plant roots can accumulate up to 100-fold higher amounts of this element than shoots [92]. The very low Zn content of crop residues in Experiment I suggests a deficit of available Zn in the soil, along with the relatively lowest initial content of total soil Zn (Table 6), under which wheat plants first stored needed Zn amounts in the grain [153].

In Experiment II, post-harvest residues showed a lower content of As and higher of Cu under PP+ than under PP-. The lower As content in residue biomass under PP+ is explained by the reduction/elimination from weed communities of those weed species that accumulate more As in their tissues (see Section 3.3.2). In turn, since no difference in Cu content in weed biomass under PP+ and PP- was found (see Table 13), the higher Cu content in post-harvest residues under PP+ likely resulted from an increase in the proportion of wheat roots in the residue biomass, which usually contain more Cu than wheat stems [107,152].

No other studies were found on the effect of recycled fertilizers on As, Cr, Cu, Ni, and Zn contents in post-harvest residues, although a previous paper by the authors found that biofertilizers from renewable raw materials, with low contents of Cd and Pb, did not alter the PTE contents in post-harvest residues of wheat fields [60]. Partial reference may be provided by studies involving PTE content in wheat straw (see Section 3.3.1), weeds (see Section 3.3.2), and roots, i.e., those plant organs whose parts (remnants) are left in fields after harvest. However, these studies are also scarce.

Considering the retention of PTE in crop residues [149], as well as their carryover between crop rotations [154], more field experiments in this matter are needed.

4. Conclusions

The recycled fertilizers tested did not increase the As, Cr, Ni, Cu, and Zn contents of the soil and plant biomass when applied in amounts up to 35.2 kg·P·ha⁻¹. The contents of these elements in soil were below the permissible levels for arable land in Poland. Their concentrations in wheat grain and straw did not exceed the permissible or suggested limits for plant material to be used for food and feed, while in the weed and post-harvest residue biomass, they usually fell within the biological plant variability ranges. Such findings may be one of the first steps toward recommending the presented recycled formulations to replace or supplement traditional commercial fertilizers from primary raw materials. However, considering the potential for PTE accumulation in (and leaching from) the soil, further long-term field studies on PTE fate with repeated applications of recycled fertilizers are needed.

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