

Article

Purpose-Oriented Antibody Libraries Incorporating Tailored CDR3 Sequences

Pauline Bonvin [†], Sophie Venet [†], Marie Kosco-Vilbois and Nicolas Fischer ^{*}

Novimmune S.A., 14 chemin des Aulx 1228 Plan-les-Ouates, Switzerland;

E-Mails: pbonvin@novimmune.com (P.B.); sophie.venet-bonnot@morphosys.com (S.V.);

mkosco-vilbois@novimmune.com (M.K.-V.)

[†] These authors contributed equally to this work.

^{*} Author to whom correspondence should be addressed; E-Mail: nfischer@novimmune.com;
Tel.: +41-79-102-62-31; Fax: +41-22-839-71-42.

Supplementary Materials

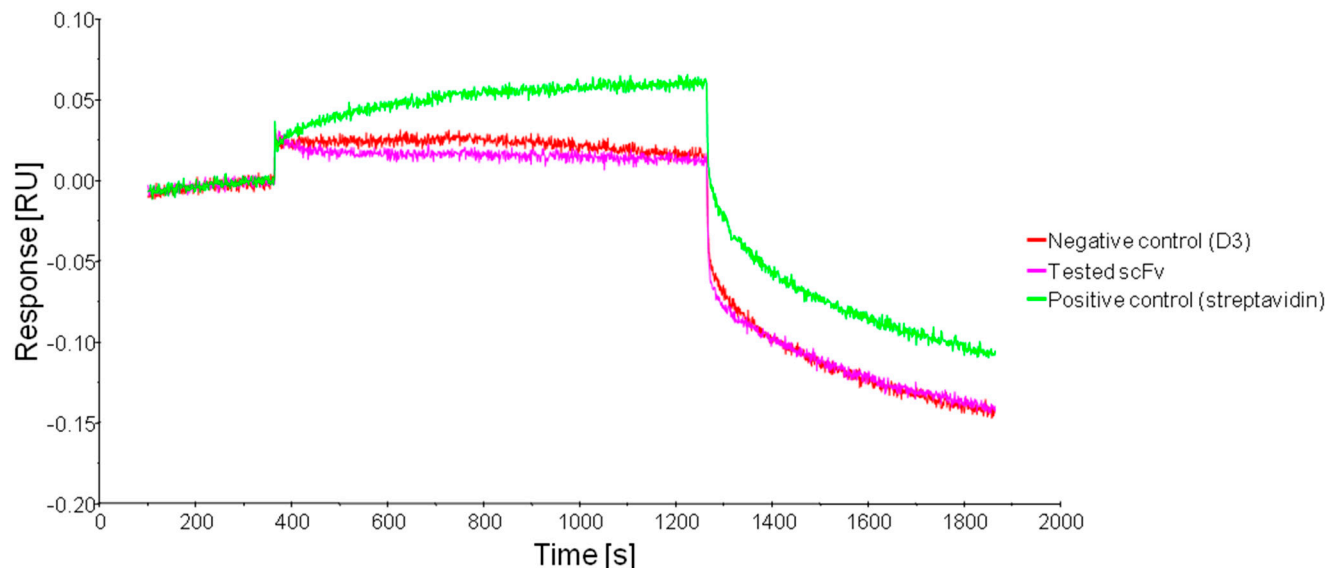


Figure S1. Competition for HRP binding by BLI. Biotin-HRP was loaded onto streptavidin biosensors. Sensors were then dipped into a solution containing the clone D3 and subsequently transfer to wells containing the indicated scFv or recombinant streptavidin as a positive control. None of the candidates tested was able to bind to the biosensors following the incubation with D3, demonstrating that all candidates have overlapping epitopes. An example of data obtained with anti-HRP candidates is shown above.

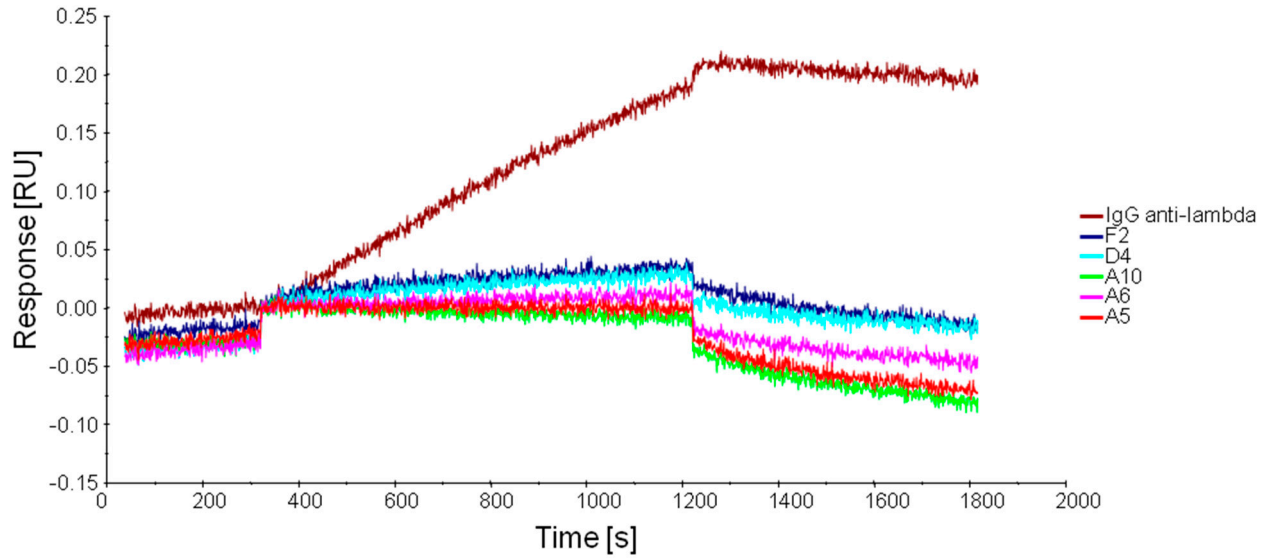


Figure S2. Competition for hFc binding by BLI. A lambda hIgG1 was loaded onto anti-human biosensors. Sensors were then dipped into a solution containing the clone A5 and subsequently transfer to wells containing the indicated scFv or a anti-lambda constant domain as a positive control. None of the candidates tested was able to bind to the biosensors following the incubation with A5, demonstrating that all candidates have overlapping epitopes.

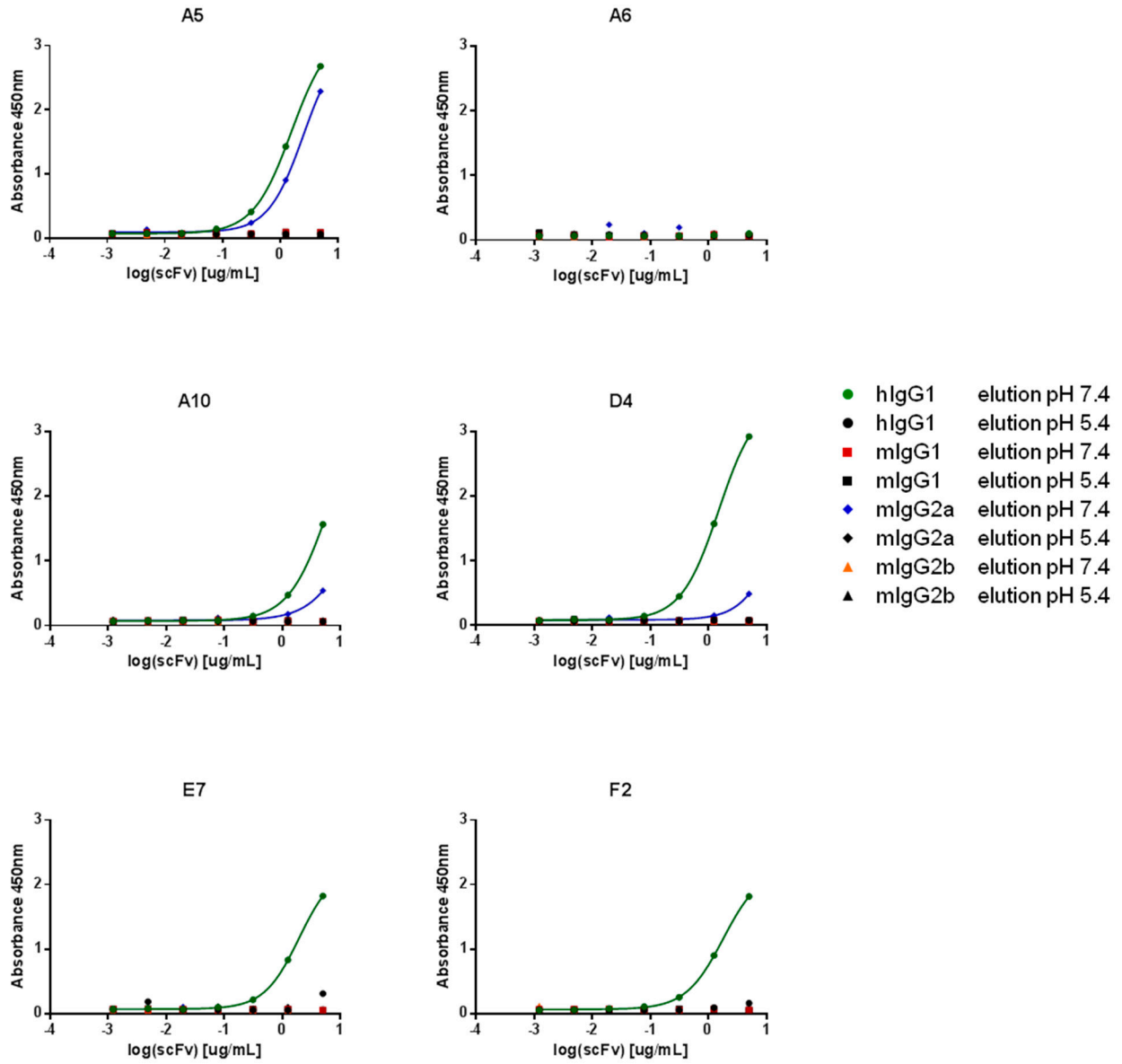


Figure S3. Specificity of hits obtained from selections against the hFc domain. The cross-reactivity of individual clones (scFv format) was assessed by pH-dependent ELISA using human and murine IgG subclasses.