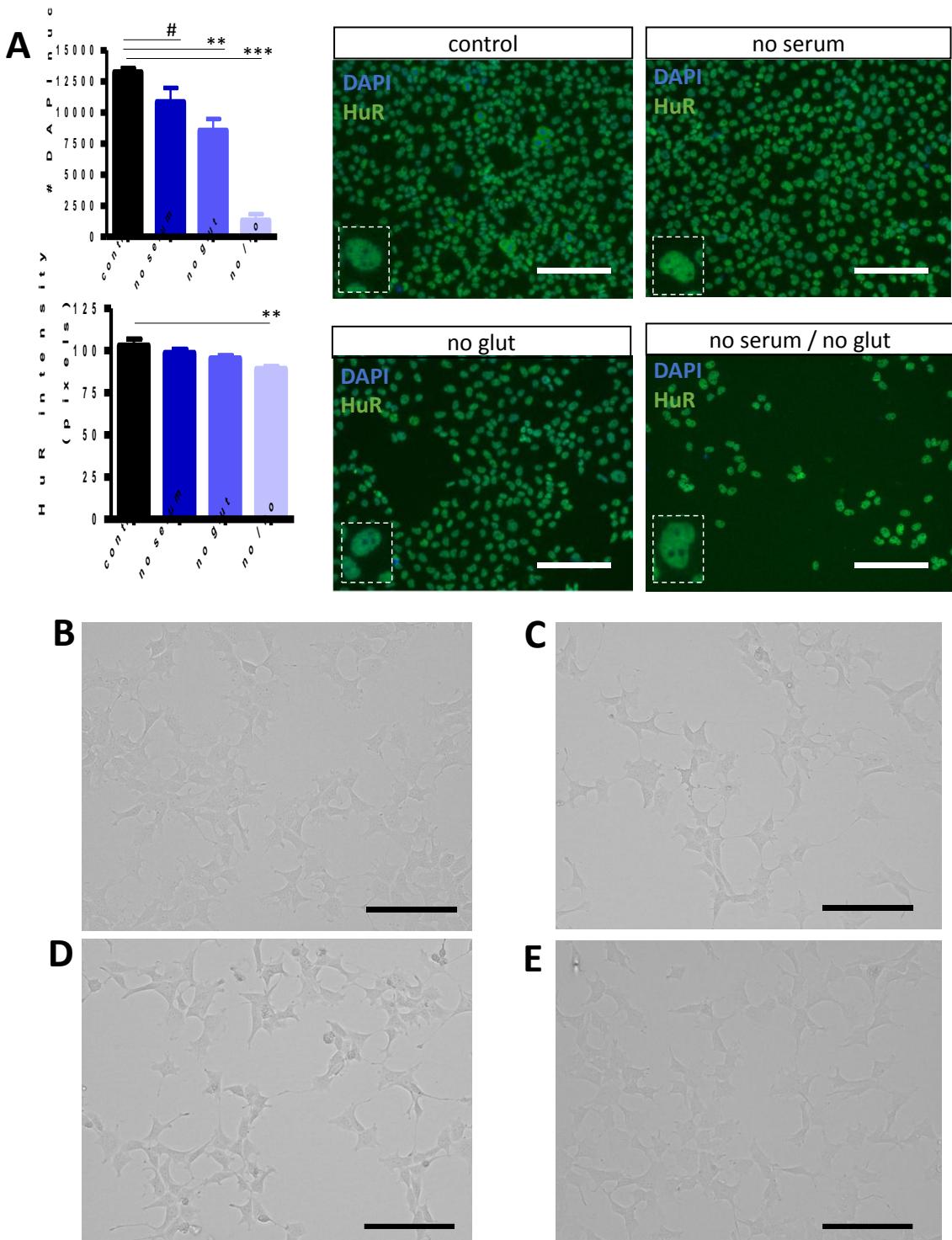
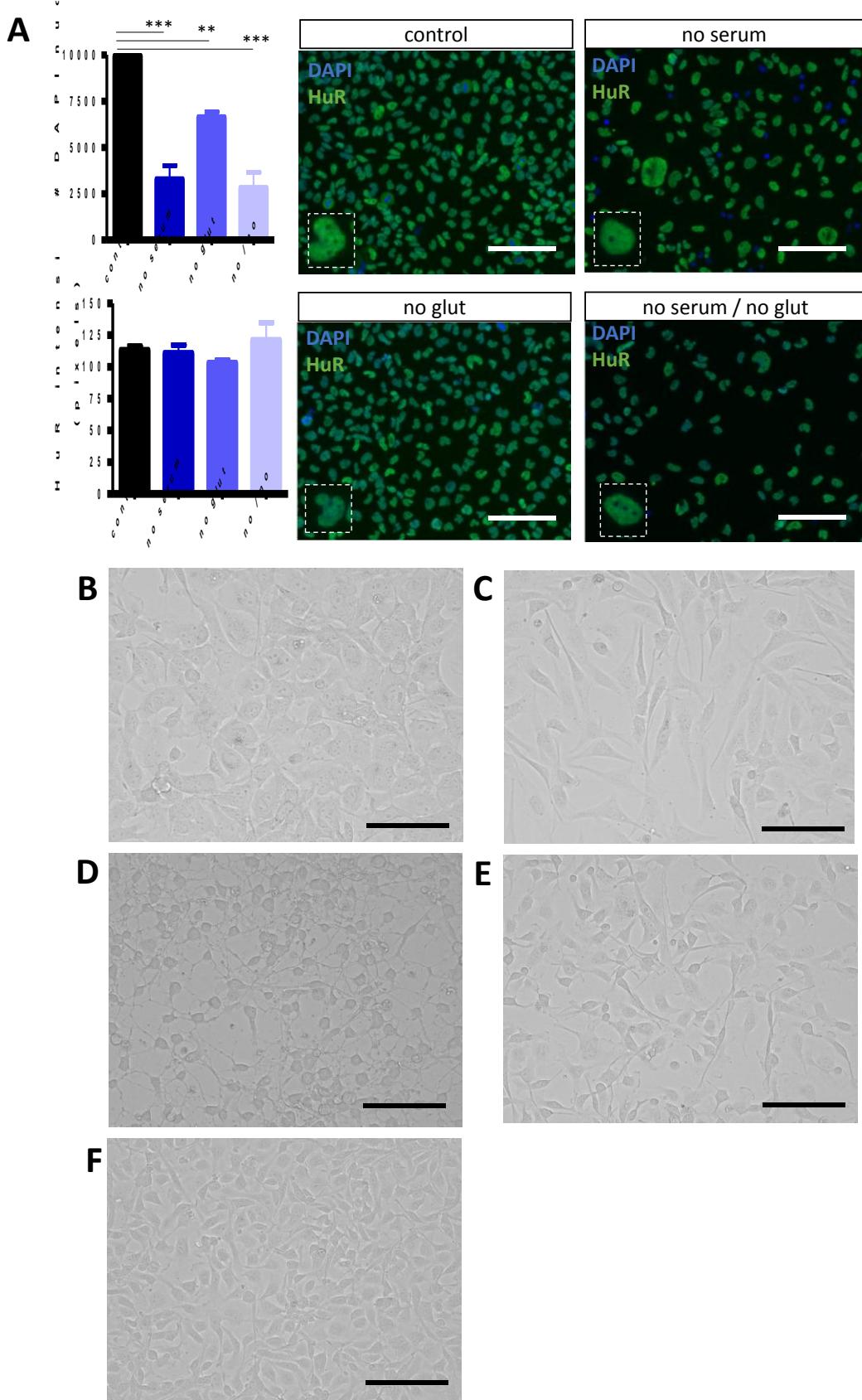


# Supplementary figures and tables

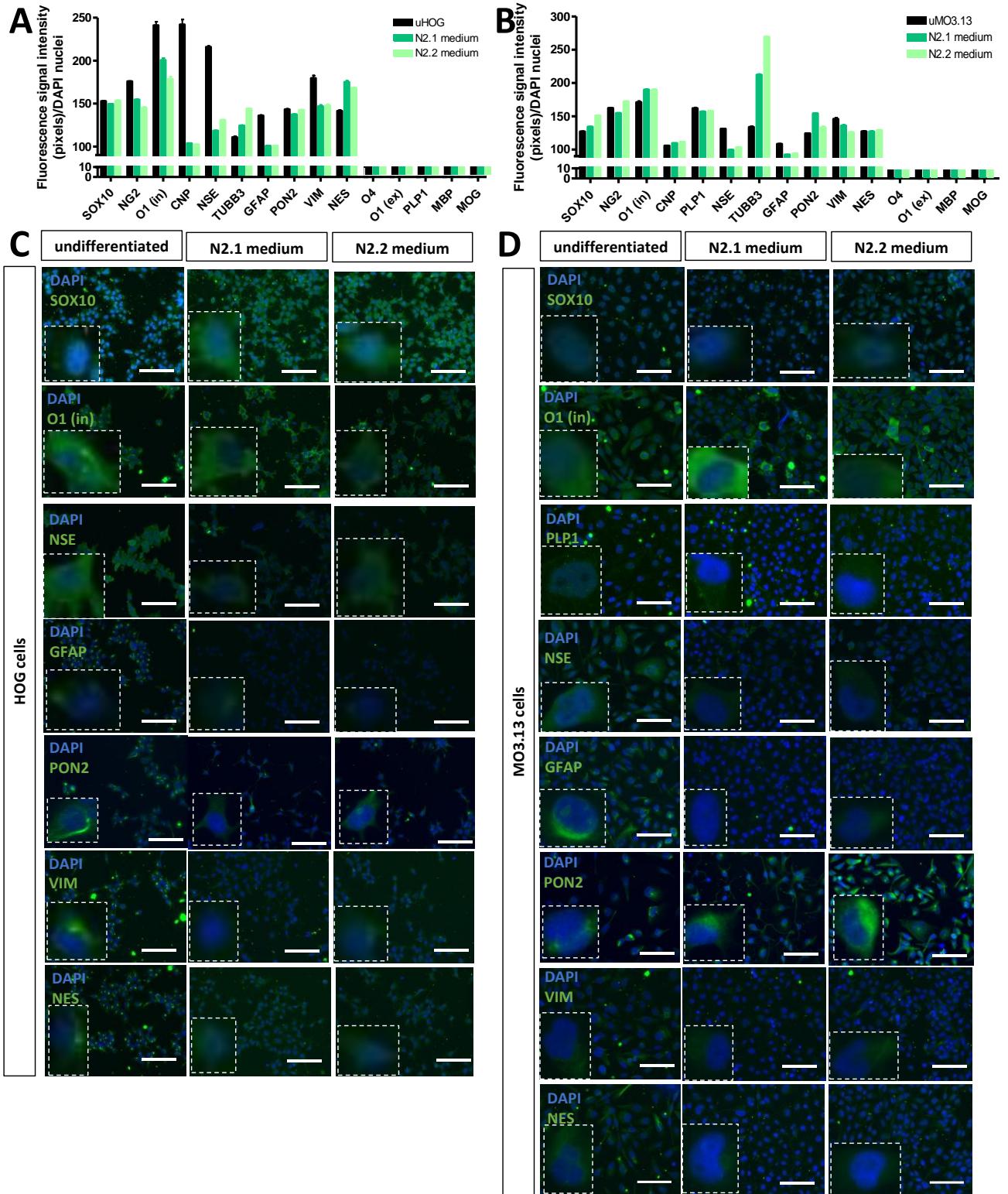
De Kleijn et al. (2019)



**Figure S1.** (A) HOG cell count (#DAPI nuclei), cytoplasmic HuR intensity quantifications and representative immunocytochemistry images of HOG cells cultured without serum, without Glutamax (glutamine) and without serum and Glutamax. Based on ANOVA with Tukey-b post-hoc comparisons on four independent experiments ( $n=4$ ). P-values: \*  $< 0.05$ , \*\*  $< 0.01$ , \*\*\*  $< 0.001$ . Morphology of (B) undifferentiated HOG and (C) HOG differentiated with N2.1 medium, (D) N2.2 medium and (E) T3. Scale bar = 50  $\mu$ m.



**Figure S2.** (A) MO3.13 cell count (#DAPI nuclei), cytoplasmic HuR intensity quantifications and representative immunocytochemistry images of MO3.13 cells cultured without serum, without Glutamax (glutamine) and without serum and Glutamax. Based on ANOVA with Tukey-b post-hoc comparisons on four independent experiments ( $n=4$ ). P-values: \*  $< 0.05$ , \*\*  $< 0.01$ , \*\*\*  $< 0.001$ . Morphology of (B) undifferentiated MO3.13 and differentiated with (C) PMA, (D) N2.1 medium, (E) N2.2 medium and (F) T3. Scale bar = 50  $\mu$ m.



**Figure S3.** Immunocytochemical analyses in undifferentiated and differentiated HOG and MO3.13 cells. **(A)** Quantification of fluorescence signal in immunocytochemistry for undifferentiated HOG cells (uHOG) and HOG cells differentiated with N2.1 or N2.2 medium. Fluorescence quantification was based on one experiment ( $n=1$ ), error bars = SEM of total number of DAPI-positive nuclei in one condition. **(B)** Quantification of fluorescence signal in immunocytochemistry for undifferentiated MO3.13 cells (uMO3.13) and MO3.13 cells differentiated with N2.1 or N2.2 medium. Fluorescence quantification was based on one experiment ( $n=1$ ), error bars = SEM of total number of DAPI-positive nuclei in one condition. **(C)** Representative immunocytochemistry images for each marker in HOG cells and **(D)** MO3.13 cells. Scale bar = 50  $\mu$ m.

**Table S1.** Variables and operationalisations used for SH-SY5Y and HOG or SH-SY5Y and MO3.13 co-culture experiments. OL: oligodendrocyte.

Variable	Operationalisation
Coating	<ul style="list-style-type: none"> <li>- Matrigel 1:100 (Corning)</li> <li>- Poly-L-ornithine (Sigma) + Laminin (Sigma)</li> </ul>
Total cell density	<ul style="list-style-type: none"> <li>- <math>2.8 \cdot 10^5</math> cells/cm<sup>2</sup></li> <li>- <math>2.25 \cdot 10^5</math> cells/cm<sup>2</sup></li> <li>- <math>1.8 \cdot 10^5</math> cells/cm<sup>2</sup></li> <li>- <math>1.125 \cdot 10^5</math> cells/cm<sup>2</sup></li> <li>- <math>5.6 \cdot 10^4</math> cells/cm<sup>2</sup></li> <li>- <math>2.8 \cdot 10^4</math> cells/cm<sup>2</sup></li> </ul>
OL to neuron ratio	<ul style="list-style-type: none"> <li>- 1:1</li> <li>- 1:2</li> <li>- 1:3</li> <li>- 1:6</li> <li>- 1:9</li> <li>- 1:18</li> </ul>
Co-culture duration	<ul style="list-style-type: none"> <li>- 8 days</li> <li>- Longer than 8 days</li> </ul>
Pre-differentiation of OLs in mono-culture	<ul style="list-style-type: none"> <li>- Yes</li> <li>- No</li> </ul>
Time-point of OL addition after neuronal differentiation initiation (day 10)	<ul style="list-style-type: none"> <li>- Day 10</li> <li>- Day 11</li> <li>- Day 14</li> <li>- Day 17</li> <li>- Day 19</li> <li>- Day 21</li> </ul>
Medium change mode	<ul style="list-style-type: none"> <li>- 100% of medium</li> <li>- 50% of medium</li> </ul>
Basal differentiation medium	<ul style="list-style-type: none"> <li>- Neurobasal (Thermo Fisher)</li> <li>- Brainphys (STEMCELL technologies)</li> </ul>
Medium supplements	<ul style="list-style-type: none"> <li>- D-Glucose (Merck)</li> <li>- Sodium pyruvate (Gibco)</li> <li>- Glutamax (Gibco)</li> <li>- B27 supplement (Thermo Fisher)</li> <li>- SM1 supplement (STEMCELL technologies)</li> <li>- Culture-one supplement (Thermo Fisher)</li> </ul>
Proteins/hormones	<ul style="list-style-type: none"> <li>- Brain-derived neurotrophic factor (BDNF) (Sigma)</li> <li>- All-trans retinoic acid (ATRA) (Sigma)</li> <li>- Apo-transferrin (Sigma)</li> <li>- Putrescine (Sigma)</li> <li>- Triiodothyronine (T3) (Sigma)</li> <li>- Sodium selenite (Sigma)</li> <li>- Dibutyryl-cAMP (Sigma)</li> <li>- Progesterone (Sigma)</li> <li>- Human insulin (Sigma)</li> <li>- Hydrocortisone (Sigma)</li> <li>- D-biotin (Sigma)</li> <li>- L-Glutamic Acid (Sigma)</li> <li>- IBMX (Sigma)</li> <li>- Ascorbic acid (Sigma)</li> <li>- N-acetyl-cysteine (NAC) (Sigma)</li> <li>- IGF-1 (Cell Guidance Systems)</li> </ul>

	- PMA (Sigma) - Lithium chloride (LiCl) - Sodium chloride (NaCl) - Potassium chloride (KCl)
Small molecules	- Miconazole (Sigma) - Bradykinin (Sigma) - Montelukast (Tebu-Bio) - Prosaptide (Sigma) - Clobetasol (Sigma) - Carbachol (Sigma) - MDL29,951 (Tebu-Bio) - ATP (Sigma)

**Table S2.** Vendors and concentrations of antibodies used in immunohistochemistry (ICC) and Western blot (WB) experiments.

Target	Vendor(clone)	Dilution for ICC	Dilution for WB
CC1	Abcam (16794)	1:500	
CNPase	Aves Labs	1:250	1:500
DBH	Abcam (ab96615)	1:500	
FluoroMyelin	Thermo Fisher Scientific	1:100	
Red			
GAPDH	Cell Signaling Technology (14C10)		1:2000
GAP43	Aves Labs	1:250	
GFAP	UC Davis/NIH NeuroMab Facility (N206A/8)	1:500	
HuR	Santa Cruz (3A2)	1:200	
Jagged1	Santa Cruz (E-12)	1:500	
L1CAM	Santa Cruz (5G3)	1:500	
MBP	Aves labs (polyclonal)	1:500	1:2000
MBP	Millipore (SMI99; monoclonal)	1:500	1:500
MOG	Home-made antibody CRICM, Pitié-Salpêtrière, Hospital, Paris	1:50	1:1000
MPZ	Aves Labs	1:250	
NEFL	Santa Cruz (DA2)	1:500	
NES	Millipore (MAB353)	1:500	
NG2	Abcam (ab129051)	1:100	
NSE	Aves Labs	1:250	
O1	Invitrogen	1:250	
O4	Home-made antibody CRICM, Pitié-Salpêtrière, Hospital, Paris	1:2	
PLP1	Abcam (ab28486)	1:500	1:2000
PON2	Santa Cruz (C-5)	1:500	1:250
SOX10	Abcam (EPR4007)	1:100	
TH	Millipore (AB152)	1:500	
TUBB3	Biolegend (801201)	1:1000	1:2000
TUBB3	Biolegend (802001)	1:1000	
VIM	DSHB (40E-C)	1:50	

**Table S3.** Sequences of primers used in HOG and MO3.13 differentiation experiments. FW: forward primer, RV: reverse primer.

Gene	Sequence	Gene	Sequence
BLBP FW	5'-GAGACAAAGTGGTCATCAGGACTC	MYRF FW	5'-CTTCAGCGTGGTGTCCATGTC
BLBP RV	5'-CCATCCAGGCTAACAAACAGACTTA	MYRF RV	5'-GCAGCAAAGAGGGCTGTATGC
CNP FW	5'-GGAGTACGCTAACAAAGATGTGA	NESFW	5'-CGGGCTACTGAAAAGTTCCAG
CNP RV	5'-CACAAAGAGGGCAGAGATGGT	NES RV	5'-ACATCTTGAGGTGCCAG
DHH FW	5'-GCCGTGCTTGACATCACTA	NEFL FW	5'-GACCCCTGAAATCGAAGCATG
DHH RV	5'-ATCAGCTTGACCGACACGTG	NEFL RV	5'-TTGATCGTGTCTGCATAGCG
EGR2 FW	5'-CACGTCGGTGACCATCTTC	PDGFRa FW	5'-GCCCCGAGGAATGGAGTTTT
EGR2 RV	5'-ATCATGCCATCTCCGGC	PDGFRa RV	5'-GCAGAAAGGTACTCCCTTCGA
EIF4A2 FW	5'-GGTGACATGGACCAGAAGGAGA	PLP FW	5'-GCTGATGCCAGAATGTATGGTG
EIF4A2 RV	5'-CCCCTCTGCCAATTCTGTGAA	PLP RV	5'-CAATCATGAAGGTGAGCAGGG
FGFR3 FW	5'- TCCTGCTCTGGGAGATCTCAC	PPIA FW	5'-CAGGGTTATGTGTCAAGGTG
FGFR3 RV	5'- TGATCATGTACAGGTGTTGTG	PPIA RV	5'- CCATTTGTTGGTCCAGC
GAPDH FW	5'-GTCATGGGTGTGAACCATGAGA	SLC1A3 FW	5'- CTTCTGGTAACACGGAAAACC
GAPDH RV	5'-GCATGGACTGTGGTCATGAGTC	SLC1A3 RV	5'- TGGGTAGGTGGCAGAACTT
GPR17 FW	5'- GAGAGATGCTAAACTCTCAGC	SOX10 FW	5'-AGGAGAAGGAGGTTGACTGTTG
GPR17 RV	5'- CAGGGAGAAGTTGGTGTACAGAC	SOX10 RV	5'-AGGTGCAGCCCCCTCATCTTC
GFAP FW	5'-CCAGGACCTGCTCAATGTCAA	TUBB3 FW	5'-GGGAAGTCATCAGTGAGCAT
GFAP RV	5'-TCCAGGCTGGTTCTCGAACATC	TUBB3 RV	5'-GAGGCACGTACTTGTGAGAAGAGG
MAG FW	5'-CCAGGGAGCCCCATCGAC	VCAN FW	5'- GAATGTCACTCTAACATCCCTGCGT
MAG RV	5'-GGTTGTCCCCCTGCGAG	VCAN RV	5'- TCACATGTCTCGGTATCTGCTC
MOBP FW	5'-CCGTTCACCTTCTCAATTCC	VEGF-A FW	5'-GCTCAGAGCGGAGAAAGCATT
MOBP RV	5'-GCTGGTTCTGGTCTTCTGGC	VEGF-A RV	5'-TCGGCTTGTACATCTGCAAG
MOG FW	5'-TTTGATCCCCACTTCTGAGG	VEGF-C FW	5'-GGGCCAACCGAGAATTGA
MOG RV	5'-CGTAGCTTCAAGGAATTGCC	VEGF-C RV	5'-GCCGTCTGTAACAGCTGCATGT
MBP FW	5'-ACCCAAGATGAAAACCCCGTA	VIM FW	5'-GAAATGGCTCGTCACCTTCGT
MBP RV	5'-TCCGTAGCAAATCCTGGTCT	VIM RV	5'-GGAAGAGGCAGAGAAATCCTGC
MPZ FW	5'-TGCAGAGGAGGCTCAGTGTAT	YWHAZ FW	5'-CGCTGGTGTGACAAGAAAGG
MPZ RV	5'-CCTTGGCCTTCTCACTGAC	YWHAZ RV	5'-GAAGTTAAGGCCAGACCCAGT

**Table S4.** Differentiation studies in HOG and MO3.13 cells. ↓: a decreased expression relative to undifferentiated cells was reported. ↑: an increased expression relative to undifferentiated cells was reported. X: an absence of expression was reported. √: a presence of expression was reported.

	HOG		MO3.13	
	Undifferentiated	Differentiated	Undifferentiated	Differentiated
NG2				↓ protein [25,34]
PDGFR $\alpha$		↓ protein [25]		↓ protein [25,12]
Gangliosides (A2B5)	√ lipid [24]	√ lipid [24]	√ lipid [73]	
Olig2			√ protein [59]	↑ mRNA [43] ↑ protein [43] √ protein [59]
CNPase	√ mRNA [14] √ protein [14] X protein [24]	√ mRNA [14] ↑ protein [24]	√ mRNA [14] √ protein [14,73,60] X mRNA [35]	√ mRNA [14] ↑ mRNA [35] ↑ protein [34]
GalC (O1)	√ lipid [25,14] X lipid [24]	↑ lipid [24] √ lipid [25]	√ lipid [14,25,73]	√ lipid [25]
Sulfatide (O4)			√ lipid [73]	
CC1				√ ? [49]
CGT				↑ protein [12]
FynKinase		↑ mRNA [7]		↑ mRNA [7]
OSP			√ protein [60]	
GST-p				↑ protein [53,54]
MBP	√ protein [14] X protein [24]	↑ mRNA [14] ↑ protein [14] X protein [24]	√ mRNA [14,35] √ protein [14,73,37,59]	↑ mRNA [14] ↑ mRNA [43,35] ↑ protein [27,34,73,60,43] √ protein [37,40,38,59,47,50] √ ? [49]
PLP1	X protein [18,21]	↑ protein [21,18,20]	√ protein [60]	↑ protein [34,12]
OMG		↑ mRNA [14]		
MOBP		↑ mRNA [7,14]		↑ mRNA [7]
MOG		↑ mRNA [14]		↑ mRNA [14] √ protein [40, 50]
MAG		↑ mRNA [14]	√ protein [60]	↑ protein [34]
VIM			√ protein [73]	
S100B				↓ protein [34]
GFAP	X protein [14]		X protein [14,59] √ protein [37]	↓ protein [34,37] X protein [59]

<b>PDGFR<math>\beta</math></b>				$\checkmark$ protein [47]
<b>FGFR1</b>				$\checkmark$ protein [47]
<b>FGFR3</b>				$\checkmark$ protein [47]
<b>MAL2</b>	$\checkmark$ protein [21]	$\uparrow$ protein [21,20]		