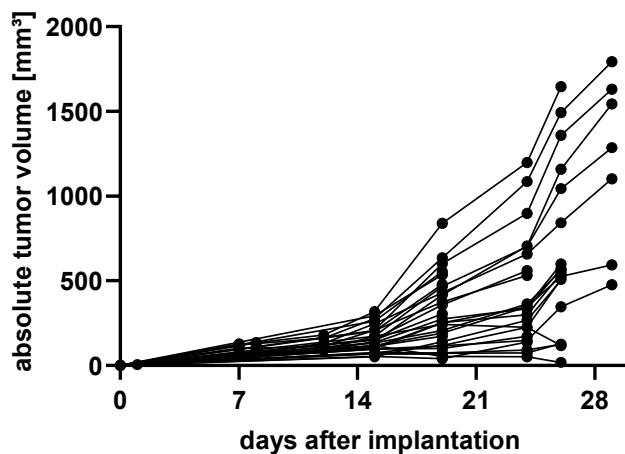


1 Article

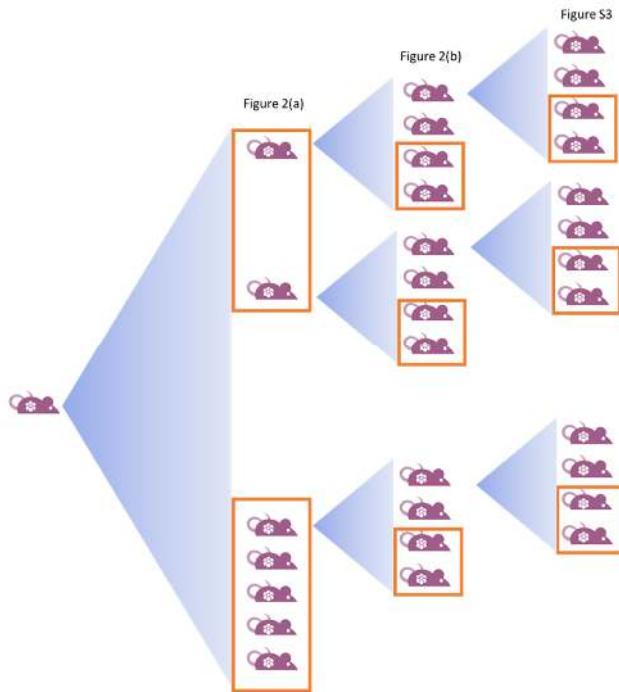
2 **Supplementary Materials:** The following are available online at www.mdpi.com/xxx/s1,

3



4

5 Supplemental Figure S1. Tumor growth behavior of subcutaneously implanted LXFA 677 *in vivo*. LXFA 677 was
6 implanted subcutaneously into NMRI nude mice. Tumor volume was measured twice weekly from the day of
7 implantation for four weeks.



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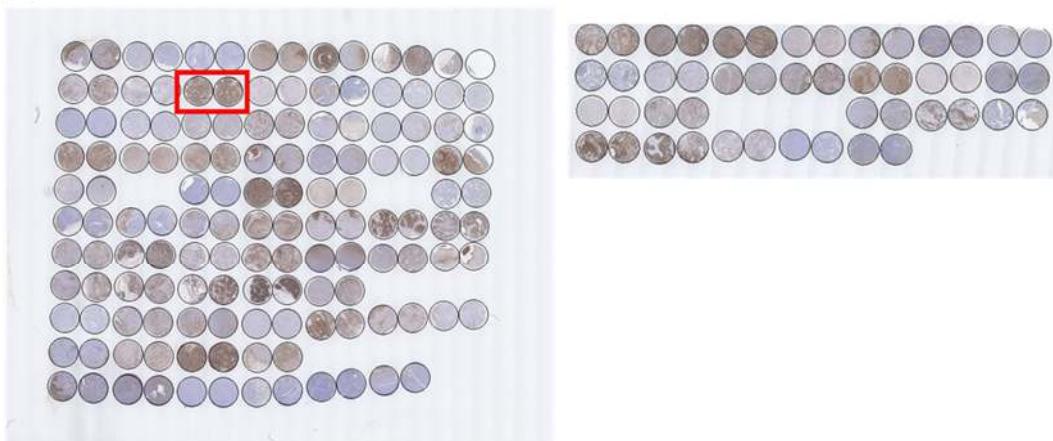
9

10 Supplemental Figure S2: Induction of resistance in NSCLC PDX LXFA 677 *in vivo*. Flowchart of the
11 transplantation and treatment process of LXFA 677 and the three resistant sublines. Untreated LXFA 677 PDX
12 material was transplanted subcutaneously into six NMRI nude mice and Gefitinib treatment was performed as

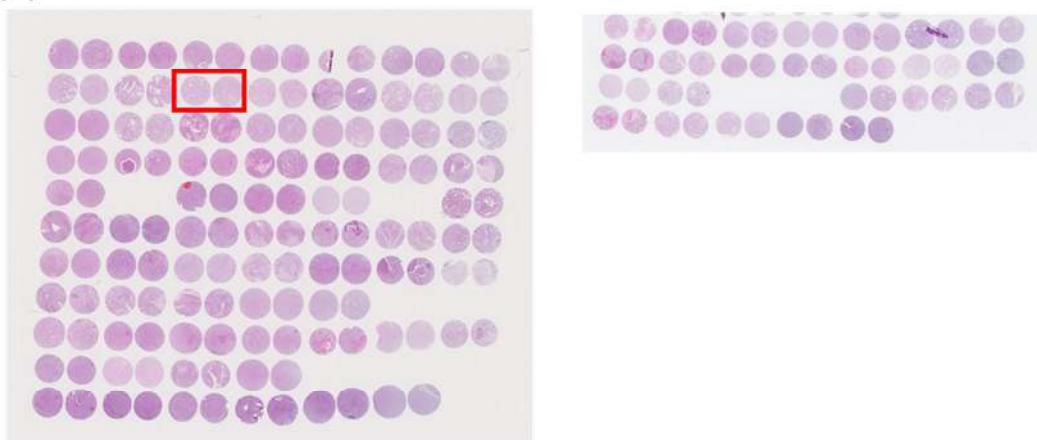
13 described in Figure 2 (a). Three tumor sublines, called LXFA 677res1, -res2 and -res3, showed progressive growth
 14 under constant treatment with Gefitinib and were re-transplanted into four NMRI nude mice, respectively. The
 15 four mice were stratified into a control group receiving the vehicle or the treatment group receiving Gefitinib as
 16 described in Figure 2 (b). One untreated control tumor of each subline served as donor material for a subsequent
 17 implantation into four NMRI nude mice. Again animals were stratified into control and treatment arm and
 18 Gefitinib was dosed as described in supplemental Figure S5. The orange box is indicating Gefitinib treatment

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(a)



(b)



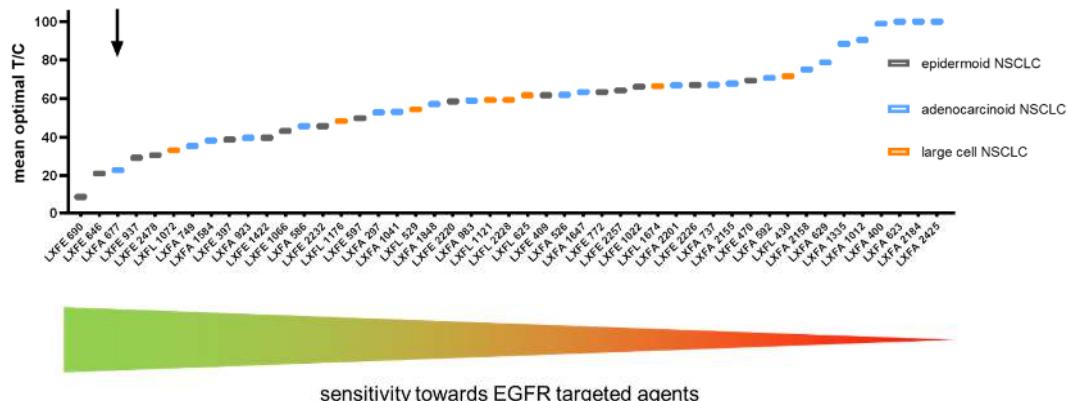
(c)

LXFA 289	LXFA 297	LXFA 400	LXFA 526	LXFA 586	LXFA 592	LXFA 623	LXA 3100	LXA 3104	LXA 3106	LXA SMTCA62	LXF 2436	LXF 2467	LXA 2157
LXFA 629	LXFA 644	LXFA 677	LXFA 737	LXFA 749	LXFA 923	LXFA 983	LXA 2160	LXA 2194	LXA 2366	LXA 2388	LXA 2403	LXA 2425	LXA 2426
LXFA 1012	LXFA 1041	LXFA 1335	LXFA 1584	LXFA 1647	LXFA 1848	LXFA 2155	LXA 2764	LXA 2889	LXA 2899	LXA 2900	LXF 386	LXE 2190	LXE 2198
LXFA 2158	LXFA 2165	LXFA 2184	LXFA 2201	LXFA 2204	LXFA 2207	LXFA 2217	LXE 2214	LXE 2220	LXE 2226	LXF 605	LXF 688		
LXFA 2250		LXF 1125	LXF 2478	LXF 2764		LXA 3109							
LXFE 211	LXFE 397	LXFE 409	LXFE 470	LXFE 597	LXFE 646	LXFE 690							
LXFE 772	LXFE 937	LXFE 1022	LXFE 1066	LXFE 1422	LXFE 2159	LXFE 2162							
LXFE 2226	LXFE 2232	LXFE 2257	LXFE 2348	LXFE 2415									
LXFL 430	LXFL 529	LXFL 625	LXFL 1072	LXFL 1121	LXFL 1176	LXFL 1674							
LXFL 2228	LXFL 2237	LXFE 2324	LXFL 2377										
LXFS 538	LXFS 573	LXFS 615	LXFS 650	LXFS 1129	LXFS 2156								

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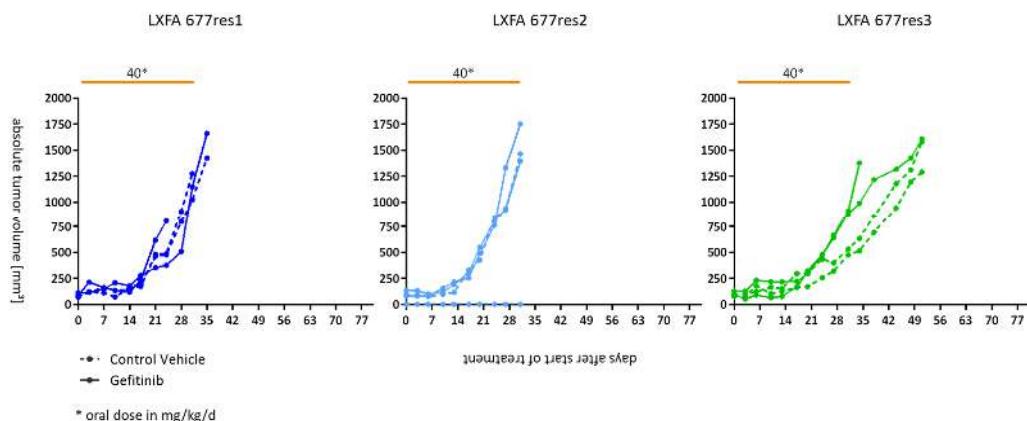
21 Figure S3: TMA of the NSCLC PDX panel comprising 85 NSCLC PDX models in duplicates. The red box is
 22 indicating tumor model LXFA 677. (a): IHC with anti-human EGFR Antibody followed by DAB staining and
 23 hematoxylin counterstaining (b): Hematoxylin& Eosin staining (c): Mapping of the individual tumor models on
 24 the TMA. The red box is indicating LXFA 677 model.

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 27 Supplemental Figure S4: Sensitivity towards EGFR targeting agents of 47 NSCLC PDX *in vivo*. Depicted are
 28 the mean optimal T/C values of one – five EGFR targeting agents per model. In total nine different compounds
 29 were tested: Osimertinib, Afatinib, Cetuximab, Erlotinib, Gefitinib, Necitumumab, Lapatinib, Sorafenib and
 30 Sunitinib. Dosing and schedule are depicted in Table 2. The respective tumor model was implanted
 31 subcutaneously in NMRI nude mice and treatment started when a group median tumor volume of 250 mm³ was
 32 reached. The group size was n = 5 -10 animals/group.

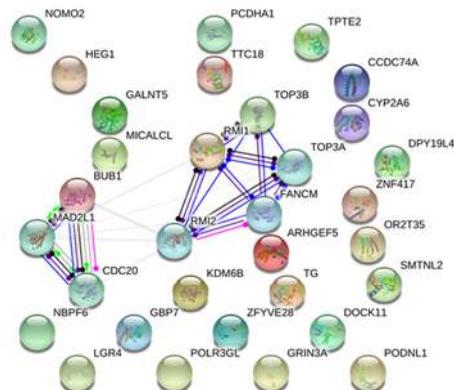
33



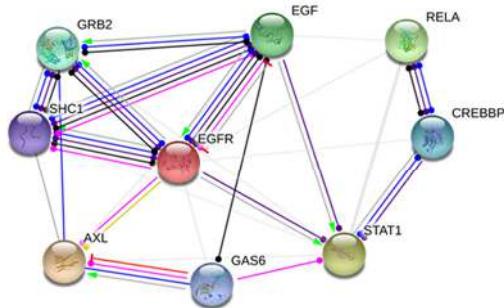
34
 35 Figure S5. The orange box is indicating the mice receiving Gefitinib treatment: Tumor growth curves for
 36 individuals tumors after re-implantation of tumors from the untreated control groups of the resistant lines LXFA
 37 677res1, -res2 and res3. The orange line is indicating the duration of the treatment. The respective dose per day
 38 is shown above the line.

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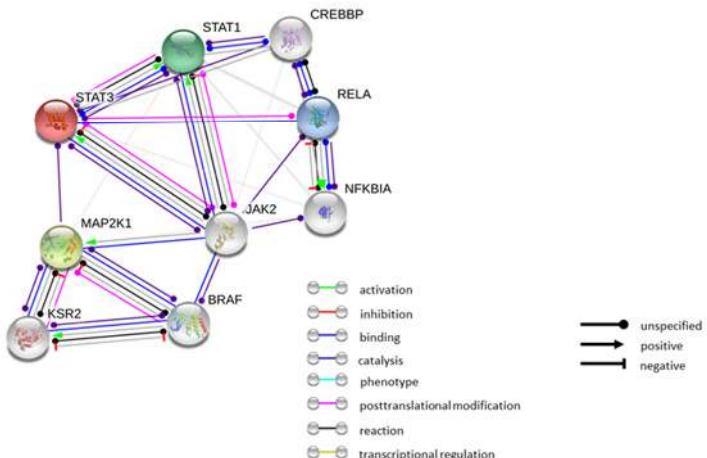
(a)



(b)



(c)



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41 Figure S6: Functional protein association networks of (a): Genes found to be mutated in all three Gefitinib-
 42 resistant LXFA 677 subclones (b): Functional protein association networks of proteins with an upregulated logFC
 43 > 0.5 in the resistant lines (c): Functional protein association networks of proteins with an upregulated logFC >
 44 0.5 in the phosphorylated vs total protein ratio. The STRING (<https://string-db.org/>) program was used to assign
 45 mutated genes and the upregulated proteins to a protein network. As a minimum required interaction score the
 46 medium confidence level was applied ($p=0.400$) allowing a maximal number of five interactors. Analysis of the
 47 involvement of the mutated genes/upregulated proteins in biological processes and functions is based on GO-
 48 term and KEGG pathways.

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51 Table S1. Number of common and unique mutations in the different sublines of NSCLC PDX LXFA 677

	LXFA 677	LXFA 677res1	LXFA 677res2	LXFA 677res3
common	504	504	504	504
in 2/3 sublines	26	30	42	38
unique	46	53	28	26
total	576	587	574	568

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Table S2. Annotation of mutations shared among all three resistant lines based on SIFT and PolyPhen. nd = dot determined

SYMBOL	Transcript	HGVSG Nomenclature	Consequence (ENSEMBL)	Amino_acid_change	SIFT prediction	PolyPhen predictiton	Mutation existent in LXFE_2478
GBP7	ENST00000294671	1:g.89133401G>C	missense_variant	Q507E	tolerated	benign	No
NBPF6	ENST00000495380	1:g.108459067G>A	missense_variant	S329N	tolerated	benign	No
OR2T35	ENST00000641268	1:g.248639168C>T	missense_variant	V31I	tolerated	benign	Yes
CCDC74A	ENST00000295171	2:g.131532868G>A	missense_variant	R294Q	tolerated	benign	No
HEG1	ENST00000311127	3:g.125013588_125013605dup	inframe_insertion	S668SSSSSS	nd	nd	Yes
ARHGEF5	ENST00000056217	7:g.144374813C>A	missense_variant	L1408I	deleterious	benign	No
MICALCL	ENST00000256186	11:g.12294836_12294841dup	inframe_insertion	A456APP	nd	nd	Yes
TPTE2	ENST00000400230	13:g.19492854T>C	missense_variant	K39E	tolerated - low confidence	benign	Yes
TPTE2	ENST00000400230	13:g.19492871A>C	missense_variant	L33R	tolerated - low confidence	benign	Yes
NOMO2	ENST00000621364	16:g.18531526C>T	missense_variant	V493M	deleterious	possibly damaging	No
SMTNL2	ENST00000389313	17:g.4592373G>A	missense_variant	D138N	deleterious	benign	No
KDM6B	ENST00000254846	17:g.7846893_7846898dup	inframe_insertion	L251LPP	nd	nd	Yes
ZNF417	ENST00000312026	19:g.57909712G>T	missense_variant	A189E	tolerated	benign	No
DOCK11	ENST00000276202	X:g.118598098A>C	missense_variant	E818D	nd	nd	Yes

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56 Table S3. Most significant biological processes and functions (based on GO, KEGG and reactome pathway) in which the mutated genes in at least 2/3 of the resistant lines are
 57 involved.

Pathway	#term ID	term description	observed gene count	background gene count	false discovery rate	matching proteins in our network
GO	GO:0003917	DNA topoisomerase type I activity	2	4	0.0069	TOP3A, TOP3B
KEGG	hsa03460	Fanconi anemia pathway	5	51	9.34e-07	FANCM, RMI1, RMI2, TOP3A, TOP3B
	hsa04110	Cell cycle	3	123	0.0148	BUB1, CDC20, MAD2L1
	hsa04114	Oocyte meiosis	3	116	0.0148	BUB1, CDC20, MAD2L1
	hsa03440	Homologous recombination	2	40	0.0165	TOP3A, TOP3B
Reactome	HSA-69620	Cell Cycle Checkpoints	6	265	0.00049	BUB1, CDC20, MAD2L1, RMI1, RMI2, TOP3A
	HSA-73894	DNA Repair	4	290	0.0065	FANCM, RMI1, RMI2, TOP3A

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Table S4. Most significant biological processes and functions in which the upregulated proteins are involved based on GO-term

#term ID	term description	observed gene count	background gene count	false discovery rate	matching proteins in our network (labels)
GO:0016032	viral process	8	571	6.59e-09	AXL, CREBBP, EGFR, GAS6, GRB2, RELA, SHC1, STAT1
GO:0038128	ERBB2 signaling pathway	4	31	3.60e-07	EGF, EGFR, GRB2, SHC1
GO:0007166	cell surface receptor signaling pathway	9	2198	7.80e-07	AXL, CREBBP, EGF, EGFR, GAS6, GRB2, RELA, SHC1, STAT1
GO:0051897	positive regulation of protein kinase B signaling	5	157	1.00e-06	AXL, EGF, EGFR, GAS6, GRB2
GO:0007173	epidermal growth factor receptor signaling pathway	4	52	1.44e-06	EGF, EGFR, GRB2, SHC1

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Table S5. Most significant biological processes and functions in which the upregulated proteins are involved based on KEGG pathway.

#term ID	term description	observed gene count	background gene count	false discovery rate	matching proteins in your network (labels)
hsa01521	EGFR tyrosine kinase inhibitor resistance	6	78	4.80e-11	AXL, EGF, EGFR, GAS6, GRB2, SHC1
hsa05215	Prostate cancer	5	97	2.39e-08	CREBBP, EGF, EGFR, GRB2, RELA
hsa05160	Hepatitis C	5	131	5.75e-08	EGF, EGFR, GRB2, RELA, STAT1
hsa05165	Human papillomavirus infection	6	317	5.75e-08	CREBBP, EGF, EGFR, GRB2, RELA, STAT1
hsa04630	Jak-STAT signaling pathway	5	160	1.09e-07	CREBBP, EGF, EGFR, GRB2, STAT1
hsa04014	Ras signaling pathway	5	228	3.87e-07	EGF, EGFR, GRB2, RELA, SHC1
hsa04917	Prolactin signaling pathway	4	69	3.87e-07	GRB2, RELA, SHC1, STAT1
hsa05200	Pathways in cancer	6	515	3.87e-07	CREBBP, EGF, EGFR, GRB2, RELA, STAT1
hsa05212	Pancreatic cancer	4	74	3.87e-07	EGF, EGFR, RELA, STAT1
hsa05214	Glioma	4	68	3.87e-07	EGF, EGFR, GRB2, SHC1
hsa04012	ErbB signaling pathway	4	83	4.55e-07	EGF, EGFR, GRB2, SHC1
hsa04066	HIF-1 signaling pathway	4	98	7.94e-07	CREBBP, EGF, EGFR, RELA

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Table S6. Most significant biological processes and functions in which the upregulated activated proteins are involved based on GO-term.

#term ID	term description	observed gene count	background gene count	false discovery rate	matching proteins in your network (labels)
GO:0035556	intracellular signal transduction	8	1528	3.96e-06	BRAF, JAK2, KSR2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
GO:0042981	regulation of apoptotic process	8	1501	3.96e-06	BRAF, CREBBP, JAK2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
GO:0070106	interleukin-27-mediated signaling pathway	3	11	4.03e-06	JAK2, STAT1, STAT3
GO:0070757	interleukin-35-mediated signaling pathway	3	11	4.03e-06	JAK2, STAT1, STAT3
GO:0009966	regulation of signal transduction	9	3033	4.73e-06	BRAF, CREBBP, JAK2, KSR2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
GO:0071354	cellular response to interleukin-6	4	30	1.10e-06	JAK2, RELA, STAT1, STAT3
GO:0035556	intracellular signal transduction	8	1528	3.96e-06	BRAF, JAK2, KSR2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
GO:0042981	regulation of apoptotic process	8	1501	3.96e-06	BRAF, CREBBP, JAK2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
GO:0070106	interleukin-27-mediated signaling pathway	3	11	4.03e-06	JAK2, STAT1, STAT3

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Table S7. Most significant biological processes and functions in which the upregulated activated proteins are involved based on KEGG pathway.

#term ID	term description	observed gene count	background gene count	false discovery rate	matching proteins in your network (labels)
hsa04062	Chemokine signaling pathway	7	181	2.84e-11	BRAF, JAK2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
hsa05167	Kaposi's sarcoma-associated herpesvirus infection	7	183	2.84e-11	CREBBP, JAK2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
hsa05200	Pathways in cancer	8	515	8.59e-11	BRAF, CREBBP, JAK2, MAP2K1, NFKBIA, RELA, STAT1, STAT3
hsa05161	Hepatitis B	6	142	4.14e-10	CREBBP, MAP2K1, NFKBIA, RELA, STAT1, STAT3
hsa05164	Influenza A	6	168	8.85e-10	CREBBP, JAK2, MAP2K1, NFKBIA, RELA, STAT1
hsa04917	Prolactin signaling pathway	5	69	1.66e-09	JAK2, MAP2K1, RELA, STAT1, STAT3
hsa05212	Pancreatic cancer	5	74	1.99e-09	BRAF, MAP2K1, RELA, STAT1, STAT3
hsa05215	Prostate cancer	5	97	6.41e-09	BRAF, CREBBP, MAP2K1, NFKBIA, RELA
hsa04659	Th17 cell differentiation	5	102	7.27e-09	JAK2, NFKBIA, RELA, STAT1, STAT3
hsa05145	Toxoplasmosis	5	109	9.03e-09	JAK2, NFKBIA, RELA, STAT1, STAT3
hsa05160	Hepatitis C	5	131	2.00e-08	BRAF, NFKBIA, RELA, STAT1, STAT3
hsa05162	Measles	5	133	2.00e-08	JAK2, NFKBIA, RELA, STAT1, STAT3
hsa05168	Herpes simplex infection	5	181	8.21e-08	CREBBP, JAK2, NFKBIA, RELA, STAT1
hsa04024	cAMP signaling pathway	5	195	1.10e-07	BRAF, CREBBP, MAP2K1, NFKBIA, RELA
hsa05221	Acute myeloid leukemia	4	66	1.48e-07	BRAF, MAP2K1, RELA, STAT3
hsa04920	Adipocytokine signaling pathway	4	69	1.64e-07	JAK2, NFKBIA, RELA, STAT3
hsa05140	Leishmaniasis	4	70	1.64e-07	JAK2, NFKBIA, RELA, STAT1
hsa05220	Chronic myeloid leukemia	4	76	2.12e-07	BRAF, MAP2K1, NFKBIA, RELA
hsa01521	EGFR tyrosine kinase inhibitor resistance	4	78	2.22e-07	BRAF, JAK2, MAP2K1, STAT3
hsa04658	Th1 and Th2 cell differentiation	4	88	3.36e-07	JAK2, NFKBIA, RELA, STAT1

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74 **Author Contributions:** Conceptualization, J.S., C.T., H.K. and W.S.; methodology, C.T., K.K., L.K. A.-L. P. and
75 E.O.; software, D.B. and H.K.; validation, J.S., E.O. L.K., H.K. and W.S.; formal analysis, J.S., L.K. and H.K.;
76 investigation, C.T., K.K., E.O. and A.-L.P.; data curation, C.T., K.K., A.-L.P. and L.K.; writing—original draft
77 preparation, J.S.; writing—review and editing, H.K., L.K. and W.S.; visualization, J.S. W.S. and H.K.; supervision,
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84 **References**

- 85 1. Wang, Z., ErbB Receptors and Cancer. *Methods in molecular biology* (Clifton, N.J.) **2017**, 1652, 3-35.
- 86 2. Antony, J.; Thiery, J. P.; Huang, R. Y., Epithelial-to-mesenchymal transition: Lessons from development,
87 insights into cancer and the potential of EMT-subtype based therapeutic intervention. *Physical biology* **2019**.
- 88 3. Siu, M. K.; Abou-Kheir, W.; Yin, J. J.; Chang, Y. S.; Barrett, B.; Suau, F.; Casey, O.; Chen, W. Y.; Fang, L.;
89 Hynes, P.; Hsieh, Y. Y.; Liu, Y. N.; Huang, J.; Kelly, K., Loss of EGFR signaling regulated miR-203 promotes
90 prostate cancer bone metastasis and tyrosine kinase inhibitors resistance. *Oncotarget* **2014**, 5 (11), 3770-84.
- 91 4. Tulchinsky, E.; Demidov, O.; Krajewska, M.; Barlev, N. A.; Imyanitov, E., EMT: A mechanism for escape
92 from EGFR-targeted therapy in lung cancer. *Biochimica et biophysica acta. Reviews on cancer* **2019**, 1871 (1), 29-39.
- 93 5. Xu, J.; Wang, J.; Zhang, S., Mechanisms of resistance to irreversible epidermal growth factor receptor
94 tyrosine kinase inhibitors and therapeutic strategies in non-small cell lung cancer. *Oncotarget* **2017**, 8 (52), 90557-
95 90578.
- 96 6. Zhu, X.; Bao, Y.; Guo, Y.; Yang, W., Proline-Rich Protein Tyrosine Kinase 2 in Inflammation and Cancer.
97 *Cancers* **2018**, 10 (5).
- 98 7. Takeda, M.; Nakagawa, K., First- and Second-Generation EGFR-TKIs Are All Replaced to Osimertinib in
99 Chemo-Naive EGFR Mutation-Positive Non-Small Cell Lung Cancer? *International journal of molecular sciences*
100 **2019**, 20 (1), 146.
- 101 8. Kobayashi, S.; Boggon, T. J.; Dayaram, T.; Jänne, P. A.; Kocher, O.; Meyerson, M.; Johnson, B. E.; Eck, M. J.;
102 Tenen, D. G.; Halmos, B., EGFR Mutation and Resistance of Non-Small-Cell Lung Cancer to Gefitinib. *New
103 England Journal of Medicine* **2005**, 352 (8), 786-792.
- 104 9. Ohashi, K.; Maruvka, Y. E.; Michor, F.; Pao, W., Epidermal Growth Factor Receptor Tyrosine Kinase
105 Inhibitor–Resistant Disease. *Journal of Clinical Oncology* **2013**, 31 (8), 1070-1080.
- 106 10. Sequist, L. V.; Waltman, B. A.; Dias-Santagata, D.; Digumarthy, S.; Turke, A. B.; Fidias, P.; Bergethon, K.;
107 Shaw, A. T.; Gettinger, S.; Cosper, A. K.; Akhavanfard, S.; Heist, R. S.; Temel, J.; Christensen, J. G.; Wain, J. C.;
108 Lynch, T. J.; Vernovsky, K.; Mark, E. J.; Lanuti, M.; Iafrate, A. J.; Mino-Kenudson, M.; Engelman, J. A., Genotypic
109 and Histological Evolution of Lung Cancers Acquiring Resistance to EGFR Inhibitors. *Science Translational
110 Medicine* **2011**, 3 (75), 75ra26-75ra26.
- 111 11. Herter-Sprie, G. S.; Kung, A. L.; Wong, K. K., New cast for a new era: preclinical cancer drug development
112 revisited. *The Journal of clinical investigation* **2013**, 123 (9), 3639-45.

- 113 12. Hidalgo, M.; Amant, F.; Biankin, A. V.; Budinska, E.; Byrne, A. T.; Caldas, C.; Clarke, R. B.; de Jong, S.;
114 Jonkers, J.; Maelandsmo, G. M.; Roman-Roman, S.; Seoane, J.; Trusolino, L.; Villanueva, A., Patient-derived
115 xenograft models: an emerging platform for translational cancer research. *Cancer discovery* **2014**, *4* (9), 998-1013.
- 116 13. Teicher, B. A., Tumor models for efficacy determination. *Molecular cancer therapeutics* **2006**, *5* (10), 2435-43.
- 117 14. Tentler, J. J.; Tan, A. C.; Weekes, C. D.; Jimeno, A.; Leong, S.; Pitts, T. M.; Arcaroli, J. J.; Messersmith, W. A.;
118 Eckhardt, S. G., Patient-derived tumour xenografts as models for oncology drug development. *Nature reviews.
119 Clinical oncology* **2012**, *9* (6), 338-50.
- 120 15. Conway, T.; Wazny, J.; Bromage, A.; Tymms, M.; Sooraj, D.; Williams, E. D.; Beresford-Smith, B., Xenome—
121 a tool for classifying reads from xenograft samples. *Bioinformatics* **2012**, *28* (12), i172-i178.
- 122 16. McLaren, W.; Gil, L.; Hunt, S. E.; Riat, H. S.; Ritchie, G. R. S.; Thormann, A.; Flicek, P.; Cunningham, F., The
123 Ensembl Variant Effect Predictor. *Genome Biology* **2016**, *17* (1), 122.
- 124 17. Sherry, S. T.; Ward, M. H.; Kholodov, M.; Baker, J.; Phan, L.; Smigelski, E. M.; Sirotnik, K., dbSNP: the
125 NCBI database of genetic variation. *Nucleic acids research* **2001**, *29* (1), 308-11.
- 126 18. Dayem Ullah, A. Z.; Oscanoa, J.; Wang, J.; Nagano, A.; Lemoine, N. R.; Chelala, C., SNPnexus: assessing
127 the functional relevance of genetic variation to facilitate the promise of precision medicine. *Nucleic acids research*
128 **2018**, *46* (W1), W109-W113.
- 129 19. Meseure, D.; Vacher, S.; Lallemand, F.; Alsibai, K. D.; Hatem, R.; Chemlali, W.; Nicolas, A.; De Koning, L.;
130 Pasman, E.; Callens, C.; Lidereau, R.; Morillon, A.; Bieche, I., Prognostic value of a newly identified MALAT1
131 alternatively spliced transcript in breast cancer. *British journal of cancer* **2016**, *114* (12), 1395-404.
- 132 20. Troncale, S.; Barbet, A.; Coulibaly, L.; Henry, E.; He, B.; Barillot, E.; Dubois, T.; Hupe, P.; de Koning, L.,
133 NormaCurve: a SuperCurve-based method that simultaneously quantifies and normalizes reverse phase protein
134 array data. *PloS one* **2012**, *7* (6), e38686.
- 135 21. Szklarczyk, D.; Gable, A. L.; Lyon, D.; Junge, A.; Wyder, S.; Huerta-Cepas, J.; Simonovic, M.; Doncheva, N.
136 T.; Morris, J. H.; Bork, P.; Jensen, L. J.; Mering, C. V., STRING v11: protein-protein association networks with
137 increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic acids
138 research* **2019**, *47* (D1), D607-d613.
- 139 22. Ritchie, M. E.; Phipson, B.; Wu, D.; Hu, Y.; Law, C. W.; Shi, W.; Smyth, G. K., limma powers differential
140 expression analyses for RNA-sequencing and microarray studies. *Nucleic acids research* **2015**, *43* (7), e47-e47.
- 141 23. Morgillo, F.; Della Corte, C. M.; Fasano, M.; Ciardiello, F., Mechanisms of resistance to EGFR-targeted
142 drugs: lung cancer. *ESMO open* **2016**, *1* (3), e000060.
- 143 24. Sebens, S.; Schafer, H., The tumor stroma as mediator of drug resistance—a potential target to improve
144 cancer therapy? *Current pharmaceutical biotechnology* **2012**, *13* (11), 2259-72.
- 145 25. Campos-Parra, A. D.; Zuloaga, C.; Manriquez, M. E.; Aviles, A.; Borbolla-Escoboza, J.; Cardona, A.;
146 Meneses, A.; Arrieta, O., KRAS mutation as the biomarker of response to chemotherapy and EGFR-TKIs in
147 patients with advanced non-small cell lung cancer: clues for its potential use in second-line therapy decision
148 making. *American journal of clinical oncology* **2015**, *38* (1), 33-40.
- 149 26. Zhang J.; Babic, A., Regulation of the MET oncogene: molecular mechanisms. *Carcinogenesis* **2016**, *37* (4),
150 345-355.
- 151 27. Yun, H. S.; Baek, J. H.; Yim, J. H.; Um, H. D.; Park, J. K.; Song, J. Y.; Park, I. C.; Kim, J. S.; Lee, S. J.; Lee, C.
152 W.; Hwang, S. G., Radiotherapy diagnostic biomarkers in radioresistant human H460 lung cancer stem-like cells.
153 *Cancer biology & therapy* **2016**, *17* (2), 208-18.
- 154 28. Haffner, C.; Frauli, M.; Topp, S.; Irmler, M.; Hofmann, K.; Regula, J. T.; Bally-Cuif, L.; Haass, C.; Nicolin
155 and its binding partner Nomo are novel Nodal signaling antagonists. *The EMBO journal* **2004**, *23* (15), 3041-50.

- 156 29. Kalyan, A.; Carneiro, B. A.; Chandra, S.; Kaplan, J.; Chae, Y. K.; Matsangou, M.; Hendrix, M. J. C.; Giles, F.,
157 Nodal Signaling as a Developmental Therapeutics Target in Oncology. *Molecular cancer therapeutics* **2017**, *16* (5),
158 787-792.
- 159 30. He, P.; Wu, W.; Wang, H.; Liao, K.; Zhang, W.; Xiong, G.; Wu, F.; Meng, G.; Yang, K., Co-expression of Rho
160 guanine nucleotide exchange factor 5 and Src associates with poor prognosis of patients with resected non-small
161 cell lung cancer. *Oncology reports* **2013**, *30* (6), 2864-70.
- 162 31. Komiya, Y.; Onodera, Y.; Kuroiwa, M.; Nomimura, S.; Kubo, Y.; Nam, J. M.; Kajiwara, K.; Nada, S.;
163 Oneyama, C.; Sabe, H.; Okada, M., The Rho guanine nucleotide exchange factor ARHGEF5 promotes tumor
164 malignancy via epithelial-mesenchymal transition. *Oncogenesis* **2016**, *5* (9), e258.
- 165 32. DiBardino, D. M.; Rawson, D. W.; Saqi, A.; Heymann, J. J.; Pagan, C. A.; Bulman, W. A., Next-generation
166 sequencing of non-small cell lung cancer using a customized, targeted sequencing panel: Emphasis on small
167 biopsy and cytology. *CytoJournal* **2017**, *14*, 7.
- 168 33. Gadea, G.; Blangy, A., Dock-family exchange factors in cell migration and disease. *European journal of cell
169 biology* **2014**, *93* (10-12), 466-77.
- 170 34. Gordon, E. A.; Whisenant, T. C.; Zeller, M.; Kaake, R. M.; Gordon, W. M.; Krotee, P.; Patel, V.; Huang, L.;
171 Baldi, P.; Bardwell, L., Combining docking site and phosphosite predictions to find new substrates: identification
172 of smoothelin-like-2 (SMTNL2) as a c-Jun N-terminal kinase (JNK) substrate. *Cellular signalling* **2013**, *25* (12),
173 2518-29.
- 174 35. Kim, B. H.; Shenoy, A. R.; Kumar, P.; Das, R.; Tiwari, S.; MacMicking, J. D., A family of IFN-gamma-
175 inducible 65-kD GTPases protects against bacterial infection. *Science (New York, N.Y.)* **2011**, *332* (6030), 717-21.
- 176 36. Miura, K., ERK2-binding domain is required for phosphorylation of EBITEIN1, a potential downstream
177 interactor of ERK2. *Biochemical and biophysical research communications* **2008**, *375* (3), 367-71.
- 178 37. Silva, M. P.; Barros-Silva, J. D.; Vieira, J.; Lisboa, S.; Torres, L.; Correia, C.; Vieira-Coimbra, M.; Martins, A.
179 T.; Jeronimo, C.; Henrique, R.; Paulo, P.; Teixeira, M. R., NCOA2 is a candidate target gene of 8q gain associated
180 with clinically aggressive prostate cancer. *Genes, chromosomes & cancer* **2016**, *55* (4), 365-74.
- 181 38. Xiao, Z. G.; Shen, J.; Zhang, L.; Li, L. F.; Li, M. X.; Hu, W.; Li, Z. J.; Cho, C. H., The Roles of Histone
182 Demethylase UTX and JMJD3 (KDM6B) in Cancers: Current Progress and Future Perspectives. *Current medicinal
183 chemistry* **2016**, *23* (32), 3687-3696.
- 184 39. Xu, G.; Zhang, M.; Zhu, H.; Xu, J., A 15-gene signature for prediction of colon cancer recurrence and
185 prognosis based on SVM. *Gene* **2017**, *604*, 33-40.
- 186 40. Ali, G.; Bruno, R.; Poma, A. M.; Affinito, O.; Monticelli, A.; Piaggi, P.; Ricciardi, S.; Lucchi, M.; Melfi, F.;
187 Chella, A.; Cocozza, S.; Fontanini, G., Whole transcriptome targeted gene quantification provides new insights
188 on pulmonary sarcomatoid carcinomas. *Scientific reports* **2019**, *9* (1), 3536.
- 189 41. Cathcart-Rake, E.; Lopez-Chavez, A., Young male with fanconi anemia and EGFR-mutant non-small-cell
190 lung cancer. *Journal of thoracic oncology : official publication of the International Association for the Study of Lung Cancer*
191 **2014**, *9* (11), e83-5.
- 192 42. Pfaffle, H. N.; Wang, M.; Gheorghiu, L.; Ferraiolo, N.; Greninger, P.; Borgmann, K.; Settleman, J.; Benes, C.
193 H.; Sequist, L. V.; Zou, L.; Willers, H., EGFR-activating mutations correlate with a Fanconi anemia-like cellular
194 phenotype that includes PARP inhibitor sensitivity. *Cancer research* **2013**, *73* (20), 6254-63.
- 195 43. Ye, H.; Zhang, X.; Chen, Y.; Liu, Q.; Wei, J., Ranking novel cancer driving synthetic lethal gene pairs using
196 TCGA data. *Oncotarget* **2016**, *7* (34), 55352-55367.

- 197 44. Garcia-Higuera, I.; Taniguchi, T.; Ganesan, S.; Meyn, M. S.; Timmers, C.; Hejna, J.; Grompe, M.; D'Andrea, A. D., Interaction of the Fanconi anemia proteins and BRCA1 in a common pathway. *Molecular cell* **2001**, *7* (2), 249-62.
- 198 45. Li, L.; Gu, X.; Yue, J.; Zhao, Q.; Lv, D.; Chen, H.; Xu, L., Acquisition of EGFR TKI resistance and EMT phenotype is linked with activation of IGF1R/NF-kappaB pathway in EGFR-mutant NSCLC. *Oncotarget* **2017**, *8* (54), 92240-92253.
- 199 46. Matusan-Ilijas, K.; Damante, G.; Fabbro, D.; Dordevic, G.; Hadzisejdic, I.; Grahovac, M.; Avirovic, M.; Grahovac, B.; Jonjic, N.; Lucin, K., EGFR expression is linked to osteopontin and Nf-kappaB signaling in clear cell renal cell carcinoma. *Clinical & translational oncology : official publication of the Federation of Spanish Oncology Societies and of the National Cancer Institute of Mexico* **2013**, *15* (1), 65-71.
- 200 47. Meylan, E.; Dooley, A. L.; Feldser, D. M.; Shen, L.; Turk, E.; Ouyang, C.; Jacks, T., Requirement for NF-kappaB signalling in a mouse model of lung adenocarcinoma. *Nature* **2009**, *462* (7269), 104-7.
- 201 48. Baud, V.; Collares, D., Post-Translational Modifications of RelB NF-kappaB Subunit and Associated Functions. *Cells* **2016**, *5* (2).
- 202 49. Lin, C.; Lu, W.; Ren, Z.; Tang, Y.; Zhang, C.; Yang, R.; Chen, Y.; Cao, W.; Wang, L.; Wang, X.; Ji, T., Elevated RET expression enhances EGFR activation and mediates EGFR inhibitor resistance in head and neck squamous cell carcinoma. *Cancer letters* **2016**, *377* (1), 1-10.
- 203 50. Wang, S. E.; Narasanna, A.; Perez-Torres, M.; Xiang, B.; Wu, F. Y.; Yang, S.; Carpenter, G.; Gazdar, A. F.; Muthuswamy, S. K.; Arteaga, C. L., HER2 kinase domain mutation results in constitutive phosphorylation and activation of HER2 and EGFR and resistance to EGFR tyrosine kinase inhibitors. *Cancer cell* **2006**, *10* (1), 25-38.
- 204 51. Guo, G.; Gong, K.; Wohlfeld, B.; Hatanpaa, K. J.; Zhao, D.; Habib, A. A., Ligand-Independent EGFR Signaling. *Cancer research* **2015**, *75* (17), 3436-41.
- 205 52. Han, W.; Carpenter, R. L.; Cao, X.; Lo, H. W., STAT1 gene expression is enhanced by nuclear EGFR and HER2 via cooperation with STAT3. *Molecular carcinogenesis* **2013**, *52* (12), 959-69.
- 206 53. Byers, L. A.; Diao, L.; Wang, J.; Saintigny, P.; Girard, L.; Peyton, M.; Shen, L.; Fan, Y.; Giri, U.; Tumula, P. K.; Nilsson, M. B.; Gudikote, J.; Tran, H.; Cardnell, R. J.; Bearss, D. J.; Warner, S. L.; Foulks, J. M.; Kanner, S. B.; Gandhi, V.; Krett, N.; Rosen, S. T.; Kim, E. S.; Herbst, R. S.; Blumenschein, G. R.; Lee, J. J.; Lippman, S. M.; Ang, K. K.; Mills, G. B.; Hong, W. K.; Weinstein, J. N.; Wistuba, II; Coombes, K. R.; Minna, J. D.; Heymach, J. V., An epithelial-mesenchymal transition gene signature predicts resistance to EGFR and PI3K inhibitors and identifies Axl as a therapeutic target for overcoming EGFR inhibitor resistance. *Clinical cancer research : an official journal of the American Association for Cancer Research* **2013**, *19* (1), 279-90.
- 207 54. Elkabets, M.; Pazarentzos, E.; Juric, D.; Sheng, Q.; Pelossof, R. A.; Brook, S.; Benzaken, A. O.; Rodon, J.; Morse, N.; Yan, J. J.; Liu, M.; Das, R.; Chen, Y.; Tam, A.; Wang, H.; Liang, J.; Gurski, J. M.; Kerr, D. A.; Rosell, R.; Teixido, C.; Huang, A.; Ghossein, R. A.; Rosen, N.; Bivona, T. G.; Scaltriti, M.; Baselga, J., AXL mediates resistance to PI3Kalpha inhibition by activating the EGFR/PKC/mTOR axis in head and neck and esophageal squamous cell carcinomas. *Cancer cell* **2015**, *27* (4), 533-46.
- 208 55. Ghiso, E.; Migliore, C.; Ciciriello, V.; Morando, E.; Petrelli, A.; Corso, S.; De Luca, E.; Gatti, G.; Volante, M.; Giordano, S., YAP-Dependent AXL Overexpression Mediates Resistance to EGFR Inhibitors in NSCLC. *Neoplasia (New York, N.Y.)* **2017**, *19* (12), 1012-1021.
- 209 56. Guo, G.; Gong, K.; Ali, S.; Ali, N.; Shallwani, S.; Hatanpaa, K. J.; Pan, E.; Mickey, B.; Burma, S.; Wang, D. H.; Kesari, S.; Sarkaria, J. N.; Zhao, D.; Habib, A. A., A TNF-JNK-Axl-ERK signaling axis mediates primary resistance to EGFR inhibition in glioblastoma. *Nature neuroscience* **2017**, *20* (8), 1074-1084.

- 239 57. Tian, Y.; Zhang, Z.; Miao, L.; Yang, Z.; Yang, J.; Wang, Y.; Qian, D.; Cai, H.; Wang, Y., Anexelekto (AXL)
240 Increases Resistance to EGFR-TKI and Activation of AKT and ERK1/2 in Non-Small Cell Lung Cancer Cells.
241 *Oncology research* **2016**, *24* (5), 295-303.
- 242 58. Qu, Y.; Wu, X.; Yin, Y.; Yang, Y.; Ma, D.; Li, H., Antitumor activity of selective MEK1/2 inhibitor AZD6244
243 in combination with PI3K/mTOR inhibitor BEZ235 in gefitinib-resistant NSCLC xenograft models. *Journal of*
244 *experimental & clinical cancer research : CR* **2014**, *33*, 52.
- 245 59. Buonato, J. M.; Lazzara, M. J., ERK1/2 blockade prevents epithelial-mesenchymal transition in lung cancer
246 cells and promotes their sensitivity to EGFR inhibition. *Cancer research* **2014**, *74* (1), 309-19.
- 247 60. Schmitt, N. C.; Trivedi, S.; Ferris, R. L., STAT1 Activation Is Enhanced by Cisplatin and Variably Affected
248 by EGFR Inhibition in HNSCC Cells. *Molecular cancer therapeutics* **2015**, *14* (9), 2103-11.
- 249 61. Cheng, C. C.; Lin, H. C.; Tsai, K. J.; Chiang, Y. W.; Lim, K. H.; Chen, C. G.; Su, Y. W.; Peng, C. L.; Ho, A. S.;
250 Huang, L.; Chang, Y. C.; Lin, H. C.; Chang, J.; Chang, Y. F., Epidermal growth factor induces STAT1 expression
251 to exacerbate the IFNr-mediated PD-L1 axis in epidermal growth factor receptor-positive cancers. *Molecular*
252 *carcinogenesis* **2018**, *57* (11), 1588-1598.
- 253 62. Phuchareon, J.; McCormick, F.; Eisele, D. W.; Tetsu, O., EGFR inhibition evokes innate drug resistance in
254 lung cancer cells by preventing Akt activity and thus inactivating Ets-1 function. *Proceedings of the National*
255 *Academy of Sciences of the United States of America* **2015**, *112* (29), E3855-63.
- 256 63. Tetsu, O.; Eisele, D. W.; McCormick, F., Resistance to EGFR-targeted therapy by Ets-1 inactivation. *Cell cycle*
257 (*Gorgetown, Tex.*) **2015**, *14* (20), 3211-3212.
- 258



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