

Review

Crosstalk between NF- κ B and Nucleoli in the Regulation of Cellular Homeostasis

Jingyu Chen and Lesley A. Stark *

Edinburgh Cancer Research Centre, Institute of Genetics and Molecular Medicine (IGMM) University of Edinburgh; Edinburgh EH42XU, Scotland, UK; Jc2037@cam.ed.ac

* Correspondence: Lesley.Stark@IGMM.ed.ac.uk; Tel.: +44-131-651-8531

Received: 4 September 2018; Accepted: 3 October 2018; Published: 5 October 2018



Abstract: Nucleoli are emerging as key sensors of cellular stress and regulators of the downstream consequences on proliferation, metabolism, senescence, and apoptosis. NF- κ B signalling is activated in response to a similar plethora of stresses, which leads to modulation of cell growth and death programs. While nucleolar and NF- κ B pathways are distinct, it is increasingly apparent that they converge at multiple levels. Exposure of cells to certain insults causes a specific type of nucleolar stress that is characterised by degradation of the PolI complex component, TIF-IA, and increased nucleolar size. Recent studies have shown that this atypical nucleolar stress lies upstream of cytosolic I κ B degradation and NF- κ B nuclear translocation. Under these stress conditions, the RelA component of NF- κ B accumulates within functionally altered nucleoli to trigger a nucleophosmin dependent, apoptotic pathway. In this review, we will discuss these points of crosstalk and their relevance to anti-tumour mechanism of aspirin and small molecule CDK4 inhibitors. We will also briefly discuss how crosstalk between nucleoli and NF- κ B signalling may be more broadly relevant to the regulation of cellular homeostasis and how it may be exploited for therapeutic purpose.

Keywords: Nucleolus; TIF-IA(RRN3); I-kappaB; stress; aspirin; CDK4; RelA; p65; cancer; neurodegenerative disorders; NF- κ B; non-steroidal anti-inflammatory drugs (NSAIDs)

1. Introduction

NF- κ B is the collective name for a family of inducible transcription factors that play a pivotal role in many cellular processes including immune response, inflammation, proliferation, and apoptosis [1,2]. In addition to classical stimuli such as cytokines and pathogens, NF- κ B is induced by a plethora of environmental and cytotoxic insults [3]. The mechanism by which these multiple insults induce the pathway, and what determines the downstream consequences on proliferation and apoptosis, has remained unclear. However, recent studies suggest that nucleoli play a role. An atypical nucleolar stress response pathway has been identified that lies upstream of NF- κ B signaling [4]. It has also been shown that following induction, the RelA component of NF- κ B can accumulate in nucleoli to trigger apoptotic pathways [5,6]. This cross talk between NF- κ B and nucleoli is important for the anti-tumour effects of aspirin and small molecule CDK4 inhibitors, suggesting therapeutic relevance. In this review, we will discuss nucleolar stress and the various levels of convergence between this and the NF- κ B pathway. We will also discuss the relevance to the anti-tumor mechanisms of aspirin, CDK4 inhibitors, and other therapeutic agents. Finally, we will touch on other processes that may be regulated by this crosstalk.

2. The Nucleolus and Stress

The nucleolus is a highly dynamic, membrane-less nuclear organelle [7]. It is primarily recognized for its role in ribosome biogenesis which is the most energy consuming process in the cell and as

such, is tightly linked to metabolic and proliferative activity. The first stage is transcription of 47S pre-ribosomal RNA (rRNA) by RNA polymerase I (PolI) (Figure 1). This transcription requires the formation of a pre-initiation complex (PIC) which includes upstream binding factor (UBF) and SL-1: a multi-protein complex consisting of TBP and at least 4 PolI specific TAFs. SL-1 confers promoter specificity while UBF acts as a de-repressor and co-activator. Another critical component of the PIC is TIF-IA (gene name RRN3). This is essential for rDNA transcription as it forms a bridge between SL-1 and Pol I, thereby tethering PolI to the rDNA promoter to generate a functional transcription pre-initiation complex [8–11]. Maturation of 47S pre-rRNA occurs co-transcriptionally through a series of covalent modifications, nucleolytic processing steps and interactions with ribosomal, and accessory proteins. This generates pre-ribosomes, which are further matured to the 40S and 60S ribosomal subunits.

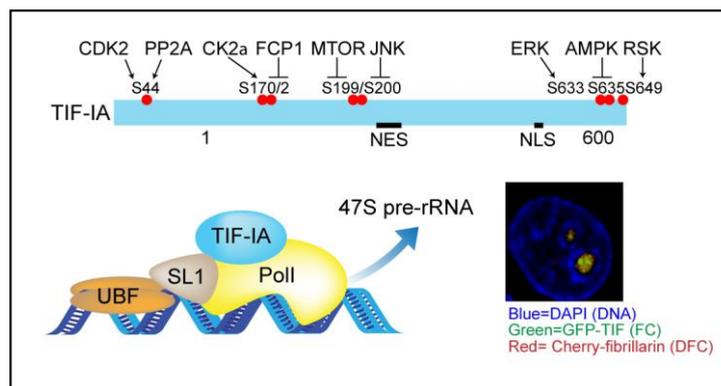


Figure 1. Top: TIF-IA is targeted by multiple kinase and phosphatase pathways. Bottom: the phosphorylation status controls the transcriptional activity of the pre-initiation complex, of which TIF-IA is a vital component. NES-nuclear export signal. NLS-nuclear localisation signal. Insert; confocal image showing the localisation of GFP-TIF-IA in fibrillar centres (FC) of nucleoli and cherry-fibrillarin in the surrounding dense fibrillar component (DFC) in fixed colorectal cancer cells. DAPI marks the DNA.

Nucleoli are divided into three sub-compartments namely; fibrillar centres (FC), dense fibrillar component (DFC), and granular component (GC). Transcription of rDNA takes place at the interface between FCs and DFCs while maturation takes place in the DFC and GC. Ribosomal DNA arrays are clustered in nucleolar organiser regions (NORs), which are present on the short arms of all five human acrocentric chromosomes [12]. NORs show various levels of activity and while nucleoli form around transcriptionally active NORs, inactive arrays are extra-nucleolar, embedded in and contributing to the heterochromatin that surrounds the organelle [13]. Generation and maintenance of the tri-partite nucleolar substructure is dependent on transcription of rDNA in active NORs and liquid-liquid phase separation (LLPS) of nucleolar components [13,14].

2.1. Nucleolar Maintenance of Cell Physiology

In addition to its role in ribosome biogenesis, the nucleolus acts as a critical stress sensor and coordinates downstream responses to stress such as altered metabolism, differentiation, cell cycle arrest, autophagy, DNA repair, senescence, and apoptosis [15,16] (Figure 2). Perturbations in nucleolar function are associated with many common diseases including ischemic heart disease, neurodegenerative disorders and cancer. Nucleolar size and number have also recently been associated with longevity in *c. elegans* and mammalian models [17,18]. Indeed, it now appears that proper dynamic control of nucleolar activity is crucial for maintaining tissue homeostasis and health.

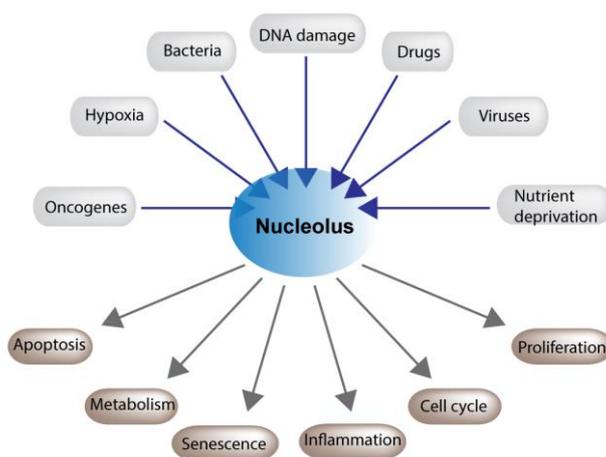


Figure 2. Nucleoli play a critical role in maintaining cellular homeostasis. Nucleolar function is altered in response to a plethora of cytotoxic and environmental stresses. This disruption modulates an array of cellular processes which allow cells to recover or, if the damage is too great, to undergo cell death.

The rate limiting step in ribosome biogenesis is transcription of the 47S pre-rRNA, which is extremely sensitive to changes in cellular environment. If cells are exposed to harmful conditions, i.e., nutrient starvation, cytotoxic agents, physical insults, or viral infections, rDNA transcription is inhibited and there is a rapid and dramatic change in nucleolar structure: the FC and DFC become segregated from the GC and form “caps” at the nucleolar periphery [19–21]. A cascade of signaling events are also initiated that influence cell phenotype [7,22]. This process is broadly termed nucleolar stress and can take different forms, dependent on cell context and the nature of the insult [23,24]. Over 4500 proteins have been isolated from mammalian nucleoli and over half of these are involved in processes out with ribosome biogenesis [25–27]. It is thought to be the dynamic flux of these proteins between nucleoli and other cellular compartments in response to stress that is ultimately responsible for the downstream effects on cell physiology [25,26].

2.2. p53 Dependent and Independent Consequences of Nucleolar Stress

The signals that link nucleolar stress to changes in cell physiology are only beginning to emerge. The most characterized pathway is the MDM2-p53 axis, which is covered in depth in some excellent reviews [28–31]. Briefly, upon stress-mediated perturbation of ribosome biogenesis, ribosomal proteins (RP) L5 and 11 are released from nucleoli in a NEDD8/PICT1 dependent manner. These proteins then accumulate in the nucleoplasm and bind to the p53 E3 ligase, MDM2. This inhibits MDM2 activity, thus preventing the ubiquitination and proteasomal degradation of p53. Consequently, p53 is stabilized and activates target genes involved in cell cycle arrest, senescence, and apoptosis. While this pathway is clearly important, recent reports from yeast, flies and mammalian cells indicate that in some contexts, perturbation of ribosome biogenesis can modulate cell growth, death, and autophagy in a p53 independent manner [23,24,32,33]. For example, Donati et al. demonstrated that specific interference with the PolII factor, *POLR1A*, induces cell cycle arrest in mammalian cells in the absence of p53 [34]. They proposed a model in which RPL11 binding to MDM2 blocks the MDM2-E2F interaction, thus causing E2F degradation and cell cycle arrest. Similarly, the Russo lab demonstrated that the apoptotic response to nucleolar stress can occur in the absence of functional p53 [35]. In this study, RPL3 was overexpressed to mimic perturbations of ribosome biogenesis. This caused the formation of an RpL3, Sp1, NPM complex at the p21 promoter and consequently, cell cycle arrest and apoptosis. In another example, it was shown that nucleolar stress destabilizes the proto-oncogene PIM1, causing increased levels of p27Kip1 and cell cycle arrest in p53^{-/-} cells [36]. Proteomic studies indicate that hundreds of proteins that shuttle from the nucleolus in response to cytotoxic stimuli have p53 independent functions, supporting the notion of important p53 independent nucleolar stress pathways [25,26].

3. TIF-IA-NF- κ B Nucleolar Stress

Like p53, NF- κ B plays a critical role in maintaining cellular homeostasis under stress and emerging evidence indicates this transcription factor pathway also lies downstream of perturbed nucleolar function [4].

3.1. Stress Activation of the NF- κ B Pathway

In mammalian cells there are five members in the NF- κ B family namely, RelA (p65), RelB, c-Rel, p105/p50 (NF- κ B1), and p100/p52 (NF- κ B2) [37,38]. These proteins homo and hetero-dimerize through their Rel homology domain to create a variety of transcription factor complexes (56). In resting cells, these complexes are retained in the cytoplasm by a family of I κ B inhibitory proteins (I κ B α , I κ B β , I κ B γ and Bcl-3). When the cell is exposed to a wide array of stimuli including inflammatory cytokines, bacterial pathogens, cytotoxic agents, nutrient deprivation, hypoxia, and physical insult, I κ B proteins are phosphorylated by inhibition of I κ B (IKK) kinase (IKK1/IKK α , IKK2/IKK β IKK γ /Nemo) complexes [37]. This phosphorylation targets I κ B for ubiquitination and degradation by the 26S proteasome. NF- κ B complexes are then free to translocate to the nucleus where they influence the expression of numerous (>150) genes including those involved in inflammation, immune response, senescence, cell cycle, and apoptosis [3].

Classic NF- κ B stimuli such as tumor necrosis factor alpha (TNF α) and interleukin-1 (IL-1) induce rapid IKK activation/I κ B degradation and the upstream pathway responsible for this rapid activation is very well documented [39,40]. In contrast, stress stimuli (including UV-C radiation, nutrient deprivation and chemopreventative/therapeutic agents) tend to activate the pathway with a much slower and delayed kinetic [3,41]. A number of mechanisms have been proposed for this delayed activation. For example, Kato et al. demonstrated that UV-C-mediated degradation of I κ B is dependent upon a p38-CK2 axis [42], while Jiang et al. demonstrated that phosphorylation of translation initiation factor 2 α (eIF2 α) is required for activation of the NF- κ B pathway by a variety of stresses [43,44]. More recently, it was shown that an atypical form of nucleolar stress, characterized by degradation of the PolII complex component, TIF-IA, lies upstream of NF- κ B signaling in response to specific stress stimuli [4].

3.2. TIF-IA Degradation—A Novel Form of Nucleolar Stress

TIF-IA is the key component of the PIC that transduces environmental signals to the PolII transcriptional machinery [22,45]. If nutrient availability is altered or the cell is under stress, the phosphorylation status of TIF-IA is modulated by a complex network of kinases and phosphatases, which ultimately activate or inactivate the protein to fine tune the transcriptional output (Figure 1) [46]. While TIF-IA is mainly known for its role in the nucleolus, the protein shuttles dynamically between this and other cellular compartments. Indeed, using a GFP-TIF-IA approach, Szymański et al. found that 48% of the protein is present in the cytoplasm while only 7% is located in the nucleolus (although the concentration in the nucleolus is higher) [47]. The mechanisms that control the cellular localization of TIF-IA are still unclear, but are known to be targeted by specific stresses [48]. It is also unclear if the protein plays a role in other compartments. What is clear is that it is an important regulator of cell proliferation and apoptosis. Genetic deletion in mice leads to embryonic lethality while deletion or depletion in mouse embryonic fibroblasts (MEFs), cancer, and neuronal cells causes cell cycle arrest and apoptosis [49,50].

Given the considerable overlap in stresses that target TIF-IA/perturb ribosome biogenesis and those that activate NF- κ B, our lab explored the connection. In doing so, we uncovered a novel pathway by which nucleolar function is altered by stress (Figure 3) [4]. We found that multiple stress stimuli, including aspirin, UV-C and the second messenger ceramide, not only alter the phosphorylation status of TIF-IA, but also induce degradation of the protein. This effect was not observed in response to TNF or the DNA damaging agent, camptothecin, indicating specificity. The mechanism by which TIF-IA is degraded in response to stress is complex and involves both proteasome and lysosomal pathways.

It is dependent on de-phosphorylation of TIF-IA at Serine 44 and the PolII complex associated factors upstream binding factor (UBF) and p14ARF. It also lies downstream of CDK4 inhibition, which is a common response to stress stimuli of the NF- κ B pathway.

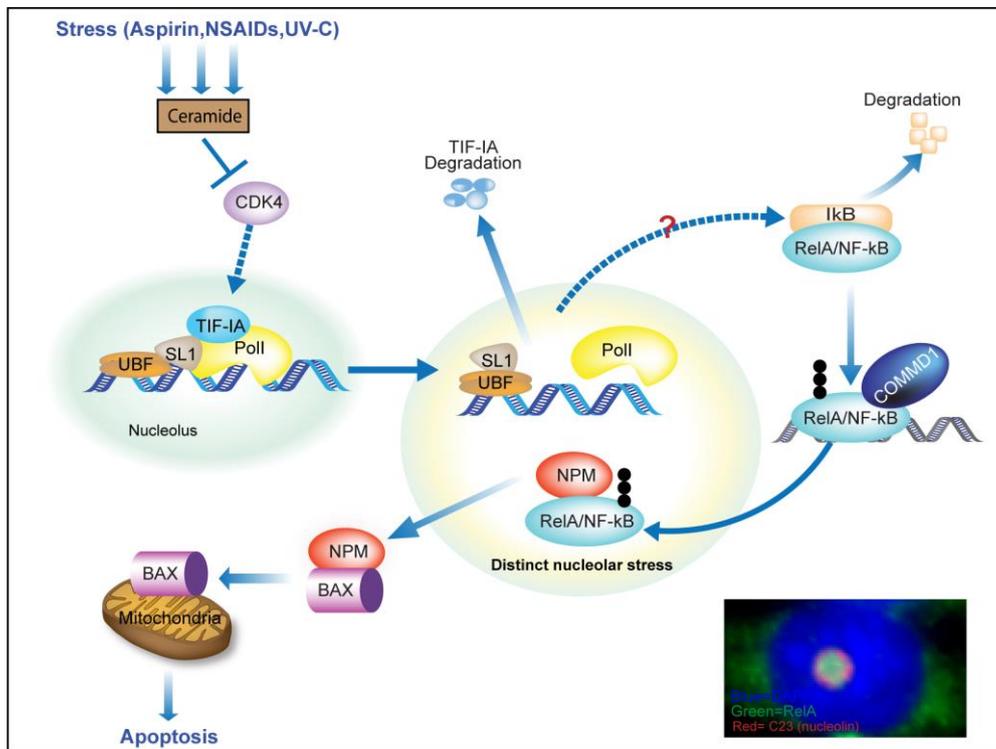


Figure 3. TIF-IA-NF- κ B-nucleolar stress response. When cells are exposed to a variety of specific stresses, ceramide is generated leading to inhibition of CDK4. This inhibition induces degradation of TIF-IA (in a manner dependent upon UBF and p14ARF), which in turn causes increased nucleolar size, gross changes in nucleolar morphology and degradation of I κ B. I κ B degradation allows RelA/NF- κ B to translocate into the nucleus and recruit a COMMD1 dependent ubiquitin ligase complex. Ubiquitination of RelA by this specific complex targets the protein to nucleoli, where it binds nucleophosmin (NPM), causing this protein to relocate out of nucleoli to the cytoplasm, where it is free to bind BAX and transport BAX to the mitochondria to mediate apoptosis. The signalling network(s) that links altered nucleolar function to I κ B degradation is unknown. Inset: Immunofluorescence showing enlarged segregated nucleoli and nucleolar accumulation of RelA in response to aspirin (5mM, 16h). DAPI depicts DNA.

As would be expected, stress-mediated degradation of TIF-IA was associated with inhibition of rDNA transcription. It was also associated with striking morphological changes to nucleolar structure and activation of the NF- κ B pathway.

3.3. Nucleolar Enlargement as a Consequence of TIF-IA Degradation

In most cases, stress-mediated segregation of the nucleolar sub-structure is associated with a significant reduction in size of the organelle. A role for TIF-IA in this phenomenon was initially suggested by genetic deletion of the gene in MEFs, which caused a loss of nucleolar morphology and a reduction in nucleolar size [49]. Stress-mediated inhibition of TIF-IA by targeted phosphorylation is also associated with decreased nucleolar size [22,48]. In contrast, we found that stress-mediated degradation of TIF-IA is paralleled by a striking increase in nucleolar size, alongside segregation of nucleolar components [4] (Figure 3). This stress-mediated increase in nucleolar size was paralleled by inhibition of rDNA transcription and was blocked when TIF-IA degradation was blocked, indicating that the two events are linked. These data question the paradigm that nucleolar size is linked to the

rate of rDNA transcription. Similar to these data, Fatyol et al. found that the MG132 proteasome inhibitor induces a significant increase in nucleolar volume while inhibiting rDNA transcription and inducing morphological changes to nucleoli [51]. Interestingly, we have found that low dose MG132 causes TIF-IA degradation in a similar manner to stress (unpublished data). The NEDD8 inhibitor MLN4924 has also been shown to cause an increase in nucleolar size alongside nucleolar stress [52].

3.4. Activation of the NF- κ B Pathway as a Consequence of TIF-IA Degradation

The first evidence that NF- κ B signaling may lie downstream of perturbation in nucleolar function came from experiments showing siRNA-mediated depletion of PolII complex components, (including TIF-IA) causes degradation of I κ B α , S536 phosphorylation of RelA (a marker of activation), nuclear translocation of RelA, increased NF- κ B transcriptional activity and increased transcription of NF- κ B target genes [4]. Interestingly, this effect was not mimicked by the PolII inhibitors actinomycinD, CX5461 or BMH-21, suggesting that unlike p53 nucleolar stress response, activation of NF- κ B signaling is not directly linked to inhibition of rDNA transcription. Kinetic studies revealed that stress-mediated degradation of TIF-IA precede cytoplasmic activation of NF- κ B, suggesting a potential link. Indeed, it was found that blocking degradation of TIF-IA, using specific siRNAs and a dominant negative TIF-IA mutant, blocked the effects of specific stresses on the NF- κ B pathway [4] (Figure 3). These data revealed a novel TIF-IA-NF- κ B nucleolar stress axis.

The TIF-IA-NF- κ B nucleolar stress response pathway was evident in multiple cell types and in tumors from colon cancer patients treated ex vivo with the chemopreventative agent aspirin (see below) indicating broad and *in vivo* relevance [4]. Multiple proteins that regulate the NF- κ B pathway reside within nucleoli, which could account for this connection. Interestingly, CK2, which has previously been shown to be involved in UV-C-mediated activation of the NF- κ B pathway [42], is bound to TIF-IA in the PolII complex [42,53]. Similarly, phosphorylation of eIF2 α in response to ER stress has been shown to both inhibit TIF-IA activity [54] and to activate NF- κ B [43,44]. NIK (NF- κ B inducing kinase), which acts upstream of the I κ B kinase (IKK) complex, shuttles through nucleoli [55]. The ribosomal proteins L3 and S3 have also been shown to complex with I κ B and modulate NF- κ B activity respectively [55–57]. L3 was found to bind to and stabilize I κ B, thus repressing NF- κ B activity, while S3 promoted activity by interacting with NF- κ B complexes in the nucleus.

3.5. TIF-IA-NF- κ B Nucleolar Stress and the Induction of Apoptosis

While stimulation of the NF- κ B pathway is generally regarded as anti-apoptotic, in particular contexts, and especially in response to cellular stress, NF- κ B acts to promote apoptosis [58,59]. Indeed, those stresses that stimulate the NF- κ B pathway through TIF-IA degradation (eg aspirin, UV-C, ceramide) are known to require nuclear translocation of NF- κ B for their pro-apoptotic activity [60–64]. In keeping with a pro-apoptotic role for the TIF-IA-NF κ B pathway, it was found that blocking TIF-IA degradation not only blocked nuclear translocation of NF- κ B/RelA in response to aspirin and CDK4 inhibition, but also blocked the apoptotic effects of the agents [4]. The mechanism by which stress-mediated nuclear translocation of NF- κ B promotes apoptosis has been the subject of debate. However, recent studies indicate nucleolar sequestration of NF- κ B proteins, particularly RelA, plays an important role [5].

4. Nucleolar Sequestration of RelA and Apoptosis

Cellular stress not only causes a dynamic flux of regulatory proteins out of nucleoli, but also sequestration of such proteins in the organelle [65–67]. This sequestration regulates gene expression, impacts nuclear structure, modulates specific apoptotic pathways, and influences autophagy [68]. Examples include nucleolar accumulation of p53, LC3II and ubiquitinated proteins in response to proteasome inhibition [65,66,69,70]. Nucleolar sequestration of NF- κ B repressing factor in response to heat stress, which causes repression of rDNA transcription [68], and nucleolar accumulation of von Hippel-Lindau protein, DNA methyltransferase 1 (DNMT1), and the DNA polymerase subunit

POLD1 (all with a specific nucleolar detention sequence) in response to heat shock, hypoxia, and acidosis [67,71]. Most recently, Gupta et al. demonstrated regulated nucleolar compartmentalization of the histone modifier, H2B [72]. Hence, sequestration of proteins within nucleoli is also emerging as an important mechanism for maintaining cellular homeostasis.

When exploring the mechanisms by which nuclear translocation of NF- κ B induces apoptosis, it was found that in response to specific pro-apoptotic stress stimuli (e.g., aspirin, serum deprivation, and UV-C radiation), the RelA component of NF- κ B translocates from the cytoplasm to the nucleoplasm and then to nucleoli, causing an accumulation of the protein in the organelle [5]. Nucleoplasmic to nucleolar translocation of RelA was found to be dependent upon an N-terminal nucleolar localization signal (NoLS). Using a dominant-negative mutant deleted for this motif, it was shown that nucleolar sequestration of RelA is causally involved in reduced basal NF- κ B transcriptional activity and the induction of apoptosis (Figure 3) [5]. Since this initial study, nucleolar sequestration of RelA has been observed in a number of other models. Loveridge et al. demonstrated that the NSAIDs sulindac, sulindac sulphone, and indomethacin induce nucleolar translocation of RelA in colon cancer cell lines, demonstrated that this was dependent on the N-terminal NoLS and showed that blocking nucleolar translocation of RelA blocked the apoptotic effects of these agents [63]. The anti-tumor agent, 2-methoxyestradiol (2ME2) (a naturally occurring derivative of estradiol), the potent Trk inhibitor and anti-tumor agent, K252a, [73] expression of the homeobox transcription factor, Hox-A5 [74], small molecule inhibitors of CDK4 [75,76] and the proteasome inhibition [77] have also been shown to induce nucleolar sequestration of RelA, which is associated with, or causally involved in, the induction of apoptosis. Nucleolar sequestration of p50 has also been reported. Dadsetan et al. demonstrated that the anti-TNF therapy, infliximab, induces “massive” nucleolar localisation of NF- κ B/p50 in the hippocampus of rats with a portacaval shunt (PCS). They also demonstrated that this nucleolar localization is associated with a decrease in transcription of NF- κ B target genes and a reduction in neuroinflammation [78]. Subsequent studies have demonstrated that nucleolar translocation of RelA, is dependent upon ubiquitination, facilitated by the multifunctional protein, COMMD1 (Figure 3) [77,79].

It was originally assumed that nucleolar translocation of RelA mediates apoptosis because the protein is sequestered away from the promoters of anti-apoptotic genes. However, it is now known that once in the nucleolus, RelA triggers a cascade of events that actively promotes apoptosis (Figure 3) [6]. That is, nucleolar RelA causes nucleophosmin (NPM)/B23 to relocate to the cytoplasm, bind BAX then transport BAX to the mitochondria to initiate apoptosis [6,80,81]. Others have also demonstrated that NPM relocation, and an NPM-BAX interaction, is critical for the pro-apoptotic effects of NF- κ B stimuli such as UV-C, [81]. Interestingly, stress stimuli such as aspirin and UV-C that utilize this pathway to induce cell death, also cause degradation of TIF-IA and atypical nucleolar stress, suggesting that these initial effects on nucleoli may prime cells for subsequent nucleolar accumulation of RelA and cytoplasmic translocation of NPM (Figure 3).

5. Therapeutic Relevance of Crosstalk between Nucleoli and the NF- κ B Pathway

High levels of nucleolar activity are a hallmark of cancer and contribute to tumor growth by allowing de-regulated protein synthesis and uncontrolled activity of nucleolar cell growth/death pathways [16,82]. Changes in nucleolar morphology and function are also common in age related neurodegenerative disorders and increasing evidence suggests that this dysfunction contributes to disease progression, as well as the normal aging process [17,18,46,83,84]. Similarly, dysregulated NF- κ B activity is common in cancer, neurodegenerative disorders and aging and contributes to the progression of these diseases/aging through promotion of a chronic inflammatory environment and modulation of genes that regulate cell growth/death [40,85,86]. Hence, both these pathways are attractive therapeutic targets.

One agent that has been found to target both these pathways simultaneously through nucleolar-NF- κ B signaling is aspirin [4,5]. Overwhelming evidence indicates that aspirin and related agents have considerable anti-tumor activity and the potential to prevent colorectal and other cancers.

Indeed, a recent meta-analysis suggested that continual aspirin use could reduce colorectal cancer risk by up to 40% [87–90]. Epidemiological and experimental evidence also suggests aspirin use protects against neurological disorders such as Alzheimer's disease [90,91]. However, the agent cannot be recommended for preventative purpose due to its side effect profile.

In experiments aimed at understanding the mechanism of action of aspirin against colorectal cancer, it was found that the agent causes degradation of TIF-IA and inhibition of rDNA transcription [4]. Furthermore, it was shown that this degradation is causally linked to stimulation of the NF- κ B pathway, nucleolar sequestration of RelA, repression of NF- κ B activity and the induction of apoptosis (Figure 3) [4,5,60]. A link between TIF-IA degradation and NF- κ B signaling was observed in multiple colon cancer cells lines, in cell lines derived from human pre-malignant intestinal lesions and in four out of seven human tumors treated ex vivo with low doses of the agent, suggesting pharmacological relevance [4]. In contrast to aspirin, the small molecule PolI inhibitor CX5461, which has shown considerable promise as an anti-cancer therapy and is currently in clinical trials for hematologic malignancies and triple negative breast cancer [82,92,93], had no effect on NF- κ B signaling. Similarly, the small molecule PolI inhibitor, BMH-21, that is also showing promise as an anti-cancer agent [94], did not stimulate the NF- κ B pathway. These data highlight the complexity of targeting nucleoli in cancer and the differential downstream consequences. They also reveal a novel and exciting mechanism of action of aspirin that warrants further investigation.

Increased CDK4 activity is a common occurrence in cancer and contributes to cancer progression by allowing unrestricted proliferation of tumor cells [76,95,96]. In keeping with this critical role, small molecule CDK4 inhibitors (CDK4i) have shown considerable promise as anti-cancer agents and are currently in phase I/II clinical trials in a variety of malignancies. However, their precise mechanism of action is unclear. Previous studies from this lab had demonstrated that small molecule CDK4 inhibitors stimulate the NF- κ B pathway and that this is essential for their pro-apoptotic activity against colorectal cancer cells [75]. More recently, Chen et al. demonstrated that CDK4 inhibition causes degradation of TIF-IA, which is causally linked to stimulation of the NF- κ B pathway and the induction of apoptosis (Figure 3) [4]. These data identify small molecule CDK4 inhibitors as another class of agents that simultaneously inhibit rDNA transcription and NF- κ B activity. Responses to aspirin are generally restricted to colon cancer cells [97]. In contrast, CDK4 inhibition caused TIF-IA dependent NF- κ B pathway stimulation in multiple cell types, suggesting crosstalk between these pathways may be broadly relevant for the maintenance of cellular homeostasis and the induction of apoptosis.

6. Summary

Both nucleolar and NF- κ B pathways play a vital role in maintaining cellular homeostasis under conditions of stress. Both pathways are also implicated in aging and are dysfunctional in age related diseases such as cancer and neurodegenerative disorders. Emerging evidence indicates that there are multiple levels of crosstalk between these two pathways that are important for maintaining cellular homeostasis and regulating apoptosis. However, this evolving field is in its infancy and there are still a number of important questions to be answered. For example, in what contexts is this novel nucleolar stress response pathway active and does it contribute to the aetiology of age related disease. With regard to this point, it is interesting to note that both nucleoli and NF- κ B are dysfunctional in senescence, a hallmark of aging. Further understanding of the mechanisms that regulate the stability of TIF-IA, and those that link altered TIF-IA levels to activation of the cytoplasmic NF- κ B pathway, would allow development of small molecules that act to specifically and simultaneously target dysfunctional rDNA transcription and NF- κ B activity. Similarly, identification of nucleolar pathways triggered by RelA would allow the development of RelA mimetics that mediate apoptosis by targeting dysfunctional nucleoli. Indeed, further understanding in this area could reveal a whole new class of targets to be exploited for therapeutic purposes. It could also reveal biomarkers of response to aspirin and CDK4 inhibitors, which have already been shown to utilize nucleolar-NF- κ B signaling to act against cancer cells.

Author Contributions: J.C. researched the topic and J.C. and L.A.S. prepared the manuscript.

Funding: Funding for the work was provided by WWCR (formally AICR) [10-0158 to L.S.]; Rosetrees Trust [A631, JS16/M225 to L.S.]; BBSRC [BB/H530362/1 to L.S.]; MRC [MR/J001481/1]; Bowel and Cancer Research [to L.S.]; University of Edinburgh scholarships (to J.C.).

Acknowledgments: We would like to thank H Thoms for critically reading the manuscript and publishing services at the IGMM for help with figure preparation.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Hayden, M.S.; Ghosh, S. NF- κ B, the first quarter-century: Remarkable progress and outstanding questions. *Genes Dev.* **2012**, *26*, 203–234. [[CrossRef](#)] [[PubMed](#)]
2. Karin, M. Nuclear factor- κ B in cancer development and progression. *Nature* **2006**, *441*, 431–436. [[CrossRef](#)] [[PubMed](#)]
3. Pahl, H.L. Activators and target genes of rel/NF- κ B transcription factors. *Oncogene* **1999**, *18*, 6853–6866. [[CrossRef](#)] [[PubMed](#)]
4. Chen, J.; Lobb, I.T.; Morin, P.; Novo, S.M.; Simpson, J.; Kennerknecht, K.; von Kriegsheim, A.; Batchelor, E.E.; Oakley, F.; Stark, L.A. Identification of a novel TIF-IA-nf- κ B nucleolar stress response pathway. *Nucleic Acids Res.* **2018**, *46*, 6188–6205. [[CrossRef](#)] [[PubMed](#)]
5. Stark, L.A.; Dunlop, M.G. Nucleolar sequestration of rela (p65) regulates nf- κ B-driven transcription and apoptosis. *Mol. Cell. Biol.* **2005**, *25*, 5985–6004. [[CrossRef](#)] [[PubMed](#)]
6. Khandelwal, N.; Simpson, J.; Taylor, G.; Rafique, S.; Whitehouse, A.; Hiscox, J.; Stark, L.A. Nucleolar nf- κ B/relA mediates apoptosis by causing cytoplasmic relocalization of nucleophosmin. *Cell Death Differ.* **2011**, *18*, 1889–1903. [[CrossRef](#)] [[PubMed](#)]
7. Boulon, S.; Westman, B.J.; Hutten, S.; Boisvert, F.M.; Lamond, A.I. The nucleolus under stress. *Mol. Cell* **2010**, *40*, 216–227. [[CrossRef](#)] [[PubMed](#)]
8. Schnapp, A.; Pfeleiderer, C.; Rosenbauer, H.; Grummt, I. A growth-dependent transcription initiation factor (TIF-IA) interacting with rna polymerase i regulates mouse ribosomal rna synthesis. *EMBO J.* **1990**, *9*, 2857–2863. [[CrossRef](#)] [[PubMed](#)]
9. Yuan, X.; Zhao, J.; Zentgraf, H.; Hoffmann-Rohrer, U.; Grummt, I. Multiple interactions between RNA polymerase I, TIF-IA and TAF(I) subunits regulate preinitiation complex assembly at the ribosomal gene promoter. *EMBO Rep.* **2002**, *3*, 1082–1087. [[CrossRef](#)] [[PubMed](#)]
10. Jin, R.; Zhou, W. TIF-IA: An oncogenic target of pre-ribosomal RNA synthesis. *Biochim. Biophys. Acta* **2016**, *1866*, 189–196. [[CrossRef](#)] [[PubMed](#)]
11. Grummt, I. Life on a planet of its own: Regulation of RNA polymerase I transcription in the nucleolus. *Genes Dev.* **2003**, *17*, 1691–1702. [[CrossRef](#)] [[PubMed](#)]
12. Wright, J.E.; Mais, C.; Prieto, J.L.; McStay, B. A role for upstream binding factor in organizing ribosomal gene chromatin. *Biochem. Soc. Symp.* **2006**, *77*–84. [[CrossRef](#)]
13. Mangan, H.; Gailin, M.O.; McStay, B. Integrating the genomic architecture of human nucleolar organizer regions with the biophysical properties of nucleoli. *FEBS J.* **2017**, *284*, 3977–3985. [[CrossRef](#)] [[PubMed](#)]
14. Wei, M.T.; Elbaum-Garfinkle, S.; Holehouse, A.S.; Chen, C.C.; Feric, M.; Arnold, C.B.; Priestley, R.D.; Pappu, R.V.; Brangwynne, C.P. Phase behaviour of disordered proteins underlying low density and high permeability of liquid organelles. *Nat. Chem.* **2017**, *9*, 1118–1125. [[CrossRef](#)] [[PubMed](#)]
15. Tsai, R.Y.; Pederson, T. Connecting the nucleolus to the cell cycle and human disease. *FASEB J.* **2014**, *28*, 3290–3296. [[CrossRef](#)] [[PubMed](#)]
16. Nunez Villacis, L.; Wong, M.S.; Ferguson, L.L.; Hein, N.; George, A.J.; Hannan, K.M. New roles for the nucleolus in health and disease. *Bioessays* **2018**, *40*, e1700233. [[CrossRef](#)] [[PubMed](#)]
17. Buchwalter, A.; Hetzer, M.W. Nucleolar expansion and elevated protein translation in premature aging. *Nat. Commun.* **2017**, *8*, 328. [[CrossRef](#)] [[PubMed](#)]
18. Tikku, V.; Jain, C.; Raz, Y.; Nakamura, S.; Heestand, B.; Liu, W.; Spath, M.; Suchiman, H.E.D.; Muller, R.U.; Slagboom, P.E.; et al. Small nucleoli are a cellular hallmark of longevity. *Nat. Commun.* **2016**, *8*, 16083. [[CrossRef](#)] [[PubMed](#)]

19. Nemeth, A.; Grummt, I. Dynamic regulation of nucleolar architecture. *Curr. Opin. Cell Biol.* **2018**, *52*, 105–111. [[CrossRef](#)] [[PubMed](#)]
20. Van Sluis, M.; McStay, B. Nucleolar reorganization in response to rDNA damage. *Curr. Opin. Cell Biol.* **2017**, *46*, 81–86. [[CrossRef](#)] [[PubMed](#)]
21. Jacob, M.D.; Audas, T.E.; Uniacke, J.; Trinkle-Mulcahy, L.; Lee, S. Environmental cues induce a long noncoding RNA-dependent remodeling of the nucleolus. *Mol. Biol. Cell* **2013**, *24*, 2943–2953. [[CrossRef](#)] [[PubMed](#)]
22. Mayer, C.; Grummt, I. Cellular stress and nucleolar function. *Cell Cycle* **2005**, *4*, 1036–1038. [[CrossRef](#)] [[PubMed](#)]
23. James, A.; Wang, Y.; Rajee, H.; Rosby, R.; DiMario, P. Nucleolar stress with and without p53. *Nucleus* **2014**, *5*, 402–426. [[CrossRef](#)] [[PubMed](#)]
24. Holmberg Olausson, K.; Nister, M.; Lindstrom, M.S. P53 -dependent and -independent nucleolar stress responses. *Cells* **2012**, *1*, 774–798. [[CrossRef](#)] [[PubMed](#)]
25. Moore, H.M.; Bai, B.; Boisvert, F.M.; Latonen, L.; Rantanen, V.; Simpson, J.C.; Pepperkok, R.; Lamond, A.I.; Laiho, M. Quantitative proteomics and dynamic imaging of the nucleolus reveal distinct responses to uv and ionizing radiation. *Mol. Cell. Proteom.* **2011**, *10*, M111. [[CrossRef](#)] [[PubMed](#)]
26. Boisvert, F.M.; Lam, Y.W.; Lamont, D.; Lamond, A.I. A quantitative proteomics analysis of subcellular proteome localization and changes induced by DNA damage. *Mol. Cell. Proteom.* **2010**, *9*, 457–470. [[CrossRef](#)] [[PubMed](#)]
27. Andersen, J.S.; Lam, Y.W.; Leung, A.K.; Ong, S.E.; Lyon, C.E.; Lamond, A.I.; Mann, M. Nucleolar proteome dynamics. *Nature* **2005**, *433*, 77–83. [[CrossRef](#)] [[PubMed](#)]
28. Woods, S.J.; Hannan, K.M.; Pearson, R.B.; Hannan, R.D. The nucleolus as a fundamental regulator of the p53 response and a new target for cancer therapy. *Biochim. Biophys. Acta* **2015**, *1849*, 821–829. [[CrossRef](#)] [[PubMed](#)]
29. Bursac, S.; Brdovcak, M.C.; Donati, G.; Volarevic, S. Activation of the tumor suppressor p53 upon impairment of ribosome biogenesis. *Biochim. Biophys. Acta* **2014**, *1842*, 817–830. [[CrossRef](#)] [[PubMed](#)]
30. Hein, N.; Hannan, K.M.; George, A.J.; Sanij, E.; Hannan, R.D. The nucleolus: An emerging target for cancer therapy. *Trends Mol. Med.* **2013**, *19*, 643–654. [[CrossRef](#)] [[PubMed](#)]
31. Rubbi, C.P.; Milner, J. Disruption of the nucleolus mediates stabilization of p53 in response to DNA damage and other stresses. *EMBO J.* **2003**, *22*, 6068–6077. [[CrossRef](#)] [[PubMed](#)]
32. Russo, A.; Russo, G. Ribosomal proteins control or bypass p53 during nucleolar stress. *Int. J. Mol. Sci.* **2017**, *18*. [[CrossRef](#)] [[PubMed](#)]
33. Al Baker, E.A.; Boyle, J.; Harry, R.; Kill, I.R. A p53-independent pathway regulates nucleolar segregation and antigen translocation in response to DNA damage induced by uv irradiation. *Exp. Cell Res.* **2004**, *292*, 179–186. [[CrossRef](#)] [[PubMed](#)]
34. Donati, G.; Brighenti, E.; Vici, M.; Mazzini, G.; Terre, D.; Montanaro, L.; Derenzini, M. Selective inhibition of rRNA transcription downregulates E2F-1: A new p53-independent mechanism linking cell growth to cell proliferation. *J. Cell Sci.* **2011**, *124*, 3017–3028. [[CrossRef](#)] [[PubMed](#)]
35. Russo, A.; Esposito, D.; Catillo, M.; Pietropaolo, C.; Crescenzi, E.; Russo, G. Human rpL3 induces G₁/S arrest or apoptosis by modulating p21^{waf1/cip1} levels in a p53-independent manner. *Cell Cycle* **2013**, *12*, 76–87. [[CrossRef](#)] [[PubMed](#)]
36. Iadevaia, V.; Caldarola, S.; Biondini, L.; Gismondi, A.; Karlsson, S.; Dianzani, I.; Loreni, F. Pim1 kinase is destabilized by ribosomal stress causing inhibition of cell cycle progression. *Oncogene* **2010**, *29*, 5490–5499. [[CrossRef](#)] [[PubMed](#)]
37. Gilmore, T.D. Introduction to NF-κB: Players, pathways, perspectives. *Oncogene* **2006**, *25*, 6680–6684. [[CrossRef](#)] [[PubMed](#)]
38. Baeuerle, P.A. Pro-inflammatory signaling: Last pieces in the NF-κB puzzle? *Curr. Biol.* **1998**, *8*, R19–R22. [[CrossRef](#)]
39. Ohtake, F.; Saeki, Y.; Ishido, S.; Kanno, J.; Tanaka, K. The k48-k63 branched ubiquitin chain regulates NF-κB signaling. *Mol. Cell* **2016**, *64*, 251–266. [[CrossRef](#)] [[PubMed](#)]
40. DiDonato, J.A.; Mercurio, F.; Karin, M. NF-κB and the link between inflammation and cancer. *Immunol. Rev.* **2012**, *246*, 379–400. [[CrossRef](#)] [[PubMed](#)]
41. Wu, Z.H.; Miyamoto, S. Many faces of NF-κB signaling induced by genotoxic stress. *J. Mol. Med.* **2007**, *85*, 1187–1202. [[CrossRef](#)] [[PubMed](#)]

42. Kato, T., Jr.; Delhase, M.; Hoffmann, A.; Karin, M. CK2 is a C-terminal I κ B kinase responsible for NF- κ B activation during the uv response. *Mol. Cell* **2003**, *12*, 829–839. [[CrossRef](#)]
43. Jiang, H.Y.; Wek, S.A.; McGrath, B.C.; Scheuner, D.; Kaufman, R.J.; Cavener, D.R.; Wek, R.C. Phosphorylation of the α subunit of eukaryotic initiation factor 2 is required for activation of NF- κ B in response to diverse cellular stresses. *Mol. Cell. Biol.* **2003**, *23*, 5651–5663. [[CrossRef](#)] [[PubMed](#)]
44. Jiang, H.Y.; Wek, R.C. GCN2 phosphorylation of eIF2 α activates NF- κ B in response to UV irradiation. *Biochem. J.* **2005**, *385*, 371–380. [[CrossRef](#)] [[PubMed](#)]
45. Mayer, C.; Zhao, J.; Yuan, X.; Grummt, I. Mtor-dependent activation of the transcription factor TIF-IA links rRNA synthesis to nutrient availability. *Genes Dev.* **2004**, *18*, 423–434. [[CrossRef](#)] [[PubMed](#)]
46. Parlato, R.; Bierhoff, H. Role of nucleolar dysfunction in neurodegenerative disorders: A game of genes. *AIMS Mol. Sci.* **2015**, *2*, 211–224. [[CrossRef](#)]
47. Szymanski, J.; Mayer, C.; Hoffmann-Rohrer, U.; Kalla, C.; Grummt, I.; Weiss, M. Dynamic subcellular partitioning of the nucleolar transcription factor TIF-IA under ribotoxic stress. *Biochim. Biophys. Acta* **2009**, *1793*, 1191–1198. [[CrossRef](#)] [[PubMed](#)]
48. Mayer, C.; Bierhoff, H.; Grummt, I. The nucleolus as a stress sensor: JNK2 inactivates the transcription factor TIF-IA and down-regulates rRNA synthesis. *Genes Dev.* **2005**, *19*, 933–941. [[CrossRef](#)] [[PubMed](#)]
49. Yuan, X.; Zhou, Y.; Casanova, E.; Chai, M.; Kiss, E.; Grone, H.J.; Schutz, G.; Grummt, I. Genetic inactivation of the transcription factor TIF-IA leads to nucleolar disruption, cell cycle arrest, and p53-mediated apoptosis. *Mol. Cell* **2005**, *19*, 77–87. [[CrossRef](#)] [[PubMed](#)]
50. Parlato, R.; Kreiner, G.; Erdmann, G.; Rieker, C.; Stotz, S.; Savenkova, E.; Berger, S.; Grummt, I.; Schutz, G. Activation of an endogenous suicide response after perturbation of rRNA synthesis leads to neurodegeneration in mice. *J. Neurosci.* **2008**, *28*, 12759–12764. [[CrossRef](#)] [[PubMed](#)]
51. Fatyol, K.; Grummt, I. Proteasomal atpases are associated with rDNA: The ubiquitin proteasome system plays a direct role in RNA polymerase I transcription. *Biochim. Biophys. Acta* **2008**, *1779*, 850–859. [[CrossRef](#)] [[PubMed](#)]
52. Bailly, A.; Perrin, A.; Bou Malhab, L.J.; Pion, E.; Larance, M.; Nagala, M.; Smith, P.; O'Donohue, M.F.; Gleizes, P.E.; Zomerdijk, J.; et al. The NEDD8 inhibitor MLN4924 increases the size of the nucleolus and activates p53 through the ribosomal-Mdm2 pathway. *Oncogene* **2016**, *35*, 415–426. [[CrossRef](#)] [[PubMed](#)]
53. Bierhoff, H.; Dunder, M.; Michels, A.A.; Grummt, I. Phosphorylation by casein kinase 2 facilitates rRNA gene transcription by promoting dissociation of TIF-IA from elongating rRNA polymerase I. *Mol. Cell. Biol.* **2008**, *28*, 4988–4998. [[CrossRef](#)] [[PubMed](#)]
54. DuRose, J.B.; Scheuner, D.; Kaufman, R.J.; Rothblum, L.I.; Niwa, M. Phosphorylation of eukaryotic translation initiation factor 2 α coordinates rRNA transcription and translation inhibition during endoplasmic reticulum stress. *Mol. Cell. Biol.* **2009**, *29*, 4295–4307. [[CrossRef](#)] [[PubMed](#)]
55. Birbach, A.; Bailey, S.T.; Ghosh, S.; Schmid, J.A. Cytosolic, nuclear and nucleolar localization signals determine subcellular distribution and activity of the NF- κ B inducing kinase I κ B kinase. *J. Cell Sci.* **2004**, *117*, 3615–3624. [[CrossRef](#)] [[PubMed](#)]
56. Wan, F.; Anderson, D.E.; Barnitz, R.A.; Snow, A.; Bidere, N.; Zheng, L.; Hegde, V.; Lam, L.T.; Staudt, L.M.; Levens, D.; et al. Ribosomal protein S3: A KH domain subunit in NF- κ B complexes that mediates selective gene regulation. *Cell* **2007**, *131*, 927–939. [[CrossRef](#)] [[PubMed](#)]
57. Russo, A.; Maiolino, S.; Pagliara, V.; Ungaro, F.; Tatangelo, F.; Leone, A.; Scalia, G.; Budillon, A.; Quaglia, F.; Russo, G. Enhancement of 5-FU sensitivity by the proapoptotic RPL3 gene in p53 null colon cancer cells through combined polymer nanoparticles. *Oncotarget* **2016**, *7*, 79670–79687. [[CrossRef](#)] [[PubMed](#)]
58. Perkins, N.D. The diverse and complex roles of NF- κ B subunits in cancer. *Nat. Rev. Cancer* **2012**, *12*, 121–132. [[CrossRef](#)] [[PubMed](#)]
59. Radhakrishnan, S.K.; Kamalakaran, S. Pro-apoptotic role of NF- κ B: Implications for cancer therapy. *Biochim. Biophys. Acta* **2006**, *1766*, 53–62. [[CrossRef](#)] [[PubMed](#)]
60. Stark, L.A.; Din, F.V.N.; Zwacka, R.M.; Dunlop, M.G. Aspirin-induced activation of the NF- κ B signalling pathway: A novel mechanism for aspirin-mediated apoptosis in colon cancer cells. *FASEB J.* **2001**, *15*, 1273–1275. [[CrossRef](#)] [[PubMed](#)]
61. Campbell, K.J.; Rocha, S.; Perkins, N.D. Active repression of antiapoptotic gene expression by RelA(p65) NF- κ B. *Mol. Cell* **2004**, *13*, 853–865. [[CrossRef](#)]

62. Fillet, M.; Bentires-Alj, M.; Deregowski, V.; Greimers, R.; Gielen, J.; Piette, J.; Bours, V.; Merville, M.P. Mechanisms involved in exogenous C2- and C6-ceramide-induced cancer cell toxicity. *Biochem. Pharmacol.* **2003**, *65*, 1633–1642. [[CrossRef](#)]
63. Loveridge, C.J.; Macdonald, A.D.; Thoms, H.C.; Dunlop, M.G.; Stark, L.A. The proapoptotic effects of sulindac, sulindac sulfone and indomethacin are mediated by nucleolar translocation of the rela(p65) subunit of NF- κ B. *Oncogene* **2008**, *27*, 2648–2655. [[CrossRef](#)] [[PubMed](#)]
64. Sansom, O.J.; Stark, L.A.; Dunlop, M.G.; Clarke, A.R. Suppression of intestinal and mammary neoplasia by lifetime administration of aspirin in Apc^{Min/+} and Apc^{min/+}, Msh2^{-/-} mice. *Cancer Res.* **2001**, *61*, 7060–7064. [[PubMed](#)]
65. Salmina, K.; Huna, A.; Inashkina, I.; Belyayev, A.; Krigerts, J.; Pastova, L.; Vazquez-Martin, A.; Erenpreisa, J. Nucleolar aggresomes mediate release of pericentric heterochromatin and nuclear destruction of genotoxicity treated cancer cells. *Nucleus* **2017**, *8*, 205–221. [[CrossRef](#)] [[PubMed](#)]
66. Latonen, L. Nucleolar aggresomes as counterparts of cytoplasmic aggresomes in proteotoxic stress. Proteasome inhibitors induce nuclear ribonucleoprotein inclusions that accumulate several key factors of neurodegenerative diseases and cancer. *Bioessays* **2011**, *33*, 386–395. [[CrossRef](#)] [[PubMed](#)]
67. Audas, T.E.; Jacob, M.D.; Lee, S. Immobilization of proteins in the nucleolus by ribosomal intergenic spacer noncoding RNA. *Mol. Cell* **2012**, *45*, 147–157. [[CrossRef](#)] [[PubMed](#)]
68. Coccia, M.; Rossi, A.; Riccio, A.; Trotta, E.; Santoro, M.G. Human NF- κ B repressing factor acts as a stress-regulated switch for ribosomal rna processing and nucleolar homeostasis surveillance. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 1045–1050. [[CrossRef](#)] [[PubMed](#)]
69. Rubbi, C.P.; Milner, J. Non-activated p53 co-localizes with sites of transcription within both the nucleoplasm and the nucleolus. *Oncogene* **2000**, *19*, 85–96. [[CrossRef](#)] [[PubMed](#)]
70. Latonen, L.; Moore, H.M.; Bai, B.; Jaamaa, S.; Laiho, M. Proteasome inhibitors induce nucleolar aggregation of proteasome target proteins and polyadenylated RNA by altering ubiquitin availability. *Oncogene* **2011**, *30*, 790–805. [[CrossRef](#)] [[PubMed](#)]
71. Audas, T.E.; Jacob, M.D.; Lee, S. The nucleolar detention pathway: A cellular strategy for regulating molecular networks. *Cell Cycle* **2012**, *11*, 2059–2062. [[CrossRef](#)] [[PubMed](#)]
72. *Cancer Registration Statistics Scotland 1986–1995*; ISD: Scotland, UK, 1998.
73. Parrondo, R.; de las Pozas, A.; Reiner, T.; Rai, P.; Perez-Stable, C. NF- κ B activation enhances cell death by antimetabolic drugs in human prostate cancer cells. *Mol. Cancer* **2010**, *9*, 182. [[CrossRef](#)] [[PubMed](#)]
74. Lee, D.H.; Forscher, C.; Di Vizio, D.; Koeffler, H.P. Induction of p53-independent apoptosis by ectopic expression of HOXA5 in human liposarcomas. *Sci. Rep.* **2015**, *5*, 12580. [[CrossRef](#)] [[PubMed](#)]
75. Thoms, H.C.; Dunlop, M.G.; Stark, L.A. P38-mediated inactivation of cyclin d1/cyclin-dependent kinase 4 stimulates nucleolar translocation of rela and apoptosis in colorectal cancer cells. *Cancer Res.* **2007**, *67*, 1660–1669. [[CrossRef](#)] [[PubMed](#)]
76. Thoms, H.C.; Dunlop, M.G.; Stark, L.A. Cdk4 inhibitors and apoptosis: A novel mechanism requiring nucleolar targeting of rela. *Cell Cycle* **2007**, *6*, 1293–1297. [[CrossRef](#)] [[PubMed](#)]
77. Thoms, H.C.; Loveridge, C.J.; Simpson, J.; Clipson, A.; Reinhardt, K.; Dunlop, M.G.; Stark, L.A. Nucleolar targeting of rela(p65) is regulated by COMMD1-dependent ubiquitination. *Cancer Res.* **2010**, *70*, 139–149. [[CrossRef](#)] [[PubMed](#)]
78. Dadsetan, S.; Balzano, T.; Forteza, J.; Agusti, A.; Cabrera-Pastor, A.; Taoro-Gonzalez, L.; Hernandez-Rabaza, V.; Gomez-Gimenez, B.; Elmlili, N.; Llansola, M.; et al. Infliximab reduces peripheral inflammation, neuroinflammation, and extracellular gaba in the cerebellum and improves learning and motor coordination in rats with hepatic encephalopathy. *J. Neuroinflamm.* **2016**, *13*, 245. [[CrossRef](#)] [[PubMed](#)]
79. O'Hara, A.; Simpson, J.; Morin, P.; Loveridge, C.J.; Williams, A.C.; Novo, S.M.; Stark, L.A. P300-mediated acetylation of COMMD1 regulates its stability, and the ubiquitylation and nucleolar translocation of the rela NF- κ B subunit. *J. Cell Sci.* **2014**, *127*, 3659–3665. [[CrossRef](#)] [[PubMed](#)]
80. Kerr, L.E.; Birse-Archbold, J.L.; Short, D.M.; McGregor, A.L.; Heron, I.; Macdonald, D.C.; Thompson, J.; Carlson, G.J.; Kelly, J.S.; McCulloch, J.; et al. Nucleophosmin is a novel bax chaperone that regulates apoptotic cell death. *Oncogene* **2007**, *26*, 2554–2562. [[CrossRef](#)] [[PubMed](#)]
81. Thompson, J.; Finlayson, K.; Salvo-Chirnside, E.; MacDonald, D.; McCulloch, J.; Kerr, L.; Sharkey, J. Characterisation of the bax-nucleophosmin interaction: The importance of the bax C-terminus. *Apoptosis* **2008**, *13*, 394–403. [[CrossRef](#)] [[PubMed](#)]

82. Quin, J.E.; Devlin, J.R.; Cameron, D.; Hannan, K.M.; Pearson, R.B.; Hannan, R.D. Targeting the nucleolus for cancer intervention. *Biochim. Biophys. Acta* **2014**, *1842*, 802–816. [[CrossRef](#)] [[PubMed](#)]
83. Kreiner, G.; Bierhoff, H.; Armentano, M.; Rodriguez-Parkitna, J.; Sowodniok, K.; Naranjo, J.R.; Bonfanti, L.; Liss, B.; Schutz, G.; Grummt, I.; et al. A neuroprotective phase precedes striatal degeneration upon nucleolar stress. *Cell Death Differ.* **2013**, *20*, 1455–1464. [[CrossRef](#)] [[PubMed](#)]
84. Evsyukov, V.; Domanskyi, A.; Bierhoff, H.; Gispert, S.; Mustafa, R.; Schlaudraff, F.; Liss, B.; Parlato, R. Genetic mutations linked to parkinson's disease differentially control nucleolar activity in pre-symptomatic mouse models. *Dis. Models Mech.* **2017**, *10*, 633–643. [[CrossRef](#)] [[PubMed](#)]
85. Osorio, F.G.; Soria-Valles, C.; Santiago-Fernandez, O.; Freije, J.M.; Lopez-Otin, C. NF- κ B signaling as a driver of ageing. *Int. Rev. Cell Mol. Biol.* **2016**, *326*, 133–174. [[PubMed](#)]
86. Shabab, T.; Khanabdali, R.; Moghadamtousi, S.Z.; Kadir, H.A.; Mohan, G. Neuroinflammation pathways: A general review. *Int. J. Neurosci.* **2017**, *127*, 624–633. [[CrossRef](#)] [[PubMed](#)]
87. Din, F.V.; Theodoratou, E.; Farrington, S.M.; Tenesa, A.; Barnetson, R.A.; Cetnarskyj, R.; Stark, L.; Porteous, M.E.; Campbell, H.; Dunlop, M.G. Effect of aspirin and nsaid on risk and survival from colorectal cancer. *Gut* **2010**, *59*, 1670–1679. [[CrossRef](#)] [[PubMed](#)]
88. Cuzick, J.; Thorat, M.A.; Bosetti, C.; Brown, P.H.; Burn, J.; Cook, N.R.; Ford, L.G.; Jacobs, E.J.; Jankowski, J.A.; La, V.C.; et al. Estimates of benefits and harms of prophylactic use of aspirin in the general population. *Ann. Oncol.* **2015**, *26*, 47–57. [[CrossRef](#)] [[PubMed](#)]
89. Burn, J.; Gerdes, A.M.; Macrae, F.; Mecklin, J.P.; Moeslein, G.; Olschwang, S.; Eccles, D.; Evans, D.G.; Maher, E.R.; Bertario, L.; et al. Long-term effect of aspirin on cancer risk in carriers of hereditary colorectal cancer: An analysis from the capp2 randomised controlled trial. *Lancet* **2011**, *378*, 2081–2087. [[CrossRef](#)]
90. Wang, J.; Tan, L.; Wang, H.F.; Tan, C.C.; Meng, X.F.; Wang, C.; Tang, S.W.; Yu, J.T. Anti-inflammatory drugs and risk of alzheimer's disease: An updated systematic review and meta-analysis. *J. Alzheimers Dis.* **2015**, *44*, 385–396. [[CrossRef](#)] [[PubMed](#)]
91. Patel, D.; Roy, A.; Kundu, M.; Jana, M.; Luan, C.H.; Gonzalez, F.J.; Pahan, K. Aspirin binds to pparalpha to stimulate hippocampal plasticity and protect memory. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E7408–E7417. [[CrossRef](#)] [[PubMed](#)]
92. Drygin, D.; Lin, A.; Bliesath, J.; Ho, C.B.; O'Brien, S.E.; Proffitt, C.; Omori, M.; Haddach, M.; Schwaebe, M.K.; Siddiqui-Jain, A.; et al. Targeting RNA polymerase i with an oral small molecule cx-5461 inhibits ribosomal RNA synthesis and solid tumor growth. *Cancer Res.* **2011**, *71*, 1418–1430. [[CrossRef](#)] [[PubMed](#)]
93. Yan, S.; Frank, D.; Son, J.; Hannan, K.M.; Hannan, R.D.; Chan, K.T.; Pearson, R.B.; Sanij, E. The potential of targeting ribosome biogenesis in high-grade serous ovarian cancer. *Int. J. Mol. Sci.* **2017**, *18*. [[CrossRef](#)] [[PubMed](#)]
94. Peltonen, K.; Colis, L.; Liu, H.; Trivedi, R.; Moubarek, M.S.; Moore, H.M.; Bai, B.; Rudek, M.A.; Bieberich, C.J.; Laiho, M. A targeting modality for destruction of rna polymerase i that possesses anticancer activity. *Cancer Cell* **2014**, *25*, 77–90. [[CrossRef](#)] [[PubMed](#)]
95. Sherr, C.J.; Beach, D.; Shapiro, G.I. Targeting cdk4 and cdk6: From discovery to therapy. *Cancer Discov.* **2015**, *6*, 353–367. [[CrossRef](#)] [[PubMed](#)]
96. Cole, A.M.; Myant, K.; Reed, K.R.; Ridgway, R.A.; Athineos, D.; van den Brink, G.R.; Muncan, V.; Clevers, H.; Clarke, A.R.; Sicinski, P.; et al. Cyclin d2-cyclin-dependent kinase 4/6 is required for efficient proliferation and tumorigenesis following apc loss. *Cancer Res.* **2010**, *70*, 8149–8158. [[CrossRef](#)] [[PubMed](#)]
97. Din, F.V.; Dunlop, M.G.; Stark, L.A. Evidence for colorectal cancer cell specificity of aspirin effects on nf kappa b signalling and apoptosis. *Br. J. Cancer* **2004**, *91*, 381–388. [[CrossRef](#)] [[PubMed](#)]

