

*Supplementary File*

# **Multi-step Enzymatic Synthesis of 1,9-Nonanedioic Acid from a Renewable Fatty Acid and Its Application for the Enzymatic Production of Biopolyesters**

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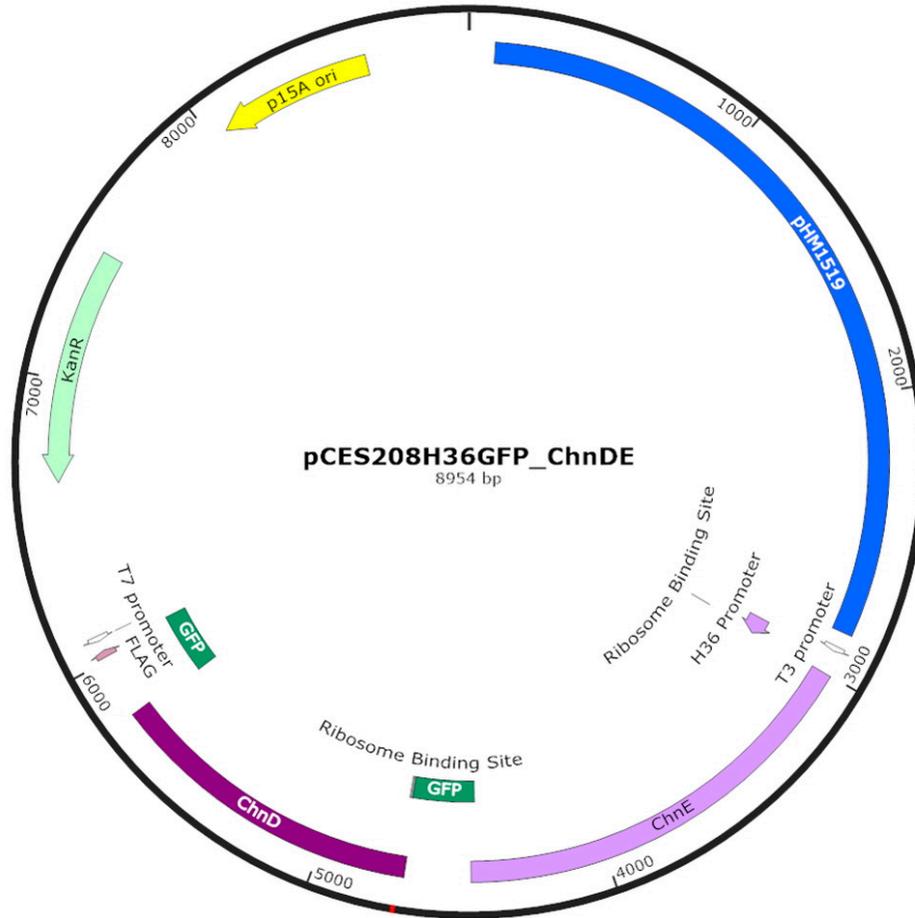
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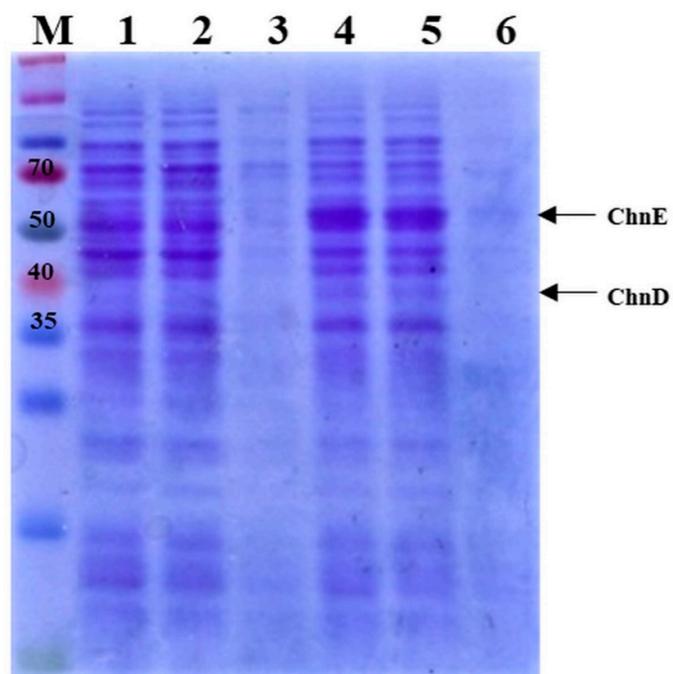
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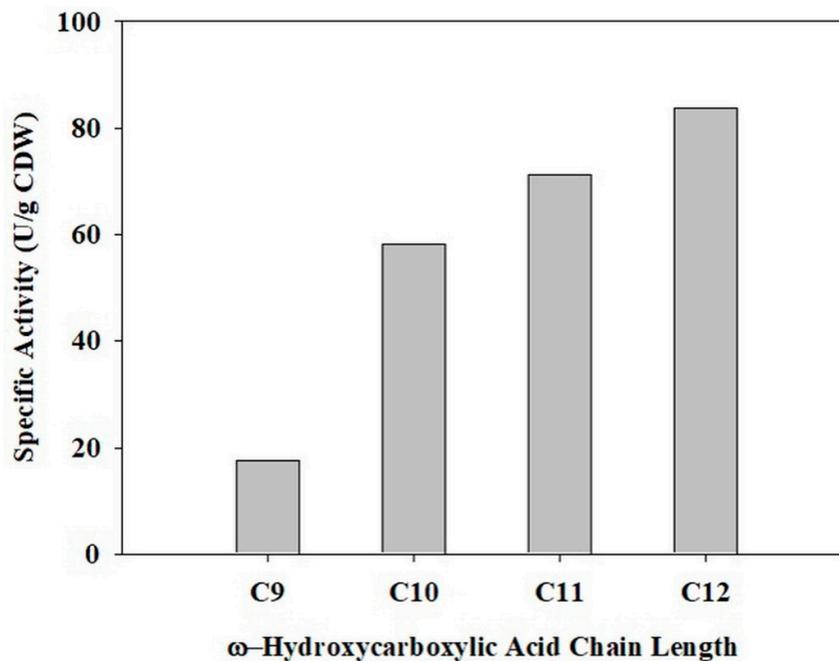
**Figure S1.** Map of pCES208H36GFP-ChnDE for the ChnDE expression in *Corynebacterium glutamicum* ATCC 13032. The alcohol/aldehyde dehydrogenase genes (chnDE) of *Acinetobacter* sp. NCIMB 9871 [1] were inserted into an *E. coli/C. glutamicum* shuttle vector, pCES208H36GFP [2, 3].



**Figure S2.** SDS-PAGE analysis of the protein extracts of *C. glutamicum* ATCC 13032 and the recombinant *C. glutamicum* ATCC 13032 pCES208H36GFP-ChnDE. The wild type (lanes 1,2,3) and recombinant cells (lanes 4,5,6) were harvested after 12 h of cultivation in CGXII medium and fractionated to total, soluble and insoluble fractions. Lane M: marker protein; lanes 1,4: total fraction; lanes 2,5: soluble fraction; lanes 3,6: insoluble fraction.

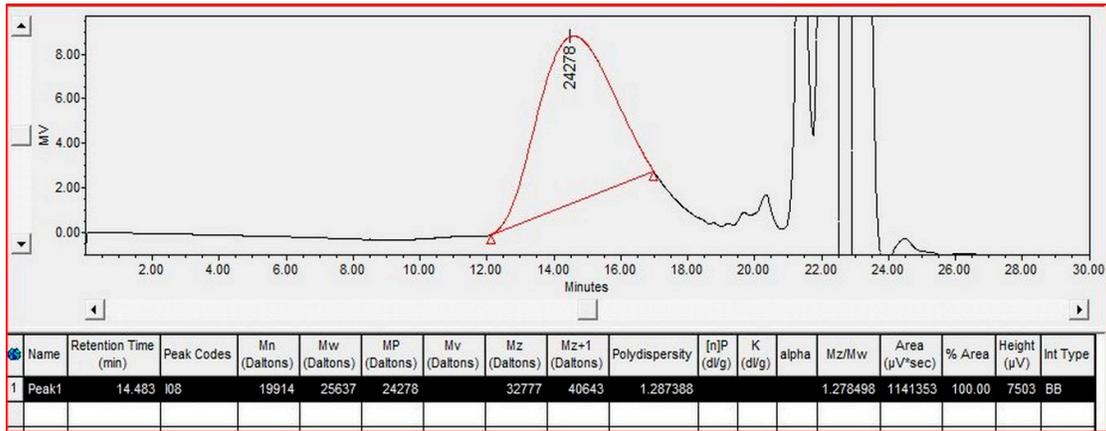


**Figure S3.** The specific oxidation rates of the recombinant *C. glutamicum* for the C9 to C12  $\omega$ -hydroxycarboxylic acids. The whole-cell bioconversion was initiated by adding 20 mM substrate (e.g., 9-hydroxynonanoic acid, 10-hydroxydecanoic acid, 11-hydroxyundecanoic acid, 12-hydroxydodecanoic acid), which were prepared in dimethyl sulfoxide (DMSO). The reaction was conducted in 50 mM Tris-HCl buffer (pH 8.0) containing 8 g dry cells/L and 0.5 g/L Tween 80 at 35°C, 200 rpm. The specific oxidation rates were determined based on the product concentrations at 10 or 30 min.



**Figure S4.** HPLC chromatogram of the biopolyester, which had been produced from azelaic acid and 1,8-octanediol by the immobilized lipase B from *Candida antarctica* (i.e., GF CalB-IM (GenoFocus (Korea))) (A). The biopolyester, which had been isolated from the reaction medium (B).

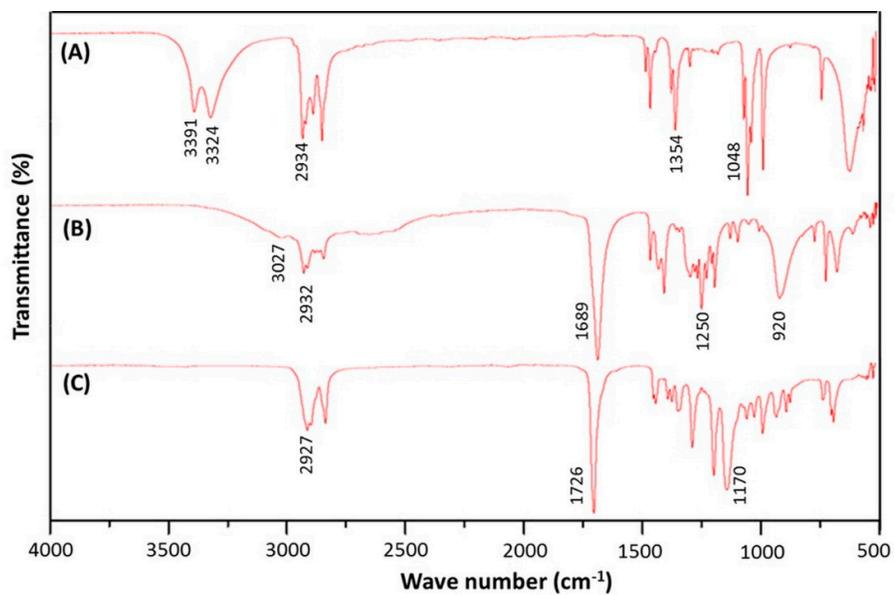
(A)



(B)



**Figure S5.** FT-IR spectra of the reaction components, (A) 1,8-octanediol and (B) azelaic acid and (C) polyester product formed in the poly-esterification in toluene at 75°C.



**Table S1.** Bacterial strains, plasmids, and oligonucleotides used in this study.

	Relevant characteristics	Reference or source
<b>Strains</b>		
<i>E. coli</i> DH5 $\alpha$	F- (80d lacZ M15) (lacZYA-argF) U169 hsdR17 (r - m +) recA1 endA1 relA1 deoR	RBC (Real Biotech)
<i>C. glutamicum</i>	Wild type	ATCC 13032
<b>Plasmids</b>		
pCES208H36GFP	6.7 kb, <i>E. coli</i> - <i>C. glutamicum</i> shuttle vector, Km <sup>r</sup> , pCES208 derivative; P <sub>H36</sub> , eGFP	[3]
pCES208H36GFP- ChnDE	8.9 kb, pCES208 derivative; P <sub>H36</sub> , eGFP	This study
<b>Primers</b>		
<i>ChnE</i> -F	5'-GAGTAGCATGGG <b>ATCC</b> ATGAACTATCCA AATATACCTTTATATATCAACGGTGAG-3'	<i>Bam</i> HI
<i>ChnE</i> -R	5'-TCATGCTGTTTCATATGCTAATTGAGTTG CGTAATAAATTTGGTTCTGAGGT-3'	<i>Nde</i> I
<i>ChnD</i> -F	5'- AATGGAATCAAAG <b>TTAGAAAGGAGG</b> AT GCACTGTTACTGCGTGACG-3'	<i>Hpa</i> I
<i>ChnD</i> -R	5'- TCTAATTTGAAG <b>TTTCAGTTTT</b> CGTGCA TAAGCACAATACG-3'	<i>Hpa</i> I

Restriction sites are shown in bold. The underlined nucleotides represent ribosome binding site.

## References

1. Iwaki, H.; Hasegawa, Y.; Teraoka, M.; Tokuyama, T.; Bergeron, H.; Lau, P.C. Identification of a Transcriptional Activator (ChnR) and a 6-Oxohexanoate Dehydrogenase (ChnE) in the Cyclohexanol Catabolic Pathway in *Acinetobacter* sp. Strain NCIMB 9871 and Localization of the Genes that Encode Them. *J. Appl. Environ. Microbiol.* **1999**, *65*, 5158-5162.
2. Park, J.-U.; Jo, J.-H.; Kim, Y.-J.; Chung, S.-S.; Lee, J.-H.; Lee, H.-H. Construction of Heat-Inducible Expression Vector of *Corynebacterium glutamicum* and *C. ammoniagenes*: Fusion of  $\lambda$  Operator with Promoters Isolated from *C. ammoniagenes*. *J. Microbiol. Biotechnol.* **2008**, *18*, 639-647.
3. Yim, S.S.; An, S.J.; Kang, M.; Lee, J.; Jeong, K.J. Isolation of Fully Synthetic Promoters for High-Level Gene Expression in *Corynebacterium glutamicum*. *Biotechnol. Bioeng.* **2013**, *110*, 2959-2969, doi:10.1002/bit.24954.