

Supporting information

Selectivity and Sustainability of Electroenzymatic Process for Glucose Conversion to Gluconic Acid

Varničić M.^a, Zasheva N.I.^b, Haak E.^c, Sundmacher K.^{a,b} and Vidaković-Koch T.^a

^a *Max Planck Institute for Dynamics of Complex Technical Systems, Sandtorstr. 1, 39106 Magdeburg, Germany*

^b *Otto-von-Guericke- University Magdeburg, Chair for Process Systems Engineering, Universitätsplatz 2, 39106 Magdeburg, Germany*

^c *Otto-von-Guericke- University Magdeburg, Institut für Chemie, 39106 Magdeburg, Germany*

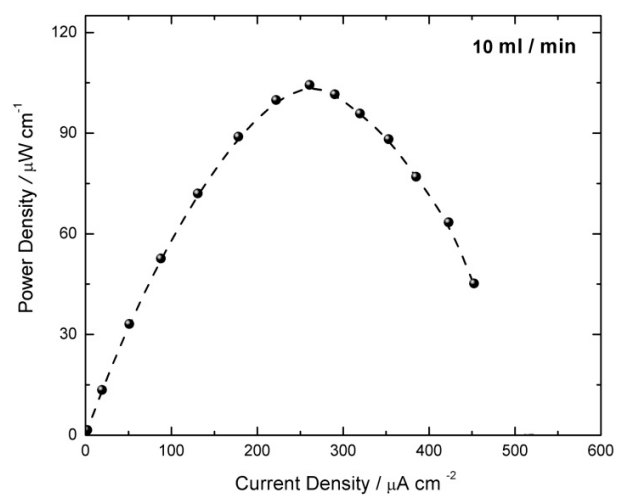
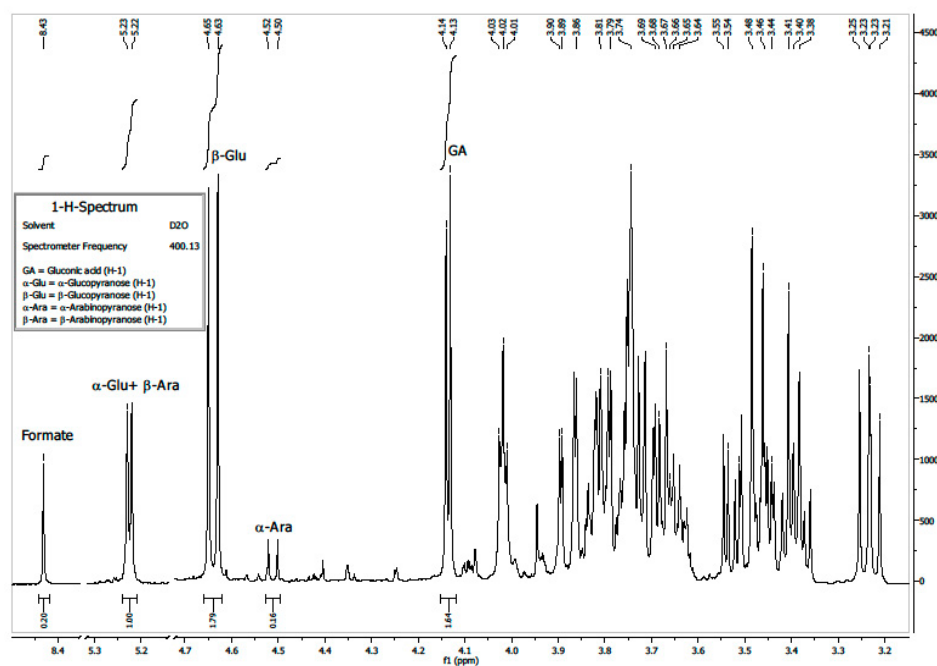
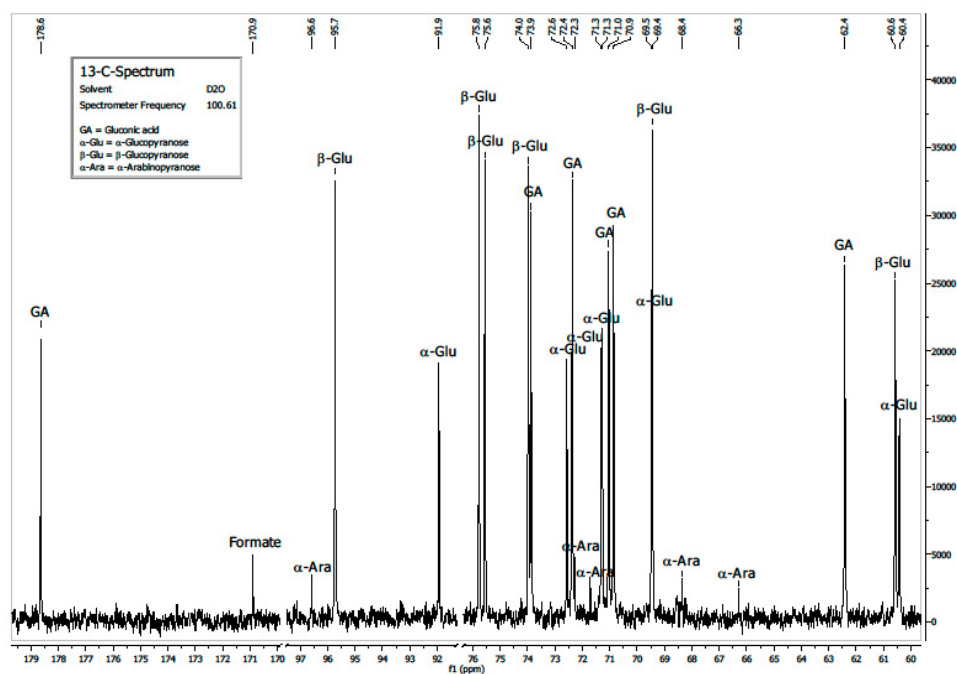


Figure S1 Power curve of the electroenzymatic reactor at 10 ml min⁻¹ flow rate, 20 mM glucose and O₂ supplied from the gas phase at the cathode side

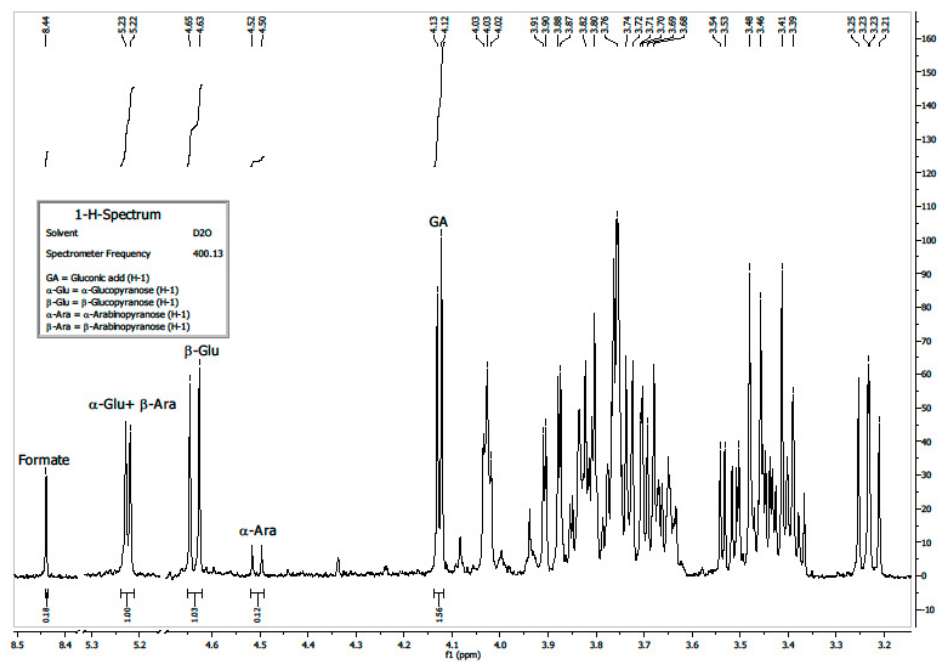


a)

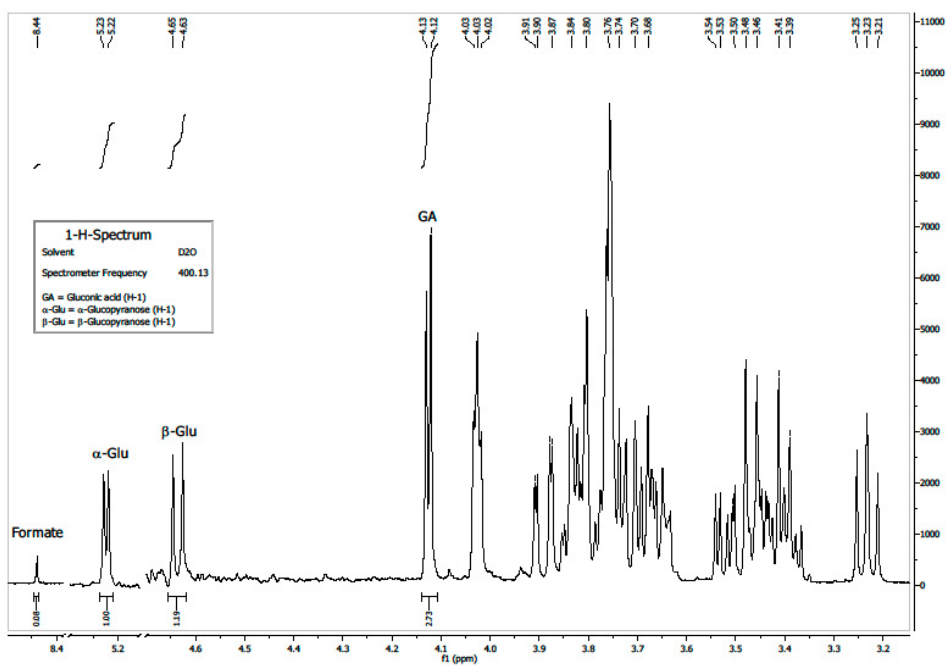


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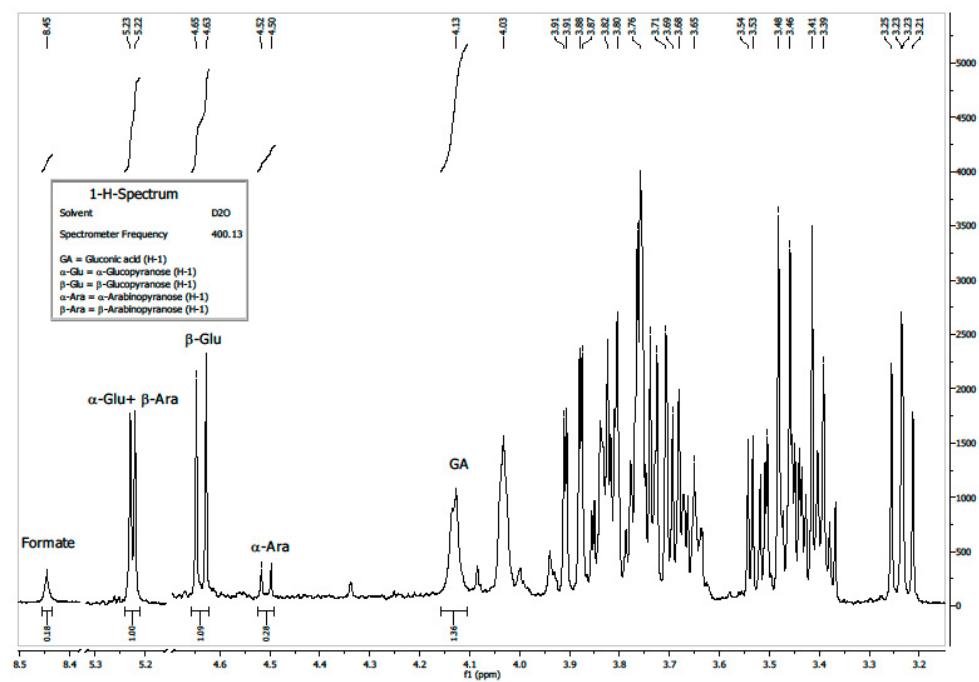
Figure S2 NMR spectra of lyophilized sample for D-arabinose presentation a) ^1H spectra b) ^{13}C spectra



a)

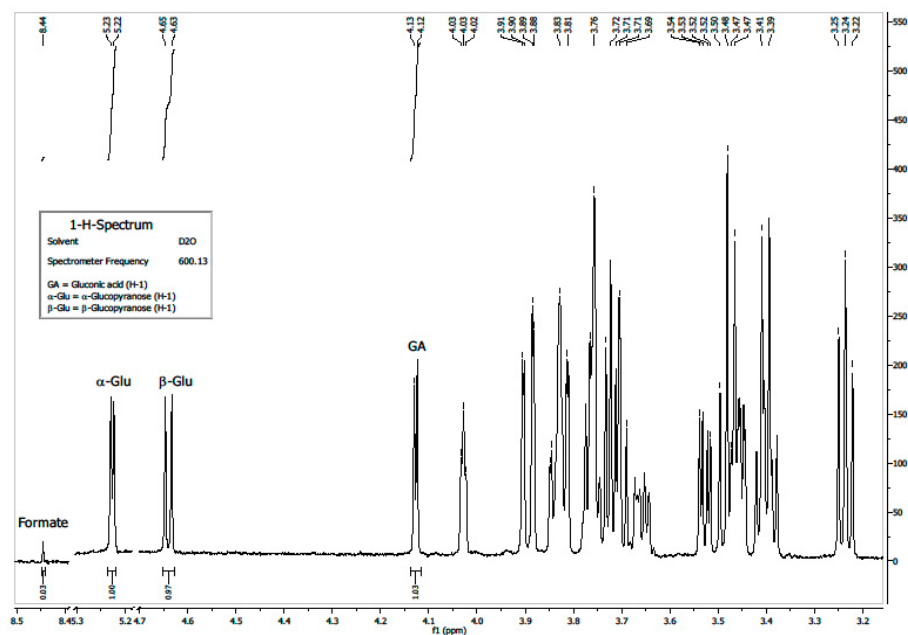


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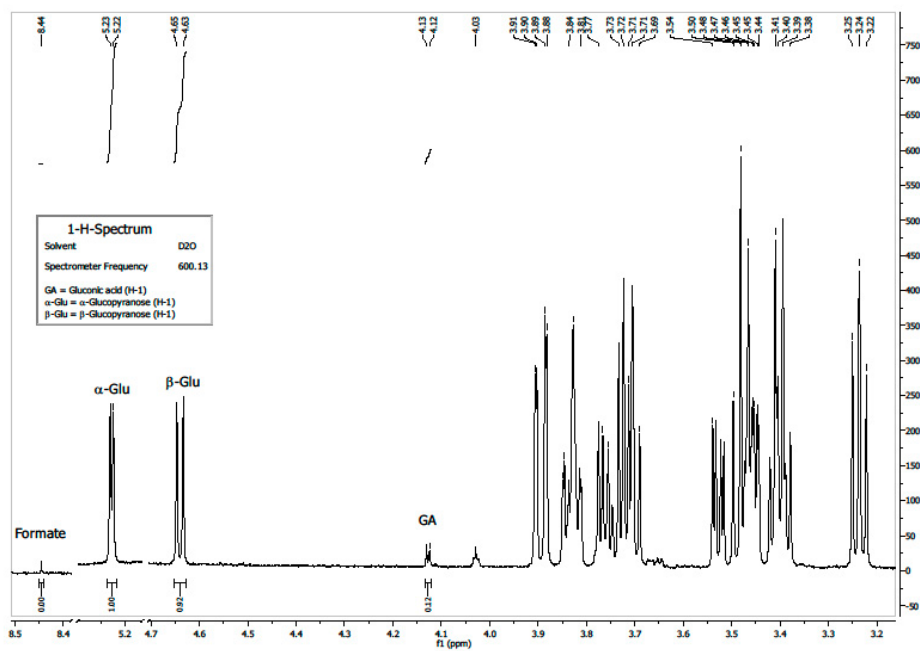


c)

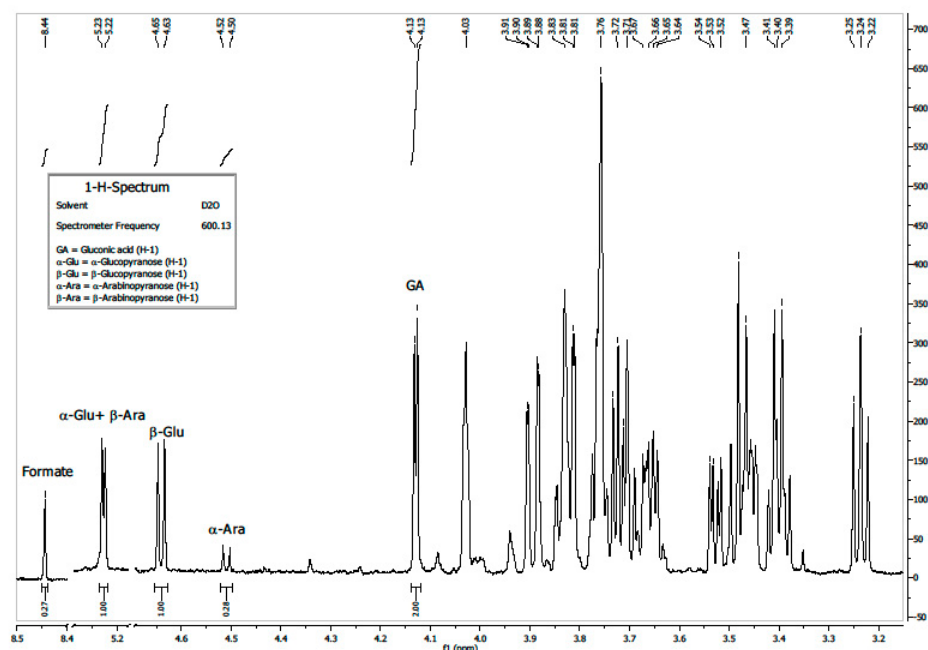
Figure S3 NMR analysis of reaction solutions after 7h in electroenzymatic reactors tested at three different flow rates a) 10 ml min⁻¹, b) 5 ml min⁻¹ and c) 2 ml min⁻¹



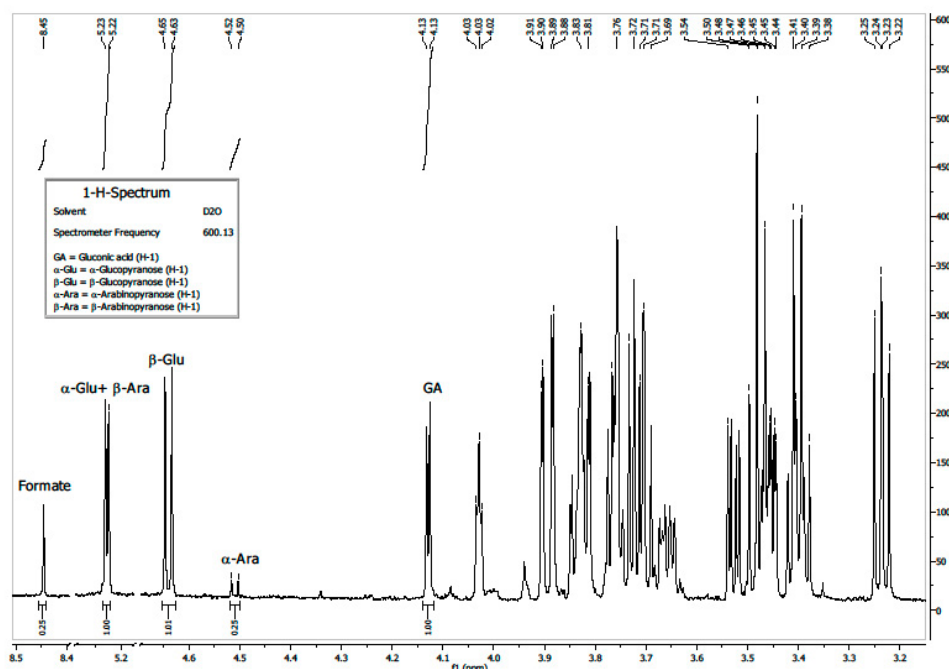
a)



b)



c)



d)

Figure S4 NMR analysis of reaction mixtures after 7h of enzymatic electrodes operation in half-cell experiments, a) and b) anode at different potentials; c) and d) cathode, at different potentials, Conditions presented in Table 2

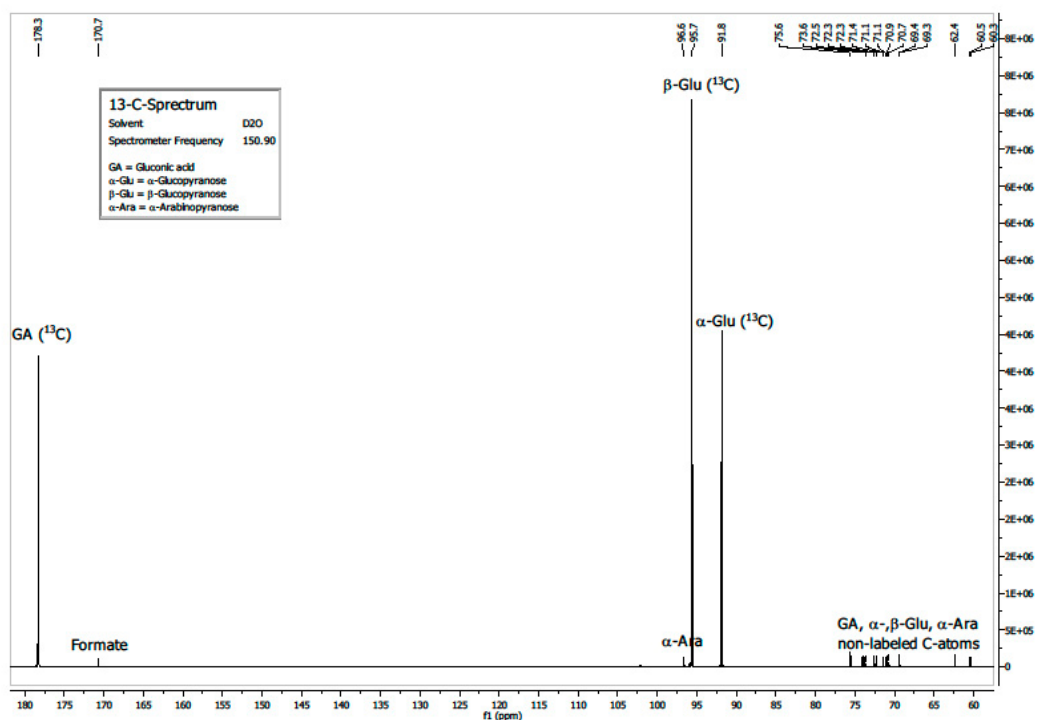
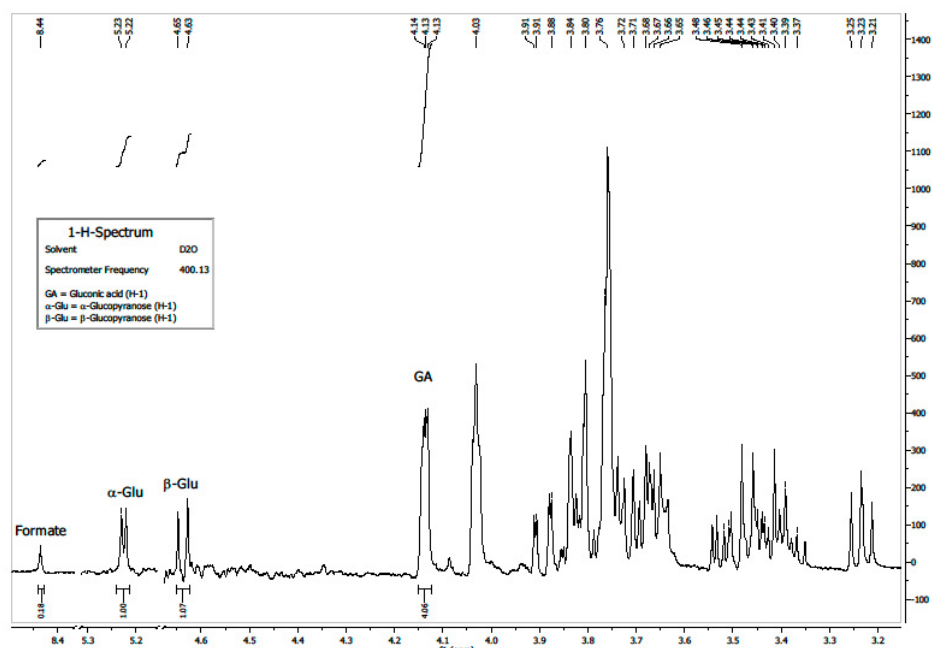
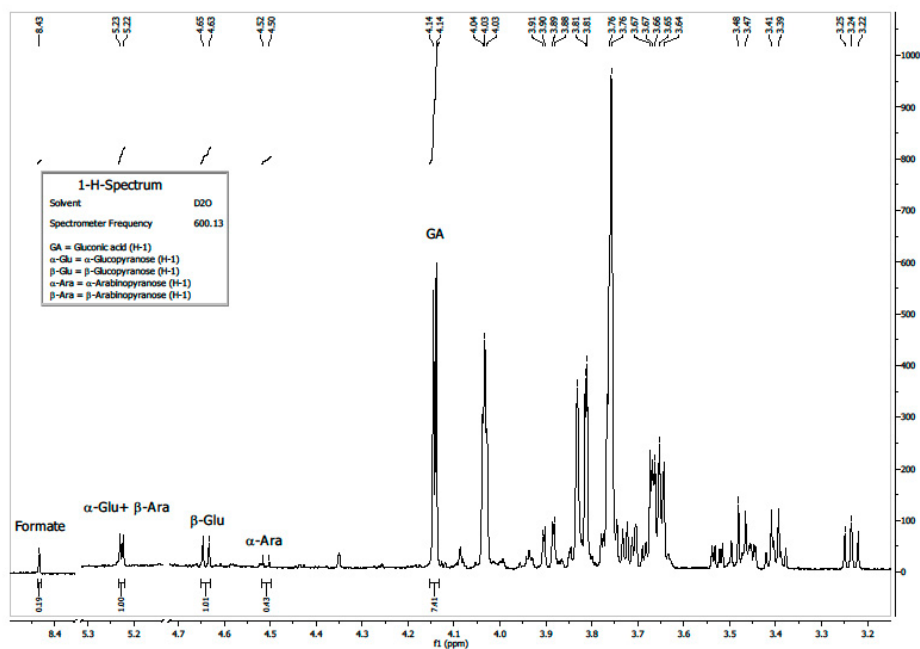


Figure S5 NMR spectra with labeled glucose (^{13}C spectra)



a)



b)

Figure S6 NMR results for electroenzymatic reactors operated at two different conditions of cell potential and time, 5 ml min⁻¹ a) cell potential -0.1 V, time 16 h and b) cell potential variation between -0.1 to 0.05, time 21h, Conditions presented in Table 3

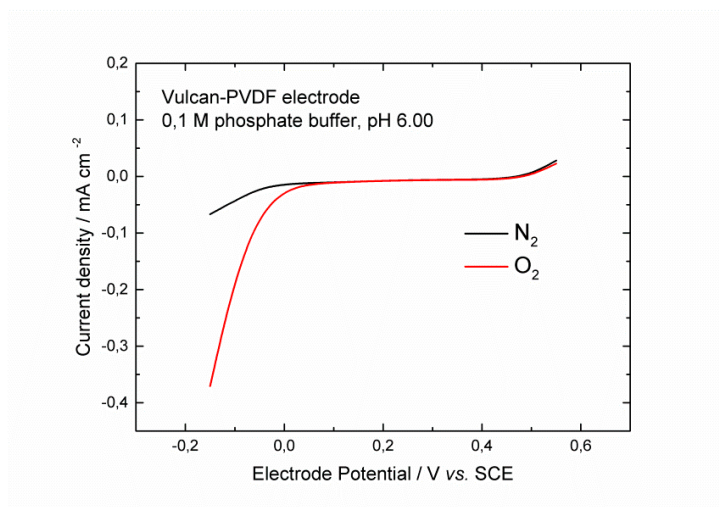


Figure S7 Polarization curves of Vulcan nanomaterial in the presence and absence of oxygen

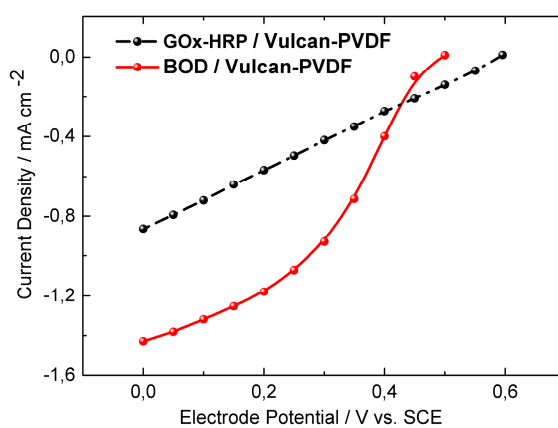


Figure S8 Comparison of BOD and GOx-HRP enzymatic electrode obtained in half-cell system, 20 mM glucose, Conditions for BOD-electrode: pH 7.00, 37 °C, for GOx-HRP: pH 6.00, 22 °C; 400 rpm, O₂