

cancers

ISSN 2072-6694

www.mdpi.com/journal/cancers

Review

Metabolic Disorder, Inflammation, and Deregulated Molecular Pathways Converging in Pancreatic Cancer Development: Implications for New Therapeutic Strategies

Yoshiharu Motoo ^{1,*}, Takeo Shimasaki ^{1,3}, Yasuhito Ishigaki ², Hideo Nakajima ¹, Kazuyuki Kawakami ³ and Toshinari Minamoto ³

- Department of Medical Oncology, Kanazawa Medical University, 1-1 Daigaku, Uchinada, Ishikawa 920-0293, Japan; E-Mails: takeo@kanazawa-med.ac.jp (T.S.); hideonak@kanazawa-med.ac.jp (H.N.)
- Medical Research Institute, Kanazawa Medical University, 1-1 Daigaku, Uchinada, Ishikawa 920-0293, Japan; E-Mail: ishigaki@kanazawa-med.ac.jp
- Division of Translational & Clinical Oncology, Cancer Research Institute, Kanazawa University, Kanazawa, Japan; E-Mails: kawakami@med.kanazawa-u.ac.jp (K.K.); minamoto@staff.kanazawa-u.ac.jp (T.M.)
- * Author to whom correspondence should be addressed; E-Mail: motoo@kanazawa-med.ac.jp; Tel.: +81-76-218-8284; Fax: +81-76-218-8283.

Received: 17 December 2010; in revised form: 18 January 2011 / Accepted: 21 January 2011 / Published: 24 January 2011

Abstract: Pancreatic cancer develops and progresses through complex, cumulative biological processes involving metabolic disorder, local inflammation, and deregulated molecular pathways. The resulting tumor aggressiveness hampers surgical intervention and renders pancreatic cancer resistant to standard chemotherapy and radiation therapy. Based on these pathologic properties, several therapeutic strategies are being developed to reverse refractory pancreatic cancer. Here, we outline molecular targeting therapies, which are primarily directed against growth factor receptor-type tyrosine kinases deregulated in tumors, but have failed to improve the survival of pancreatic cancer patients. Glycogen synthase kinase-3 β (GSK3 β) is a member of a serine/threonine protein kinase family that plays a critical role in various cellular pathways. GSK3 β has also emerged as a mediator of pathological states, including glucose intolerance, inflammation, and various cancers (e.g., pancreatic cancer). We review recent studies that demonstrate the anti-tumor effects of GSK3 β inhibition alone or in combination with chemotherapy and radiation. GSK3 β

inhibition may exert indirect anti-tumor actions in pancreatic cancer by modulating metabolic disorder and inflammation.

Keywords: pancreatic cancer; glucose intolerance; inflammation; oncogenic pathways; therapeutic target; $GSK3\beta$

Abbreviations

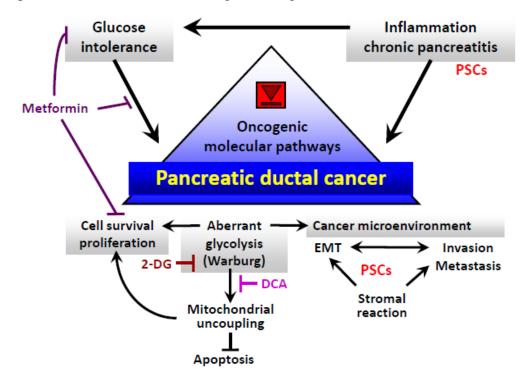
AMPK, 5'-AMP-activated protein kinase; ATP, adenosine triphosphate; CDK, cyclin-dependent kinase; COX-2, cyclooxygenase-2; CT, computed tomography; DCA, dichloroacetate; 2-DG, 2-deoxy-D-glucose; DM, diabetes mellitus; DPC, deleted in pancreatic carcinoma; EGF, epidermal growth factor; EGFR, EGF receptor; EMT, epithelial-mesenchymal transition; FDG, [¹⁸F] fluoro-2-D-deoxyglucose; FGF, fibroblast growth factor; GSK3β, glycogen synthase kinase-3β; HGF, hepatocyte growth factor; hTERT, human telomerase reverse transcriptase; IGF-1, insulin-like growth factor-1; LKB1, liver kinase B1; MAPK, mitogen-activated protein kinase; mTOR, mammalian target of rapamycin; NADPH, nicotinamide adenine dinucleotide phosphate; NF-κB, nuclear factor-κB; NO, nitric oxide; iNOS, inducible NO synthase; PDGF, platelet-derived growth factor; PDGFR, PDGF receptor; PDH, pyruvate dehydrogenase; PDK1, PDH kinase 1; PET, positron emission tomography; PI3K, phosphatidylinositol 3'-kinase; PPARγ, peroxisome proliferator-activated receptor-γ; PSC, pancreatic stellate cell; ROS, reactive oxygen species; TCA, tricarboxylic acid; TGF, transforming growth factor; VEGF, vascular endothelial growth factor; VEGFR, VEGF receptor.

1. Introduction

Pancreatic cancer is a devastating disease and a major unresolved health problem due to its late clinical diagnosis and predisposition towards metastasis [1,2]. Pancreatic cancer is characterized by highly proliferative and invasive tumor cells [3]. Despite progress in approaches to treatment [4], such aggressive biological behavior thwarts early diagnosis and curative surgical intervention and renders tumors resistant to conventional chemotherapy, radiation therapy, and their combination [5-7], leading to a poor prognosis [8]. Therefore, understanding the detailed molecular and biological basis of pancreatic cancer pathogenesis facilitates advances in the diagnosis, treatment, and prevention of this disease.

Pancreatic cancer develops through a series of complex, cumulative biological processes involving metabolic disorder and chronic local inflammation in association with stromal changes and deregulated molecular pathways (Figure 1). Based on these pathologic properties, new therapeutic strategies are being developed to reverse the refractory stage of pancreatic cancer. Here, we review the multidimensional processes leading to pancreatic cancer development and progression, and discuss biology-based therapeutic alternatives to conventional cancer treatments. In addition, we highlight an emerging strategy for cancer treatment that targets glycogen synthase kinase 3β (GSK3 β), focusing on the effect of its deregulation on pancreatic cancer.

Figure 1. Molecular and biological pathways involved in the development and progression of pancreatic cancer and putative mechanisms underlying the anticancer effects of metformin, 2-deoxyglucose (2-DG), and dichloroacetate (DCA). The gray triangle in a box indicates a molecular target. Abbreviations: EMT, epithelial-to-mesenchymal transition; PSCs, pancreatic stellate cells; Warburg, Warburg effect.



2. Metabolic Disorder Coincides with Pancreatic Cancer

Although systemic metabolic disorders such as obesity and type II diabetes mellitus (DM) influence the risk of developing pancreatic cancer and clinical outcomes (reviewed in [9,10]), the abnormal metabolic profile of cancer cells dictates their survival, proliferation, and invasion, as well as susceptibility to chemotherapy and radiation [11,12]. Thus, metabolic disorder and altered tumor cell metabolism are potential targets for cancer treatment and (chemo) prevention [13,14].

2.1. Obesity, Glucose Intolerance, and Pancreatic Cancer

The association between obesity/DM and pancreatic cancer risk has long been controversial. However, recent reviews and meta-analyses of prospective observational studies have demonstrated that obesity, defined by an increased body mass index, is significantly associated with a risk of pancreatic cancer development [15,16]. DM is also a clinical manifestation of pancreatic cancer, and case-control and prospective studies have demonstrated an increased risk of pancreatic cancer in patients with long-term DM [17-19]. Obesity is associated with the early manifestations of pancreatic cancer and lower overall patient survival [20], although the influence of DM on pancreatic cancer progression or morbidity is not clear [10]. Possible mechanisms behind the association between obesity and worse clinical outcomes may include increased risk of DM, thrombosis, or other comorbidities; impaired immune response leading to aggressive tumor behavior; and poor response to conventional anticancer therapies [10]. The effect of obesity and DM on the development and

progression of pancreatic cancer appears to be mediated by adipokines, reactive oxygen species (ROS), inflammatory cytokines, and insulin resistance, which result in activation of insulin-like growth factor-1 (IGF-1) and mammalian target of rapamycin (mTOR) pathways (reviewed in [10]).

A number of studies have reported the potential role of glucose-lowering therapies in reducing the risk of pancreatic cancer (reviewed in [21]). Metformin is a biguanide that is most frequently prescribed for diabetes [22]. Recently, a comprehensive review and meta-analysis of epidemiologic studies demonstrated an inverse correlation between the use of metformin and incidence of pancreatic cancer and overall survival of patients with diabetes [23]. Increasing evidence also suggests that metformin exerts a therapeutic effect against cancer [24,25]. Metformin decreases insulin resistance and indirectly reduces levels of insulin and IGF-1, which promote cancer cell proliferation [26]. Further, metformin activates the tumor suppressor pathway mediated by liver kinase B1 (LKB1) and 5' AMP-activated protein kinase (AMPK), an important sensor of cellular energy status [27], thereby inactivating mTOR signaling [14]. IGF-1 receptor and G protein-coupled receptor signaling is implicated in the autocrine-paracrine stimulation of a variety of malignant tumors, including exocrine pancreatic cancer. Recent studies demonstrated that metformin-induced activation of AMPK disrupts the crosstalk between insulin/IGF-1 receptor and G protein-coupled receptor signaling pathways in pancreatic cancer cells and inhibits proliferation of these cells in xenograft models, suggesting this crosstalk as a target for treatment of pancreatic cancer by metformin [28].

2.2. Distinct Metabolic Properties of Cancer Cells

The fundamental metabolic characteristics of cancer cells include increased glucose uptake, aerobic glycolysis even under normoxic condition (Warburg effect) [29], and impaired oxidative phosphorylation in the tricarboxylic acid (TCA) cycle, which results in mitochondrial uncoupling [30]. These properties could explain the ability of cancer cells to survive, invade host tissues, and resist the induction of apoptosis by chemotherapeutic agents and ionizing radiation [30,31]. However, the glycolytic phenotype of cancer cells is a potential target for cancer diagnosis and treatment [32]. For example, enhanced glucose uptake by cancer cells can be used to visualize cancer by positron emission tomography (PET) using the radioisotopelabeled glucose analogue 2-[18F]-fluoro-2-deoxy-D-glucose (FDG). FDG-PET in combination with computed tomography (PET-CT) enables detection of metastatic lesions of most cancers with both sensitivity and specificity greater than 90% [33]. Pharmacologic agents targeting the glycolytic phenotype of cancer cells include 2-deoxy-D-glucose (2-DG) and dichloroacetate (DCA).

A glucose analogue, 2-DG, is the most attractive agent for targeting aberrant glucose metabolism in cancer cells [34]. 2-DG inhibits glucose transport by competing with glucose transporters and is subsequently phosphorylated by hexokinase to form 2-DG-6-phospate. Phosphorylated 2-DG is not further metabolized by inhibiting glucose-6-phosphate isomerase, thereby reducing the production of adenosine triphosphate (ATP) and nicotinamide adenine dinucleotide phosphate (NADPH) from glycolysis coupled with the pentose phosphate pathway. Thus, 2-DG exerts antitumor effects by starving cancer cells [34].

The association between the glycolytic phenotype (*i.e.*, TCA cycle defects) and resistance to apoptosis is attributed to decreased mitochondrial hydrogen peroxide production and cytochrome C release [30,31]. Pyruvate dehydrogenase (PDH) plays a crucial role in triggering the TCA cycle by

converting pyruvate to citric acid. PDH kinase 1 (PDK1), which phosphorylates and inactivates PDH, is frequently overactivated in cancer cells, resulting in an impaired TCA cycle and mitochondrial hyperpolarization. Thus, inhibiting PDK1 would re-activate PDH and restore mitochondrial membrane polarity, thereby facilitating cancer cell apoptosis in response to chemotherapeutic agents and radiation. DCA, an orally bio-available small molecule, is a well characterized PDK1 inhibitor. The ability of DCA to inhibit lactate production (by stimulating PDH and the TCA cycle) has been long used to treat lactic acidosis, which complicates inherited mitochondrial disorders [35,36]. A recent study demonstrated that DCA induces cancer cell apoptosis by selectively inhibiting PDK1 in cancer cells, leading to metabolic remodeling from glycolysis to glucose oxidation and normalization of mitochondrial function [37]. A recent clinical trial of oral DCA in children with congenital lactic acidosis reported that DCA was well tolerated and safe [36]. Thus, orally available DCA is a promising selective anticancer agent.

3. Inflammation and Stromal Reactions in Pancreatic Cancer

The well recognized link between chronic inflammation and tumor development in many organs [38] is consistent with the reported causative association and interaction between chronic pancreatitis and pancreatic cancer [39-41]. In addition to the etiologic role of inflammation in carcinogenesis, systemic and local inflammation are frequent manifestations of pancreatic cancer and have been implicated in tumor progression and clinical outcomes [39].

3.1. Chronic Pancreatitis and Pancreatic Cancer

Chronic pancreatitis is a risk factor for developing pancreatic cancer [39-41]. This association is supported by a recent meta-analysis of 22 well-performed epidemiologic studies [42]. The risk of developing pancreatic cancer for patients with hereditary pancreatitis is much higher than for patients with sporadic chronic pancreatitis [43-45]. The incidence of chronic pancreatitis in the general population is only about 5 to 10 per 100,000 persons a year, as estimated from hospitalization data. In particular, hereditary pancreatitis caused by germline mutations in the cationic trypsinogen gene [46] accounts for less than 1% of all chronic pancreatitis cases. Therefore, chronic pancreatitis does not constitute a main precursor of pancreatic cancer [9,42].

Despite this low incidence, both forms of chronic pancreatitis have provided substantial evidence for putative inflammatory mechanisms contributing to pancreatic cancer development and progression, including proinflammatory cytokines, nuclear factor-κB (NF-κB), cyclooxygenase-2 (COX-2), peroxisome proliferator-activated receptor-γ (PPARγ), nitric oxide (NO) synthesized by inducible NO synthase (iNOS), DNA damage caused by release of proteolytic enzymes and ROS, and somatic mutations in oncogenes (e.g., K-ras) and tumor suppressor genes (e.g., p53, p16, DPC4/Smad) [47-49]. Pancreatic cancer progression shares these molecular alterations, which are promising targets for early molecular diagnosis, treatment, and prevention of the disease [47-49].

Histopathologic findings of chronic pancreatitis include marked fibrosis, in which pancreatic stellate cells (PSCs) play a crucial role [50,51]. In the normal pancreas, quiescent PSCs produce vitamin A in the periacinar and interlobular space. In response to pancreatic inflammation, PSCs are activated and transformed to a myofibroblast-like phenotype; they proliferate, migrate and produce

extracellular matrix components (e.g., collagens, laminin, fibronectin), matrix metalloproteinases, and tissue inhibitors of matrix metalloproteinases. This phenotypic change is induced by inflammatory cytokines (e.g., tumor necrosis factor, interleukin-1, and interleukin-6), growth factors such as transforming growth factor (TGF)- β 1, TGF- α , platelet-derived growth factor (PDGF), and fibroblast growth factor (FGF)-2; and ROS [52]. In addition to their roles in the pathogenesis of chronic pancreatitis and cancer, these factors also promote pancreatic cancer progression [50,51].

3.2. Cancer-Stromal Interaction and Tumor Microenvironment

Chronic inflammation, desmoplastic stromal reaction, and neovascularization associated with pancreatic cancer combine to produce a distinct tissue microenvironment where cancer cell proliferation and invasion are facilitated by cancer-stromal interactions [53,54]. Activated PSCs are primarily responsible for the desmoplastic reaction and tumor angiogenesis in response to various growth factors such as TGF-β, FGF, HGF, and IGF-1. Results from *in vitro* and *in vivo* studies suggest that cancer cells recruit PSCs to tumors, where PSCs promote cancer cell proliferation and facilitate their invasion and metastasis, and that FGF and PDGF mediate these interactions between the two cell types (reviewed in [55]).

Phenotypic changes also occur in cancer cells, as represented by epithelial-mesenchymal transition (EMT) at the interface between tumor and stroma, in which epithelial cells undergo morphologic changes characterized by a transition from epithelial to fibroblastic (mesenchymal cell) phenotypes. Most factors involved in pancreatic cancer-stromal interactions have the potential to induce EMT in cancer cells. This process involves loss of cell-to-cell adhesion and E-cadherin expression, actin cytoskeleton reorganization, and increased expression of mesenchymal molecules (e.g., vimentin, fibronectin, α -smooth muscle actin, N-cadherin). In this way, EMT facilitates the invasion and metastasis of cancer cells and renders them resistant to chemotherapy and radiation [56,57]. Accordingly, growth factors such as TGF- β and hepatocyte growth factor (HGF, or its receptor c-Met) that are involved in cancer-stromal interactions and EMT have been well studied in order to develop therapeutic strategies targeting these factors [53,54].

4. Targeting Molecular Pathways Deregulated in Pancreatic Cancer

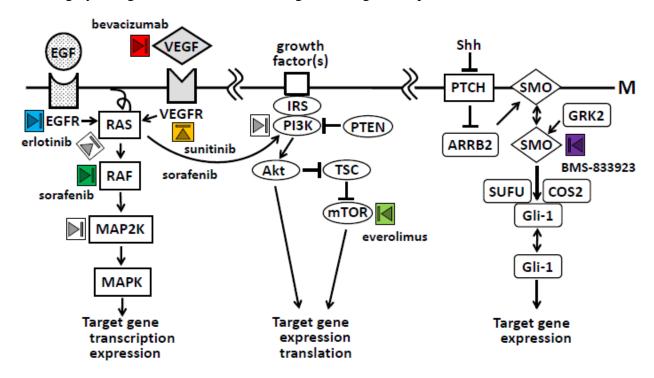
Most cases of pancreatic cancer are resistant to conventional chemotherapy and radiation therapy [5-7]; therefore, new strategies are needed to enhance the antitumor effects of gemcitabine, which is the standard chemotherapeutic agent used to treat pancreatic cancer [58]. These new classes of biology-based treatment modalities include molecular target-directed therapies.

4.1. Deregulated Molecular Pathways Mediated by Receptor-Type Tyrosine Kinases

Molecular studies have investigated the complex genetic mechanisms of cancer, which involve multidirectional signal transduction pathways [3,59,60]. As shown in Figure 2, the major signal transduction pathways in pancreatic cancer pathogenesis and progression are RAS/mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI3K)/Akt/mTOR, and hedgehog pathways [3]. The receptor tyrosine kinase family and their ligands, which include epidermal growth factor (EGF)

receptor (EGFR), vascular endothelial growth factor (VEGF) receptor (VEGFR), and PDGF receptor (PDGFR), are targets of therapy because they are overexpressed in many tumor types, including pancreatic cancer [61].

Figure 2. Critical molecular pathways leading to the development and progression of pancreatic cancer. Abbreviations: ARRB2, arrestin β2; COS2, kinesin-related protein Costal 2; DUSP6, dual specificity phosphatase 6; EGF, epidermal growth factor; EGFR, EGF receptor; GRK2, G protein-coupled receptor kinase-2; IRS, insulin receptor substrate 1; M, cell membrane; MAPK, mitogen-activated protein kinase; MAP2K, MAP kinase kinases; mTOR, mammalian target of rapamycin; PI3K, phosphatidylinositol 3-kinase; PTCH, patched; PTEN, phosphatase and tensin homolog deleted in chromosome 10; Shh, sonic hedgehog; SMO, smoothened; SUFU, suppressor of fused; TSC, tuberous sclerosis complex; VEGF, vascular endothelial growth factor; VEGFR, VEGF receptor. The gray triangle in a box indicates a target for drug development.



4.2. Pharmacologic Agents that Target Deregulated Kinases

Currently available agents that target these factors include anti-EGFR antibodies (cetuximab, panitumumab), small molecule EGFR inhibitors (gefitinib, erlotinib), an anti-VEGF antibody (bevacizumab), and a small molecule VEGFR inhibitor (axitinib). A number of phase III clinical trials have tested kinase inhibitors as monotherapy or in combination therapy with gemcitabine for pancreatic cancer, but other than the combination of erlotinib and gemcitabine [62], these approaches have produced few therapeutic benefits [63]. Characterization of new molecular targets is necessary in order to develop strategies that enhance the effect of gemcitabine and improve the survival rate. Recent studies have pursued potential kinases as targets for new anticancer agents [64] and evaluated agents targeting known kinases (e.g., EGFR, check-point kinase 1) to combine with gemcitabine in order to improve its antitumor effects [65].

5. GSK3ß, an Emerging Therapeutic Target in Cancer

GSK3 β has emerged as a critical factor that plays distinct pathologic roles in glucose intolerance, inflammation, and in various cancer types (e.g., pancreatic cancer). Here, we briefly review recent studies, including our own, that demonstrate the direct anticancer effects of GSK3 β inhibition, alone or in combination with chemotherapy and radiation.

5.1. Outline of GSK3\beta and its Involvement in Chronic Progressive Diseases

GSK3 β was first identified as a serine/threonine protein kinase that regulates glucose/glycogen metabolism under the control of insulin signaling. Unlike most protein kinases, GSK3 β is active in normal cells, and its activity is controlled by subcellular localization, differential phosphorylation at serine 9 (S9) and tyrosine 216 (Y216) residues, and different binding partners. In addition to regulating its primary target, glycogen synthase, GSK3 β is involved in other fundamental cellular pathways depending on its substrates and binding partners [66-68]. GSK3 β is a potential therapeutic target for common chronic diseases including type 2 DM and Alzheimer's disease, given the causative associations with glucose intolerance, neurodegenerative disorders, and inflammation [69-71].

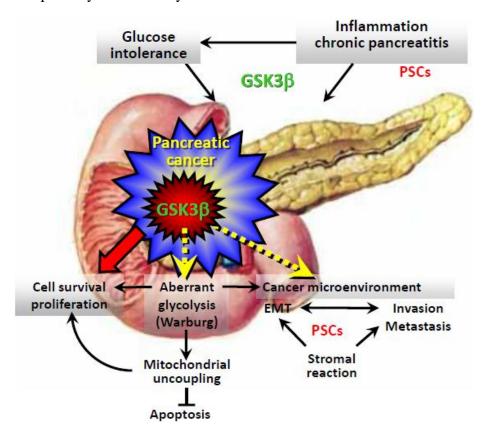
5.2. Pathologic Role of GSK3β in Various Cancer Types

Under physiologic conditions, GSK3 β phosphorylates several transcription factors (e.g., c-Jun, c-Myc), cell cycle regulators (e.g., cyclin D1), and proto-oncoproteins (e.g., β -catenin), thereby triggering their degradation via the ubiquitin-proteasome system. GSK3 β is, therefore, hypothesized to inhibit tumor development by interfering with oncogenic signaling (e.g., Wnt, hedgehog) [72]. However, there is little evidence that links GSK3 β inactivation or loss of GSK3 β expression with tumor development.

In the last five years, we demonstrated that deregulated expression, phosphorylation of S9 and Y216, and GSK3β activity are distinct features of gastrointestinal cancers including pancreatic cancers and glioblastoma, and that GSK3β sustains the survival and proliferation of these tumor cells. A pathologic role for GSK3β is supported by observations that inhibition of its activity reduced the survival and proliferation of different cancer cell types, predisposing them to apoptosis both *in vitro* and in tumor xenografts [73-76]. We also found that GSK3β inhibition in cancer cells was accompanied by restoration of p53 and Rb tumor suppressor pathways [75,76] and downregulation of human telomerase reverse transcriptase (hTERT), resulting in cell senescence [76]. This led us to propose GSK3β as a potential target for cancer treatment and to apply for domestic and international patents [77].

Simultaneously and following our studies on the antitumor effects of GSK3β inhibition, similar observations from other laboratories were reported for various cancer types with underlying mechanisms that included regulation by GSK3β inhibition of several pathways mediated by p53, Rb, p27^{Kip1}, cyclin-dependent kinase (CDK), cyclin D1, c-Myc, and NF-κB [78]. Although the putative role of GSK3β in cancer is still debated [79,80], the overall results indicate that aberrant expression and activity of GSK3β is likely to be a common and fundamental characteristic of a broad spectrum of cancers (Figure 3) [78].

Figure 3. Systemic and local effects of aberrant GSK3 β on risk factors (glucose intolerance and chronic inflammation) and progression of pancreatic cancer. Molecular mechanisms leading to the pathways indicated by the dotted arrows are not well characterized.



Based on the potential involvement of GSK3β in NF-κB-mediated cell survival [81,82], a number of studies demonstrated that GSK3β is involved in pancreatic cancer cell survival via the NF-κB pathway [83-85]. Few studies had focused on the role of GSK3β in the cellular response to chemotherapy until we reported that GSK3β inhibition sensitizes glioblastoma cells to chemotherapeutic agents (e.g., temozolomide, ACNU) and ionizing radiation [75]. However, a recent study failed to demonstrate that disrupting NF-κB activity by inhibiting GSK3β sensitizes PANC-1 pancreatic cancer cells to gemcitabine [85]. We previously found that GSK3β inhibition did not affect endogenous NF-κB transcriptional activity in tumor cells established from pancreatic cancers and glioblastoma [75,76]. In a preliminary study, we found that a small-molecule GSK3β inhibitor increased pancreatic cancer cell sensitivity to gemcitabine in cell culture and tumor xenografts when its dose and treatment protocol were optimized, and have identified the molecular mechanisms underlying the increased sensitivity [86]. Our findings indicate that GSK3β inhibition combined with chemotherapy is a novel and promising strategy to sensitize pancreatic cancer cells to gemcitabine.

5.3. Putative Antitumor Effects of GSK3β Inhibition via Modulation of Tumor Biology

Increasing evidence suggests that GSK3β participates in a wide range of physiological processes that determine cell fate, including cell motility, energy metabolism, and transcriptional control (reviewed in [78]). In addition to the pathologic roles of GSK3β in cancer cell survival and proliferation [73-78,83-85] as discussed above, GSK3β may influence critical biological properties of

cancer cells, such as their dependence on glycolysis and invasive ability associated with EMT (Figure 3). Modulation of these biological properties by pharmacologic inhibition of GSK3 β may sensitize pancreatic cancer cells to standard chemotherapy and radiation. Given the systemic pathology caused by aberrant GSK3 β activity in glucose intolerance and chronic inflammation [71], GSK3 β inhibition may decrease the risk of developing pancreatic cancer by improving these conditions. Accordingly, investigating the functions and pathologic roles of GSK3 β should establish a firm molecular basis for future cancer treatments that target this kinase.

6. Perspectives

Here, we reviewed recent studies on the epidemiologic characteristics of pancreatic cancer and the molecular and biological mechanisms contributing to its development and progression, and highlight advances in alternatives to conventional treatments. Although molecular target-directed therapy is currently attracting considerable attention, especially for cancer refractory to standard chemotherapy and radiation, this therapy produces therapeutic effects that are still far from sufficient for most patients with advanced and recurrent pancreatic cancer. For many cancer patients, including those with pancreatic cancer, resistance to currently available therapeutics presents a major obstacle and is due to the aberrant metabolism of cancer cells, their microenvironment, their ability to invade and metastasize, and the acquisition of gene mutations. Multidisciplinary approaches directed to a more complete understanding of pancreatic cancer pathogenesis hold great promise in improving the outlook of this disease.

Acknowledgments

This work was supported in part by Grants-in-Aids for Scientific Research from the Japanese Ministry of Education, Science, Sports, Technology and Culture (to Y.M., T.M.), Grant for Collaborative Research from Kanazawa Medical University (C2008-3), and by a Project Research Grant from the High-Tech Research Center of Kanazawa Medical University (H2010 -11).

References

- 1. Li, D.; Xie, K.; Wolff, R.; Abbruzzese, J.L. Pancreatic cancer. *Lancet* **2004**, *363*, 1049-1057.
- 2. Schneider, G.; Siveke, J.T.; Eckel, F.; Schmid, R.M. Pancreatic cancer: Basic and clinical aspects. *Gastroenterology* **2005**, *128*, 1606-1625.
- 3. Bardeesy, N.; DePinho, R.A. Pancreatic cancer biology and genetics. *Nat. Rev. Cancer* **2002**, *2*, 897-909.
- 4. Lockhart, A.C.; Rothenberg, M.L.; Berlin, J.D. Treatment for pancreatic cancer: Current therapy and continued progress. *Gastroenterology* **2005**, *128*, 1642-1654.
- 5. Wray, C.J.; Ahmad, S.A.; Matthews, J.B.; Lowy, A.M. Surgery for pancreatic cancer: Recent controversies and current practice. *Gastroenterology* **2005**, *128*, 1626-1641.
- 6. Ben-Josef, E.; Lawrence, T.S. Chemotherapy for unresectable pancreatic cancer. *Int. J. Clin. Oncol.* **2008**, *13*, 121-126.

7. Okusaka, T.; Ito, Y.; Furuse, J.; Yamada, S.; Ishii, H.; Shibuya, K.; Ioka, T.; Shinchi, H. Current status of chemoradiotherapy for locally advanced pancreatic cancer in Japan. *Int. J. Clin. Oncol.* **2008**, *13*, 127-131.

- 8. Jamel, A.; Siegel, R.; Ward, E.; Hao, Y.; Xu, J.; Thun, M.J. Cancer statistics, 2009. *CA Cancer J. Clin.* **2009**, *59*, 225-249.
- 9. Raimondi, S.; Maisonneuve, P.; Lowenfels, A.B. Epidemiology of pancreatic cancer: An overview. *Nat. Rev. Gastroenterol. Hepatol.* **2009**, *6*, 699-708.
- 10. Li, D.; Abbruzzese, J.L. New strategies in pancreatic cancer: Emerging epidemiologic and therapeutic concepts. *Clin. Cancer Res.* **2010**, *16*, 4313-4318.
- 11. Smallbone, K.; Gatenby, R.A.; Gillies, R.J.; Maini, P.K.; Gavaghan, D.J. Metabolic changes during carcinogenesis: Potential impact on invasiveness. *J. Theor. Biol.* **2007**, 244, 703-713.
- 12. DeBerardinis, R.J.; Sayed, N.; Ditsworth, D.; Thompson, C.B. Brick by brick: Metabolism and tumor cell growth. *Curr. Opin. Genet. Dev.* **2008**, *18*, 54-61.
- 13. Gatenby, R.A.; Gillies, R.J. Glycolysis in cancer: A potential target for therapy. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 1358-1366.
- 14. Engelman, J.A.; Cantley, L.C. Chemoprevention meets glucose control. *Cancer Prev. Res.* **2010**, *3*, 1049-1052.
- 15. Larsson, S.C.; Orsini, N.; Wolk, A. Body mass index and pancreatic cancer risk: A meta-analysis of prospective studies. *Int. J. Cancer* **2007**, *120*, 1993-1998.
- 16. Renehan, A.G.; Tyson, M.; Egger, M.; Heller, R.F.; Zwahlen, M. Body-mass index and incidence of cancer: A systemic review and meta-analysis of prospective observation studies. *Lancet* **2008**, *371*, 569-578.
- 17. Huxley, R.; Ansary-Moghaddam, A.; Berrington de Gonz ález, A.; Barzi, F.; Woodward, M. Type-II diabetes and pancreatic cancer: A meta-analysis of 36 studies. *Br. J. Cancer* **2005**, *92*, 2076-2083.
- 18. Luo, J.; Iwasaki, M.; Inoue, M.; Sasazuki, S.; Otani, T.; Ye, W.; Tsugane, S. JPHC Study Group. Body mass index, physical activity and the risk of pancreatic cancer in relation to smoking status and history of diabetes: A large-scale population-based cohort study in Japan—the JPHC study. *Cancer Causes Contr.* **2007**, *18*, 603-612.
- 19. Li, D.; Tang, H.; Hassan, M.M.; Holly, E.A.; Bracci, P.M.; Silverman, D.T. Diabetes and risk of pancreatic cancer: A pooled analysis of three large case-control studies. *Cancer Causes Contr.* **2010**, doi: 10.1007/s10552-010-9686-3.
- 20. Li, D.; Morris, J.S.; Liu, J.; Hassan, M.M.; Day, R.S.; Bondy, M.L.; Abbruzzese, J.L. Body mass index and risk, age of onset, and survival in patients with pancreatic cancer. *JAMA* **2009**, *301*, 2553-2562.
- 21. Li, D.; Yeung, S.C.; Hassan, M.M.; Konopleva, M.; Abbruzzese, J.L. Antidiabetic therapies affect risk of pancreatic cancer. *Gastroenterology* **2009**, *137*, 482-488.
- 22. Bolen, S.; Feldman, L.; Vassy, J.; Wilson, L.; Yeh, H.C.; Marinopoulos, S.; Wiley, C.; Selvin, E.; Wilson, R.; Bass, E.B.; Brancati, F.L. Systematic review: Comparative effectiveness and safety of oral medications for type 2 diabetes mellitus. *Ann. Intern. Med.* **2007**, *147*, 386-399.

23. DeCensi, A.; Puntoni, M.; Goodwin, P.; Cazzaniga, M.; Gennari, A.; Bonanni, B.; Gandini, S. Metformin and cancer risk in diabetic patients: A systemic review and meta-analysis. *Cancer Prev. Res.* **2010**, *3*, 1452-1461.

- 24. Ben Sahra, I.; Le Marchand-Brustel, Y.; Tanti, J.-F.; Bost, F. Metformin in cancer therapy: A new perspective for an old antidiabetic drug? *Mol. Cancer Ther.* **2010**, *9*, 1092-1099.
- 25. Pollak, M. Metformin and other biguanides in oncology: Advancing the research agenda. *Cancer Prev. Res.* **2010**, *3*, 1060-1065.
- 26. Pollak, M. Insulin and insulin-like growth factor signalling in neoplasia. *Nat. Rev. Cancer* **2008**, 8, 915-928.
- 27. Shackelford, D.B.; Shaw, R.J. The LKB1-MAPK pathway: Metabolism and growth control in tumor suppression. *Nat. Rev. Cancer* **2009**, *9*, 563-575.
- 28. Rozengurt, E.; Sinnett-Smith, J.; Kisfalvi, K. Crosstalk between insulin/insulin-like growth factor-1 receptors and G protein-coupled receptor signaling systems: A novel target for the antidiabetic drug metformin in pancreatic cancer. *Clin. Cancer Res.* **2010**, *16*, 2505-2511.
- 29. Kim, J.; Dang, C.V. Cancer's molecular sweet tooth and the Warburg effect. *Cancer Res.* **2006**, 66, 8927-8930.
- 30. Samudio, I.; Fiegl, M.; Andreeff, M. Mitochondrial uncoupling and the Warburg effect: Molecular basis for the reprogramming of cancer cell metabolism. *Cancer Res.* **2009**, *69*, 2163-2166.
- 31. Vander Heiden, M.G.; Cantley, L.C.; Thompson, C.B. Understanding the Warburg effect: The metabolic requirements of cell proliferation. *Science* **2010**, 324, *1029-1033*.
- 32. Kroemer, G.; Pouyssegur, J. Tumor cell metabolism: Cancer's Achilles' heel. *Cancer Cell* **2008**, *13*, 472-482.
- 33. Mankoff, D.A.; Eary, J.F.; Link, J.M.; Muzi, M.; Rajendran, J.G.; Spence, A.M.; Krohn, K.A. Tumor-specific positron emission tomography imaging in patients: [¹⁸F] Fruolodeoxyglucose and beyond. *Clin. Cancer Res.* **2007**, *13*, 3460-3469.
- 34. Dwarakanath, B.S.; Jain, V. Targeting glucose metabolism with 2-deoxy-D-glucose for improving cancer therapy. *Future Oncol.* **2009**, *5*, 581-585.
- 35. Stacpoole, P. W.; Nagaraja, N.V.; Hutson, A.D. Efficacy of dichloroacetate as a lactate-lowering drug. *J. Clin. Pharmacol.* **2003**, *43*, 683-691.
- 36. Stacpoole, P.W.; Kerr, D.S.; Barnes, C.; Bunch, S.T.; Carney, P.R.; Fennell, E.M.; Felitsyn, N.M.; Gilmore, R.L.; Greer, M.; Henderson, G.N.; Hutson, A.D.; Neiberger, R.E.; O'Brien, R.G.; Perkins, L.A.; Quisling, R.G.; Shroads, A.L.; Shuster, J.J.; Silverstein, J.H.; Theriaque, D.W.; Valenstein, E. Controlled clinical trial of dichloroacetate for treatment of congenital lactic acidosis in children. *Pediatrics* **2006**, *117*, 1519-1531.
- 37. Bonnet, S.; Archer, S.L.; Allalunis-Turner, J.; Haromy, A.; Beaulieu, C.; Thompson, R.; Lee, C.T.; Lopaschuk, G.D.; Puttagunta, L.; Bonnet, S.; Harry, G.; Hashimoto, K.; Porter, C.J.; Andrade, M.A.; Thebaud, B.; Michelakis, E.D. A mitochondria-K+ channel axis is suppressed in cancer and its normalization promotes apoptosis and inhibits cancer growth. *Cancer Cell* **2007**, *11*, 37-51.
- 38. Coussens, L.M.; Werb, Z. Inflammation and cancer. *Nature* **2002**, 420, 860-867.

39. McKay, C.J.; Glen, P.; McMillan, D.C. Chronic inflammation and pancreatic cancer. *Best Pract. Res. Clin. Gastroenterol.* **2008**, 22, 65-73.

- 40. Hart, A.R.; Kennedy, H.; Harvey, I. Pancreatic cancer: A review of the evidence on causation. *Clin. Gastroenterol. Hepatol.* **2008**, *6*, 275-282.
- 41. Greer, J.B.; Whitcomb, D.C. Inflammation and pancreatic cancer: An evidence-based review. *Curr. Opin. Pharmacol.* **2009**, *9*, 411-418.
- 42. Raimondi, S.; Lowenfels, A.B.; Morselli-Labate, A.M.; Maisonneuve, P.; Pezzilli, R. Pancreatic cancer in chronic pancreatitis: Aetiology, incidence, and early detection. *Best Practice Res. Clin. Gastroenterol.* **2010**, *24*, 349-358.
- 43. Howes, N.; Lerch, M.M.; Greenhalf, W.; Stocken, D.D.; Ellis, I.; Simon, P.; Truninger, K.; Ammann, R.; Cavallini, G.; Charnley, R.M.; Uomo, G.; Delhaye, M.; Spicak, J.; Drumm, B.; Jansen, J.; Mountford, R.; Whitcomb, D.C.; Neoptolemos, J.P.; European Registry of Hereditary Pancreatitis and Pancreatic Cancer (EUROPAC). Clinical and genetic characteristics of hereditary pancreatitis in Europe. *Clin. Gastroenterol. Hepatol.* **2004**, *2*, 252-261.
- 44. Rebours, V.; Boutron-Ruault, M.C.; Schnee, M.; Férec, C.; Maire, F.; Hammel, P.; Ruszniewski, P.; Lévy, P. Risk of pancreatic adenocarcinoma in patients with hereditary pancreatitis: A national exhaustive series. *Am. J. Gastroenterol.* **2008**, *103*, 111-119.
- 45. Greer, J.B.; Lynch, H.T.; Brand, R.E. Herediatry pancreatic cancer: A clinical perspective. *Best Pract. Res. Clin. Gastroenterol.* **2009**, *23*, 159-170.
- 46. Whitcomb, D.C.; Gorry, M.C.; Preston, R.A.; Furey, W.; Sossenheimer, M.J.; Ulrich, C.D.; Martin, S.P.; Gates, L.K., Jr.; Amann, S.T.; Toskes, P.P.; Liddle, R.; McGrath, K.; Uomo, G.; Post, J.C.; Ehrlich, G.D. Herediatry pancreatitis is caused by a mutation in the cationic trypsinogen gene. *Nat. Genet.* **1996**, *14*, 141-145.
- 47. Farrow, B.; Evers, B.M. Inflammation and the development of pancreatic cancer. *Surg. Oncol.* **2002**, *10*, 153-169.
- 48. Garcea, G.; Dennison, A.R.; Steward, W.P.; Berry, D.P. Role of inflammation in pancreatic carcinogenesis and the implications for future therapy. *Pancreatology* **2005**, *5*, 514-529.
- 49. Uomo, I.; Miraglia, S.; Pastorello, M. Inflammation and pancreatic ductal adenocarcinoma: A potential scenario for novel drug targets. *J. Pancreatol. (JOP)* **2010**, *11*, 199-202.
- 50. Alg ül, H.; Treiber, M.; Lesina, M.; Schmid, R.M. Mechanisms of disease: Chronic inflammation and cancer in the pancreas—a potential role for pancreatic stellate cells? *Nat. Clin. Pract. Gastroenterol. Hepatol.* **2007**, *4*, 454-462.
- 51. Apte, M.; Pirola, R.; Wilson, J. New insights into alcoholic pancreatitis and pancreatic cancer. *J. Gastroenterol. Hepatol.* **2009**, 24, S51-S56.
- 52. Masamune, A.; Shimosegawa, T. Signal transduction in pancreatic stellate cells. *J. Gastroenterol.* **2009**, *44*, 249-260.
- 53. Mahadevan, D.; Von Hoff, D.D. Tumor-stroma interactions in pancreatic ductal adenocarcinoma. *Mol. Cancer Ther.* **2007**, *6*, 1186-1197.
- 54. Joyce, A.C. Therapeutic targeting of the tumor microenvironment. *Cancer Cell* **2005**, 7, 513-520.
- 55. Vonlaufen, A.; Phillips, P.A.; Xu, Z.; Goldstein, D.; Pirola, R.C.; Wilson, J.S.; Apte, M.V. Pancreatic stellate cells and pancreatic cancer cells: An unholy alliance. *Cancer Res.* **2008**, *68*, 7707-7710.

56. Yilmaz, M.; Christofori, G. EMT, the cytoskeleton, and cancer cell invasion. *Cancer Metastasis Rev.* **2009**, 28, 15-33.

- 57. Polyak, K.; Weiberg, R.A. Transition between epithelial and mesenchymal states: Acquisition of malignant and stem cell traits. *Nat. Rev. Cancer* **2009**, *9*, 265-273.
- 58. Giovannetti, E.; Mey, V.; Nannizzi, S.; Pasqualetti, G.; Tacca, M.D.; Danesi, R. Pharmacogenetics of anticancer drug sensitivity in pancreatic cancer. *Mol. Cancer Ther.* **2006**, *5*, 1387-1395.
- 59. Hruban, R.H.; Goggins, M.; Parsons, J.; Kern, S.E. Progression model for pancreatic cancer. *Clin. Cancer Res.* **2000**, *6*, 2969-2972.
- 60. Furukawa, T.; Sunamura, M.; Horii, A. Moecular mechanisms of pancreatic carcinogenesis. *Cancer Sci.* **2006**, *97*, 1-7.
- 61. Giroux, V.; Dagorn, J.C.; Iovanna, J.L. A review of kinases implicated in pancreatic cancer. *Pancreatology* **2009**, *9*, 738-754.
- 62. Moore, M.J.; Goldstein, D.; Hamm, J.; Figer, A.; Hecht, J.R.; Gallinger, S.; Au, H.J.; Murawa, P.; Walde, D.; Wolff, R.A.; Campos, D.; Lim, R.; Ding, K.; Clark, G.; Voskoglou-Nomikos, T.; Ptasynski, M.; Parulekar, W. Erlotinib plus gemcitabine compared with gemcitabine alone in patients with advanced pancreatic cancer: A phase III trial of the National Cancer Institute of Canada Clinical Trials Group. *J. Clin. Oncol.* **2007**, *25*, 1960-1966.
- 63. Furukawa, T. Molecular targeting therapy for pancreatic cancer: Current knowledge and perspectives from bench to bedside. *J. Gastroenterol.* **2008**, *43*, 905-911.
- 64. Giroux, V.; Iovanna, J.; Dagorn, J.-C. Probing the human kinome for kinases involved in pancreatic cancer cell survival and gemcitabine resistance. *FASEB J.* **2006**, *20*, 1982-1991.
- 65. Morgan, M.A.; Parsels, L.A.; Maybaum, J.; Lawrence, T.S. Improving gemcitabine-mediated radiosensitization using molecularly targeted therapy: A review. *Clin. Cancer Res.* **2008**, *14*, 6744-6750.
- 66. Harwood, A.J. Regulation of GSK-3: A cellular multiprocessor. Cell 2001, 105, 821-824.
- 67. Doble, B.W.; Woodgett, J.R. GSK-3: Tricks of the trade for a multi-tasking kinase. *J. Cell. Sci.* **2003**, *116*, 1175-1186.
- 68. Jope, R.S.; Johnson, G.V. The glamour and gloom of glycogen synthase kinase-3. *Trends Biochem. Sci.* **2004**, *29*, 95-102.
- 69. Cohen, P.; Goedert, M. GSK3 inhibitors: Development and therapeutic potential. *Nat. Rev. Drug Discov.* **2004**, *3*, 479-487.
- 70. Meijer, L.; Flajolet, M.; Greengard, P. Pharmacological inhibitors of glycogen synthase kinase 3. *Trends Pharmacol. Sci.* **2004**, *25*, 471-480.
- 71. Jope, R.S.; Yuskaitis, C.J.; Beurel, E. Glycogen synthase kinase-3 (GSK3): Inflammation, diseases, and therapeutics. *Neurochem. Res.* **2007**, *32*, 577-595.
- 72. Manoukian, A.S.; Woodgett, J.R. Role of GSK-3 in cancer: Regulation by Wnts and other signaling pathways. *Adv. Cancer Res.* **2002**, *84*, 203-229.
- 73. Shakoori, A.; Ougolkov, A.; Yu, Z.W.; Zhang, B.; Modarressi, M.H.; Billadeau, D.D.; Mai, M.; Takahashi, Y.; Minamoto, T. Deregulated GSK3β activity in colorectal cancer: Its association with tumor cell survival and proliferation. *Biochem. Biophys. Res. Commun.* **2005**, *334*, 1365-1373.

74. Shakoori, A.; Mai, W.; Miyashita, K.; Yasumoto, K.; Takahashi, Y.; Ooi, A.; Kawakami, K.; Minamoto, T. Inhibition of GSK-3β activity attenuates proliferation of human colon cancer cells in rodents. *Cancer Sci.* **2007**, *98*, 1388-1393.

- 75. Miyashita, K.; Kawakami, K.; Nakada, M.; Mai, W.; Shakoori, A.; Fujisawa, H.; Hayashi, Y.; Hamada, J.; Minamoto, T. Potential therapeutic effect of glycogen synthase kinase 3β inhibition against human glioblastoma. *Clin. Cancer Res.* **2009**, *15*, 887-897.
- 76. Mai, W.; Kawakami, K.; Shakoori, A.; Kyo, S.; Miyashita, K.; Yokoi, K.; Jin, M.; Shimasaki, T.; Motoo, Y.; Minamoto, T. Deregulated GSK3β sustains gastrointestinal cancer cells survival by modulating human telomerase reverse transcriptase and telomerase. *Clin. Cancer Res.* **2009**, *15*, 6810-6819.
- 77. Minamoto, T. Suppression of cancer and method for evaluating anticancer agent based on the effect of inhibiting GSK3β. International patent WO2006/073202. 2006 Jul 13; United States patent US 11/794,716, 4 January 2006; European patent EP1845094, 17 October 2007; Japan patent 2006-550915, 21 June 2007.
- 78. Miyashita, K.; Nakada, M.; Shakoori, A.; Ishigaki, Y.; Shimasaki, T.; Motoo, Y.; Kawakami, K.; Minamoto, T. An emerging strategy for cancer treatment targeting aberrant glycogen synthase kinase 3β. *Anti-Cancer Agents Med. Chem.* **2009**, *9*, 1114-1122.
- 79. Patel, S.; Woodgett, J. Glycogen synthase kinase-3 and cancer: Good cop, bad cop? *Cancer Cell* **2008**, *14*, 351-353.
- 80. Luo, J. Glycogen synthase kinase 3β (GSK3β) in tumorigenesis and cancer chemotherapy. *Cancer Lett.* **2009**, *273*, 194-200.
- 81. Hoeflich, K.P.; Luo, J.; Rubie, E.A.; Tsao, M.S.; Jin, O.; Woodgett, J.R. Requirement for glycogen synthase kinase-3β in cell survival and NF-κB activation. *Nature* **2000**, *406*, 86-90.
- 82. Schwabe, R.F.; Brenner, D.A. Role of glycogen synthase kinase-3 in TNF-α-induced NF-κB activation and apoptosis in hepatocytes. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2002**, 283, G204-G211.
- 83. Ougolkov, A.V.; Fernandez-Zapico, M.E.; Savoy, D.N.; Urrutia, R.A.; Billadeau, D.D. Glycogen synthase kinase-3β participates in nuclear factor κB-mediated gene transcription and cell survival in pancreatic cancer cells. *Cancer Res.* **2005**, *65*, 2076-2081.
- 84. Wilson, W., 3rd.; Baldwin, A.S. Maintenance of constitutive IκB kinase activity by glycogen synthase kinase-3α/β in pancreatic cancer. *Cancer Res.* **2008**, *68*, 8156-8163.
- 85. Mamaghani, S.; Patel, S.; Hedley, D.W. Glycogen synthase kinase-3 inhibition disrupts nuclear factor-κB activity in pancreatic cancer, but fails to sensitize to gemcitabine chemotherapy. *BMC Cancer* **2009**, *9*, 132-132.
- 86. Shimasaki, T.; Ishigaki, Y.; Nakamura, Y.; Takata, T.; Nakaya, N.; Nakajima, H.; Sato, I.; Zhao, X.; Kitano, A.; Kawakami, K.; Tanaka, T.; Takegami, T.; Tomosugi, N.; Minamoto, T.; Motoo, Y. Glycogen synthase kinase 3β inhibition sensitizes pancreatic cancer cells to gemcitabine. *J. Gastroenterol.* **2011**, submitted.
- © 2011 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).