

Review

Nature's Green Potential: Anticancer Properties of Plants of the Euphorbiaceae Family

Víctor Jiménez-González ^{1,*} , Tomasz Kowalczyk ^{2,*} , Janusz Piekarski ³, Janusz Szemraj ⁴ , Patricia Rijo ^{5,6}  and Przemysław Sitarek ⁷ 

¹ Department of Pharmacology, Faculty of Pharmacy, University of Seville, 41012 Seville, Spain

² Department of Molecular Biotechnology and Genetics, Faculty of Biology and Environmental Protection, University of Lodz, Banacha 12/16, 90-237 Lodz, Poland

³ Department of Surgical Oncology, Medical University in Lodz, 93-513 Lodz, Poland; janusz.piekarski@umed.lodz.pl

⁴ Department of Medical Biochemistry, Medical University of Lodz, 92-215 Lodz, Poland; janusz.szemraj@umed.lodz.pl

⁵ CBIOS-Lusófona University's Research Center for Biosciences and Health Technologies, 1749-024 Lisbon, Portugal; p1609@ulusofona.pt

⁶ Instituto de Investigação do Medicamento (iMed.U LISBOA), Faculdade de Farmácia, Universidade de Lisboa, 1649-003 Lisbon, Portugal

⁷ Department of Medical Biology, Medical University of Lodz, 90-151 Lodz, Poland; przemyslaw.sitarek@umed.lodz.pl

* Correspondence: vjimenez3@us.es (V.J.-G.); tomasz.kowalczyk@biol.uni.lodz.pl (T.K.)

Simple Summary: Euphorbiaceae is a large family of flowering plants that includes a wide spectrum of useful plants, from edible plants to toxic and medicinal plants. They are cosmopolitan plants that have very different shapes, from little herbaceous plants to big trees and cactus-like forms. This review article focuses on the potential anticancer activity of extracts, isolated compounds, and nanoparticles generated from the plants of the Euphorbiaceae family based on in vitro and in vivo experiments. Possible mechanisms of action are also discussed.



Citation: Jiménez-González, V.; Kowalczyk, T.; Piekarski, J.; Szemraj, J.; Rijo, P.; Sitarek, P. Nature's Green Potential: Anticancer Properties of Plants of the Euphorbiaceae Family. *Cancers* **2024**, *16*, 114. <https://doi.org/10.3390/cancers16010114>

Academic Editors: David Wong, Anupam Bishayee and Rajeev K. Singla

Received: 20 November 2023

Revised: 17 December 2023

Accepted: 21 December 2023

Published: 25 December 2023

Abstract: The number of cancer cases will reach 24 million in 2040, according to the International Agency for Research on Cancer. Current treatments for cancer are not effective and selective for most patients; for this reason, new anticancer drugs need to be developed and researched enough. There are potentially useful drugs for cancer isolated from plants that are being used in the clinic. Available information about phytochemistry, traditional uses, in vitro and in vivo experiments with plants, and pure compounds isolated from the Euphorbiaceae family indicates that this family of plants has the potential to develop anticancer drugs. This review examines selected species from the Euphorbiaceae family and their bioactive compounds that could have potential against different types of cancer cells. It reviews the activity of crude extracts, isolated compounds, and nanoparticles and the potential underlying mechanisms of action.

Keywords: Euphorbiaceae family; in vitro; in vivo; anticancer; extracts; pure compounds; nanoparticles



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Nowadays, cancer is a serious health problem that represents a great cost to national health systems around the world. Usually cells live, repair errors, divide, and die, and new cells replace the old ones. But sometimes, a mutation in the DNA of a cell that can not repair itself through reparation mechanisms leads to multiple abnormal divisions. These multiple divisions could activate oncogenes (inducing cell growth) or/and deactivate tumor suppressor genes (repressing cell growth), leading to an uncontrolled cell cycle. All these fast divisions, without control, accumulate different mutations that could imply fast

growth and cancer cell death evasion. These mutations could also favor the loss of adhesion of tumor cells, which would facilitate their movement to other areas of the body through epithelial-mesenchymal transition, promoting tumor invasion and metastasis. In addition, it has been established that tumors release some angiogenic cytokines that affect vessel formation, tumor development, invasion, and metastasis. The most important angiogenic cytokines are vascular endothelial growth factor (VEGF) and basic fibroblast growth factor (bFGF), which are also poor markers of the prognostic and aggressiveness of the illness in patients [1,2].

More than 19 million new cases of cancer and 10 million deaths from cancer were reported in 2020 [3]. The most frequently diagnosed cancer was breast cancer (11.7%), lung cancer (11.6%), colorectal cancer (10%), and prostate cancer (7.3%). Furthermore, estimates made by experts expect that in 2040 there will be 24.8 million cases of cancer (47% more than in 2020) [4]. With this future perspective, new treatments and the development of new drugs will be difficult to achieve if governments do not increase the share devoted to cancer, and this lack of financing may be a problem in receiving quality care [5]. New therapies for cancer are usually directed toward developing more effective drugs and trying to avoid the side effects of the actual anticancer drugs. But all these strategies are usually very complex or hard and expensive to achieve, using new molecular targets, nanomedicine, and bioengineering [6]. However, the plant kingdom is still being unexplored; nonetheless, several very useful anticancer drugs are of plant origin, such as Paclitaxel, Vincristine, Vinblastine, Irinotecan, Topotecan, and Etoposide [7–9].

The Euphorbiaceae family of plants is a group of plants that has gained the interest of the scientific community due to their traditional uses in ethnomedicine, their high diversity of compounds, the potential toxicity of these compounds, and their easy access to these cosmopolitan plants [10]. Different plants from the genus of the family, such as *Euphorbia*, *Croton*, *Jatropha*, and *Cnidoscolus*, have been tested in vitro for their anticancer activity with excellent results [11–13]. This gives us an opportunity to continue delving deeper into the compounds present in these plants and their mechanisms of action as potential anticancer drugs. Several secondary metabolites have been described in the Euphorbiaceae family; for example, there is a high diversity of terpenoids with different types of original skeleton configurations [14–16]. Some common flavonoids, like quercetin and apigenin, have been isolated from this plant family [17–20]. Additionally, it has been documented that these plants contain certain tryptamine-derived alkaloids [21].

The main purpose of this review is to bring together all the information available about the anticancer effect in vitro or in vivo of plant extracts or pure compounds isolated from the Euphorbiaceae family published in the last ten years.

2. Criteria for the Selection of Experimental Papers

This review is based on primary literature published between 2012 and 2023 (to date). The papers were selected from different electronic databases: PubMed, Google Scholar, Scopus, and Web of Science. The following terms were used to achieve the search: Euphorbiaceae with: family, anticancer, antiproliferative, cytotoxic, plant extract, pure compound, in vitro, in vivo, and nanoparticles. The articles reporting extracts or pure compounds from the Euphorbiaceae family with some anticancer, antiproliferative, or cytotoxic activity in vitro or in vivo were included. Other articles reporting different types of reviews, articles in different languages than English, articles without full text access, lacking specific plant names, without reports of clear objectives and methodology, published more than ten years ago, using plant species other than Euphorbiaceae, were excluded. Duplicate articles from different database searching results were also excluded. All the inclusion/exclusion criteria were checked again after the removal of these articles. Each selected research paper was examined, and the following data were selected and presented in the tables: scientific plant name, parts of the plants used for extract preparation or pure compound isolation, type of extract, class of compounds or different compounds identified in the extract, cancer cell lines used or animal model/cell line inoculated with

cancer-inducing compounds, activity or mechanism of action, and reference. Articles explaining the mechanisms of action of Euphorbiaceae plant extracts or isolated compounds were discussed before the tables in the main text.

3. The Euphorbiaceae Family of Plants

The Euphorbiaceae family of plants has about 228 plant genera accepted according to Plants of the World Online by Royal Botanic Garden Kew [22] and more than 7000 species of plants according to the Global Biodiversity Information Facility [23]. This family of green plants has a very different overall shape. There are habits from herbaceous plants (for example, *Euphorbia peplus* L.) to shrubs (like *Ricinus communis* L.) or woody trees (*Hevea brasiliensis* (Willd. ex A.Juss.) Müll.Arg.) and cactus-like shapes (*Euphorbia ingens* (E.Mey. ex Boiss.)). These plants can be annuals or perennials, monoics or dioics, and usually present some type of latex. The leaves are opposite (sometimes alternatives) and have stipules that could be transformed into spines or glands. The flowers gather in an inflorescence called cyathium, and sometimes they are inconspicuous flowers that lack a corolla. Sometimes the cyathium contains several masculine flowers and only one feminine flower. Androecium with one or numerous stamens, and gynoecium with two or three styles and carpels. The fruits are usually contained in capsules, and they separate easily from each other [24–26]. Among the best-known genera of this family, we find *Euphorbia*, *Jatropha*, *Croton*, *Acalypha*, *Cnidoscolus*, and *Ricinus* (Figure 1) [22]. Some plants in the family are very important for humans; for example, *Mannihot sculenta* L. is an edible plant that is cultivated in many countries and is considered a basic food. This crop is easy to grow in a variety of environments, is undemanding, and has very good nutritional properties [27,28]. *Mannihot sculenta* L. was grown as a crop on more than one million hectares in countries such as Ghana, Angola, Cote d’Ivoire, Brazil, Thailand, the Democratic Republic of the Congo, and Nigeria in 2021, according to the Food and Agriculture Organization of the United Nations [29]. Other plants are considered toxic to be consumed but still useful; for example, in the proper dosage, *Ricinus communis* L., commonly known as Castor oil, has been used in the past as a purgative [30] and is approved by the US Food and Drug Administration as an antisticking additive agent to produce hard candy [31]. Many other uses of castor’s oil have been reported, including uterine contraction [32], antiviral [33], antibacterial [34], and antiinflammatory [30]. In addition, the oils extracted from the seeds are very useful for industrial applications [31,35].

Plants of the most numerous genera, *Euphorbia*, have been used traditionally around the world for different purposes, for example, to feed animals, as additives in food, as fuels, or for environmental uses, but the two main purposes have been to poison different animals and as folk medicines [10]. These two main purposes take advantage of the toxicological properties of the diverse phytochemistry present in these plants. For example, it is well known that different species of Euphorbiaceae, for example, *Euphorbia tirucalli* L., have been used in Africa to poison fish. This is a very common way of fishing with the help of plant metabolites that spread through water and poison animals [36]. On the other hand, when used as folk medicines, they are known to be used mainly for digestive disorders, skin or subcutaneous tissue disorders, infections, inflammation, or respiratory disorders [10]. For example, for digestive disorders, decoctions of *Euphorbia hirta* L. are recorded to be used in distant places such as Burundi, the Philippines, and China as antidiarrheal [37–39]. *Euphorbia lathyris* L. is recorded as being used as a purgative in different countries in Europe. For skin disorders, *Euphorbia maculata* L. is well known to be used in folk medicine against warts [40]. To treat infections, for example, *Euphorbia hirta* L. had been used in the past for gonorrhoea in Africa [41]. The same plant has been reported to be used in Australia for bronchitis (inflammation) [42]. For respiratory disorders, it is used in Nepal for the treatment of asthma [43].

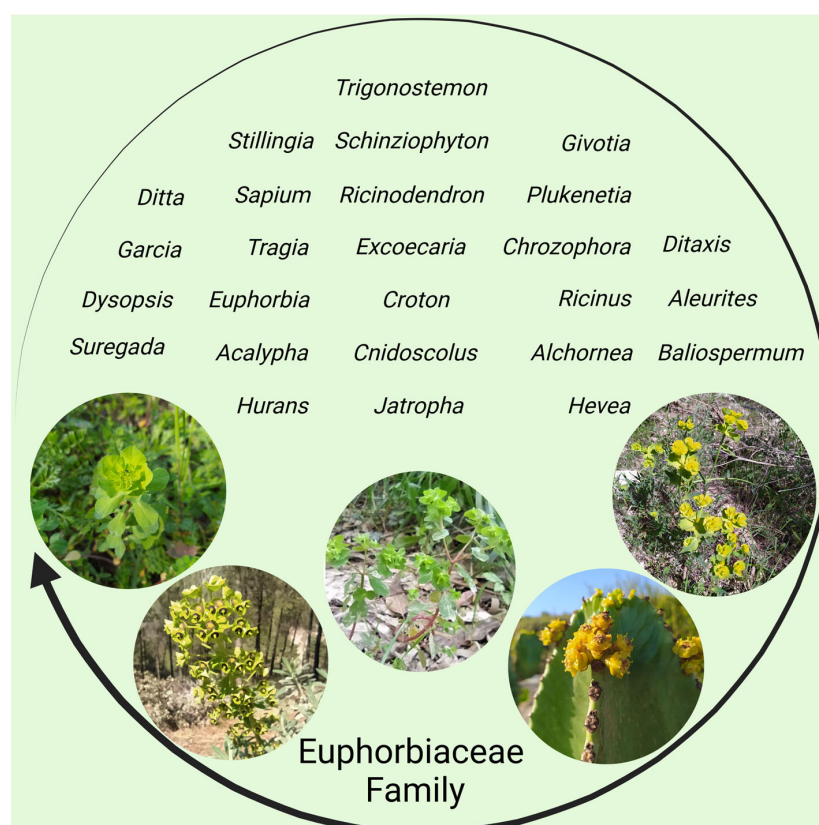


Figure 1. The Euphorbiaceae family of green plants. (created by BioRender).

3.1. Phytochemistry of the Euphorbiaceae Family

This family of plants is very diverse and rich in secondary metabolites. Different compounds have been isolated, for example, terpenoids and flavonoids. The first class of compounds in Euphorbiaceae are terpenoids, classified into diterpenoids, triterpenoids, and sesquiterpenoids. These metabolites have been studied in this family for a long time for their different biological activities. The diterpenoids are cyclized from the precursor geranyl-geranyl-phosphate to the corresponding diterpenoid skeleton. In the Euphorbiaceae family, compounds have been isolated with different types of diterpenoid skeletons [44–46].

For example, in the genus *Euphorbia*, different types of diterpenoids have been described, such as labdane, abietane, atisane, kaurane, isopimarane, rosane, dolabrane, casbane, cembrane, rhamnofolane, gaditanone, ingenane, ingol, jatrophone, jatrophone, lathyrane, cyclomyrsinol, myrsinol, premysinol, paralane, pepluane, presegetane, segetane, tiglane, cyclojatrophone, and epoxyjatrophone [44,45]. Due to the high diversity of plants and metabolites in this genus, every year new diterpenoids are discovered. Zhu et al. [47] using the roots *Euphorbia fischeriana* L. described two new *ent*-abietane diterpenoids, euphonoids H and I. Zhang et al. [48] using the same plant species have reported for the first time the presence of a new *ent*-rosane diterpene, named ebracteolatas D. On the other hand, in the *Croton* genus, labdanes, clerodanes, trachylobane, kaurene, cembrane, and isopimarane diterpenoids have been identified [46]. Wang et al. [49] reported the isolation of a new *ent*-abietane diterpenoid (7b,13a,15-tri-hydroxy-*ent*-abieta-8(14)-en-3-one) from the leaves of *Croton lachnocarpus* Benth. In the genus *Jatropha*, different diterpenoids' skeletons have also been described, such as tiglane, casbene, daphnane, lathyrane, jatrophone, podocarpene, and rhamnofolane [50]. For example, the team led by Yuan et al. [51] isolated jatropodagins A, a new lathyrane-type diterpenoid.

Triterpenoids are also very common and diverse in the family. In the genus *Euphorbia*, different types of skeletons have been described, for example, tirucallane, euphane, lanostane, cycloartanes, lupane, oleanane, taraxarane, friedoursane, friedelane, and ursane [52].

And still, nowadays, new compounds are being discovered. A new cycloartane-type triterpene (23 R/S-3b-hydroxycycloart-24-ene-23-methyl ether) was isolated from the aerial parts of *Euphorbia dendroides* L. [53]. In the genus *Croton*, several triterpenoids have been described [46]. For example, lupeol has been isolated from different parts of the species *Croton sylvaticus* Hochst. and *Croton zambezicus* Müll. Arg. [54,55]. β -Sitosterol has been isolated from *Croton zambezicus* Müll. Arg. and *Croton steenkampianus* Gerstner [15,56]. Betulinic acid, betulin, and lupenone have been isolated from the fruits of *Croton zambezicus* [57]. Jatrogrossidione, a triterpenoid isolated from the branches and leaves of *Jatropha gossypifolia* L., was isolated by Zhan et al. [58].

Also, a few sesquiterpenoids have been reported from *Euphorbia*, *Croton*, *Jatropha*, etc. [46,59,60]. Aryanin, a new sesquiterpene lactone, was isolated from the aerial parts of *Euphorbia microsphaera* Boiss by Azizi et al. [61]. Another sesquiterpene, caryophyllene, was reported to be present in *Croton* species as a volatile constituent [46].

The presence of flavonoids in this family is widespread. These compounds could have an impact on health, for example, as potential anticancer agents that have been previously reported [62]. For example, quercitrin has been isolated from *Euphorbia hirta* L. [63], and apigenin has been isolated from *Croton betulaster* Mull, *Jatropha gossypifolia* L., and *Macaranga gigantifolia* Merr. [17,20,64].

Some alkaloids have been isolated from this family also, from latex, roots, or other parts of the plants. Alkaloids are important chemicals; some of them have been isolated or developed for cancer treatment, such as the alkaloids of vinca [65]. For example, Novello et al. [66] have isolated, from *Croton echinoides* Baill, a new alkaloid (*N-trans*-feruloyl-3,5-dihydroxyindolin-2-one) as a mixture with other already known alkaloids (*N-trans-p*-coumaroyl-tryptamine, *N-trans-p*-coumaroyl-5-hydroxytryptamine, *N-trans*-4-methoxycinnamoyl-5-hydroxytryptamine, and *N-trans*-feruloyl-5-hydroxytryptamine).

3.2. Cytotoxic and Anticancer Effects of Euphorbiaceae Extracts In Vitro Studies

Testing plant extracts is one of the first steps in the search for new compounds with potential biological properties, including an anticancer effect. Many articles confirm the preliminary cytotoxic properties of extracts from many medicinal plant families, which can induce apoptosis in many cancer lines. The Euphorbiaceae family is a source of many interesting compounds with broad medicinal applications. The first step of our analysis was to analyze the effect of plant extracts on cytotoxic effects in in vitro studies. Mesas et al. [67] showed that an ethanolic extract of *Euphorbia lathyris* seeds rich in polyphenols such as esculetin, euphorbetin, gaultherin, and kaempferol-3-rutinoside had an antiproliferative effect against colon cancer cell lines (T84 and HCT-15) and glioblastoma multiforme. The authors demonstrated that the induction of apoptosis is associated with overexpression of caspase-9 (casp-9), caspase-3 (casp-3), and caspase-8 (casp-8) and activation of autophagy. In another study, Sultana et al. [68] showed that an aqueous leaf extract of *Excoecaria agallocha* (L.), where the head compound was bergenin, effectively reduced the proliferation of SiHa cervical cancer cells by inducing autophagy and apoptosis in a coordinated manner, with simultaneous stimulation of mitophagy and cell cycle arrest in the G2/M phase. In contrast, Kwan et al. [69] showed that a methanolic extract of *Euphorbia hirta* exhibited significant inhibition of MCF-7 breast cancer cell survival through induction of apoptosis via a casp-3-independent pathway, activation of caspase-2, caspase-6, casp-8, and casp-9, accumulation of cells in the S and G2/M phases, and DNA fragmentation. Similarly, Mfotie Njoya et al. [70] showed that *Croton gratissimus* leaf extract exhibited cytotoxic effects on various cancer lines (A549, Caco-2, HeLa, MCF-7) and inhibited cancer cell growth through induction of caspase-3 (casp-3)/caspase-7 (casp-7) activation, with the highest induction (1.83-fold change) obtained on HeLa cells. Vargas-Madriz et al. [71] presented that *Cnidioscolus aconitifolius* and *Porophyllum ruderale*, rich in polyphenols, reduced the metabolic activity of human SW480 colon adenocarcinoma cells. In addition, both extracts increased the total number of apoptotic cells and arrested the cell cycle in the G0/G1 phases. Other studies are presented in Table 1.

Table 1. Cytotoxic properties of selected extracts from the Euphorbiaceae family against cancer cells.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Acalypha fruticosa</i> Forssk.	Aerial parts	Methanol	-	MCF-7, HCT-116, and HepG-2	12.2 ± 0.6 µg/mL, 4.81 ± 0.4 µg/mL, and 5.21 ± 0.7 µg/mL, respectively.	Cytotoxic	[72]
<i>Acalypha indica</i> L.	Leaves	Hexane	-	MCF-7	50 µg/mL	Cytotoxic	[73]
<i>Acalypha monostachya</i> Cav.	Aerial parts	Distilled water, absolute methanol, and <i>n</i> -hexane	Phenols, coumarins, lactones, flavonoids, saponins, aromatic compounds, carbohydrates, and carbonyl groups. Methanol and hexane: steroids and terpenoids. Aqueous: alkaloids.	HeLa and MDA-MB-231	-	Cytotoxic	[74]
<i>Baliospermum montanum</i> (Willd.) Müll.Arg.	Leaves	Methanol	-	Jurkat	298 µg/mL	Cytotoxic	[75]
<i>Baliospermum montanum</i> (Willd.) Müll.Arg.	Roots	Ethanol	Propiophenones	HepG2 and KKU M156	HepG2 (0.06 ± 0.02 µg/mL) and KKU M157 (0.16 ± 0.02 µg/mL)	Cytotoxic	[76]
<i>Blumeodendron toxbrai</i> (Blume.)	Stem Bark	Hexane, Dichloromethane, and Methanolic	-	MCF-7	Hexane extract 121.24 ± 0.15 µg/mL, Dichloromethane extract 55 ± 0.48 µg/mL, and methanolic extract 70.71 ± 0.15 µg/mL	Cytotoxic	[77]
<i>Croton sylvaticus</i> Hochst.	Leaves	Acetone and ethanol	-	A549, Caco-2, HeLa, and MCF-7	Acetone extract: A549 (32.78 ± 2.55 µg/mL), Caco-2 (150.63 ± 8.79 µg/mL), HeLa (169.09 ± 13.0 µg/mL), MCF-7 (13.13 ± 2.76 µg/mL). Ethanol extract: A549 (1.75 ± 0.62 µg/mL), Caco-2 (103.73 ± 1.47 µg/mL), HeLa (106.52 ± 4.50 µg/mL), MCF-7 (6.02 ± 1.60 µg/mL).	Cytotoxic. casp-3/casp7 pathway.	[70]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Chrozophora oblongifolia</i> (Delile) Spreng.	Root Bark	Aqueous methanol, fractions with <i>n</i> -hexane, methylene chloride, and ethyl acetate	Carbohydrates and/or glycosides, sterols and/or triterpenes, tannins, alkaloids, and saponins	MCF-7 and Huh-7	Cytotoxic activity (%) of the methanolic extracts MCF-7 (15.82 ± 0.66) and Huh-7 (31.28 ± 0.68). <i>n</i> -hexane MCF-7 (59.55 ± 0.76) and Huh-7 (54.17 ± 0.46). Methylene chloride, MCF-7 (83.83 ± 0.37) and Huh-7 (82.79 ± 0.55). Ethyl acetate MCF-7 (22.25 ± 0.13) and Huh-7 (72.52 ± 0.23).	Cytotoxic	[78]
<i>Chrozophora plicata</i> (Vahl) A.Juss. ex Spreng.	Leaves	Petroleum ether, chloroform, hexane, ethyl acetate, methanol, and water	flavonoids, alkaloids, glycosides, and lignans.	DAL	25–50 µg/mL	Cytotoxic	[79]
<i>Cnidoscopus aconitifolius</i> (Mill.) I.M. Johnst.	Leaves	Methanol	Saponins, tannins, terpenes, and flavonoids	MCF-7 and NCI-H460	4.50 ± 0.58 µg/mL and 3.29 ± 4.57 µg/mL, respectively.	Cytotoxic	[80]
<i>Cnidoscopus chayamansa</i> McVaugh.	Leaves	Ethanolic	-	HT-29	CTC50 (µg/mL) ≥ 1000 ± 0.00	Cytotoxic	[81]
<i>Cnidoscopus multilobus</i> (Pax) I.M. Johnst	Leaves	Ethanol-water (70:30)		HeLa	130–160 µg/mL	Cytotoxic	[82]
<i>Cnidoscopus quercifolius</i> Pohl	Bark root	Methanol, chloroform fraction	Faveline, Faveline methyl ether, Deoxofaveline, and Neofavelanone	OVCAR-8, HCT-116, and HL-60	OVCAR-8 15.23 ± 2.0 µg/mL, HCT-116 7.07 ± 0.59 µg/mL, and HL-60 4.95 ± 0.19 µg/mL	Cytotoxic	[83]
<i>Croton acutifolius</i> Esser	Twigs and leaves	Hexane, ethyl acetate, and methanol	Retusin	KKU-M213, FaDu, HT-29, MDA-M, B-231, SH-SY5Y, A 549 and MMNK-2	Ethyl acetate extract against KKU-M213, MDA-MB-231, A-549, and MMNK-1 with the ED50 at 3.31 µg/mL, 0.58 µg/mL, 0.58 µg/mL, and 0.65 µg/mL, respectively.	Cytotoxic	[84]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Croton bonplandianus</i> Baill.	Leaves	Acetone	-	A549	15.68 ± 0.006 µg/mL	Cytotoxic, apoptosis, and G2M phase arrest	[85]
<i>Croton caudatus</i> Geiseler	Leaves	Chloroform, ethanol and aqueous	-	HeLa	80 µg/mL	Cytotoxic, increased DNA damage	[86]
<i>Croton caudatus</i> Geiseler	Leaves	Chloroform, ethanol, and aqueous	Alkaloids, saponins, tannins, and cardiac glycosides.	Dalton's lymphoma (DL)	28.36 µg/mL	Cytotoxic	[87]
<i>Croton caudatus</i> Geiseler	Leaves	Methanol		HeLa	59.70 µg/mL	Cytotoxic	[88]
<i>Croton fluviatilis</i> Esser	Stems	Hexane, ethyl acetate, and methanol	β-amyrin (1), stigmasterol, and β-sitosterol	KKU-M213, FaDu, HT-29, MDA-M, B-231, SH-SY5Y, A-549, and MMNK-1	Hexane extract cytotoxicity against KKU-M213, MDA-MB-231, and A-549 at the ED50 values of 1.70 µg/mL, 2.62 µg/mL, and 0.60 µg/mL, respectively	Cytotoxic	[84]
<i>Croton heliotropiifolius</i> Kunth	Leaves	Methanol	Gallic acid	NCI-H292, MCF-7, Hep-2, and HL-60	% inhibitor activity in NCI-H292 (46.5 ± 2.6), MCF-7 (21.7 ± 3.7), Hep-2 (26.7 ± 7.1), and HL-60 (59.5 ± 2.9)	Inhibitory activity	[89,90]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Croton membranaceus</i> Müll. Arg.	Roots	Hydroethanolic	5-Hydroxypipelicolic acid, Phenol, 3, 5-bis(1, 1-dimethylethyl)-, 2-Octenoic acid, 5, 5, 7-trihydroxy, 9, 10-Secocholeasta-5,7,10(19)-tiene-1,3-diol, 25-[(trimethylsilyl)oxy]-, Phenol, 4-(3-hydroxy-1-propenyl)-2-methoxy, <i>n</i> -Hexadecanoic acid, Benzene, 1,2,4,5-tetrakis(1-methylethyl)-, Prednisolone acetate, 1H, 4H-Pyrazolo[3,4-b]pyran-5-carbonitrile, 6-amino-4-(2, 4, 5-trimethoxyphenyl)-3-methyl and Astaxanthin	22Rv1	3.809 µg/mL	Cytotoxic, inhibit colony-forming and migration abilities	[91]
<i>Croton sphaerogynus</i> Baill.	Leaves	Hexane, dichloromethane, and methanol	Abieta-8,11-diene-3-one, Podocarp-7-ene,13-methyl-13-vinyl-3-one, Abieta-8,11,13-trien12-ol, Podocarp-7-ene-3-ol, 13-methyl-13-vinyl, 13-Hydroxy-abieta8,11-dien-7-one, and Crotonin derivative	786-0, HT-29, K562, NCI-ADR/RES, NCI-H460, MCF-7, PC-3, OVCAR-3, U251, and UACC-62.	Hexane and dichloromethane against NCI-H460 (GI50 0.26 µg/mL and 0.33 µg/mL, respectively) and K562 (GI50 0.60 µg/mL and <0.25 µg/mL, respectively).	Antiproliferative	[92]
<i>Croton thorelii</i> Gagnep.	Stems	Hexane, ethyl acetate, and methanol	5-hydroxy-7,4'-dimethoxyflavone	KKU-M213, FaDu, HT-29, MDA-M, B-231, SH-SY5Y, A-549, and MMNK-3	KKU-M213, MDA-MB-231, and A-549 had ED50 values of 0.55 µg/mL, 0.72 µg/mL, and 1.75 µg/mL, respectively.	Cytotoxic	[84]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Croton tiglium</i> L.	Seeds	Ethylether and methanol	Isoguanosine, 12-O-Acetylphorbol-13-tigliate, and 13-O-Acetylphorbol-20-linoleate	A549	-	Apoptosis via apoptosis regulator or bcl-2-like protein 4/B-cell lymphoma 2 protein (Bax/Bcl-2) Pathways	[13]
<i>Croton urucurana</i> Baill	-	Hydroalcoholic	-	U937 and THP1	402.2 µg/mL and 360.3 µg/mL, respectively.	Cytotoxic and Apoptosis	[93]
<i>Drypetes sepiaria</i> (Wight & Arn.) Pax & K.Hoffm.	Leaves	Methanol	Phenolics and flavonoids	SiHa	10 µg/mL	Cytotoxic, apoptosis, or necrosis. casp-3 activation.	[94]
<i>Euphorbia cactus</i> Ehrenb. ex Boiss.		Methanol	Phenols, flavonoids, diterpenes, sesquiterpenoids, terpenoids, anthocyanins, tannins, steroids, cerebrosides, anthraquinones, phloracetophenones, glycerols, alkaloids, and carbohydrates	A549, LoVo, and MCF-7	A549 (20.1 ± 0.5 µg/mL), LoVo (53.2 ± 0.4 µg/mL), and MCF-7 (58.80 ± 1.83 µg/mL)	Cytotoxic, in A549, G2M cell cycle arrest. changes in the level of gene expression of Bax, Bcl-2, and casp-3	[95]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/ Mechanism/ Effect	Ref.
<i>Euphorbia caducifolia</i> Haines	Aerial parts	Ethanol and fractions: aqueous, ethyl acetate, and petroleum ether.	Friedooleanane-3- ol, (3 α)-, 1,2-Benzendicarboxylic acid, mono (2- ethylhexyl) ester, Docosanoic acid, methyl ester; methyl behenate, Hexadecanoic acid methyl ester, methyl palmitate, 9, 12-Octadecadienoic Acid (Z,Z)-, methyl ester; methyl Linoleate (Z,Z)- isomer 9-Octadecenoic acid (z)-, methyl ester; methyl oleate	MCF-7, NCI-H460, PC-3, and HeLa	MCF-7 (61 $\pm$ 1.0 μ g/mL), NCI-H460 (19 $\pm$ 6.3 μ g/mL), PC-3 (135 $\pm$ 3 μ g/mL), and HeLa (80 $\pm$ 2 μ g/mL)	Cytotoxic	[96]
<i>Euphorbia davidii</i> Subils	Whole plant	<i>n</i> -Hexane and chloroform	kaempferol 3-O-rhamnoside, myricetin 3-O-rhamnoside, and quercetin 3-O-rhamnoside.	HeLa, MCF7, A2781, and A431	GI %: <i>n</i> -Hexane extract against HeLa (22.44 $\pm$ 2.66), MCF7 (45.88 $\pm$ 1.58), A2780 (26.72 $\pm$ 0.91) and A431 (25.65 $\pm$ 2.99) and chloroform extract against HeLa (22.28 $\pm$ 2.74), MCF7 (52.63 $\pm$ 0.88), A2781 (47.10 $\pm$ 0.68) and A431 (21.64 $\pm$ 0.37)	Cytotoxic	[97]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Euphorbia dendroides</i> L	Aerial parts	Ethanol	Ellagic acid, pyrogallol, e-vanillic acid, benzoic acid, catechin, epi-catechin, alpha-coumaric acid, and salicylic acid	HepG2, HCT116 and MCF	HepG2 (9.5 mg/mL), HCT-116 (13.6 mg/mL), and MCF-7 (20.9 mg/mL).	Antiproliferative	[98]
<i>Euphorbia graminea</i> Jacq.	Leaves	Aqueous Methanol, fractions with <i>n</i> -hexane, ethyl acetate, chloroform, and water	Tannins, flavonoids, saponins, cardiac glycosides, and terpenes	MCF7 and NCI-H460	At 250 µg/mL, the extract recorded −3.1 and +75% cytotoxic and growth inhibitory effects on MCF-7 and NCI-H460 cell lines, respectively. While the cytotoxic effect became more pronounced on MCF-7 cell lines as −2.19 and −9.6% cytotoxicities were recorded by the chloroform fraction at 75 and 100 µg/mL, the ethyl acetate fraction recorded +58.72 and +89.33% inhibitory effects at similar concentrations.	Cytotoxic	[99]
<i>Euphorbia grandicornis</i> Blanc	Aerial parts	Dichloromethane	Methyl 2,5-dihydroxybenzoate, <i>n</i> -octylbenzoate, Friedelanol, Germanicol, β-glutininol, β-amyrin, Stigmasterol, β-Sitosterol, (24R)-tirucalla-8,25-diene-3β, 24-diol, Euphorbol, Hexyl (E)-3-(4-hydroxy-3-methoxyphenyl)-2 propenoate	HeLa	83.84 ± 2.94 µg/mL	-	[100]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Euphorbia grandicornis</i> Blanc	Roots and aerial parts	Dichloromethane	β -glutinin, β -amyrin, 24-methylenetirucalla-8-en-3 β -ol, (–)-tirucalla-8, 25-diene-3 β -24R-diol, Stigmasterol, Sitosterol, Hexyl (E)-3-(4-hydroxy-3-methoxyphenyl)-2-propenoate	MCF-7 and HCC70	For roots, extract MCF-7 ($0.83 \pm 1.14 \mu\text{g/mL}$) and HCC70 ($0.83 \pm 1.14 \mu\text{g/mL}$), and for aerial parts, extract MCF-7 ($1.03 \pm 1.15 \mu\text{g/mL}$), and HCC70 ($0.31 \pm 1.06 \mu\text{g/mL}$)	Cytotoxic	[101]
<i>Euphorbia grantii</i> Oliv.	Aerial parts	Methanol and dichloromethane fraction	Friedelin, 3-b-friedelinol, epifriedelanol, euphol, cycloartenol, cycloartenyl acetate, epifriedelinyl acetate, and euphylbenzoate	MCF-7 and MCF-7ADR	Methanol extracts MCF-7 ($16.47 \mu\text{g/mL}$) and MCF-7ADR ($19.55 \mu\text{g/mL}$) Dichloromethane fraction, MCF-7 ($10.31 \mu\text{g/mL}$) and MCF-7ADR ($10.41 \mu\text{g/mL}$)	Cytotoxic	[102]
<i>Euphorbia granulata</i> Forssk.	Whole plant	Methanol	t-cinnamic acid, p-hydroxybenzoic acid, vanillic acid, 3,4-dihydroxybenzoic acid, syringic acid, p-coumaric acid, gallic acid, ferulic acid, caffeic acid, and sinapic acid	MCF-7, A2780 and HT-29	MCF-7 ($16.23 \pm 4.50 \mu\text{g/mL}$), A2780 ($22.80 \pm 1.55 \mu\text{g/mL}$), and HT-29 ($41.89 \pm 0.07 \mu\text{g/mL}$)	Cytotoxic	[103]
<i>Euphorbia helioscopia</i> L.	Powdered	Hexane, acetone, methanol, and water		DLD-1	$140.83 \pm 0.31 \mu\text{g/mL}$	Cytotoxic	[104],
<i>Euphorbia helioscopia</i> L.	Whole plant	Ethanol	-	Hep-2, T-47D, HT-29, and PC-3	%GI Hep-2 (27), T-47D (7), HT-29 (0), and PC-3 (11)	Cytotoxic	[12]
<i>Euphorbia heterophylla</i> Desf.	-	Methanol		HepG2	-	cell cycle arrest	[105]
<i>Euphorbia hierosolymitana</i> Boiss.	Whole plant	Methanol		MCF-7, HepG-2, HCT-116, PC-3	HCT-116 (4.22 ppm)	Cytotoxic. Changed the cell cycle and affected the gene expression of her2, Bax, and Bcl-2.	[106]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Euphorbia hierosolymitana</i> Boiss.	Aerial parts	Ethyl acetate and <i>n</i> -butanol fraction	2-Myristynoyl acid pantethene, Palmitic acid, methyl ester, Pyrrolidine,1-bicyclo 3,2,1Oct-2-En-3-Yl, Desulphosinigrin	MCF-7, PC3, A549 and Caco-2.	MCF-7 (93 µg/mL), PC3 (100 µg/mL), A549 (103 µg/mL), and Caco-2 (170 µg/mL). For the <i>n</i> -butanol fraction, for MCF-7, PC3, and A549 (>500 µg/mL), and for Caco-2 (152 µg/mL).	Cytostatic	[107]
<i>Euphorbia hirta</i> L.	Leaves	Ethanol	Anthroquinone, terpenoids, alkaloids, phenolic compounds, tannins, flavonoids, steroids, coumarins, and saponins	DLA and EAC	DLA (560.83 µg/mL) and EAC (384.7 µg/mL)	Cytotoxic	[108]
<i>Euphorbia hirta</i> L.	Roots	Ethanolic	-	MCF-7	From 10 µg/mL (61.57 ± 0.16 µg/mL) to 100 µg/mL (48.08 ± 0.30 µg/mL)	Cytotoxic	[109]
<i>Euphorbia hirta</i> L.	Whole plant	Ethanol	9,12,15-Octadecatrien-1-ol, Pentadecylic acid, Ethyl palmitate, Methyl linoleate, 5-Hydroxymethyl-2-furancarboxaldehyde, Ethyl linoleolate	HL-60	50–100 µg/mL	Anticancer	[110]
<i>Euphorbia hirta</i> L.	Whole plant	Petroleum Ether and Chloroform	-	HepG2	Petroleum ether 200 µg/mL and Chloroform 150 µg/mL	Cytotoxic	[111]
<i>Euphorbia hirta</i> L.	Whole plant	Methanol and distilled water	-	HCT-15	510.66 µg/mL	Cytotoxic	[112]
<i>Euphorbia hyssopifolia</i> L.	Aerial parts	Ethanol	Mono and sesquiterpenes, triterpenes, steroids, flavonoids, cinnamic derivatives, hydrolysable tannins, and saponins	HepG2	-	Cytotoxic and genotoxic	[113]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/ Mechanism/ Effect	Ref.
<i>Euphorbia inarticulata</i> Schweinf.	-	Ethanolic	Catechol, syringic acid, cinnamic acid, caffeic acid, gallic acid, ellagic acid, and benzoic acid	Huh-7 and HeLa	Huh-7 104.52 ± 2.74 µg/mL and HeLa 145.11 ± 6.21 µg/mL	Cytotoxic	[114]
<i>Euphorbia ingens</i> E.Mey. ex Boiss.	Root	Dichloromethane:methanol (36.25%) and ethyl acetate fraction	Mayor constituents: 6-pentylidene-4,5-secoandrostane-4,17 beta-diol, 2-bornanol, 1-octadecene, 1-tridecene, and 1-dodecene	DU-145	9.71 ± 0.40 µg/mL	Cytotoxic, Regulation of the Phospho-inositide 3-kinases/Protein kinase B (PI3K/ Akt), MAPK, and tumor protein p53 (p53) signalling pathways	[115]
<i>Euphorbia lactea</i> Haw.	Stems	Hydroalcoholic	-	HN22	250–500 µg/mL	Cytotoxic and anti-migratory activity	[116]
<i>Euphorbia macroclada</i> Boiss.	Leaves, flower and body	Acetone	-	MCF-7 and L-929	Leaves (8.91 ± 0.10 µg/mL)	Cytotoxic	[117]
<i>Euphorbia milii</i> Des Moul.	Aerial parts	Methanol	-	HepG-2	HepG-2 (87.1 ± 9.4 µg/mL)	Cytotoxic	[118]
<i>Euphorbia nivulia</i> Buch.-Ham	Aerial parts	Aqueous ethanol	Phenols, flavonoids, terpenoids, glycosides, alkaloids, saponins, steroids, and tannins	HeLa cells	-	Cytotoxic	[119]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/ Mechanism/ Effect	Ref.
<i>Euphorbia paralias</i> L.	Aerial parts	Methanol, fractions with dichloromethane, and ethyl acetate	Gallic acid, ellagic acid, Kaempferol-3-O-(600-O-galloyl-b-D-glucopyranoside), Quercetin-3-O-b-D-glucopyranoside, and Quercetin-3-O-b-D-arabinopyranoside	HepG2	HepG2 (26.4 ± 1.2 mg/mL)	Cytotoxic	[120]
<i>Euphorbia platyphyllos</i> L.	Whole plant	Diethyl ether, petroleum ether, ethyl acetate, methanol, water infusion, and water decoction	-	MCF-7	>300 µg/mL, 98.07 ± 0.58 µg/mL, 46.24 ± 0.57 µg/mL, 97.16 ± 0.51 µg/mL, 38.29 ± 0.57 µg/mL and 27.79 ± 0.58 µg/mL respectively.	Cytotoxic, Apoptosis	[121]
<i>Euphorbia pulcherrima</i> Willd. ex Klotzsch	-	Methanol		HepG2	-	cell cycle arrest	[105]
<i>Euphorbia rigida</i> M.Bieb.	Aerial parts	Methanol	-	Hep3B and HepG2	-	Cytotoxic	[122]
<i>Euphorbia royleana</i> Boiss.	Aerial parts	Methanol	-	HepG-2, HCT-116 and MCF-7	HepG-2 (0.42 ± 0.7), and HCT-116 (285.1 ± 19.2)	Cytotoxic	[118]
<i>Euphorbia tirucalli</i> L.	Aerial parts	Ethanol and fractions with ethanol, hexane, dichloromethane, ethyl acetate, and aqueous fractions AGS	Aqueous and ethyl acetate fractions (ellagic acid, 1-o-Galloyl-β-d-glucose, sucrose or isomers, Quercitrin, 2,3-hexahydroxydiphenoyl-d-glucose or isomers, rutin, corilagin or isomers, and pedunculagin/casuariin)	AGS	AGS fractions: ethanol (11.73 ± 0.31 µg/mL), hexane (10.33 ± 2.01 µg/mL), dichloromethane (85.00 ± 9.5 µg/mL), ethyl acetate (120.9 ± 2.21 µg/mL), and aqueous (13.08 ± 0.99 µg/mL)	Cytotoxic	[123]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/ Mechanism/ Effect	Ref.
<i>Euphorbia tirucalli</i> L.	Aerial parts	Hexanic and hydroalcoholic	-	HCT-116	25.26 ± 0.18 µg/mL	Cytotoxic, Increase in the expression of casp-3 and p53	[124]
<i>Euphorbia triaculeata</i> Forssk.	Whole plant	Methanol	-	MCF-7, PC-3, HEPG2 and MCF-10A	0–50 µg/mL	Cytotoxicity and genotoxicity	[125]
<i>Euphorbia turcomanica</i> Boiss.	Aerial parts	Heptane, ethyl acetate, dichloromethane, acetone, methanol, and methanol-water (70–30)	Flavonoid, alkaloid, anthraquinone, and tannin.	Hela and HT-29	Methanol-water (50 µg/mL), acetone (90 µg/mL), dichloromethane (230 µg/mL), methanol (420 µg/mL), and heptane (450 µg/mL) in HeLa cells. Methanol-water (43 µg/mL), acetone (115 µg/mL), dichloromethane (125 µg/mL), methanol (250 µg/mL), and heptane (390 µg/mL) in HT-29 cells.	Cytotoxic	[126]
<i>Euphorbia umbellata</i> (Pax) Bruyns	Fresh stems and leaves	Methanol	-	U-251, MCF-7, 786-0, NCI-H460	Stems extract (GI50 between 8.1 and 30.3 mg/mL) Leaf extract (GI50 between 35.6 and >250 mg/mL). Acetone fraction (GI50, between 0.37 and 2.9 mg/mL)	Antiproliferative	[127]
<i>Euphorbia umbellata</i> (Pax) Bruyns	Bark	Ethanol:water (70:30), chloroform fraction	Euphol, sitosterol, lanosterol, lupeol, cycloartenol, friedelin-3b-ol, friedelin	Jurkat clone E6-1	29.00 ± 1.49 µg/mL, 10.06 ± 1.48 µg/mL and 4.83 ± 2.25 µg/mL for 24, 48 and 72 h	Cytotoxic, apoptosis and cell cycle arrest	[128]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/ Mechanism/ Effect	Ref.
<i>Euphorbia umbellata</i> (Pax) Bruyns	Green branches and leaves	Water, methanol, ethanol, and chloroform	Flavonoids, phenols and tannins, terpenoids, steroids, carbohdraates, alkaloids and saponins.	A549	Water (90.15 ± 2.50 µg/mL), ethanol (125.27 ± 2.00 µg/mL), chloroform (197.66 ± 2.40 µg/mL), and methanol (4.30 ± 0.44 µg/mL)	Cytotoxic	[129]
<i>Excoecaria agallocha</i> L.	Leaves	Methanol and Chloroform	-	Hep 2	Methanol extract range between 125–200 µg/mL and chloroform extract, 15–30 µg/mL	Cytotoxic	[130]
<i>Excoecaria agallocha</i> L.	Leaves	Methanol	-	M14, SKMEL5, SKMEL2, SKMEL28, MALME3M, UACC62, UACC257, U251, SNB19, MDA-MB-231, MCF-7, T47D, BT549, Ovcар3, Ovcар5, HCC2998, Colo205, HCT15, KM12, HeLa, SIHA and C33A	M14 (47.5 ± 4.12 µg/mL), SKMEL5 (89.2 ± 4.62 µg/mL), SKMEL2 (30.0 ± 2.12 µg/mL), SKMEL28 (29.8 ± 2.46 µg/mL), MALME3M (35.0 ± 3.12 µg/mL), UACC62 (34.0 ± 2.90 µg/mL), UACC257 (26.0 ± 3.12 µg/mL), U251 (19.44 ± 1.14 µg/mL), SNB19 (19.84 ± 1.60 µg/mL), MDA-MB-231 (15.56 ± 1.14 µg/mL), MCF-7 (20.24 ± 1.16 µg/mL), T47D (19.20 ± 0.15 µg/mL), BT549 (63.42 ± 5.12 µg/mL), Ovcар3 (39.44 ± 1.14 µg/mL), Ovcар5 (19.84 ± 0.60 µg/mL), HCC2998 (20.28 ± 0.56 µg/mL), Colo205 (12.0 ± 2.12 µg/mL), HCT15 (39.0 ± 3.82 µg/mL), KM12 (20.0 ± 1.70 µg/mL), HeLa (18.60 ± 2.20 µg/mL), SIHA (23.42 ± 1.90 µg/mL) and C33A (39.9 ± 3.10 µg/mL)	Cytotoxic	[131]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Flueggea leucopyrus</i> Willd.	Leaves and bark	Ethyl acetate and methanol.	Bergenin and bergenin isomers	A2780	Methanol extract of bark 12.58 ± 1.02 µg/mL, Ethyl acetate extract of leaves 36.35 ± 0.17 µg/mL	Cytotoxic and antiproliferative effect	[132]
<i>Hura crepitans</i> L.	Leaves	Methanol and <i>n</i> -butanol	-	HepG2	-	Cytotoxic	[133]
<i>Jatropha curcas</i> L.	Leaves	Methanol	Hexadecanoic acid, hexadecanoic acid methyl ester, anethole, estragol, oleic acid, phytol, and carvacrol, octadecanoic acid methyl ester, and thymol	HepG2	47.2 ± 2.48 µg/mL	Cytotoxic	[11]
<i>Jatropha curcas</i> L.	Leaves	Ethanol	-	T-47D, SiHa, and OVCAR-7	%GI T-47D (0), SiHa (47), and OVCAR-7 (30)	Cytotoxic	[12]
<i>Jatropha dioica</i> Sesse ex Cerv.	Roots	Aqueous	Alkaloids, flavonoids, saponins, phenols, tannins, and carbohydrates		-		
<i>Jatropha gossypifolia</i> L.	Leaves	Methanol	-	HepG2	15.3 ± 0.95 µg/mL	Cytotoxic	[11]
<i>Jatropha multifida</i> L.	Leaves	Methanol	-	HepG2	29.6 ± 1.27 µg/mL	Cytotoxic	[11]
<i>Jatropha podagrica</i> Hook.	Leaves	Methanol and distilled water (80:20)	-	A549 and PC12	GI > 80% at 100 µg/mL	Antiproliferative	[11]
<i>Jatropha zeyheri</i> Sond.	Roots	Ethyl acetate	Mayor contents: Hexadecanoic acid, Octadecanoic acid, (Z)-9-Octadecenamide, 11- <i>n</i> -Decylheneicosane, Octacosane, 9-Hexacosene, Ethyl vallesiachotamate, Cyclooctacosane, Cyclotetracosane, and Tricosane	Caco-2, A547 and MCF-7	8.83 ± 0.00 µg/mL, 224.48 ± 0.01 µg/mL, and 102.88 ± 2.17 µg/mL respectively	Cytotoxic	[134]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Mallotus cumingii</i> Müll.Arg.	Leaves	Methanol, fractions with hexane and ethyl acetate	Phenolic compounds, flavonoids, terpenoids, cardiac glycosides and saponins.	HCT-116	Methanol (31.51 µg/mL), hexane fraction (17.49 µg/mL) and ethyl acetate fraction (7.75 µg/mL)	Cytotoxic	[135]
<i>Mallotus philippensis</i> (Lam.) Müll.Arg.	Leaves	Methanol	alkaloids, flavonoids, tannins, diterpenes, steroids, and phenolic compounds	MCF-7	190 g/mL	Cytotoxic and apoptosis through the intrinsic pathway	[136]
<i>Manihot esculenta</i> Crantz	Aerial parts	Ether and chloroform fraction	-	A-549	Inhibition ratio % (69.71 ± 1.18) at 50 µg/mL	Cytotoxic	[137]
<i>Mercurialis annua</i> L.	Aerial parts	Ethanolic	Kaempferol, Isorhamnetin, Quercetin, and Rutin	K562, MCF-7, HeLa, and A562	%GI 100 µM K562 (28.52 ± 0.57), MCF-7 (20.74 ± 6.96), HeLa (14.44 ± 2.16), and A562 (10.33 ± 2.75)	Antiproliferative	[138]
<i>Plukenetia volubilis</i> L.	Leaves	Methanol, ethanol, chloroform, hexane, and water	terpenoids, saponins, and flavonoids	HeLa, and A549	Inhibition of 40–50% rate.	Antiproliferative effect	[139]
<i>Ricinus communis</i> L.	Leaves	Aqueous	-	A375	48 µg/mL.	Cytotoxic	[140]
<i>Ricinus communis</i> L.	Stems and seeds	Ethanol	-	A549, HT-29, SW-20, SiHa, Hep-2, T-47D, OVCAR-5 and PC-3	Seed extract activity was 41%, 11%, 12%, and 14% against A549, OVCAR-5, PC-5 respectively. Stem extract activity was 9%, 31% and 40% activity against Hep-2, HT-29, and SiHa cell lines (100 µg/mL).	Cytotoxic	[141]

Table 1. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Cell Lines	IC ₅₀	Activity/Mechanism/Effect	Ref.
<i>Ricinus communis</i> L.	Stems	Ethanol	-	Hep-2, HT-29, SiHa, and OVCAR-6	%GI Hep-2 (9), HT-29 (31), SiHa (47), and OVCAR-6 (30).	Cytotoxic	[12]
<i>Ricinus communis</i> L.	Seeds	Ethanol	-	502713, A549, OVCAR-5, and PC-6	%GI 502713 (41), A549 (11), OVCAR-5 (12) and PC-5 (14).	Cytotoxic	[12]
<i>Ricinus communis</i> L.	Different part of the seeds (testa, tegmen, embryo, endosperm, etc)	Methanol	Phorbol esters	THP-1	testa extract in THP-1 (109.9 µg/mL)	Cytotoxic	[142]
<i>Schinziophyton rautanenii</i> (Schinz) Radcl.-Sm.	Root and bark	Aqueous and methanolic	Alkaloids, flavonoids, anthraquinones, coumarins and triterpenes	TK10, MCF-7, and UACC-62	Aqueous (100–150 µg/mL) and methanolic (70–120 µg/mL) for both cell lines.	Cytotoxic	[143]
<i>Tragia involucrata</i> L.	Whole plant	Ethanol	Clionasterol, Squalene, 2-Ethylhexyl phthalate, Phytol, neophytadiene, ethyl palmitate, ethyl linolate, linolenic acid, viminalol, etc	YAC-1	-	Cytotoxic	[144]

Data not reported are represented by “-”.

3.3. Cytotoxic and Anticancer Effects of Euphorbiaceae Pure Compounds In Vitro Studies

Many studies have focused on investigating the induction of apoptosis through various signaling pathways in cancer cells as a result of isolated plant compounds. In their study, Wisniewski et al. [145] showed that, of the compounds derived from *Euphorbia* sp., latilagascene B is an effective P-glycoprotein inhibitor capable of increasing doxorubicin accumulation in resistant cells (human colon carcinoma LoVo cells). In contrast, Li et al. [146] showed that Trigothyoid N, a natural diterpenoid isolated from *Trigonostemon thyrsoides*, revealed a strong ability to inhibit the proliferation of A549 lung cancer cells through cell cycle arrest. In addition, the compound can inhibit tumor proliferation and migration by targeting mitochondria, regulating the signal transducer and activator of transcription 3/focal adhesion kinase) signaling pathway (STAT3/FAK), and inhibiting angiogenesis. In another study, Lin et al. [147] showed that *Euphorbia* L2, a lathyrane diterpenoid isolated from the seeds of *Euphorbia lathyris* L., possessed potent cytotoxicity against A549 lung cancer cells and induced apoptosis through the mitochondrial pathway via an increase in reactive oxygen species (ROS), loss of mitochondrial potential, release of cytochrome c, activation of casp-9 and casp-3, and cleavage of poly(ADP-ribose) polymerase. In turn, Fan et al. [148] showed that the 8,9-seco-*ent*-kaurane diterpenoid isolated from *Croton kongensis* induced apoptosis, autophagy, and metastasis suppression in triple-negative breast cancer (TNBC) cells by inhibiting Akt. Additionally, in in vivo studies, it significantly inhibited TNBC tumor growth without causing side effects. Wongprayoon et al. [149] noted that *Euphorbia lactea* triterpenoid friedelan-3 β -ol was cytotoxic to several cancer cell lines, including HN22, HepG2, HCT116, and HeLa. Furthermore, the authors showed that the compound induced S-phase cell cycle arrest in HN22 cells without inducing apoptosis at the same concentration and exposure time. Studies about isolated compounds are presented in Table 2 below.

Table 2. Cytotoxic properties of isolated compounds from the Euphorbiaceae family of plants against cancer cells.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Croton crassifolius</i> Geiseler	Roots	Ethanol	Crotonpyrone A	HeLa and NCI-446	HeLa (10.21 µg/mL) and NCI-446 (6.59 µg/mL)	Cytotoxic	[150]
<i>Croton crassifolius</i> Geiseler	Roots	Ethanol	Crotonpyrone B	HeLa and NCI-446	HeLa (9.54 µg/mL) and NCI-446 (6.52 µg/mL)	Cytotoxic	[150]
<i>Croton damayeshu</i> Y.T.Chang	Twigs and leaves	Ethanol	Crodamoid H	A549	A549 (0.9 ± 0.6 µM)	Cytotoxic	[151]
<i>Croton damayeshu</i> Y.T.Chang	Twigs and leaves	Ethanol	Crodamoid I	A549 and HL-60	A549 (1.3 ± 0.2 µM) and HL-60 (2.4 ± 1.0 µM)	Cytotoxic	[151]
<i>Croton damayeshu</i> Y.T.Chang	Twigs and leaves	Ethanol	4α-deoxyphorbol-12-tiglate- 13-isobutyrate	A549 and HL-60	A549 (1.9 ± 0.1 µM) and HL-60 (1.8 ± 0.8 µM)	Cytotoxic	[151]
<i>Croton echioides</i> Baill.	Stem bark	EtOH/H ₂ O	<i>N-trans</i> -4-Methoxy- cinnamoyl-5- hydroxytryptamine	HCT-116	HCT-116 (86.8 µmol L ^{−1})	Cytotoxic	[21]
<i>Croton floribundus</i> SPRENG.	Root bark	Hexane	<i>ent</i> -kaur-16-en-6a,19-diol	HCT-116, HL60, MDA-MB-435, and HCT-8	HCT-116 (12.1 µg/mL), MDA-MB-435 (14.3 µg/mL), and HCT-9 (13.5 µg/mL)	Cytotoxic	[152]
<i>Croton lachnocarpus</i> Benth.	Leaves	Ethanol	7b,15-dihydroxy- <i>ent</i> -abieta- 8,11,13-trien-3-one	A549, BGC-823, HepG2, HL-60, MCF-7,	A549 (56.3 µM), BGC-823 (68.7 µM), HepG2 (66.9 µM), HL-60 (52.3 µM), MCF-7 (56.2 µM), W480 (59.4 µM)	Cytotoxic	[49]
<i>Croton lachnocarpus</i> Benth.	Leaves	Ethanol	2b,15-dihydroxy- <i>ent</i> -abieta- 8,11,13-triene	A549, BGC-823, HepG2, HL-60, MCF-7, W480	A549 (52.3 µM), BGC-823 (52.2 µM), HepG2 (59.7 µM), HL-60 (49.4 µM), MCF-7 (60.1 µM), W480 (57.6 µM)	Cytotoxic	[49]
<i>Croton lachnocarpus</i> Benth.	Leaves	Ethanol	7b,13a,15-tri-hydroxy- <i>ent</i> - abieta-8(14)-en-3-one	A549, BGC-823, HepG2, HL-60, MCF-7, W480	A549 (28.7 µM), BGC-823 (26.6 µM), HepG2 (25.3 µM), HL-60 (31.7 µM), MCF-7 (27.1 µM), W480 (24.9 µM)	Cytotoxic	[49]
<i>Croton laui</i> Merr. & F.P.Metcalf			crotonolide A	HL-60 and P-388	HL-60 (9.42 µM) and P-388 (7.45 µM)	Cytotoxic	[153]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Croton tiglium</i> L.	Seeds	Ethanol	4-deoxy-20-oxophorbol 12-tiglyl 13-acetate	K562, A549, and Huh-7	K562 (0.03 µM), A549 (6.88 µM), and Huh-7 (3.85 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Ethanol	7-oxo-5-ene-phorbol-13-(2- methylbutyrate	K562, A549, and Huh-7	K562 (0.03 µM), A549 (6.33 µM), and Huh-7 (20.9 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Ethanol	crotignoid C	K562, A549, and Huh-7	K562 (0.07 µM), A549 (8.86 µM), and Huh-7 (11.6 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Ethanol	13-O-(2-metyl)butyryl- phorbol	K562, A549, and Huh-7	K562 (0.05 µM), A549 (43.5 µM), and Huh-7 (34.2 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Ethanol	12-O-tiglylphorbol-13- acetate	K562, A549, and Huh-7	K562 (0.07 µM), A549 (8.50 µM), and Huh-7 (3.36 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Ethanol	crotignoid F	K562, A549, and Huh-7	K562 (0.05 µM), A549 (3.11 µM), and Huh-7 (4.41 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Ethanol	phorbol	K562, A549, and Huh-7	K562 (0.10 µM), A549 (15.3 µM), and Huh-7 (5.93 µM)	Cytotoxic	[154]
<i>Croton tiglium</i> L.	Seeds	Methanol	13-O-acetylphorbol-20- oleate.	SNUB387	SNUB387 (1.2 ± 0.1 µM)	Cytotoxic	[155]
<i>Croton tiglium</i> L.	Seeds	Methanol	13-O-acetyl-4-deoxy-4α- phorbol-20-linoleate.	SNUB387	SNUB387 (8.1 ± 1.3 µM)	Cytotoxic	[155]
<i>Croton tiglium</i> L.	Seeds	Methanol	13-O-acetyl-4-deoxy-4α- phorbol-20-oleate.	SNUB387	SNUB387 (6.6 ± 0.9 µM)	Cytotoxic	[155]
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	12-O-acetylphorbol-13- isobutyrate	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (4.0 µM), MOLT-4 (2.4 µM), U937 (6.8 µM), MCF-7 (13 µM), Hela (3.9 µM), DU145 (7.2 µM), A549 (5.8 µM) SGC-7091 (13 µM), H1975 (10 µM), HL60 (12 µM)	Cytotoxic	[156]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	12-O-benzoylphorbol-13-(2-methyl)butyrate	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (15 µM), MOLT-4 (12 µM), U937 (17 µM), MCF-7 (20 µM), Hela (4.6 µM), DU145 (4.3 µM), A549 (6.9 µM), SGC-7091 (10 µM), H1975 (3.3 µM), HL60 (6.8 µM)	Cytotoxic	[156]
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	12-O-tiglyl-7-oxo-5-ene-phorbol-13-(2-methylbutyrate)	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (17 µM), MOLT-4 (4.8 µM), U937 (21 µM), MCF-7 (20 µM), Hela (5.0 µM), DU145 (10 µM), A549 (19 µM), SGC-7091 (23 µM), H1975 (10 µM), HL60 (10 µM)	Cytotoxic	[156]
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	13-O-(2-methyl)butyryl-4-deoxy-4a-phorbol.	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (8 µM), MOLT-4 (9.9 µM), U937 (18 µM), MCF-7 (24 µM), Hela (10 µM), DU145 (10 µM), A549 (4.5 µM), SGC-7091 (5.4 µM), H1975 (3.3 µM), HL60 (9.8 µM)	Cytotoxic	[156]
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	12-O-tiglylphorbol-13-propionate	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (4.4 µM), MOLT-4 (1.1 µM), U937 (5.5 µM), MCF-7 (>50 µM), Hela (9.2 µM), DU145 (1.1 µM), A549 (32 µM), SGC-7091 (43 µM), H1975 (10 µM), HL60 (1.2 µM)	Cytotoxic	[156]
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	12-O-tiglylphorbol-13-isobutyrate	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (2.2 µM), MOLT-4 (1.0 µM), U937 (2.6 µM), Hela (10 µM), DU145 (5.0 µM), H1975 (10 µM), HL60 (1.2 µM)	Cytotoxic	[156]
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	12-O-tiglylphorbol-13-(2-methyl)butyrate	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (7.2 µM), MOLT-4 (10 µM), U937 (14 µM), MCF-7 (>50 µM), Hela (10 µM), DU145 (11 µM), A549 (>50 µM), SGC-7091 (>50 µM), H1975 (10 µM), HL60 (9.9 µM)	Cytotoxic	[156]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Croton tiglium</i> L.	Branches and leaves	Ethanol	tiglin A	K562, MOLT-4, U937, MCF-7, Hela, DU145, A549, SGC-7091, H1975, HL60	K562 (15 µM), MOLT-4 (12 µM), U937 (17 µM), MCF-7 (20 µM), Hela (4.6 µM), DU145 (4.3 µM), A549 (6.9 µM), SGC-7091 (10 µM), H1975 (3.3 µM), HL60 (6.8 µM)	Cytotoxic	[156]
<i>Croton tiglium</i> L.	Seeds	Acetone	7-Keto-12-O-tiglylphorbol-13-acetate	HL60 and A549	HL60 (6.22 ± 3.24 µg/mL) and A549 (18.0 ± 9.48 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	Phorbol-12-isobutyrate	HL60 and A549	HL60 (0.22 ± 0.15 µg/mL) and A549 (0.74 ± 0.48 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	12-O-Tiglylphorbol-13-acetate	HL60 and A549	HL60 (0.02 ± 0.01 µg/mL) and A549 (0.10 ± 0.03 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	12-O-(2-methyl)-butyrylphorbol-13-aetate	HL60 and A549	HL60 (<0.01 µg/mL) and A549 (0.01 ± 0.00 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	12-O-tiglyl-phorbol-13-isobutyrate	HL60 and A549	HL60 (<0.01 µg/mL) and A549 (<0.01 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	phorbol-12-tiglate	HL60 and A549	HL60 (83.1 ± 2.89 µg/mL) and A549 (38.6 ± 30.1 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	phorbol-12-tetradecanoate	HL60 and A549	HL60 (3.14 ± 2.17 µg/mL) and A549 (4.71 ± 1.92 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	phorbol-13-aetate	HL60 and A549	HL60 (80.9 ± 11.6 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	phorbol-13-decanoate	HL60 and A549	HL60 (0.02 ± 0.02 µg/mL) and A549 (0.94 ± 0.01 µg/mL)	Cytotoxic	[157]
<i>Croton tiglium</i> L.	Seeds	Acetone	4-deoxy-4α-phorbol-13-acetate	HL60 and A549	HL60 (89.0 ± 0.76 µg/mL)	Cytotoxic	[157]
<i>Croton urucurana</i> Baill.	Bark	Ethanol	Orbitide [1–9-NαC]-crouorb A1	786-O, HT29, MCF7, ADR-RES, Hep-G2, and PC-03	786-O (18.69 ± 0.82 µg/mL), HT29 (37.28 ± 0.57 µg/mL), MCF7 (35.49 ± 2.59 µg/mL), ADR-RES (3.98 ± 0.20 µg/mL), Hep-G2 (41.31 ± 2.70 µg/mL) and PC-03 (29.80 ± 0.34 µg/mL)	Cytotoxic	[158]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Croton velutinus</i> Baill.	Roots	Methanol	(E)-1-(7,8-epoxypropen) phenyl benzoate	B16F10, HL-60, HCT116, MCF-7, and HepG2	B16F10 ($14.4 \pm 0.5 \mu\text{M}$), HL-60 ($9.8 \pm 2.6 \mu\text{M}$), HCT116 ($12.9 \pm 1.8 \mu\text{M}$), MCF-7 ($6.8 \pm 1.59 \mu\text{M}$) and HepG2 ($16.7 \pm 0.7 \mu\text{M}$)	Cytotoxic	[159]
<i>Croton velutinus</i> Baill.	Roots	Methanol	sellovicine B	B16F10, HL-60, HCT116, MCF-7, and HepG2	B16F10 ($13.8 \pm 01 \mu\text{M}$), HL-60 ($11.4 \pm 3.6 \mu\text{M}$), HCT116 ($13.2 \pm 1.0 \mu\text{M}$), MCF-7 ($11.1 \pm 1.4 \mu\text{M}$) and HepG2 ($18.3 \pm 1.8 \mu\text{M}$)	Cytotoxic	[159]
<i>Drypetes hainanensis</i> Merr.	Leaves and stems	Ethanol	4 β -hydroxy-23-nor-friedel- 3-one	BEL-7402, A549, HL60	GI rates: BEL-7402 (3.0%), A549 (9.7%), HL60 (4.1%)	Cytotoxic	[160]
<i>Drypetes hainanensis</i> Merr.	Leaves and stems	Ethanol	friedelin	BEL-7402, A549, HL60	HL60 (1.3%)	Cytotoxic	[160]
<i>Drypetes hainanensis</i> Merr.	Leaves and stems	Ethanol	friedelane-3,7-dione	BEL-7402, A549, HL60	BEL-7402 (4.6%), A549 (21.1%), HL60 (43.1%)	Cytotoxic	[160]
<i>Euphorbia ammak</i> Schweinf.	Leaves	Ethanol	euphol	HeLa	HeLa (9.25 mg/mL)	Cytotoxic	[161]
<i>Euphorbia ammak</i> Schweinf.	Leaves	Ethanol	α -glutinol	HeLa	HeLa (7.6 mg/mL)	Cytotoxic	[161]
<i>Euphorbia ammak</i> Schweinf.	Leaves	Ethanol	stigmasterol	HeLa	HeLa (10 mg/mL)	Cytotoxic	[161]
<i>Euphorbia balsamifera</i> Aiton	Aerial parts	Ethanol	Kampferol-3,40-dimethyl ether	HCT-116, HePG2, and MCF7	HCT116 (111.46 mM), HePG2 (42.67 mM) and MCF7 (44.90 mM)	Cytotoxic	[162]
<i>Euphorbia connata</i> Boiss.	Aerial flowering parts	Dichlorom- ethane/ Acetone	3,7,14,15-tetraacetyl-5- propanoyl-13(17)-epoxy- 8,10(18)-myrsinadiene	MDA-MB-231 and MCF-7	MDA-MB-231 ($24.53 \pm 3.39 \mu\text{M}$) and MCF-7 ($37.73 \pm 3.41 \mu\text{M}$)	Cytotoxic	[163]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia connata</i> Boiss.	Aerial flowering parts	Dichloro- methane/Acetone	3,7,10,14,15-pentaacetyl-5- butanoyl-13,17-epoxy-8- myrsinene	MDA-MB-231 and MCF-7	MDA-MB-231 ($26.67 \pm 1.41 \mu\text{M}$) and MCF-7 ($34.57 \pm 2.12 \mu\text{M}$)	Cytotoxic	[163]
<i>Euphorbia dendroides</i> L.	Aerial parts	Methanol	23 R/S-3b-hydroxycycloart- 24-ene23-methyl ether	HepG2, Huh-7, KLM-1, 1321N1, HeLa	HepG2 ($20.67 \pm 0.72 \mu\text{M}$), Huh-7 ($16.24 \pm 0.53 \mu\text{M}$), KLM-1 ($22.59 \pm 0.94 \mu\text{M}$), 1321N1 ($25.99 \pm 0.31 \mu\text{M}$), HeLa ($40.50 \pm 3.14 \mu\text{M}$)	Cytotoxic	[164]
<i>Euphorbia dendroides</i> L.	Aerial parts	Methanol	24-methylene cycloartan-3b-ol	HepG2, Huh-7, KLM-1, 1321N1, HeLa	HepG2 ($10.93 \pm 0.21 \mu\text{M}$), Huh-7 ($7.42 \pm 0.16 \mu\text{M}$), KLM-1 ($21.48 \pm 0.60 \mu\text{M}$), 1321N1 ($12.32 \pm 0.58 \mu\text{M}$), HeLa ($13.68 \pm 0.16 \mu\text{M}$)	Cytotoxic	[164]
<i>Euphorbia dendroides</i> L.	Aerial parts	Methanol	cycloart-23-ene-3b,25-diol monoacetate	HepG2, Huh-7, KLM-1, 1321N1, HeLa	HepG2 ($12.81 \pm 0.73 \mu\text{M}$), Huh-7 ($<0.47 \mu\text{M}$), KLM-1 ($22.48 \pm 0.64 \mu\text{M}$), 1321N1 ($25.17 \pm 0.32 \mu\text{M}$), HeLa ($54.05 \pm 1.11 \mu\text{M}$)	Cytotoxic	[164]
<i>Euphorbia dendroides</i> L.	Aerial parts	Methanol	3b-hydroxy-cycloart-23-ene- 25 methyl ether	HepG2, Huh-7, KLM-1, 1321N1, HeLa	HepG2 ($12.72 \pm 2.38 \mu\text{M}$), Huh-7 ($<0.44 \mu\text{M}$), KLM-1 ($<0.44 \mu\text{M}$), 1321N1 ($0.63 \pm 0.15 \mu\text{M}$), HeLa ($3.7 \pm 0.39 \mu\text{M}$)	Cytotoxic	[164]
<i>Euphorbia dendroides</i> L.	Aerial parts	Methanol	24 R/S-3b-hydroxy-25- methylene Figure 1. Chemical structures of compounds 1–11, isolated from <i>Euphorbia dendroides</i> L. aerial parts. 830 A. R. HASSAN ET AL. cycloartan-24-ol	HepG2, Huh-7, KLM-1, 1321N1, HeLa	HepG2 (15.54 ± 1.95), Huh-7 (16.33 ± 1.22), KLM-1 (22.38 ± 1.29), 1321N1 (13.53 ± 0.33), HeLa (>4.52)	Cytotoxic	[164]
<i>Euphorbia denticulata</i> Lam	-	Acetone	taraxast-12-ene-3 β ,20,21(α)- triol	DU-145	DU-145 ($12.2 \pm 2.9 \mu\text{M}$)	Cytotoxic	[165]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia denticulata</i> Lam	-	Acetone	cycloartane-3 β ,25-diol	DU-145	DU-145 (27.5 $\pm$ 4.9 μ M)	Cytotoxic	[165]
<i>Euphorbia denticulata</i> Lam	-	Acetone	cycloartane-3 β ,24,25-triol	DU-145	DU-145 (18.3 $\pm$ 1.4 μ M)	Cytotoxic	[165]
<i>Euphorbia ebracteolata</i> Hayata	Roots	Ethanol	Ebracteolata A	HL60, A549, SMMC-7721, MCF-7, and SW480	HL60 (17.5 μ M), A549 (11.0 μ M), SMMC-7721 (16.8 μ M), MCF-7 (17.5 μ M), and SW480 (18.0 μ M)	Cytotoxic	[166]
<i>Euphorbia ebracteolata</i> Hayata	Roots	Ethanol	Yuexiandajisu F	HL60, A549, SMMC-7721, MCF-7, and SW480	HL60 (16.8 μ M), A549 (19.7 μ M), SMMC-7721 (18.4 μ M), MCF-7 (15.3 μ M), and SW480 (15.3 μ M)	Cytotoxic	[166]
<i>Euphorbia ebracteolata</i> Hayata	Roots	Ethanol	jolkinol B	HL60, A549, SMMC-7721, MCF-7, and SW480	HL60 (5.0 μ M), A549 (11.5 μ M), SMMC-7721 (3.5 μ M), MCF-7 (15.8 μ M), and SW480 (9.5 μ M)	Cytotoxic	[166]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	ebracteolatas D	HCT116, A549, HeLa, SW620, MCF-7, HepG-2	HCT116 (>40 μ M), A549 (22.03 μ M), HeLa (>40 μ M), SW620 (>40 μ M), MCF-7 (>40 μ M), HepG-2 (>40 μ M)	Cytotoxic	[48]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	ebractenoid Q	HCT116, A549, HeLa, SW620, MCF-7, HepG-2	HCT116 (33.18 μ M), A549 (2.81 μ M), HeLa (>40 μ M), SW620 (>40 μ M), MCF-7 (>40 μ M), HepG-2 (27.24 μ M)	Cytotoxic	[48]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	Euphonoid H	MDA-MB-231, HCT-15, RKO, C4-2B, C4-2B/ENZR	MDA-MB-231 (21.80 $\pm$ 2.35 μ M), HCT-15 (28.57 $\pm$ 1.16 μ M), RKO (20.46 $\pm$ 1.43 μ M), C4-2B (5.52 $\pm$ 0.65 μ M), C4-2B/ENZR (4.16 $\pm$ 0.42 μ M)	Cytotoxic	[47]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	Euphonoid I	MDA-MB-231, HCT-15, RKO, C4-2B, C4-2B/ENZR	MDA-MB-231 (7.95 $\pm$ 0.82 μ M), HCT-15 (12.45 $\pm$ 3.24 μ M), RKO (8.78 $\pm$ 2.45 μ M), C4-2B (4.49 $\pm$ 0.78 μ M), C4-2B/ENZR (5.74 $\pm$ 0.45 μ M)	Cytotoxic	[47]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia fischeriana</i> Steud	Roots	EtOH	17-Hydroxyljolkinolide B	MCF-10A, MCF-7, ZR-75-1 and MDA-MB-231	MCF-10A (3.4 ± 0.1 $\mu\text{g/mL}$), MCF-7 (4.7 ± 0.2 $\mu\text{g/mL}$), ZR-75-1 (2.2 ± 0.1 $\mu\text{g/mL}$) and MDA-MB-231 (1.1 ± 0.1 $\mu\text{g/mL}$)	Cytotoxic	[167]
<i>Euphorbia fischeriana</i> Steud	Roots	EtOH	17-Acetyljolkinolide B	MCF-10A, MCF-7, ZR-75-1 and MDA-MB-231	MCF-10A (4.3 ± 0.1 $\mu\text{g/mL}$), MCF-7 (3.4 ± 0.1 $\mu\text{g/mL}$), ZR-75-1 (1.2 ± 0.1 $\mu\text{g/mL}$) and MDA-MB-231 (1.7 ± 0.1 $\mu\text{g/mL}$)	Cytotoxic	[167]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	Euphorfiatnoid B	HepG2, H460, and MCF-7	H460 (9.97 μM)	Cytotoxic	[168]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	Euphorfiatnoid A	HepG2, H460, and MCF-7	HepG2 (11.64 μM), H460 (28.54 ± 1.20 μM), and MCF-7 (40.02 ± 0.47 μM)	Cytotoxic	[168]
<i>Euphorbia fischeriana</i> Steud	Roots	Ethanol	Euphorfiatnoid C	HepG2, H460, and MCF-7	HepG2 (13.10 ± 0.35 μM), H460 (14.88 ± 0.57 μM), and MCF-7 (32.95 ± 0.40 μM)	Cytotoxic	[168]
<i>Euphorbia fischeriana</i> Steud.	Roots	Ethanol	Euphorfischerin A	HeLa, N460 and Namalwa	HeLa (4.6 ± 0.11 μM), N460 (11.5 ± 0.04 μM) and Namalwa (16.4 ± 0.07 μM)	Cytotoxic	[169]
<i>Euphorbia fischeriana</i> Steud.	Roots	Ethanol	Euphorfischerin B	HeLa, N460 and Namalwa	HeLa (9.5 ± 0.16 μM), N460 (17.4 ± 0.34 μM) and Namalwa (13.3 ± 0.19 μM)	Cytotoxic	[169]
<i>Euphorbia fischeriana</i> Steud.	Aerial parts	Methanol	3 α -acetoxy-14-hydroxy- <i>ent</i> - abieta-8(9),13(15)-dien-16,12- olide	HL-60, SMMC-7721 and SGC-7901	HL-60 (15.3 μM), SMMC-7721 (23.2 μM) and SGC-7901 (29.0 μM)	Cytotoxic	[170]
<i>Euphorbia helioscopia</i> L.	Aerial parts	Ethanol aqueous	Euphoheliphane A	OS-RC-2, Ketr-3, 769-P, G401, GRC-1 and ACHN	OS-RC-2 (47 μM), Ketr-3 (45 μM), 769-P (43 μM), G401 (38 μM), GRC-1 (41 μM) and ACHN (40 μM)	Cytotoxic	[171]
<i>Euphorbia helioscopia</i> L.	Aerial parts	Ethanol aqueous	Euphoheliphane B	OS-RC-2, Ketr-3, 769-P, G401, GRC-1 and ACHN	OS-RC-2 (31 μM), Ketr-3 (32 μM), 769-P (30 μM), G401 (34 μM), GRC-1 (33 μM) and ACHN (35 μM)	Cytotoxic	[171]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia helioscopia</i> L.	Aerial parts	Ethanol aqueous	Euphoheliphane C	OS-RC-2, Ketr-3, 769-P, G401, GRC-1 and ACHN	OS-RC-2 (35 μ M), Ketr-3 (41 μ M), 769-P (39 μ M), G401 (32 μ M), GRC-1 (38 μ M) and ACHN (36 μ M)	Cytotoxic	[171]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	Euphohelinoid A	HepG2, Hela, HL-60, SMMC-7721	HepG2 (24.3 $\pm$ 1.5 μ M), Hela (28.4 $\pm$ 1.8 μ M), HL-60 (18.6 $\pm$ 1.1 μ M), SMMC-7721 (29.6 $\pm$ 1.5 μ M)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	Euphohelinoid B	HepG2, Hela, HL-60, SMMC-7721	HepG2 (10.2 $\pm$ 1.4 μ M), Hela (9.3 $\pm$ 1.2 μ M), HL-60 (8.1 $\pm$ 0.7 μ M), SMMC-7721 (9.8 $\pm$ 1.3 μ M)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	Euphohelinoid C	HepG2, Hela, HL-60, SMMC-7721	HepG2 (>50 μ M), Hela (>50 μ M), HL-60 (>50 μ M), SMMC-7721 (>50 μ M)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	Euphohelinoid D	HepG2, Hela, HL-60, SMMC-7721	HepG2 (>50 μ M), Hela (34.5 $\pm$ 2.3 μ M), HL-60 (34.1 $\pm$ 1.6 μ M), SMMC-7721 (30.1 $\pm$ 1.9 μ M)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	Euphohelinoid F	HepG2, Hela, HL-60, SMMC-7721	HepG2 (12.5 $\pm$ 1.6 μ M), Hela (14.1 $\pm$ 0.8 μ M), HL-60 (13.3 $\pm$ 1.2 μ M), SMMC-7721 (11.1 $\pm$ 1.7 μ M)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	euphornin L	HepG2, Hela, HL-60, SMMC-7721	HepG2 (22.8 $\pm$ 1.7 μ M), Hela (25.7 $\pm$ 2.2 μ M), HL-60 (13.1 $\pm$ 1.8 μ M), SMMC-7721 (14.3 $\pm$ 2.2 μ M)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	helioscopianoid O	HepG2, Hela, HL-60, SMMC-7721	HepG2 (>50 μ M), Hela (26.2 $\pm$ 1.4 μ M), HL-60 (18.2 $\pm$ 1.9 μ M), SMMC-7721 (19.5 $\pm$ 1.2 μ M)	Cytotoxic	[172]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	euphoscopin I	HepG2, Hela, HL-60, SMMC-7721	HepG2 ($24.1 \pm 1.2 \mu\text{M}$), Hela ($29.7 \pm 2.1 \mu\text{M}$), HL-60 ($14.3 \pm 1.1 \mu\text{M}$), SMMC-7721 ($18.7 \pm 1.1 \mu\text{M}$)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	euphoscopin J	HepG2, Hela, HL-60, SMMC-7721	HepG2 ($14.9 \pm 1.3 \mu\text{M}$), Hela ($13.7 \pm 1.4 \mu\text{M}$), HL-60 ($12.4 \pm 1.2 \mu\text{M}$), SMMC-7721 ($15.0 \pm 1.7 \mu\text{M}$)	Cytotoxic	[172]
<i>Euphorbia helioscopia</i> L	Whole plant	Methanol	euphoscopin B	HepG2, Hela, HL-60, SMMC-7721	HepG2 ($23.3 \pm 1.3 \mu\text{M}$), Hela ($29.2 \pm 1.6 \mu\text{M}$), HL-60 ($20.2 \pm 1.5 \mu\text{M}$), SMMC-7721 ($27.1 \pm 1.4 \mu\text{M}$)	Cytotoxic	[172]
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	13-epicupressic acid	EJ-138, HepG2, A549, MCF-7 and PC3	EJ-138 ($>200 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($157.4 \pm 3.24 \mu\text{g/mL}$), MCF-7 ($139.1 \pm 2.14 \mu\text{g/mL}$) and PC3 ($>200 \mu\text{g/mL}$)	Cytotoxic	[173]
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	imbricatholic acid	EJ-138, HepG2, A549, MCF-7, and PC3	EJ-138 ($>200 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($>200 \mu\text{g/mL}$), MCF-7 ($173.5 \pm 4.34 \mu\text{g/mL}$) and PC3 ($>200 \mu\text{g/mL}$)	Cytotoxic	[173]
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	α -hydroxy sandaracopimaric acid	EJ-138, HepG2, A549, MCF-7, and PC3	EJ-138 ($173.3 \pm 2.37 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($>200 \mu\text{g/mL}$), MCF-7 ($>250 \mu\text{g/mL}$) and PC3 ($>250 \mu\text{g/mL}$)	Cytotoxic	[173]
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	13-epicupressic acid	EJ-138, HepG2, A549, MCF-7, and PC3	EJ-138 ($182.2 \pm 1.18 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($>200 \mu\text{g/mL}$), MCF-7 ($>200 \mu\text{g/mL}$) and PC3 ($>250 \mu\text{g/mL}$)	Cytotoxic	[173]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	β -hydroxy sandaracopimaric acid 13-epicupressic acid	EJ-138, HepG2, A549, MCF-7, and PC3	EJ-138 ($>200 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($67.4 \pm 2.45 \mu\text{g/mL}$), MCF-7 ($111.7 \pm 3.75 \mu\text{g/mL}$) and PC3 ($>200 \mu\text{g/mL}$)	Cytotoxic	[173]
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	5,7,3',4'-pentahydroxyflavone	EJ-138, HepG2, A549, MCF-7, and PC3	EJ-138 ($>200 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($135.8 \pm 7.41 \mu\text{g/mL}$), MCF-7 ($117.4 \pm 3.71 \mu\text{g/mL}$) and PC3 ($>200 \mu\text{g/mL}$)	Cytotoxic	[173]
<i>Euphorbia heterophylla</i> L.	Roots	<i>n</i> -hexane, DCM and MeOH	quercitrin	EJ-138, HepG2, A549, MCF-7, and PC3	EJ-138 ($>250 \mu\text{g/mL}$), HepG2 ($>200 \mu\text{g/mL}$), A549 ($138.1 \pm 4.62 \mu\text{g/mL}$), MCF-7 ($105.3 \pm 6.19 \mu\text{g/mL}$), and PC3 ($>200 \mu\text{g/mL}$)	Cytotoxic	[173]
<i>Euphorbia hypericifolia</i> L	Whole herb	Ethanol	euphyphenoid A	HCT-116	HCT-116 ($12.8 \pm 1.6 \mu\text{M}$)	Cytotoxic	[174]
<i>Euphorbia hypericifolia</i> L	Whole herb	Ethanol	20(S),24(R)-20,24-epoxy-24-methyldammaran-3 β -ol	HCT-116	HCT-116 ($26.8 \pm 4.6 \mu\text{M}$)	Cytotoxic	[174]
<i>Euphorbia hypericifolia</i> L	Whole herb	Ethanol	3 β -hydroxycycloart-24-one	HCT-116	HCT-116 ($7.4 \pm 0.2 \mu\text{M}$)	Cytotoxic	[174]
<i>Euphorbia hypericifolia</i> L	Whole herb	Ethanol	isomotirol	HCT-116	HCT-116 ($10.6 \pm 1.2 \mu\text{M}$)	Cytotoxic	[174]
<i>Euphorbia kansui</i> S.L.Liou ex S.B.Ho	Roots	Ethanol	Euphorikanin A	HeLa and NCI-446	HeLa ($28.85 \pm 1.41 \mu\text{M}$) and NCI-446 ($20.89 \pm 1.67 \mu\text{M}$)	Cytotoxic	[175]
<i>Euphorbia lactea</i> Haw.	Aerial parts	Ethanol, <i>n</i> -hexane fraction	friedelin	HN22, HepG2, and HCT116	-	Cytotoxic	[149]
<i>Euphorbia lactea</i> Haw.	Aerial parts	Ethanol, <i>n</i> -hexane fraction	taraxerol	HN22, HepG2, and HCT116	-	Cytotoxic	[149]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia lactea</i> Haw.	Aerial parts	Ethanol, <i>n</i> -hexane fraction	friedelan-3 α -ol	HN22, HepG2, and HCT116	-	Cytotoxic	[149]
<i>Euphorbia lagascae</i> Spreng.	Seeds	Methanol	Esculetin	LoVo and LoVo/Dx	LoVo (56.81 $\pm$ 5.42%) and LoVo/Dx (68.42 $\pm$ 7.56%)	Cytotoxic	[145]
<i>Euphorbia microsphaera</i> Boiss	Aerial parts	Hexane, chloroform and methanol	Aryanin (3aR,4S,4aS,5R,7aS,9aS)-5-hydroxy-5,8-dimethyl-3-methylene-2-oxo2,3,3a,4,4a,5,6,7,7a, 9a decahydroazuleno [6,5-b] furan-4-yl acetate)	MCF-7	MCF-7 (13.81 μ g/mL)	Cytotoxic	[61]
<i>Euphorbia nematocypha</i> Hand.-Mazz.	Roots	Ethanol	16-O-caffeoyl-16-hydroxyldodecanoic acid	MCF-7 and HeLa	MCF-7 (20.22 $\pm$ 1.2 μ mol/L) and HeLa (27.8 $\pm$ 1.4 μ mol/L)	Cytotoxic	[176]
<i>Euphorbia nematocypha</i> Hand.-Mazz.	Aerial parts	Methylene chloride	<i>trans, trans</i> -2',4'-hexadienedioicacid-1'- β -amyrin ester	MCF7 and HeLa	MCF7 (29.5 $\pm$ 3.4 μ mol /L) and HeLa (23.2 $\pm$ 4.2 μ mol /L)	Cytotoxic	[177]
<i>Euphorbia nematocypha</i> Hand.-Mazz.	Roots	Ethanol	Nematocynine	HCC1806, ST486, CT26, HeLa, and A549	HCC1806 (16.96 $\pm$ 0.16 μ M), ST486 (60.94 $\pm$ 0.74 μ M), CT26 (52.04 $\pm$ 1.96 μ M), and HeLa (52.70 $\pm$ 0.52 μ M)	Cytotoxic	[178]
<i>Euphorbia neriifolia</i> Linn.	Aerial parts	Ethanol	Phonerilin B	A549 and HL60	A549 (8.6 $\pm$ 1.7 μ M) and HL60 (9.1 $\pm$ 0.02 μ M)	Cytotoxic	[179]
<i>Euphorbia neriifolia</i> Linn.	Aerial parts	Ethanol	Phonerilin E	A549 and HL60	A549 (4.9 $\pm$ 0.06 μ M) and HL60 (9.2 $\pm$ 0.09 μ M)	Cytotoxic	[179]
<i>Euphorbia neriifolia</i> Linn.	Aerial parts	Ethanol	Phonerilin F	A549 and HL60	A549 (3.8 $\pm$ 0.2 μ M) and HL60 (4.5 $\pm$ 0.7 μ M)	Cytotoxic	[179]
<i>Euphorbia neriifolia</i> Linn.	Aerial parts	Ethanol	Phonerilin H	A549 and HL60	A549 (7.5 $\pm$ 0.8 μ M) and HL60 (5.7 $\pm$ 1.0 μ M)	Cytotoxic	[179]
<i>Euphorbia neriifolia</i> Linn.	Aerial parts	Ethanol	20-O-diacetyl-ingenol	A549 and HL60	HL60 (3.1 $\pm$ 0.2 μ M)	Cytotoxic	[179]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia neriifolia</i> Linn.	Aerial parts	Ethanol	7,12-O-diacetyl-8-O- tigloylingol	A549 and HL60	A549 ($6.4 \pm 0.2 \mu\text{M}$) and HL60 ($9.5 \pm 0.04 \mu\text{M}$)	Cytotoxic	[179]
<i>Euphorbia osyridea</i> Bioss.	Aerial flowering parts	Dichlorome- thane/Acetone	2,7,9,14-tetraacetyl-3- benzoyl-8-butanoyl-5,15- dihydroxy-6(17),11(E)- jatrophadiene	Caov-4, and OVCAR	Caov-4 ($46.27 \pm 3.86 \mu\text{M}$) and OVCAR ($38.81 \pm 3.30 \mu\text{M}$)	Cytotoxic	[180]
<i>Euphorbia osyridea</i> Bioss.	Aerial flowering parts	Dichlorome- thane/Acetone	2,7,9,14-tetraacetyl-3- benzoyl-propionyl ester-5,15-dihydroxy- 6(17),11(E)-jatrophadiene	Caov-4, and OVCAR	Caov-4 (36.48 ± 3.18), and OVCAR ($42.59 \pm 4.50 \mu\text{M}$)	Cytotoxic	[180]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Pekinenin G (11a,12b-epoxy- 18-hydroxy-1bH, 2aH-casba-3E and 7E-dien-5-one)	BGC823, A549, HT-29, and MCF-7	BGC823 ($42.7 \mu\text{M}$), A549 ($40.8 \mu\text{M}$), HT-29 ($47.8 \mu\text{M}$), and MCF-7 ($48.5 \mu\text{M}$)	Cytotoxic	[181]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Pekinenin G (11a,12b-epoxy- 18-hydroxy-1bH, 2aH-casba-3E and 7E-dien-5-one) 2	BGC823, A549, HT-29, and MCF-8	BGC823 ($53.9 \mu\text{M}$), A549 ($88.8 \mu\text{M}$), HT-29 ($70.1 \mu\text{M}$), and MCF-7 ($70.5 \mu\text{M}$)	Cytotoxic	[181]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Pekinenin G (11a,12b-epoxy- 18-hydroxy-1bH, 2aH-casba-3E and 7E-dien-5-one) 3	BGC823, A549, HT-29, and MCF-9	BGC823 ($15.6 \mu\text{M}$), A549 ($21.9 \mu\text{M}$), HT-29 ($25.1 \mu\text{M}$), and MCF-7 ($22.3 \mu\text{M}$)	Cytotoxic	[181]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Pekinenin G (11a,12b-epoxy- 18-hydroxy-1bH, 2aH-casba-3E and 7E-dien-5-one) 4	BGC823, A549, HT-29, and MCF-10	BGC823 ($25.1 \mu\text{M}$), A549 ($35.1 \mu\text{M}$), HT-29 ($30.2 \mu\text{M}$), and MCF-7 ($32.3 \mu\text{M}$)	Cytotoxic	[181]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Pekinenin G (11a,12b-epoxy- 18-hydroxy-1bH, 2aH-casba-3E and 7E-dien-5-one) 5	BGC823, A549, HT-29, and MCF-11	BGC823 ($54.8 \mu\text{M}$), A549 ($90.2 \mu\text{M}$), HT-29 ($110.7 \mu\text{M}$), and MCF-7 ($87.9 \mu\text{M}$)	Cytotoxic	[181]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Pekinenin G (11a,12b-epoxy- 18-hydroxy-1bH, 2aH-casba-3E and 7E-dien-5-one) 6	BGC823, A549, HT-29, and MCF-12	BGC823 (12.1 μ M), A549 (15.6 μ M), HT-29 (11.3 μ M), and MCF-7 (21.2 μ M)	Cytotoxic	[181]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	(–)-(1S)-15-hydroxy-18- carboxycembrene	Hela, PC-3, HT1080, A375-S2, and MDA23	Hela (35.3 $\pm$ 3.6 μ M), PC-3 (53.9 $\pm$ 6.2 μ M), HT1080 (37.3 $\pm$ 2.0 μ M), A375-S2 (28.7 $\pm$ 3.8 μ M), and MDA24 (43.5 $\pm$ 5.1 μ M)	Cytotoxic	[182]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Jolkinol B	U-937 and LOVO	U-937 (3.60 $\pm$ 0.02 μ M) and LOVO (8.44 $\pm$ 0.03 μ M)	Cytotoxic	[183]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Euphodane A	U-937	U-937 (5.92 $\pm$ 0.38 μ M)	Cytotoxic	[183]
<i>Euphorbia pekinensis</i> Rupr.	Roots	Ethanol	Isopimara-7,15-dien-3 β -ol	K-562	K-562 (0.87 $\pm$ 0.02 μ M)	Cytotoxic	[183]
<i>Euphorbia pseudocactus</i> A.Berger	Aerial parts	Methanol	Gallic acid	LS-174T	LS-174T (18.27 μ g/mL)	Cytotoxic	[184]
<i>Euphorbia pseudocactus</i> A.Berger	Aerial parts	Methanol	Ethyl gallate	LS-174T	LS-174T (25.42 μ g/mL)	Cytotoxic	[184]
<i>Euphorbia royleana</i> Boiss.	Whole plant	Ethanol	(3b,23Z)-9,19-cyclolanost-23- ene-3,25-diol	A549	A549 (4.84 $\pm$ 0.56 μ M)	Cytotoxic	[185]
<i>Euphorbia royleana</i> Boiss.	Whole plant	Ethanol	taraxerol	A549	A549 (7.11 $\pm$ 1.65 μ M)	Cytotoxic	[185]
<i>Euphorbia sanctae-catharinae</i> Fayed	Aerial parts	Dichlorometh- ane/Methanol (1:1)	4,12,20-trideoxyphorbol-13- (2,3-dimethyl) butyrate	A549 and Caco-2	A549 (3.3 (0.996) μ M) and Caco-2 (29.4 (0.972) μ M)	Cytotoxic	[175]
<i>Euphorbia schimperi</i> C.Presl	Aerial parts	MeOH/H ₂ O (70:30 V/V)	Cycloschimperol A	MCF-7, HepG2, and HCT-116	MCF-7 (55.4 $\pm$ 3 μ M), HepG2 (19.7 $\pm$ 3 μ M), and HCT-116 (20.25 $\pm$ 5 μ M)	Cytotoxic	[186]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia schimperi</i> C.Presl	Aerial parts	MeOH/H ₂ O (70:30 V/V)	Cycloschimperol B	MCF-7, HepG2, and HCT-116	MCF-7 ($2.1 \pm 0.01 \mu\text{M}$), HepG2 ($1.4 \pm 0.1 \mu\text{M}$), and HCT-116 ($1.8 \pm 0.1 \mu\text{M}$)	Cytotoxic	[186]
<i>Euphorbia schimperi</i> C.Presl	Aerial parts	MeOH/H ₂ O (70:30 V/V)	Cycloart-25-en-3-one	MCF-7, HepG2, and HCT-116	MCF-7 ($4.7 \pm 0.1 \mu\text{M}$), HepG2 ($2.3 \pm 0.2 \mu\text{M}$), and HCT-116 ($1.9 \pm 0.4 \mu\text{M}$)	Cytotoxic	[186]
<i>Euphorbia schimperiana</i> Scheele	Aerial parts	Ethanol	3,30-di-O-methylellagic acid	PC3	PC3 (5.5 mg/mL)	Cytotoxic	[187]
<i>Euphorbia sogdiana</i> Popov	Aerial parts	Acetone/Dichloro- methane (1:2)	Tiglane diterpene	EJ-138 and Jurkat T	EJ-138 (12.1 μM) and Jurkat T (16.1 μM)	Cytotoxic	[188]
<i>Euphorbia stracheyi</i> Boiss	Whole plant	Methanol	3-O-benzoyl-20- deoxymgenol	HL-60, A-549, SMMC-7721, MCF-7, and SW480	HL-60 ($0.5 \pm 0.18 \mu\text{M}$), A-549 ($21.47 \pm 0.17 \mu\text{M}$), SMMC-7721 ($18.36 \pm 1.17 \mu\text{M}$), MCF-7 ($18.82 \pm 0.84 \mu\text{M}$), and SW481 ($16.25 \pm 0.71 \mu\text{M}$)	Cytotoxic	[189]
<i>Euphorbia stracheyi</i> Boiss.	Roots	Methanol	Euphstrachenol A	MV4-11	MV4-11 (12.29 μM)	Cytotoxic	[190]
<i>Euphorbia stracheyi</i> Boiss.	Roots	Methanol	Euphstrachenol B	MV4-11	MV4-11 (14.80 μM)	Cytotoxic	[190]
<i>Euphorbia stracheyi</i> Boiss.	Roots	Methanol	Euphstrachenol C	MV4-11	MV4-11 (5.92 μM)	Cytotoxic	[190]
<i>Euphorbia taurinensis</i> All.	Whole plant	MeOH	Ingenane diterpene	L5178Y mouse T-lymphoma cells parent and MDR L5178Y	L5178Y mouse T-lymphoma cells parent (82.47 μM) and MDR L5178Y (62.81 μM)	Cytotoxic	[191]
<i>Euphorbia tirucalli</i> L.	Sap	Hexane	euphol	73 human cancer lines from 15 tumor types	Range from 1.41 to 38.89 μM	Cytotoxic	[192]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Euphorbia tithymaloides</i> L.	Stems	Methanol	friedelane-3 β -ol, 3-oxo-friedelane, euphane-7, 24-diene, 3 β -ol (butyrospermol), and euphane -7, 25-diene, 3, 24- β -diols in addition to the diterpene derivative 1 α , 13 β , 14 α -trihydroxy-3 β , 7 β -dibenzenzoyloxy-9 β , 15 β -diacetoxylatropha-5, 11-E-diene and the phytosterol β -sitosterol	HepG2, HCT-116, MCF-7 and PC-3	Only for compound: 1 α , 13 β , 14 α -trihydroxy-3 β , 7 β -dibenzenzoyloxy-9 β , 15 β -diacetoxylatropha-5, 11-E-diene, HepG2 (12.99 $\pm$ 0.9 μ M), HCT-116 (18.63 $\pm$ 1.4 μ M), MCF-7 (24.40 $\pm$ 1.9 μ M), and PC-3 (37.12 $\pm$ 2.3 μ M)	Cytotoxic	[164]
<i>Euphorbia umbellata</i> (Pax) Bruyns	Stems and leaves	Methanol	3,4,12,13-tetraacetylphorbol-20-phenylacetate	U251, MCF-7, NCI-ADR/RES, 786-0, NCI-H460, HT29, and K562	U251 (25.2 mg/mL), MCF-7 (>250 mg/mL), NCI-ADR/RES (>250 mg/mL), 786-0 (24.1 mg/mL), NCI-H460 (31.1 mg/mL), HT29 (>250 mg/mL), and K563 (65.3 mg/mL)	Cytotoxic	[127]
<i>Excoecaria agallocha</i> L.	Leaves and twigs	Ethanol	excagallonoid A	RKO	RKO (8.7 $\pm$ 1.98 μ M)	Cytotoxic	[193]
<i>Excoecaria agallocha</i> L.	Leaves and twigs	Ethanol	2-hydroxy-atis-1,16-diene-3,14-dione	RKO	RKO (2.6 $\pm$ 2.81 μ M)	Cytotoxic	[193]
<i>Jatropha gossypifolia</i> L.	Stem bark	-	Jatrophone	HepG2, WiDr, HeLa, and AGS	HepG2 (3.2 μ M), WiDr (8.97 μ M), HeLa (5.13), and AGS (2.5 μ M)	Cytotoxic	[194]
<i>Jatropha gossypifolia</i> L.	Branches and leaves	Ethanol	Jatrogrossidione	RKO	2.6 μ M	Cytotoxicity Apoptosis associated with G2/M-phase cell cycle arrest.	[58]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Jatropha tanjorensis</i> J.L.Ellis & Saroja	Leaves	Hexane, chloroform and methanol	R (+) 4-hydroxy-2-pyrrolidinone	HEP-2, B16F10, A549, and NRK 49F	HEP-2 ($42.26 \pm 0.03 \mu\text{g/mL}$), B16F10 ($44.56 \pm 0.02 \mu\text{g/mL}$), A549 ($48.26 \pm 0.03 \mu\text{g/mL}$), and NRK 49F ($47.28 \pm 0.03 \mu\text{g/mL}$).	Cytotoxic	[195]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	macabarteбенes A	MCF7, HeLa, A549, and PC3	MCF7 ($0.68 \pm 0.01 \mu\text{M}$), HeLa ($0.60 \pm 0.01 \mu\text{M}$), A549 ($0.79 \pm 0.01 \mu\text{M}$), and PC3 ($0.66 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	macabarteбенes B	MCF7, HeLa, A549, and PC3	MCF7 ($0.71 \pm 0.02 \mu\text{M}$), HeLa ($0.72 \pm 0.01 \mu\text{M}$), A549 ($0.74 \pm 0.01 \mu\text{M}$), and PC3 ($0.69 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	macabarteбенes C	MCF7, HeLa, A549, and PC3	MCF7 ($1.73 \pm 0.01 \mu\text{M}$), HeLa ($1.67 \pm 0.01 \mu\text{M}$), A549 ($1.81 \pm 0.00 \mu\text{M}$), and PC3 ($1.61 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	vedelianin	MCF7, HeLa, A549, and PC3	MCF7 ($0.32 \pm 0.03 \mu\text{M}$), HeLa ($0.51 \pm 0.01 \mu\text{M}$), A549 ($0.54 \pm 0.02 \mu\text{M}$), and PC3 ($0.39 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	schweinfurthin G	MCF7, HeLa, A549, and PC3	MCF7 ($0.95 \pm 0.02 \mu\text{M}$), HeLa ($1.18 \pm 0.01 \mu\text{M}$), A549 ($1.10 \pm 0.09 \mu\text{M}$), and PC3 ($0.91 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	8-prenylkaempferol	MCF7, HeLa, A549, and PC3	MCF7 ($6.22 \pm 0.13 \mu\text{M}$), HeLa ($6.88 \pm 0.16 \mu\text{M}$), A549 ($6.61 \pm 0.21 \mu\text{M}$), and PC3 ($6.53 \pm 0.11 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	mappain	MCF7, HeLa, A549, and PC3	MCF7 ($0.71 \pm 0.02 \mu\text{M}$), HeLa ($0.71 \pm 0.01 \mu\text{M}$), A549 ($0.81 \pm 0.02 \mu\text{M}$), and PC3 ($0.77 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	brousoflavonol F	MCF7, HeLa, A549, and PC3	MCF7 ($4.13 \pm 0.00 \mu\text{M}$), HeLa ($4.10 \pm 0.01 \mu\text{M}$), A549 ($3.83 \pm 0.01 \mu\text{M}$), and PC3 ($3.99 \pm 0.01 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga barteri</i> Müll.Arg.	Leaves	N-hexane, dichloromethane and methanol	isomacarangin	MCF7, HeLa, A549, and PC3	MCF7 ($8.43 \pm 0.26 \mu\text{M}$), HeLa ($8.49 \pm 0.21 \mu\text{M}$), A549 ($8.72 \pm 0.21 \mu\text{M}$), and PC3 ($8.5 \pm 0.31 \mu\text{M}$)	Cytotoxic	[196]
<i>Macaranga gigantea</i> (Rchb.f. & Zoll.) Müll.Arg.	Leaves	Methanol	Glyasperin	P-388	P-388 ($3.44 \mu\text{g/mL}$)	Cytotoxic	[197]
<i>Macaranga gigantea</i> (Rchb.f. & Zoll.) Müll.Arg.	Leaves	Methanol	Meliternatin	P-388	P-388 ($30.04 \mu\text{g/mL}$)	Cytotoxic	[197]
<i>Macaranga gigantifolia</i> Merr.	Leaves	Methanol, ethyl acetate fraction	Apigenin	P-388	P-388 ($14.13 \mu\text{g/mL}$)	Cytotoxic	[64]
<i>Macaranga hispida</i> (Blume) Mull.Arg	Leaves	Methanol	5,7,3',4'-tetrahydroxy-6- geranyl flavonol	P388	P388 ($0.22 \mu\text{g/mL}$)	Cytotoxic	[19]
<i>Macaranga hispida</i> (Blume) Mull.Arg	Leaves	Methanol	kaemferol 7-O- β -glucoside	P388	P388 ($101.5 \mu\text{g/mL}$)	Cytotoxic	[19]
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	kurzphenol A	HepG2	HepG2 ($30.14 \mu\text{g/mL}$)	Cytotoxic	[198]
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	kurzphenol C	A549	A549 ($17.11 \mu\text{g/mL}$)	Cytotoxic	[198]
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	8-prenylnaringenin	A549	A549 ($9.76 \mu\text{g/mL}$)	Cytotoxic	[198]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	glepidotin B	A549	A549 (15.32 µg/mL)	Cytotoxic	[198]
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	acetyltractylodinol	A549	A549 (18.22 µg/mL)	Cytotoxic	[198]
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	blumenol A	A549	A549 (18.23 µg/mL)	Cytotoxic	[198]
<i>Macaranga kurzii</i> (Kuntze) Pax & K.Hoffm.	Twigs	Ethanol	alicylic acid	A549	A549 (12.01 µg/mL)	Cytotoxic	[198]
<i>Macaranga recurvata</i> Gage	Leaves	Methanol	Macarecurvatin A	MCF7 and MCF7/TAMR	MCF7 (5.26 µM) and MCF7/TAMR (5.66 µM)	Cytotoxic	[199]
<i>Macaranga recurvata</i> Gage	Leaves	Methanol	Macarecurvatin B	MCF7 and MCF7/TAMR	MCF7 (0.96 µM) and MCF7/TAMR (1.25 µM)	Cytotoxic	[199]
<i>Macaranga recurvata</i> Gage	Leaves	Methanol	6,8- diisoprenylaromadendrin	MCF7 and MCF7/TAMR	MCF7 (5.03 µM) and MCF7/TAMR (5.83 µM)	Cytotoxic	[199]
<i>Mallotus conspurcatus</i> Croizat	Aerial parts	Methanol	6-Prenylnaringenin	HeLa and A549	HeLa (30.12 ± 1.21 µM) and A549 (70.25 ± 0.89 µM)	Cytotoxic	[200]
<i>Mallotus conspurcatus</i> Croizat	Aerial parts	Methanol	8-Prenylnaringenin	HeLa and A549	HeLa (60.16 ± 0.91 µM) and A549 (99.36 ± 1.94 µM)	Cytotoxic	[200]
<i>Mallotus conspurcatus</i> Croizat	Aerial parts	Methanol	7-O-Methyl-8- prenylnaringenin	HeLa and A549	HeLa (45.03 ± 0.82 µM) and A549 (89.16 ± 0.61 µM)	Cytotoxic	[200]
<i>Mallotus conspurcatus</i> Croizat	Aerial parts	Methanol	7-O-Methyl-6- prenylnaringenin	HeLa and A549	HeLa (19.69 ± 0.65 µM) and A549 (55.26 ± 1.87 µM)	Cytotoxic	[200]
<i>Mallotus conspurcatus</i> Croizat	Aerial parts	Methanol	4'-O-Methyl-6- prenylnaringenin	HeLa and A549	HeLa (10.08 ± 1.06 µM) and A549 (47.26 ± 0.82 µM)	Cytotoxic	[200]
<i>Manniophyton fulvum</i> Müll.Arg.	Twigs	Methanol	Betulinic acid	HeLa	4% at 62.5 µg/mL	Cytotoxic	[201]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Mareya micrantha</i> Müll. Arg.	Leaves	Ethanol/Water	29-nor-2β,15α,20β- trihydroxy-16α-acetyl- 3,1,22-trioxo-cucurbita-4,23- diene	Hep3B	Hep3B (0.12 ± 0.05 μM)	Cytotoxic	[202]
<i>Mareya micrantha</i> Müll. Arg.	Leaves	Ethanol/Water	29-nor-2β,15α,20β- trihydroxy-16α-acetyl- 3,1,22-trioxo-cucurbita-4,23- diene 29-nor-1,2,3,4,5,10-dehydro- 3,15α,20β-trihydroxy-16α- acetyl-11,22-dioxo- cucurbita-23-ene 2-O-β-D-glucopyranoside	Hep3B	Hep3B (43.8 ± 5.7 μM)	Cytotoxic	[202]
<i>Mareya micrantha</i> Müll. Arg.	Leaves	Ethanol/Water	29-nor-2β,15α,20β- trihydroxy-16α-acetyl- 3,11,22 trioxo-cucurbita-4,23-diene 2-O-β-D glucopyranoside	Hep3B	Hep3B (>50 μM)	Cytotoxic	[202]
<i>Mareya micrantha</i> Müll. Arg.	Leaves	Ethanol/Water	dihydro-epi-isocucurbitacin D	Hep3B	Hep3B (18.2 ± 2.8 μM)	Cytotoxic	[202]
<i>Mareya micrantha</i> Müll. Arg.	Leaves	Ethanol/Water	tetrahydro-cucurbitacin I	Hep3B	Hep3B (14.9 ± 3.3 μM)	Cytotoxic	[202]
<i>Mareya micrantha</i> Müll. Arg.	Leaves	Ethanol/Water	cucurbitacin L	Hep3B	Hep3B (11.3 ± 6.2 μM)	Cytotoxic	[202]
<i>Margaritaria discoidea</i> (Baill.) G. L. Webste	Stem bark	Dichloromethane and methanol (1:1).	Securinine	OVCAR-8, A2780, and A2780cis	OVCAR-8 (16.2 ± 0.5 μM), A2780 (2.7 ± 0.7 μM), and A2780cis (6.5 ± 0.4 μM)	Cytotoxic	[203]
<i>Margaritaria discoidea</i> (Baill.) G. L. Webste	Stem bark	Dichloromethane and methanol (1:1).	Gallic acid	OVCAR-8, A2780, and A2780cis	OVCAR-8 (5.2 ± 0.1 μM), A2780 (6.2 ± 0.3 μM), and A2780cis (5.4 ± 0.3 μM)	Cytotoxic	[203]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Ricinodendron heudelotii</i> (Baill.) Heckel	Leaves	Ethanol	Corilagin	HL-60, SMMC-7721, A-549, MCF-7, and SW-480	HL-60 ($25.81 \pm 0.67 \mu\text{g/mL}$), MCF-7 ($33.18 \pm 0.76 \mu\text{g/mL}$), and SW-480 ($37.04 \pm 1.06 \mu\text{g/mL}$)	Cytotoxic	[204]
<i>Suregada zanzibariensis</i> Baill.	Stem bark	Dichloromethane/Methanol (1:1)	Mangiolide	TK10, UACC62, and MCF7	TK10 ($0.02 \mu\text{g/mL}$), UACC62 ($0.03 \mu\text{g/mL}$), and MCF7 ($0.05 \mu\text{g/mL}$)	Cytotoxic	[205]
<i>Suregada zanzibariensis</i> Baill.	Stem bark	Dichloromethane/Methanol (1:1)	Jolkinolide B	TK10, UACC62, and MCF8	TK10 ($3.31 \mu\text{g/mL}$), UACC62 ($0.94 \mu\text{g/mL}$), and MCF7 ($2.99 \mu\text{g/mL}$)	Cytotoxic	[205]
<i>Trewia nudiflora</i> L.	Fruits	Ethanol	N-methyltreflorine	HeLa, MV-4-11, MCF-7, and MCF-7/ADR	HeLa (0.54 nM), MV-4-11 (3.6 nM), MCF-7 (8.6 nM), and MCF-7/ADR (13 nM)	Cytotoxic and inhibited tubulin polymerization in vitro	[206]
<i>Trewia nudiflora</i> L.	Fruits	Ethanol	Methyltrewiasine	HeLa, MV-4-11, MCF-7, and MCF-7/ADR	HeLa (1.6 nM), MV-4-11 (3.1 nM), and MCF-7 (10 nM)	Cytotoxic and inhibited tubulin polymerization in vitro	[206]
<i>Trewia nudiflora</i> L.	Fruits	Ethanol	Treflorine	HeLa, MV-4-11, MCF-7, and MCF-7/ADR	HeLa (0.74 nM), MV-4-11 (0.12 nM), and MCF-7 (5.5 nM)	Cytotoxic	[206]
<i>Trewia nudiflora</i> L.	Fruits	Ethanol	Trenudine	HeLa, MV-4-11, MCF-7, and MCF-7/ADR	HeLa (0.41 nM), MV-4-11 (4.8 nM), MCF-7 (11 nM), and MCF-7/ADR (28)	Cytotoxic	[206]
<i>Trewia nudiflora</i> L.	Fruits	Ethanol	Colubrinol	HeLa, MV-4-11, MCF-7, and MCF-7/ADR	HeLa (0.28 nM), MV-4-11 (0.21 nM), and MCF-7 (3.2 nM)	Cytotoxic	[206]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Trigonostemon heterophyllus</i> Merr.	Stems and leaves	Ethanol	Trigoheterophines A	HL60, SMMC-7721, A-549, MCF-7, and SW480	HL60 ($0.58 \pm 0.06 \mu\text{M}$), SMMC-7721 ($1.42 \pm 0.07 \mu\text{M}$), A-549 ($3.18 \pm 0.11 \mu\text{M}$), MCF-7 ($0.28 \pm 0.02 \mu\text{M}$), and SW480 ($0.93 \pm 0.05 \mu\text{M}$)	Antiproliferative	[207]
<i>Trigonostemon heterophyllus</i> Merr.	Stems and leaves	Ethanol	Trigoheterophines B	HL60, SMMC-7721, A-549, MCF-7, and SW480	HL60 ($0.66 \pm 0.04 \mu\text{M}$), SMMC-7721 ($1.98 \pm 0.08 \mu\text{M}$), A-549 ($0.52 \pm 0.03 \mu\text{M}$), MCF-7 ($0.75 \pm 0.05 \mu\text{M}$), and SW480 ($2.08 \pm 0.11 \mu\text{M}$)	Antiproliferative	[207]
<i>Trigonostemon heterophyllus</i> Merr.	Stems and leaves	Ethanol	(Z, Z, E, E)-1, 4-epoxy-16-hydroxyheneicos-1, 3, 12, 14-tetraene	HL60, SMMC-7721, A-549, MCF-7, and SW480	HL60 ($2.12 \pm 0.10 \mu\text{M}$), SMMC-7721 ($3.26 \pm 0.09 \mu\text{M}$), A-549 ($2.20 \pm 0.07 \mu\text{M}$), MCF-7 ($1.68 \pm 0.06 \mu\text{M}$), and SW480 ($2.72 \pm 0.08 \mu\text{M}$)	Antiproliferative	[207]
<i>Trigonostemon heterophyllus</i> Merr.	Stems and leaves	Ethanol	(Z, Z, E, E, E)-1, 4-epoxy-16-hydroxyheneicos-1, 3, 12, 14, 18-pentaene	HL60, SMMC-7721, A-549, MCF-7, and SW480	HL60 ($3.98 \pm 0.12 \mu\text{M}$), SMMC-7721 ($1.42 \pm 0.07 \mu\text{M}$), A-549 ($3.18 \pm 0.11 \mu\text{M}$), MCF-7 ($0.45 \pm 0.05 \mu\text{M}$), and SW480 ($2.23 \pm 0.10 \mu\text{M}$)	Antiproliferative	[207]
<i>Trigonostemon heterophyllus</i> Merr.	Stems and leaves	Ethanol	2-(hexadecyl)furan	HL60, SMMC-7721, A-549, MCF-7, and SW480	HL60 ($2.07 \pm 0.06 \mu\text{M}$), SMMC-7721 ($1.83 \pm 0.03 \mu\text{M}$), A-549 ($4.86 \pm 0.10 \mu\text{M}$), MCF-7 ($1.78 \pm 0.06 \mu\text{M}$), and SW480 ($4.28 \pm 0.09 \mu\text{M}$)	Antiproliferative	[207]
<i>Trigonostemon heterophyllus</i> Merr.	Stems and leaves	Ethanol	2-(octadecyl)furan	HL60, SMMC-7721, A-549, MCF-7, and SW480	HL60 ($1.05 \pm 0.06 \mu\text{M}$), SMMC-7721 ($2.97 \pm 0.13 \mu\text{M}$), A-549 ($6.32 \pm 0.15 \mu\text{M}$), MCF-7 ($3.02 \pm 0.07 \mu\text{M}$), and SW480 ($12.06 \pm 0.11 \mu\text{M}$)	Antiproliferative	[207]

Table 2. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/ Compounds Identified in Extract	Cell Lines	IC50	Activity/ Mechanism/Effect	Ref.
<i>Trigonostemon xyphophylloides</i> (Croizat) L.K.Dai & T.L.Wu	Twigs	Ethanol	Trigoxyphin P	SPC-A-1, BEL-7402, SGC-7901, and K-562	SPC-A-1 (1.70 µM) and K-562 (2.24 µM)	Cytotoxic	[208]
<i>Trigonostemon xyphophylloides</i> (Croizat) L.K.Dai & T.L.Wu	Twigs	Ethanol	Trigoxyphin Q	SPC-A-1, BEL-7402, SGC-7901, and K-562	SPC-A-1 (1.42 µM), SGC-7901 (2.88 µM), and K-562 (0.37 µM)	Cytotoxic	[208]
<i>Trigonostemon xyphophylloides</i> (Croizat) L.K.Dai & T.L.Wu	Twigs	Ethanol	Trigoxyphin R	SPC-A-1, BEL-7402, SGC-7901, and K-562	SPC-A-1 (12.42 µM) and K-562 (17.18 µM)	Cytotoxic	[208]
<i>Trigonostemon xyphophylloides</i> (Croizat) L.K.Dai & T.L.Wu	Twigs	Ethanol	Trigoxyphin T	SPC-A-1, BEL-7402, SGC-7901, and K-562	SPC-A-1 (0.24 µM), BEL-7402 (3.89 µM), and SGC-7901 (5.59 µM)	Cytotoxic	[208]

3.4. Anticancer Effects of Euphorbiaceae Extracts and Pure Compounds In Vivo Studies

The anticancer effects of both extracts and pure compounds have been tested in vivo using various animal models. Many studies are available on in vivo experiments on a number of plants from the Euphorbiaceae family. de Abrantes et al. [209] showed that Tonantzitlone B (TNZ-B), a diterpene from *Stillingia loranthea*, exhibits antitumor activity (1.5 or 3 mg/kg i.p.) in a mouse model of Ehrlich ascites carcinoma. The LD50 was estimated to be approximately 25 mg/kg (i.p.). TNZ-B reduced Ehrlich tumor volume and the total number of viable tumor cells. In addition, TNZ-B reduced the density of peri-tumor microvessels, suggesting an anti-angiogenic effect. Gowrav Adiga et al. [210] investigated the anticancer activity of a methanol extract of the stem bark of *Croton oblongifolius* in Sprague-Dawley rats. The rats were tumor-induced with dimethylbenz(a)anthracene (DMBA), administered orally and intramammarily, and with a plant extract. After obtaining the appropriate tumor mass, the extract was administered to the rats by gavage at 200, 500, and 800 mg/kg, which showed a dose-dependent reduction in mammary tumor volume, as confirmed by histopathological observations in the treated groups. In other studies, using a mouse model of breast cancer induced by 4T1 cells, Majumder et al. [211] demonstrated the effect of *Ricinus communis* fruit extract, rich in ricinine, *p*-coumaric acid, epigallocatechin, and ricinoleic acid, on breast cancer progression in vivo. Tumors were induced in female Balb/c mice by injecting 4T1 cells subcutaneously into the mammary fat pad. After 10 days, one group of animals received 4 i.p. doses of the extract (5 mg/kg body weight), while the other group received only the vehicle (0.9% saline). The tumor in the control group continued to grow, and animals treated with the extract showed a significant reduction in tumor volume over time. Tumors removed from the animals after 22 days showed a reduction in tumor volume of over 88% compared to the control group. Table 3 shows in vivo studies.

Table 3. Cytotoxic properties of selected extracts or pure compounds from the Euphorbiaceae family against in vivo models.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Animal Model	Treatment	Activity/Mechanism/Effect	Ref.
<i>Cnidoscopus quercifolius</i> Pohl	Root bark	Methanol, chloroform fraction (Favelin rich fraction)	Favelin, Methyl-faveline, Deoxofavelin, Neofavelanone and Coumarin	Mice	250 and 500 mg/kg/day of favelin rich fraction.	Inhibition rates of tumor growth were 58.08 and 48.71% for the 250 mg/kg and 500 mg/kg treatment groups, respectively.	[212]
<i>Croton crassifolius</i> Geiseler	Roots	Ethanol	Penduliflaworosin	Mice and Rats	12.5–50 μ M	Exerts its anti-angiogenic effect via the VEGF receptor-2 signaling pathway	[213]
<i>Euphorbia fischeriana</i> S. + <i>Ziziphus jujuba</i> M.	-	Water	Jokinolide B and 2,4-dihydroxy-6-methoxy-acetophenone	Mice	2.5, 5.0, and 10.0 g/kg groups of ESZM extract	PI3k/Akt pathway regulation of apoptosis.	[214]
<i>Euphorbia helioscopia</i> L.	Whole plant	Ethyl acetate	-	Mice	50 μ g/mL, 100 μ g/mL, and 200 μ g/mL	Growth inhibition and Cyclin D1 protein expression decreased. Cell apoptosis by changing Bcl-2, Bax, and caspase-3 protein expressions.	[215]
<i>Euphorbia royleana</i> Boiss.	-	Hexane	-	Mice	10 mg/kg.	Tumor volumes were decreased. Necrotic areas in tumor tissue.	[216]

Table 3. Cont.

Name of the Species	Part of the Plant	Type of Extract	Class of Compounds/Compounds Identified in Extract	Animal Model	Treatment	Activity/Mechanism/Effect	Ref.
<i>Excoecaria agallocha</i> L.	Bark	Methanol	Quercetin-3-O-rutinoside, Quercetin 3-O- α -L-rhamnoside, Kaempferol-3-O-(2-O-acetyl- α -L-rhamnopyranoside), Kaempferol 3-O- α -L-rhamnopyranoside, Excoecarin A, Excoecarin G1, Excoecarin G2, Taraxerone, 3beta-[(2E,4E)-6-oxo-decadienoyloxy]-olean-12-ene, 2,3-secoatisane, Exoecarin B, Exoecarin C, Exoecarin D, Excoecarin E, Exoecarin F, Exoecarin H	Mice	Different doses up to 200 mg/kg	Normal hemaetological values.	[217]
<i>Tragia involucrata</i> L.	Whole plant	Ethanol	Phenylacetaldehyde- diethylacetal, Neophytadiene, (E)-Phytol, Ethyl palmitate, Phytol, Ethyl linolate, Ethyl elaidate, Linolenic acid, Ethyl octadecanoate, 2-Ethylhexyl phthalate, Squalene, Vitamin E, Clionasterol, Viminalol, Agathisflavone, Loquatoside, Leufolin A, Quercetin, Echinacin, Apigetrin, Cynaroside, 1,2,36-tetrakis-O-galloyl-B-D-glucose, Isoquercetin and Corilagin	Mice	200 mg/kg and 400 mg/kg	Reduction of tumors	[144]

Data not reported are represented by “-”.

3.5. Anticancer and Cytotoxic Effects of Nanoparticles Prepared from Euphobiaceae Extracts and Pure Compounds

Nanoparticles (NP) combined with extracts or pure compounds of plant origin are currently a well-known method for biological research and enhancing the effects of various substances in in vitro or in vivo models. In the research by Ghramh et al. [218], an ethanolic leaf extract of *Ricinus communis* with gold nanoparticles (AuNPs) demonstrated a cytotoxic effect. Studies have shown that the extract in combination with AuNPs has a stronger effect on HeLa and HepG2 lines than the *Ricinus communis* extract alone. In turn, Alqahtani et al. [219] also demonstrated a stronger cytotoxic effect of *Jatropha pelargonifolia* extract in combination with chitosan nanoparticles against human lung adenocarcinoma (A549) than the extract alone. The same author revealed that *Euphorbia retusa* with a combination of zinc oxide NPs (ZnONPs) exhibited cytotoxic activity against A549 ($IC_{50} = 22.3 \mu\text{g/mL}$), HepG2 ($IC_{50} = 25.6$), Huh-7 ($IC_{50} = 25.7$), MCF-7 ($IC_{50} = 37.7$), and MDA-MB-231 ($IC_{50} = 37$) [220]. Similarly, Aboulthana et al. [221] showed that the cytotoxic activity of *Croton tiglium* seed extract increased after the incorporation of ZnONPs against human colon cancer cells (CACO-2). Additionally, the extract combined with ZnONPs stopped the increase of CACO-2 in G2/M and increased the percentage of total apoptotic cells and necrosis. A novel approach using *Excoecaria agallocha* leaf extract for the synthesis of silver NPs (AgONPs) was demonstrated by Banerjee et al. [222]. AgONPs exerted initial cytotoxicity specifically against all experimental malignant cells (murine melanoma (B16F10), murine colon cancer (CT26), murine lung adenocarcinoma (3LL), and murine Ehrlich ascites carcinoma (EAC)), while sparing normal cell lines. Furthermore, both in vitro and ex vivo, AgONPs are equally effective in inducing apoptosis in all cancer cells. Studies on nanoparticles are presented in Table 4 below.

Table 4. Nanoparticles made using constituents from the plants of the Euphobiaceae family.

Tested Plant	Components	Type of Nanoparticles	Cells	Effect	Ref.
<i>Acalypha wilkesiana</i> Müll.Arg.	Flowers	Ag NPs	MCF-7 (4.00 $\mu\text{g/mL}$) and PC-3 (3.60 $\mu\text{g/mL}$)	Cytotoxicity	[223]
<i>Alchornea cordifolia</i> (Schumacher & Thonn.) Müll.Arg.	Leaves	CuO–ZnO, ZnO, and CuO NPs	HeLa treatment with 100 $\mu\text{g/mL}$ —CuO–ZnO (39.94 $\pm$ 5.01), ZnO (44.05 $\pm$ 0.91) and CuO NPs (63.64 $\pm$ 8.34)	Cytotoxicity	[224]
<i>Baliospermum montanum</i> (Willd.) Müll.Arg.	Roots	Nanoparticles	Aqueous NPs (22%) and Ethanol NPs (6%)	Cytotoxicity	[225]
<i>Croton sparsiflorus</i> Morong	Leaves	AuNPs	HepG2 (116.7 $\mu\text{g/mL}$)	Cytotoxicity	[226]
<i>Euphorbia dendroides</i> L.	Aerial parts	AuNPs	HepG2 (41.72 $\pm$ 1.26 mg/mL) and HCT-116 (44.96 $\pm$ 3.23 mg/mL)	Cytotoxicity	[227]
<i>Euphorbia heterophylla</i> L.	Leaves	rGO	A549 (297.81 mg/mL) and HepG2 (357.53 mg/mL)	Cytotoxicity	[228]
<i>Euphorbia peplus</i> L.	Leaves	AuNPs	HepG2 and Hela cells	Inhibitory effect	[229]
<i>Euphorbia royleana</i> Boiss.	Pulp	Ag NPs and Cu ₂ O NPs	HCT-116 Ag NPs (50.12 $\mu\text{g/mL}$) and Cu ₂ O NPs (61.93 $\mu\text{g/mL}$)	Cytotoxicity	[230]
<i>Excoecaria agallocha</i> L.	Leaves	AgNPs	1.00 lg/mL AgNPs in MCF-7 (8.00% viability)	Cytotoxicity	[231]

4. Potential Anticancer Mechanism of Action of Euphorbiaceae Extracts and Isolated Compounds

The cytotoxic activity reported by several authors in cancer cell lines treated with Euphorbiaceae extracts or isolated compounds may be due to different mechanisms of action. It is known that these extracts and compounds can activate the intrinsic pathway and the extrinsic pathway of apoptosis. The chemical structures of the main isolated compounds with anticancer activity and their underlying mechanisms of action are presented in Figure 2 below.

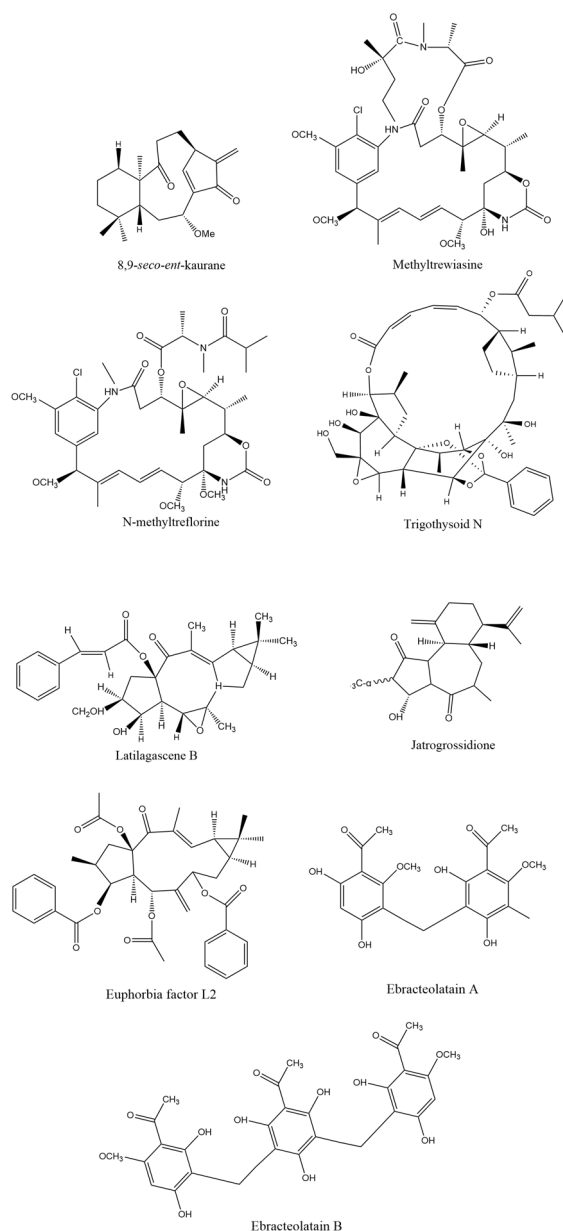


Figure 2. Chemical structure of isolated compounds from Euphorbiaceae plants with anticancer potential.

To activate the intrinsic pathway, the extract or compound could act as a ROS inducer. For example, *Croton gratissimus* Burch, *Drypetes sepiaria* (Wight & Arn.) Pax & K. Hoffm., *Euphorbia cactus* Ehrenb. ex Boiss., *Euphorbia hirta* L., *Euphorbia ingens* E. Mey. ex Boiss., *Euphorbia lathyris* L., *Excoecaria agallocha* L., *Euphorbia helioscopia* L., and *Ricinus communis* L. extracts [67,68,70,94,95,115,211,215], and the pure compounds Euphorbia Factor L2, Ebracteolatin A, and Ebracteolatin B [147,232], activate the intrinsic pathway at some

point. DNA damage promotes the liberation of cytochrome C by mitochondria. Then, cytochrome c and the APAF-1 (Apoptotic Protease Activating Factor-1) protein join together to activate casp-9. In this stage, it becomes a cascade release of different caspases activating each other from an inactive form to an active form and, at the end, activating casp-3, leading, finally, to apoptosis [67,70,94,136]. During this process, mitochondria also release SMAC/DIABLO (Second Mitochondria-Derived Activator of Caspase), an inhibitor of the XIAP (X-linked inhibitor of apoptosis) protein [233,234]. The caspases also cleave PARP (Poly ADP-Ribose Polymerase); this process is considered a hallmark of apoptosis [233].

The extrinsic pathway could be activated too, increasing the expression of casp-2/casp-8. For example, some authors have reported this activity with *Croton gratissimus* Burch, *Drypetes sepiaria* (Wight & Arn.) Pax & K.Hoffm, and *Euphorbia lathyris* L extracts [67,70,94].

The expression of p53 could also be affected by this family of plants. Several authors have reported higher levels of p53 after treatment with *Euphorbia pulcherrima* Willd. ex Klotzsch, *Euphorbia heterophylla* L., *Euphorbia ingens* E.Mey. ex Boiss, and *Excoecaria agallocha* L. in cancer cell lines [68,105,115]. p53 is a tumor-suppressive protein with the ability to induce cell death, including through apoptotic mechanisms and other transcription-dependent mechanisms or independent mechanisms [235]. Zhou et al. [232] also found that the activity of the pure compounds Ebracteolatin A and Ebracteolatin B increased p53 levels and decreased survivin levels. Survivin is a survival protein that inhibits caspases and blocks cell death [236]. Sultana et al. [68] found an altered expression on the levels of p21, the main target protein of p53. This protein, also known as CDKN1A (cyclin-dependent kinase inhibitor 1A), regulates the progression of the cell cycle with cyclin B. The balance of Bax/Bcl family protein expression can also be regulated through p53.

Changing the Bcl2/Bax (antiapoptotic/proapoptotic) protein balance could also be regulated by Euphorbiaceae plants through the STAT3 gene transcription pathway [146]. This balance is reported to be higher for Bax proteins than Bcl2 proteins when cancer cells are treated with *Croton tiglium* L., *Euphorbia cactus* Ehrenb. ex Boiss., *Euphorbia pulcherrima* Willd. ex Klotzsch, *Euphorbia heterophylla* L., *Euphorbia helioscopia* L., *Euphorbia hierosolymitana* Boiss., and *Ricinus communis* L. extracts [13,95,105,106,211,215]. Proapoptotic protein Bax is higher when cancer cells receive treatment with Trigothyoid N, Ebracteolatin A, and Ebracteolatin B also [146,232].

Li et al. [146] reported that cell invasion and migration could be regulated by Trigothyoid N through the FAK pathway. Once the compound inhibits FAK, the transcription of metalloproteinase (MMP)-2 and MMP-9 is downregulated, and the cells are not able to invade or migrate to other tissues.

The Euphorbiaceae extracts and isolated compounds could modulate the level of cytoplasmatic proteins, acting as inhibitors or downregulating/upregulating them. For example, Okpako et al. [115] reported a downregulation in androgen receptors after the treatment of prostate cancer cells DU-145 with *Euphorbia ingens* E.Mey. ex Boiss extract. Lei et al. [206] reported inhibited tubulin polymerization after treatment with Methyltrewhiasine and N-methyltrewhlorine. Tubulin polymerization/despolymerization plays a critical role in the cell cycle, and some important anticancer drugs are microtubule stabilizers of vegetal origin, for example, paclitaxel and docetaxel. Another example of protein regulation was reported by Shi et al. [237]. Using 8,9-seco-ent-kaurane diterpenoid isolated from *Croton kongensis* Gagnep in TNBC cells, the compound acted like an Akt inhibitor, which could induce apoptosis, autophagy, cell G2/M circle arrest, and inhibit cell migration. Latilagascene B was reported to be a p-glycoprotein inhibitor (Multidrug-resistant receptor) [145]. The cell cycle is regulated differently depending on the extract or pure compound used in cancer cells. In such a way that we can find stops in phase G1 [58,146], in phase G2/M [68,85], or in phase S [58] after the treatment with Euphorbiaceae. The anticancer potential mechanism of action of extracts and compounds from the Euphorbiaceae family is shown in Figure 3 below.

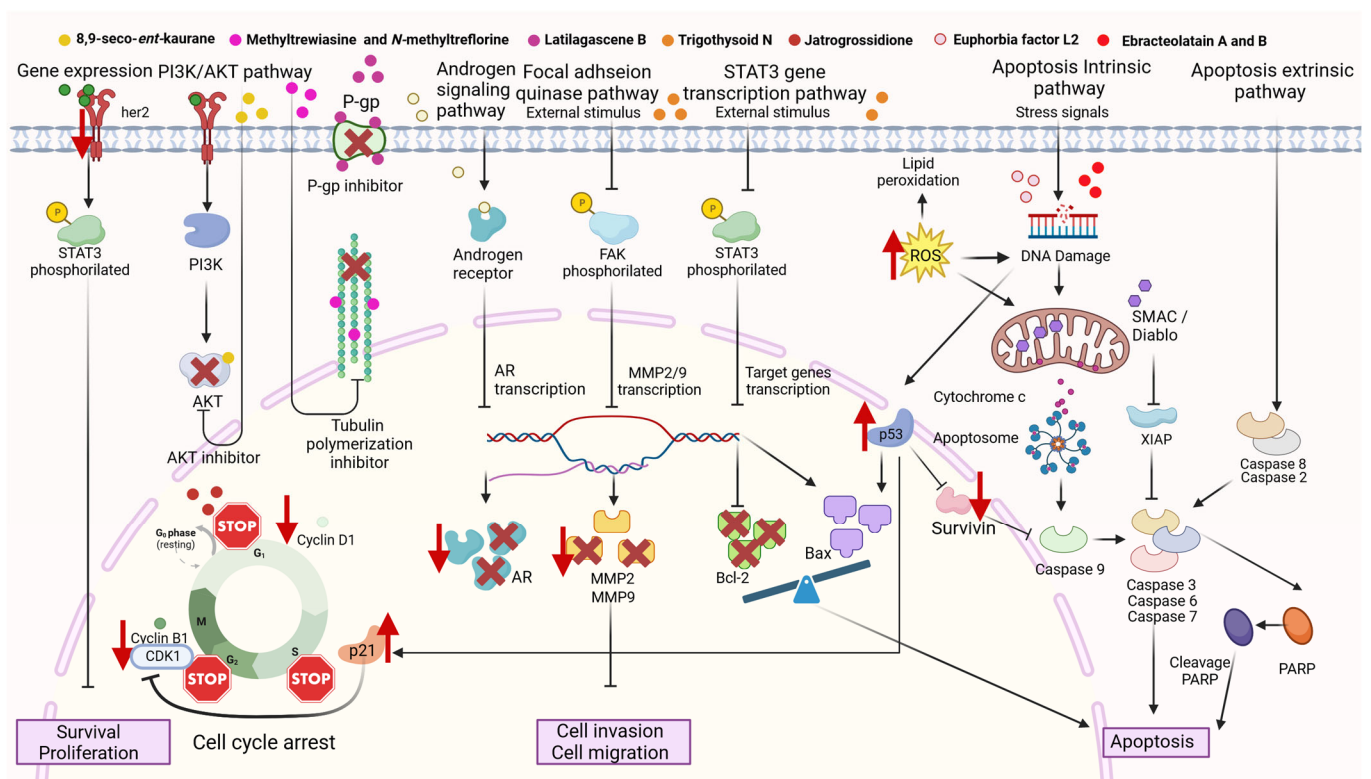


Figure 3. Potential anticancer mechanisms of action of Euphorbiaceae extracts and isolated compounds (created by BioRender).

5. Conclusions and Future Perspectives

Research into the anticancer properties of plants in the Euphorbiaceae family shows promising potential. Cancer continues to be a global health problem, both in terms of health and economy. The lack of effectiveness and selectivity of the currently most used anticancer drugs makes it necessary to continue the search for new drugs. Nature continues to be an inexhaustible source of useful molecules to cure or alleviate diseases. It is for this reason that it is necessary to continue investigating the molecules present in living beings that surround us, including plants, algae, fungi, and marine organisms. In this review, we emphasize the usefulness of the various compounds present in the Euphorbiaceae family. This family of vascular plants is a very diverse family, both in genera and species and in secondary metabolites. The presence of different types of terpenoids, flavonoids, and alkaloids with cytotoxic activity against cancer makes us think about the number of possibilities that these compounds could offer alone or through the synthesis of different derivatives from them. All these active compounds represent an opportunity for the treatment of cancer; however, from a future perspective, it is necessary to continue researching all these molecules. Greater funding is necessary for the research of new molecules to be able to advance them more quickly from the process of discovery to the regulation and approval process for the market and patients. In this context, preclinical and clinical studies in animal models, followed by human clinical trials to evaluate their efficacy, safety, and dosage in different types of cancer, seem to play a very important role in this field.

Author Contributions: Conceptualization, V.J.-G. and P.S.; methodology, V.J.-G., P.S. and T.K.; investigation, V.J.-G., P.S. and T.K.; writing—original draft preparation, V.J.-G., P.S., T.K., J.S. and J.P.; writing—review and editing, V.J.-G., P.S., T.K., J.P., J.S. and P.R.; visualization, V.J.-G.; supervision, P.S. and P.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: The authors would like to express their gratitude to Alexander Harrison for reviewing scientific English.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

AgONPs	Silver oxide nanoparticles
Akt	Protein kinase B
APAF	Apoptotic Protease-Activating Factor-1
AuNPs	Gold nanoparticles
Bax	Apoptosis regulator or bcl-2-like protein 4
Bcl-2	B-cell lymphoma 2 protein
Casp-3	Caspase 3
Casp-6	Caspase 6
Casp-7	Caspase 7
Casp-8	Caspase 8
Casp-9	Caspase 9
CuONPs	Copper oxide nanoparticles
MAPK	Mitogen-activated protein kinase
MMP-2	Metalloproteinase 2
MMP-9	Metalloproteinase 9
NP	Nanoparticles
P53	Tumor protein P53
PARP	Poly ADP-Ribose Polymerase
PI3K	Phosphoinositide 3-kinases
ROS	Reactive oxygen species
SMAC/DIABLO	Second Mitochondria-Derived Activator of Caspase
STAT3	Signal transducer and activator of transcription 3
TNBC	Triple negative breast cancer
TNB-Z	Tonantzitlolone B
XIAP	X-linked inhibitor of apoptosis
ZnONPs	Zinc oxide nanoparticles

References

1. Sarkar, S.; Horn, G.; Moulton, K.; Oza, A.; Byler, S.; Kokolus, S.; Longacre, M. Cancer Development, Progression, and Therapy: An Epigenetic Overview. *Int. J. Mol. Sci.* **2013**, *14*, 21087–21113. [\[CrossRef\]](#)
2. Bremnes, R.M.; Camps, C.; Sirera, R. Angiogenesis in Non-Small Cell Lung Cancer: The Prognostic Impact of Neoangiogenesis and the Cytokines VEGF and BFGF in Tumours and Blood. *Lung Cancer* **2006**, *51*, 143–158. [\[CrossRef\]](#)
3. World Health Organization (WHO). *WHO-Cancer Report-2020-Global Profile*; World Health Organization (WHO): Geneva, Switzerland, 2020.
4. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* **2021**, *71*, 209–249. [\[CrossRef\]](#)
5. Voda, A.I.; Bostan, I. Public Health Care Financing and the Costs of Cancer Care: A Cross-National Analysis. *Cancers* **2018**, *10*, 117. [\[CrossRef\]](#)
6. Pucci, C.; Martinelli, C.; Ciofani, G. Innovative Approaches for Cancer Treatment: Current Perspectives and New Challenges. *Ecancermedicalscience* **2019**, *13*, 961. [\[CrossRef\]](#)
7. Newman, D.J.; Cragg, G.M. Natural Products as Sources of New Drugs over the Nearly Four Decades from 01/1981 to 09/2019. *J. Nat. Prod.* **2020**, *83*, 770–803. [\[CrossRef\]](#)
8. Cragg, G.M.; Newman, D.J. Plants as a Source of Anti-Cancer Agents. *J. Ethnopharmacol.* **2005**, *100*, 72–79. [\[CrossRef\]](#)
9. Kingston, D.G.I.; Newman, D.J. The Search for Novel Drug Leads for Predominately Antitumor Therapies by Utilizing Mother Nature's Pharmacophoric Libraries. *Curr. Opin. Drug Discov. Dev.* **2005**, *8*, 207–227.
10. Ernst, M.; Grace, O.M.; Saslis-Lagoudakis, C.H.; Nilsson, N.; Simonsen, H.T.; Rønsted, N. Global Medicinal Uses of Euphorbia L. (Euphorbiaceae). *J. Ethnopharmacol.* **2015**, *176*, 90–101. [\[CrossRef\]](#)
11. Saleh, Z.M.; Abdel Azeiz, A.Z.; Mehany, A.B.M.; El-Swaify, Z.A.S. Anticancer and Antimicrobial Activity of Jatropha's Leaves Extracts. *Egypt. J. Bot.* **2023**, *63*, 621–634. [\[CrossRef\]](#)
12. Prakash, E.; Gupta, D.K. Cytotoxic Activities of Extracts of Medicinal Plants of Euphorbiaceae Family Studied on Seven Human Cancer Cell Lines. *Univ. J. Plant Sci.* **2013**, *1*, 113–117. [\[CrossRef\]](#)

13. Li, C.; Wu, X.; Sun, R.; Zhao, P.; Liu, F.; Zhang, C. *Croton tiglium* Extract Induces Apoptosis via Bax/Bcl-2 Pathways in Human Lung Cancer A549 Cells. *Asian Pac. J. Cancer Prev.* **2016**, *17*, 4893–4898. [CrossRef]
14. Vasas, A.; Hohmann, J. Euphorbia Diterpenes: Isolation, Structure, Biological Activity, and Synthesis (2008–2012). *Chem. Rev.* **2014**, *114*, 8579–8612. [CrossRef]
15. Block, S.; Baccelli, C.; Tinant, B.; Van Meervelt, L.; Rozenberg, R.; Habib Jiwan, J.-L.; Llabrès, G.; De Pauw-Gillet, M.-C.; Quetin-Leclercq, J. Diterpenes from the Leaves of *Croton zambesicus*. *Phytochemistry* **2004**, *65*, 1165–1171. [CrossRef]
16. Kemboi, D.; Siwe-noundou, X.; Krause, R.W.M.; Langat, M.K.; Tembu, V.J. Euphorbia Diterpenes: An Update of Isolation, Structure, Pharmacological Activities and Structure—Activity Relationship. *Molecules* **2021**, *26*, 5055. [CrossRef]
17. Subramanian, S.S.; Nagarajan, S.; Sulochana, N. Flavonoids of the Leaves of *Jatropha gossypifolia*. *Phytochemistry* **1971**, *10*, 1690. [CrossRef]
18. Šmejkal, K. Cytotoxic Potential of C-Prenylated Flavonoids. *Phytochem. Rev.* **2014**, *13*, 245–275. [CrossRef]
19. Megawati, M.; Saepudin, E.; Hanafi, M.; Darmawan, A.; Lotulung, P.D.N. Identification and Bioactivity Studies of Flavonoid Compounds from *Macaranga hispida* (Blume) Mull.Arg. *Makara J. Sci.* **2015**, *19*, 96–100. [CrossRef]
20. Coelho, P.L.C.; Oliveira, M.N.; da Silva, A.B.; Pitanga, B.P.S.; Silva, V.D.A.; Faria, G.P.; Sampaio, G.P.; Costa, M.d.F.D.; Braga-de-Souza, S.; Costa, S.L. The Flavonoid Apigenin from *Croton betulaster* Mull Inhibits Proliferation, Induces Differentiation and Regulates the Inflammatory Profile of Glioma Cells. *Anticancer Drugs* **2016**, *27*, 960–969. [CrossRef]
21. Novello, C.R.; Marques, L.C.; Pires, M.E.; Kutschenco, A.P.; Nakamura, C.V.; Nocchi, S.; Sarragiotto, M.H.; Mello, J.C.P. Bioactive Indole Alkaloids from *Croton echioides*. *J. Braz. Chem. Soc.* **2016**, *27*, 2203–2209. [CrossRef]
22. Euphorbiaceae. *Plants of the World Online*; Kew Royal Botanical Garden: Richmond, UK, 2023.
23. GBIF. *Euphorbiaceae in Global Biodiversity Information Facility*; GBIF: Copenhagen, Denmark, 2023. [CrossRef]
24. Valdés, B.; Talavera, S.; Fernández-Galiano, E. (Eds.) *Flora Vascular de Andalucía Occidental*; Flora Vascular: Barcelona, Spain, 1987.
25. Blanca, G.; Cabezudo, B.; Cueto, M.; Salazar, C.; Morales Torres, C. (Eds.) *Flora Vascular de Andalucía Oriental*, 2nd ed.; Corregida y Aumentada; Conserjería de Medio Ambiente, Junta de Andalucía: Granada, Spain, 2011.
26. Cueto, M.; Melendo, M.; Giménez, E.; Fuentes, J.; Carrique, E.L.; Blanca, G. *First Updated Checklist of the Vascular Flora of Andalusia (S of Spain)*, *One of the Main*; Biodiversity Centres in the Mediterranean Basin: Auckland, New Zealand, 2018; Volume 339; ISBN 9781776703104.
27. FAO. *Save and Grow: Cassava*; FAO: Rome, Italy, 2013.
28. Mohidin, S.R.N.S.P.; Moshawih, S.; Hermansyah, A.; Asmuni, M.I.; Shafqat, N.; Ming, L.C. Cassava (*Manihot esculenta* Crantz): A Systematic Review for the Pharmacological Activities, Traditional Uses, Nutritional Values, and Phytochemistry. *J. Evid.-Based Integr. Med.* **2023**, *28*, 2515690X231206227. [CrossRef] [PubMed]
29. FAOSTAT Cassava. Available online: <https://www.fao.org/faostat/es/#data/qv> (accessed on 15 September 2023).
30. Marwat, S.K.; Rehman, F.; Khan, E.A.; Baloch, M.S.; Sci, P.J.P.; Sadiq, M.; Ullah, I.; Javaria, S.; Shaheen, S. *Ricinus communis*: Ethnomedicinal Uses and Pharmacological Activities. *Pak. J. Pharm. Sci.* **2017**, *30*, 1815–1827. [PubMed]
31. Johnson, W. Final Report on the Safety Assessment of *Ricinus communis* (Castor) Seed Oil, Hydrogenated Castor Oil, Glyceryl Ricinoleate, Glyceryl Ricinoleate SE, Ricinoleic Acid, Potassium Ricinoleate, Sodium Ricinoleate, Zinc Ricinoleate, Cetyl Ricinoleate, Ethyl Ricinoleate, Glycol Ricinoleate, Isopropyl Ricinoleate, Methyl Ricinoleate, and Octyldodecyl Ricinoleate. *Int. J. Toxicol.* **2007**, *26*, 31–77.
32. Kelly, A.J.; Kavanagh, J.; Thomas, J. Castor Oil, Bath and/or Enema for Cervical Priming and Induction of Labour. *Cochrane Database Syst. Rev.* **2013**, *2013*, CD0030992013. [CrossRef] [PubMed]
33. Elkousy, R.H.; Said, Z.N.A.; El-Baseer, M.A.A.; El Wafa, S.A.A. Antiviral Activity of Castor Oil Plant (*Ricinus communis*) Leaf Extracts. *J. Ethnopharmacol.* **2021**, *271*, 113878. [CrossRef] [PubMed]
34. Fitrandi, M.I.; Sutrisno; Marfu'ah, S. Physicochemical Properties and Antibacterial Activity of Castor Oil and Its Derivatives. In *Proceedings of the IOP Conference Series: Materials Science and Engineering, Malang, Indonesia, 2–3 November 2019*; Institute of Physics Publishing: Bristol, UK, 2020; Volume 833.
35. Ogunniyi, D.S. Castor Oil: A Vital Industrial Raw Material. *Bioresour. Technol.* **2006**, *97*, 1086–1091. [CrossRef] [PubMed]
36. Neuwinger, H.D. Plants Used for Poison Fishing in Tropical Africa. *Toxicon* **2004**, *44*, 417–430. [CrossRef]
37. Lai, X.Z.; Yang, Y.B.; Shan, X.L. The Investigation of Euphorbiaceous Medicinal Plants in Southern China. *Econ. Bot.* **2004**, *58*, S307–S320. [CrossRef]
38. Polygenis-bigendako, M.J.; Lejoly, J. Plantes employées dans le traitement des diarrhées en médecine traditionnelle au burundi occidental. *Bull. Soc. R. Bot. Belg./Bull. Van K. Belg. Bot. Ver.* **1989**, *122*, 87–97.
39. Gaioni, D.T. Medical Choices in a Philippine Highland Community. *Ethnomedical and Biomedical Dimensions of Bauko Clinical Reality. Anthropos* **2002**, *97*, 505–518.
40. Bard, C.L. *A Contribution to the History of Medicine in Southern California*; Hansebooks: Norderstedt, Germany, 1894.
41. Hargreaves, B.J. The Spurge of Botswana. *Botsw. Notes Rec.* **1991**, *23*, 115–130.
42. Safford, W.E. *The Useful Plants of the Island of Guam*; Bulletin (United States National Museum); U.S. Government Printing Office: Washington, DC, USA, 1905; ISBN 9780598582195.
43. Manandhar, N.P. An Inventory of Some Herbal Drugs of Myagdi District, Nepal. *Econ. Bot.* **1995**, *49*, 371–379. [CrossRef]
44. Zhan, Z.J.; Li, S.; Chu, W.; Yin, S. Euphorbia Diterpenoids: Isolation, Structure, Bioactivity, Biosynthesis, and Synthesis (2013–2021). *Nat. Prod. Rep.* **2022**, *39*, 2132–2174. [CrossRef] [PubMed]

45. Xu, Y.; Tang, P.; Zhu, M.; Wang, Y.; Sun, D.; Li, H.; Chen, L. Diterpenoids from the Genus *Euphorbia*: Structure and Biological Activity (2013–2019). *Phytochemistry* **2021**, *190*, 112846. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Moremi, M.P.; Makolo, F.; Viljoen, A.M.; Kamatou, G.P. A Review of Biological Activities and Phytochemistry of Six Ethnomedicinally Important South African Croton Species. *J. Ethnopharmacol.* **2021**, *280*, 114416. [\[CrossRef\]](#)
47. Zhu, Q.F.; Xu, G.B.; Liao, S.G.; Yan, X.L. Ent-Abietane Diterpenoids from *Euphorbia fischeriana* and Their Cytotoxic Activities. *Molecules* **2022**, *27*, 7258. [\[CrossRef\]](#)
48. Zhang, Z.; Zhu, P.; Ma, J.; Li, X.; Yuan, W. A New Diterpenoid with Cytotoxic Activities from the Roots of *Euphorbia fischeriana*. *Chem. Nat. Compd.* **2023**, *59*, 701–705. [\[CrossRef\]](#)
49. Wang, W.; Zhang, X.J. Cytotoxic Ent-Abietane Diterpenoids from the Leaves of *Croton lachnocarpus* Benth. *J. Asian Nat. Prod. Res.* **2023**, *25*, 309–315. [\[CrossRef\]](#)
50. Sabandar, C.W.; Ahmat, N.; Jaafar, F.M.; Sahidin, I. Medicinal Property, Phytochemistry and Pharmacology of Several *Jatropha* Species (Euphorbiaceae): A Review. *Phytochemistry* **2013**, *85*, 7–29. [\[CrossRef\]](#)
51. Yuan, H.-T.; Li, Q.-F.; Tian, T.; Zhang, C.-Y.; Huang, Z.-Q.; Fan, C.-X.; Mei, K.; Zhou, J.; Zhai, X.-X.; Li, S.-B.; et al. Lathyrane Diterpenoids from *Jatropha podagrica* and Their Antitumor Activities in Human Osteosarcoma Cells. *Nat. Prod. Res.* **2021**, *35*, 5089–5095. [\[CrossRef\]](#)
52. Kemboi, D.; Peter, X.; Langat, M.; Tembu, J. A Review of the Ethnomedicinal Uses, Biological Activities, and Triterpenoids of *Euphorbia* Species. *Molecules* **2020**, *25*, 4019. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Hassan, A.R.; Ashour, A.; Amen, Y.; Nagata, M.; El-Toumy, S.A.; Shimizu, K. A New Cycloartane Triterpene and Other Phytoconstituents from the Aerial Parts of *Euphorbia dendroides*. *Nat. Prod. Res.* **2022**, *36*, 828–836. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Ngadjui, B.T.; Folefoc, G.G.; Keumedjio, F.; Dongo, E.; Sondengam, B.L.; Connolly, J.D. Crotonadiol, a Labdane Diterpenoid from the Stem Bark of *Croton zambesicus*. *Phytochemistry* **1999**, *51*, 171–174. [\[CrossRef\]](#)
55. Kapingu, M.C.; Mbwapbo, Z.H.; Moshi, M.J.; Magadula, J.J. Brine Shrimp Lethality of Alkaloids from *Croton sylvaticus* Hoechst. *East Cent. Afr. J. Pharm. Sci.* **2012**, *15*, 35–37.
56. Prozesky, E.A. *Antiplasmodial-and Chloroquine Resistance Reversal Properties of a New Diterpene from Croton steenkampianus*; University of Pretoria (South Africa) ProQuest Dissertations Publishing: Ann Arbor, MI, USA, 2005.
57. Mohamed, I.E.; El Nur, E.B.E.; Choudhary, M.I.; Khan, S.N. Bioactive Natural Products from Two Sudanese Medicinal Plants *Diospyros mespiliformis* and *Croton zambesicus*. *Rec. Nat. Prod.* **2009**, *3*, 198–203.
58. Zhang, C.-Y.; Zhang, R.-R.; Zhu, J.Y. New Tetracyclic Triterpenoids from *Jatropha gossypifolia* Induce Cell-Cycle Arrest and Apoptosis in RKO Cells. *Fitoterapia* **2018**, *130*, 145–151. [\[CrossRef\]](#)
59. Cavalcante, N.B.; da Conceição Santos, A.D.; da Silva Almeida, J.R.G. The Genus *Jatropha* (Euphorbiaceae): A Review on Secondary Chemical Metabolites and Biological Aspects. *Chem. Biol. Interact.* **2020**, *318*, 108976. [\[CrossRef\]](#)
60. Salehi, B.; Iriti, M.; Vitalini, S.; Antolak, H.; Pawlikowska, E.; Kregiel, D.; Sharifi-Rad, J.; Oyeleye, S.I.; Ademiluyi, A.O.; Czopek, K.; et al. *Euphorbia*-Derived Natural Products with Potential for Use in Health Maintenance. *Biomolecules* **2019**, *9*, 337. [\[CrossRef\]](#)
61. Azizi, K.; Hamed, A.; Azarpira, N.; Hamed, A.; Shahini, M.; Pasharan, A. A New Cytotoxic Sesquiterpene Lactone from *Euphorbia microsphaera* Boiss against Human Breast Cancer (MCF-7) and Human Fibrosarcoma (HT1080) Cells. *Toxicon* **2021**, *202*, 60–66. [\[CrossRef\]](#)
62. Kopustinskiene, D.M.; Jakstas, V.; Savickas, A.; Bernatoniene, J. Flavonoids as Anticancer Agents. *Nutrients* **2020**, *12*, 457. [\[CrossRef\]](#)
63. Gálvez, J.; Crespo, M.E.; Jiménez, J.; Suárez, A.; Zarzuelo, A. Antidiarrhoeic Activity of Quercitrin in Mice and Rats. *J. Pharm. Pharmacol.* **1993**, *45*, 157–159. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Fajriah, S.; Darmawan, A. Apigenin, an Anticancer Isolated from *Macaranga gigantifolia* Leaves. *J. Trop. Life Sci.* **2016**, *6*, 7–9.
65. Mondal, A.; Gandhi, A.; Fimognari, C.; Atanasov, A.G.; Bishayee, A. Alkaloids for Cancer Prevention and Therapy: Current Progress and Future Perspectives. *Eur. J. Pharmacol.* **2019**, *858*, 172472. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Vendruscolo, I.; Venturella, S.R.T.; Bressiani, P.A.; Marco, I.G.; Novello, C.R.; Almeida, I.V.; Vicentini, V.E.P.; Mello, J.C.P.; Düsman, E. Cytotoxicity of Extracts and Compounds Isolated from *Croton echinoides* in Animal Tumor Cell (HTC). *Braz. J. Biol.* **2022**, *82*, e264356. [\[CrossRef\]](#)
67. Mesas, C.; Martínez, R.; Ortiz, R.; Galisteo, M.; López-Jurado, M.; Cabeza, L.; Perazzoli, G.; Melguizo, C.; Porres, J.M.; Prados, J. Antitumor Effect of the Ethanolic Extract from Seeds of *Euphorbia lathyris* in Colorectal Cancer. *Nutrients* **2021**, *13*, 566. [\[CrossRef\]](#)
68. Sultana, T.; Mitra, A.K.; Das, S. Evaluation of Anti-Cancer Potential of *Excoecaria agallocha* (L.) Leaf Extract on Human Cervical Cancer (SiHa) Cell Line and Assessing the Underlying Mechanism of Action. *Futur. J. Pharm. Sci.* **2022**, *8*, 3. [\[CrossRef\]](#)
69. Kwan, Y.P.; Saito, T.; Ibrahim, D.; Al-Hassan, F.M.S.; Oon, C.E.; Chen, Y.; Jothy, S.L.; Kanwar, J.R.; Sasidharan, S. Evaluation of the Cytotoxicity, Cell-Cycle Arrest, and Apoptotic Induction by *Euphorbia hirta* in MCF-7 Breast Cancer Cells. *Pharm. Biol.* **2016**, *54*, 1223–1236. [\[CrossRef\]](#)
70. Njoya, E.M.; Eloff, J.N.; McGaw, L.J. *Croton gratissimus* Leaf Extracts Inhibit Cancer Cell Growth by Inducing Caspase 3/7 Activation with Additional Anti-Inflammatory and Antioxidant Activities. *BMC Complement. Altern. Med.* **2018**, *18*, 305. [\[CrossRef\]](#)
71. Vargas-Madriz, Á.F.; Luzardo-Ocampo, I.; Moreno-Celis, U.; Roldán-Padrón, O.; Chávez-Servín, J.L.; Vergara-Castañeda, H.A.; Martínez-Pacheco, M.; Mejía, C.; García-Gasca, T.; Kuri-García, A. Comparison of Phytochemical Composition and Untargeted

- Metabolomic Analysis of an Extract from *Cnidoscopus aconitifolius* (Mill.) I. I. Johnst and *Porophyllum ruderale* (Jacq.) Cass. and Biological Cytotoxic and Antiproliferative Activity In Vitro. *Plants* **2023**, *12*, 1987. [\[CrossRef\]](#)
72. Al-Massarani, S.; El-Sayed, M.-I.K.; El-Shaibany, A. Antioxidant and Anti-Proliferative Activities of *Acalypha fruticosa*: Possible Elucidated Mechanism. *Pak. J. Pharm. Sci.* **2019**, *32*, 2041–2050.
 73. Chekuri, S.; Panjala, S.; Anupalli, R.R. Cytotoxic Activity of *Acalypha Indica* L. Hexane Extract on Breast Cancer Cell Lines (MCF-7). *J. Phytopharm.* **2017**, *6*, 264–268. [\[CrossRef\]](#)
 74. Guillén-Meléndez, G.A.; Villa-Cedillo, S.A.; Pérez-Hernández, R.A.; Castillo-Velázquez, U.; Salas-Treviño, D.; Saucedo-Cárdenas, O.; Montes-De-oca-luna, R.; Gómez-Tristán, C.A.; Garza-Arredondo, A.J.; Zamora-ávila, D.E.; et al. Cytotoxic Effect in Vitro of *Acalypha monostachya* Extracts over Human Tumor Cell Lines. *Plants* **2021**, *10*, 2326. [\[CrossRef\]](#) [\[PubMed\]](#)
 75. Radhakrishna, S. In-Vitro Anticancer Activity of *Baliospermum montanum* on T Cell Leukemia-Jurkat Cell Line and Human Breast Cancer-MCF-7. *IAETSD J. Adv. Sci. Appl. Sci.* **2018**, *5*, 173–179.
 76. Pipatrattanaseree, W.; Itharat, A.; Mukkasombut, N.; Saesiw, U. Potential in Vitro Anti-Allergic, Anti-Inflammatory and Cytotoxic Activities of Ethanolic Extract of *Baliospermum montanum* Root, Its Major Components and a Validated HPLC Method. *BMC Complement. Altern. Med.* **2019**, *19*, 45. [\[CrossRef\]](#) [\[PubMed\]](#)
 77. Adi, A.S.; Elya, B.; Hanafi, M. Antioxidant and Cytotoxic Bioassay on *Blumeodendron toxbrai* (Blume.) Stem Bark Hexane, Dichloromethane, and Methanolic Ekstract. *Pharmacogn. J.* **2021**, *13*, 139–141. [\[CrossRef\]](#)
 78. Ramadan Kamel, M.; Nafady, A.; Hassanein, A.; Ragaey, R.; Haggag, E. Phytochemical Investigation and Assessment of Antioxidant, Antimicrobial and Cytotoxic Activities of the Root Bark *Chrozophora oblongifolia* (Delile) Spreng. (Euphorbiaceae). *J. Adv. Pharm. Res.* **2019**, *3*, 200–208. [\[CrossRef\]](#)
 79. Sunil Kumar, K.; Rao, A.S. Evaluation of Hepatoprotective and Invitro Cytotoxic Activity of Leaves of *Chrozophora plicata* Linn. *Res. J. Pharmacol. Pharmacodyn.* **2017**, *9*, 19–26. [\[CrossRef\]](#)
 80. Ikpefan, E.O.; Ayinde, B.A.; Mudassir, A.; Farooq, A.D. Comparative in Vitro Assessment of the Methanol Extracts of the Leaf, Stem, and Root Barks of *Cnidoscopus aconitifolius* on Lung and Breast Cancer Cell Lines. *Turk. J. Pharm. Sci.* **2019**, *16*, 375–379. [\[CrossRef\]](#)
 81. Kumarasamy, K.P.; Nallaperumal, N.; Natarajan, C.; Nallamadan, J. An in vitro cytotoxicity study of *Cnidoscopus chayamansa* McVaugh on selected cell lines. *World J. Pharm. Pharm. Sci.* **2014**, *3*, 1110–1116.
 82. Sánchez-Aguirre, O.A.; Juárez-Aguilar, E.; Montoya-Hernández, E.L.; Vázquez-Hernández, M.; Colorado-Peralta, R.; Sánchez-Medina, A.; Márquez-López, M.E.; Hernández-Romero, D. Antioxidant Potential of *Cnidoscopus multilobus* (Pax) I.M. Johnst and Its Antiproliferative and Cytotoxic Effect on Cervical Cancer Cells. *Eur. J. Integr. Med.* **2022**, *53*, 102134. [\[CrossRef\]](#)
 83. Paredes, P.F.M.; De Moraes, S.M.; Brito, F.C.R.; Moura, L.F.W.G.; De Rodrigues, P.A.; Benjamin, S.R.; Magalhães, F.E.A.; Florean, E.O.P.T.; Guedes, M.I.F. Characterization of *Cnidoscopus quercifolius* Pohl Bark Root Extract and Evaluation of Cytotoxic Effect on Human Tumor Cell Lines. *Asian Pac. J. Trop. Biomed.* **2018**, *8*, 345–351. [\[CrossRef\]](#)
 84. Worarat, C.; Pompimon, W.; Udomputtimekakul, P.; Sukdee, S.; Sombutsiri, P.; Kuanmuang, N.; Suwan, I.; Khamyong, Y.; Suksabai, C.; Artkla, W.; et al. In Vitro Screening for Cytotoxic, Anti-Bacterial, Anti-HIV1-RT Activities and Chemical Constituents of *Croton fluviatilis*, *Croton acutifolius*, and *Croton thorelii*. *Nat. Prod. J.* **2021**, *12*, 92. [\[CrossRef\]](#)
 85. Bhavana, J.; Mk, K.; Sumathy, A. Cytotoxic and Pro-Apoptotic Activities of Leaf Extract of *Croton bonplandianus* Baill. against Lung Cancer Cell Line A549; NISCAIR-CSIR: New Delhi, India, 2016; Volume 54.
 86. Shantabi, L.; Jagetia, G.C.; Moirangthem, D.S.; Nongalleima, K. Anticancer Activity of an Ehnomedicinal Plant *Croton caudatus* Geiseler, Kam Sabut in Cultured HeLa Cells. *Biocatal. Agric. Biotechnol.* **2020**, *23*, 101500. [\[CrossRef\]](#)
 87. Rosangkima, G.; Jagetia, G. Anticancer, Antioxidant and Analgesic Properties of *Croton caudatus*. *Int. J. Curr. Res.* **2015**, *7*, 20640–20646.
 88. De, B.; Mitra, I.; Choudhury, M.D.; Paul, S.B.; Deb, L.; De, A.; Nath, R. In Vitro Cytotoxic Study of *Croton caudatus* Geiseler—A Traditionally Known Anticancerous Plant in North-East India. *Pleione* **2018**, *12*, 165. [\[CrossRef\]](#)
 89. Jéssica, d.A.G.S.; Gibbelly, C.d.S.; Marília, G.d.F.S.; Vanessa, F.d.S.; Jaciana, d.S.A.; Teresinha, G.d.S.; Sônia, P.L. Physicochemical Characteristics and Cytotoxic Effect of the Methanolic Extract of *Croton heliotropiifolius* Kunth (Euphorbiaceae). *Afr. J. Pharm. Pharmacol.* **2017**, *11*, 321–326. [\[CrossRef\]](#)
 90. Ernandes, P.A.D.S.; Silva, J.C.P.D.; Sales, D.L.; Ribeiro, P.R.V.; de Brito, E.S.; Kerntopf, M.R.; Delmondes, G.D.A.; Pinheiro, J.C.A.; Salazar, G.J.T.; Batista, F.L.A.; et al. Chemical Constituents and Biological Activities of *Croton heliotropiifolius* Kunth. *Antibiotics* **2021**, *10*, 1074. [\[CrossRef\]](#)
 91. Yeboah, K.O.; Emosivbe, M.; Ntim, E.A.; Sam, G.H.; Ainooson, G. The Antiproliferative, Antimigratory and Anticlonogenic Effects of *Croton membranaceus* Müll. Arg. (Euphorbiaceae) Hydroethanolic Root Extract in Human 22Rv1 Castration-Resistant Prostate Cancer Cells. *Acta Pharm. Sci.* **2023**, *61*, 121–140. [\[CrossRef\]](#)
 92. Motta, L.B.; Furlan, C.M.; Santos, D.Y.A.C.; Salatino, M.L.F.; Negri, G.; de Carvalho, J.E.; Monteiro, P.A.; Ruiz, A.L.T.G.; Caruzo, M.B.; Salatino, A. Antiproliferative Activity and Constituents of Leaf Extracts of *Croton sphaerogynus* Baill. (Euphorbiaceae). *Ind. Crops Prod.* **2013**, *50*, 661–665. [\[CrossRef\]](#)
 93. Vieira, G.T.; Oliveira, T.T.d.; Monteiro, L.P.; Kanashiro, M.M.; Costa, M.R.d.; Pereira, W.L. Atividade citotóxica do extrato de *Croton urucurana* Baill contra linhagens de células leucêmicas humanas U937 e THP1. *Ciênc. Nat.* **2017**, *39*, 512. [\[CrossRef\]](#)

94. Gadamsetty, G.; Maru, S.; Tyagi, A.; Chakravarthula, S.N. Anti-Inflammatory, Cytotoxic and Antioxidant Effects of Methanolic Extracts of *Drypetes sepiaria* (Euphorbiaceae). *Afr. J. Tradit. Complement. Altern. Med. AJTCAM/Afr. Netw. Ethnomed.* **2013**, *10*, 274–282. [\[CrossRef\]](#) [\[PubMed\]](#)
95. Al-Hamoud, G.A.; Fantoukh, O.I.; Amina, M.; Nasr, F.A.; Al Musayeib, N.M.; Ahmed, M.Z.; Noman, O.M.; Al-Sharidah, R.E.; Alasmari, F.; Alqahtani, A.S. Unprecedented Insights on Chemical and Biological Significance of *Euphorbia cactus* Growing in Saudi Arabia. *Plants* **2022**, *11*, 681. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Bano, S.; Siddiqui, B.S.; Farooq, A.D.; Begum, S.; Siddiqui, F.; Kashif, M. In vitro growth inhibition and cytotoxicity of *Euphorbia caducifolia* against four human cancer cell lines and its phytochemical characterisation. *Nat. Prod. Res.* **2017**, *31*, 2936–2940. [\[CrossRef\]](#) [\[PubMed\]](#)
97. Rédei, D.; Kúsz, N.; Szabó, M.; Pinke, G.; Zupkó, I.; Hohmann, J. First Phytochemical Investigation of Secondary Metabolites of *Euphorbia davidii* Subils. and Antiproliferative Activity of Its Extracts Short Communication. *Acta Biol. Hung.* **2015**, *66*, 464–467. [\[CrossRef\]](#) [\[PubMed\]](#)
98. Ali, F.T.; Yousef, A.K.; Ahmed, F.A.; Elgneady, F.M.; El-Adl, K.; Elhady, M.M. In Silico ADMET, Docking, Anti-Proliferative and Antimicrobial Evaluations of Ethanolic Extract of *Euphorbia dendroides* L. *S. Afr. J. Bot.* **2022**, *150*, 607–620. [\[CrossRef\]](#)
99. Ikpefan, E.O.; Ayinde, B.A.; Mudassar, A.; Farooq, A.D. In Vitro Antiproliferative and Antioxidant Assessment of the Extract and Partitioned Fractions of Leaves of *Euphorbia graminea* Jacq. (Euphorbiaceae). *Niger. J. Pharm.* **2021**, *55*, 26–31. [\[CrossRef\]](#)
100. Kemboi, D.; Langat, M.K.; Siwe-Noundou, X.; Krause, R.W.M.; Isaacs, M.L.; Tembu, V.J. In Vitro Antibacterial and Cytotoxic Effects of *Euphorbia grandicornis* Blanc Chemical Constituents. *BMC Complement. Med. Ther.* **2022**, *22*, 90. [\[CrossRef\]](#)
101. Magozwi, D.K.; Peter, X.; Langat, M.K.; Mhlanga, R.; Vukea, N.; Mare, J.A.d.L.; Siwe-Noundou, X.; Krause, R.W.M.; Tembu, V.J. In Vitro Cytotoxic Effects of Chemical Constituents of *Euphorbia grandicornis* Blanc against Breast Cancer Cells. *Sci. Afr.* **2021**, *14*, e01002. [\[CrossRef\]](#)
102. Radi, M.H.; El-Shiekh, R.A.; El-Halawany, A.M.; Al-Abd, A.M.; Abdel-Sattar, E. In Vitro Cytotoxic Study of *Euphorbia grantii* Oliv. Aerial Parts against MCF-7 and MCF-7ADRBreast Cancer Cell Lines: A Bioactivity-Guided Isolation. *ACS Omega* **2023**, *8*, 18299–18305. [\[CrossRef\]](#)
103. Al-Robai, S.A.; Ahmed, A.A.; Ahmed, A.A.E.; Zabin, S.A.; Mohamed, H.A.; Alghamdi, A.A.A. Phenols, Antioxidant and Anticancer Properties of *Tagetes minuta*, *Euphorbia granulata* and *Galinsoga parviflora*: In Vitro and in Silico Evaluation. *J. Umm Al-Qura Univ. Appl. Sci.* **2023**, *9*, 15–28. [\[CrossRef\]](#)
104. Deveci, E.; Çayan, G.T.; Karakurt, S.; Duru, M.E. Anti-Colorectal Cancer Effects of Medicinal Plants: *Euphorbia helioscopia*, *Ferula elaeochytris*, and *Sideritis albiblora*. *Commagene J. Biol.* **2021**, *5*, 73–77. [\[CrossRef\]](#)
105. El-Hallouty, S.M.; Gohar, L.H.; Elgazzar, E.M. Enhancement of Radiation-Induced Cytotoxicity in Hepatocellular Carcinoma Cells by Some Plant Extracts In Vitro. *Egypt. J. Radiat. Sci. Appl.* **2022**. [\[CrossRef\]](#)
106. EL-Hallouty, S.; Batawi, A.; Shafi, M.; Rashwan, E.; Elhawary, E.; Abdelaal, N. Cytotoxicity of five plant extracts against different human cancer cell lines and their molecular mechanism. *Asian J. Pharm. Clin. Res.* **2020**, *13*, 194–198. [\[CrossRef\]](#)
107. Al-Saireh, Y.M.; Youssef, A.M.M.; Za'al Alsarayreh, A.; Al Hujran, T.A.; Al-Sarayreh, S.; Al-Shuneigat, J.M.; Alrawashdeh, H.M. Phytochemical and Anti-Cancer Properties of *Euphorbia hierosolymitana* Boiss. Crude Extracts. *J. Pharm. Pharmacogn. Res.* **2021**, *9*, 13–23. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Anitha, P.; Geegi, P.G.; Yogeswari, J.; Anthoni, S.A. In Vitro Anticancer Activity of Ethanolic Extract of *Euphorbia hirta* (L.). *Sci. Technol. Arts Res. J.* **2014**, *3*, 1. [\[CrossRef\]](#)
109. Rashmi; Gupta, P. In Vitro Anti-Inflammatory and Anti-Proliferative Potential of Roots Extract of *Euphorbia hirta* Linn. *J. Phytopharm.* **2021**, *10*, 7–9. [\[CrossRef\]](#)
110. Sharma, N.; Samarakoon, K.W.; Gyawali, R.; Park, Y.H.; Lee, S.J.; Oh, S.J.; Lee, T.H.; Jeong, D.K. Evaluation of the Antioxidant, Anti-Inflammatory, and Anticancer Activities of *Euphorbia hirta* Ethanolic Extract. *Molecules* **2014**, *19*, 14567–14581. [\[CrossRef\]](#)
111. Senniappan, P.; Srinivas, K.; Venkata, M.; Rao, B. In-Vitro Antioxidant and Cytotoxicity Activity of *Euphorbia hirta* Petroleum Ether, Chloroform Extracts. *Eur. Chem. Bull.* **2023**, *12*, 2949–2960.
112. Mali, P. Evaluation of Cytotoxicity of Aqueous Extract of *Euphorbia hirta* Using Human Colon Adenocarcinoma and Vero Cell Line. *Toxicol. Int.* **2017**, *24*, 17–21. [\[CrossRef\]](#)
113. De Sousa, A.S.; Fernandes, T.C.C.; Cardona, Y.T.; Almeida, P.M.D.; Marin-Morales, M.A.; Dos Santos, A.V.; Randau, K.P.; Benko-Iseppon, A.M.; Brasileiro-Vidal, A.C. Cytotoxic and Genotoxic Effects of Ethanolic Extract of *Euphorbia hyssopifolia* L. on HepG2 Cells. *J. Ethnopharmacol.* **2015**, *170*, 16–19. [\[CrossRef\]](#)
114. Alghamdi, A.A.A.; Ali, N.M.; Alam, M.M.; Nazreen, S.; Mahzari, A. HPLC Profile, Anticancer, Anti-Inflammatory and Antioxidant Activities of *Euphorbia inarticulata* Ethanolic Extract. *Adv. Pharmacol. Pharm.* **2023**, *11*, 156–167. [\[CrossRef\]](#)
115. Okpako, I.O.; Ng, F.A.; Kyama, C.M.; Oyem, J.C.; Njeru, S.N. Antiproliferative Activity of *Euphorbia ingens* Extract against Prostate Cancer Cell Line: An in Silico and in Vitro Analysis. *Preprints* **2023**. [\[CrossRef\]](#)
116. Wongprayoon, P.; Charoensuksai, P. Cytotoxic and anti-migratory activities from hydroalcoholic extract of *Euphorbia lactea* Haw. Against HN22 cell line. *Thai Bull. Pharm. Sci.* **2018**, *13*, 69–77.
117. Taş, A.; Şahin-Bölükbaşı, S.; Çevik, E.; Özmen, E.; Gümüş, E.; Siliğ, Y. An in Vitro Study of Cytotoxic Activity of *Euphorbia macroclada* Boiss on MCF-7 Cells. *Indian J. Pharm. Educ. Res.* **2018**, *52*, S119–S123. [\[CrossRef\]](#)
118. Ebrahim, H.Y.; Osman, S.A.; Haffez, H.R.; Hassan, Z.A. In-vitro screening of some plant extracts for their potential anticancer activity. *Afr. J. Tradit. Complement. Altern. Med.* **2020**, *17*, 1–8. [\[CrossRef\]](#)

119. Younus, M.; Hasan, M.M.; Hanif, M.; Sarwar, G.; Ahmad, K.; Ur Rehman, S.M.; Shirazi, J.H.; Jamil, Q.A.; Khan, K.R.; Syed, S.K.; et al. Botanical Features and Anti-Cancerous Potential of *Euphorbia nivulia* Buch.-Ham. *Trop. J. Nat. Product. Res.* **2021**, *5*, 1591–1596. [\[CrossRef\]](#)
120. Al-Yousef, H.M.; Alqahtani, A.S.; Ghani, A.S.A.; El-Toumy, S.A.; El-DougDoug, W.I.A.; Hassan, W.H.B.; Hassan, H.M. Nephroprotective, Cytotoxic and Antioxidant Activities of *Euphorbia paralias*. *Saudi J. Biol. Sci.* **2021**, *28*, 785–792. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Aslantürk, Ö.S.; Aşkın Çelik, T. Antioxidant, Cytotoxic and Apoptotic Activities of Extracts from Medicinal Plant *Euphorbia platyphyllos* L. *J. Med. Plants Res.* **2013**, *7*, 1293–1304. [\[CrossRef\]](#)
122. Aslantürk, Ö.S.; Yılmaz, E.Ş.; Aşkın Çelik, T.; Güzel, Y. Evaluation of the Antioxidant and Cytotoxic Potency of *Euphorbia rigida* and *Arbutus andrachne* Methanol Extracts in Human Hepatocellular Carcinoma Cell Lines In Vitro. *Beni Suef Univ. J. Basic. Appl. Sci.* **2021**, *10*, 51. [\[CrossRef\]](#)
123. de Souza, L.S.; Tosta, C.L.; de Oliveira, B.J.R.P.; Varricchio, M.C.B.N.; Kitagawa, R.R.; Filgueiras, P.R.; Kuster, R.M. Chemical Profile and Cytotoxic Evaluation of Aerial Parts of *Euphorbia tirucalli* L. on Gastric Adenocarcinoma (AGS Cells). *Nat. Prod. Res.* **2023**, *37*, 4267–4273. [\[CrossRef\]](#)
124. Archanjo, A.B.; De, F.; Careta, P.; Costa, A.V.; Nunes, C. Evaluation of cytotoxicity and expression of caspase-3 and p53 in HCT-116 cells of lineage treated with different extracts of *Euphorbia tirucalli* L. *Arch. Vet. Sci.* **2016**, *21*, 35–45. [\[CrossRef\]](#)
125. Al-Faifi, Z.I.A.; Masrahi, Y.S.; Aly, M.S.; Al-Turki, T.A.; Dardeer, T. Evaluation of Cytotoxic and Genotoxic Effects of *Euphorbia triaculeata* Forssk. Extract. *Asian Pac. J. Cancer Prev.* **2017**, *18*, 771–777. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Aliomrani, M.; Jafarian, A.; Zolfaghari, B. Phytochemical Screening and Cytotoxic Evaluation of *Euphorbia turcomanica* on HeLa and HT-29 Tumor Cell Lines. *Adv. Biomed. Res.* **2017**, *6*, 68. [\[CrossRef\]](#)
127. Campos, A.; Vendramini-Costa, D.B.; Longato, G.B.; Zermiani, T.; Ruiz, A.L.T.G.; De Carvalho, J.E.; Pandiella, A.; Filho, V.C. Antiproliferative Effect of *Synadenium grantii* Hook f. Stems (Euphorbiaceae) and a Rare Phorbol Diterpene Ester. *Int. J. Toxicol.* **2016**, *35*, 666–671. [\[CrossRef\]](#) [\[PubMed\]](#)
128. Kanunfre, C.C.; Leffers, T.; Cruz, L.S.; Luz, L.E.C.; Crisma, A.R.; Wang, M.; Avula, B.; Khan, I.A.; Beltrame, F.L. *Euphorbia umbellata* Bark Extracts—An in Vitro Cytotoxic Study. *Rev. Bras. Farmacogn.* **2017**, *27*, 206–213. [\[CrossRef\]](#)
129. Ali, M.M.; Khattab, M.; Mohamed, M.A.; Abdelsalam, R.M.; Mahmoud, K.; Soliman, S.; Barghash, M.F. The Cytotoxic Effect of *Synadenium grantii* Extract against Human Lung Carcinoma A549 Cells and Its Role in Improvement of Histopathological and Biomarkers Changes in Benzo(a)Pyrene-Induced Lung Cancer in Rats. *Int. J. Res. Pharm. Sci.* **2020**, *11*, 962–971. [\[CrossRef\]](#)
130. Periyasamy, K.; Pharm, I.J.; Sci, B.; Batsa, A.J.S. Anticancer activity of *Excoecaria agallocha* leaf extract in cell line model. *Int. J. Pharm. Biol. Sci.* **2013**, *3*, 392–398.
131. Mohapatra, R. The in Vitro Anti-Tumor Efficacy of Methanolic Leaf Extract of *Excoecaria agallocha* in Cancer Cell Lines of Different Tissue Origin. *Int. J. Innov. Res. Technol.* **2021**, *8*, 1348.
132. Hettihewa, L.M.; Munasinghe, M.M.A.B.; Bulugahapitiya, V.B.; Kihara, N. Dose dependent anti proliferative and cytotoxic effects of *Flueggea leucopyrus* Willd against human ovarian carcinoma; MTS and human telomerase enzyme inhibition. *EJBPS* **2015**, *2*, 14–18.
133. Vassallo, A.; Armentano, M.F.; Miglionico, R.; Caddeo, C.; Chirillo, C.; Gualtieri, M.J.; Ostuni, A.; Bisaccia, F.; Faraone, I.; Milella, L. *Hura crepitans* L. Extract: Phytochemical Characterization, Antioxidant Activity, and Nanoformulation. *Pharmaceutics* **2020**, *12*, 553. [\[CrossRef\]](#)
134. Mongalo, N.; Soyingbe, O.; Makhafola, T. Antimicrobial, Cytotoxicity, Anticancer and Antioxidant Activities of *Jatropha zeyheri* Sond. Roots (Euphorbiaceae). *Asian Pac. J. Trop. Biomed.* **2019**, *9*, 307–314. [\[CrossRef\]](#)
135. Cruz, A.J.D.P.; Raymundo, J.K.G.; Jacinto, S.D.; Tolentino, J.E.; Ipuan-Colet, L.A.D.G. Combining in vitro and in ovo assays to screen for anti-cancer and anti-angiogenic effects of the leaf extracts of *Mallotus cumingii* Müll.Arg. (Euphorbiaceae). *Malays. Appl. Biol.* **2022**, *51*, 73–82. [\[CrossRef\]](#)
136. Benny, B.; Krishna, A.S.; Samraj, S.; John, P.; Radhakrishnan, U. Cytotoxic and Antiproliferative Potential of Methanolic Extract of *Mallotus philippensis* in MCF-7 Cell Line. *J. Phytopharm.* **2022**, *11*, 60–63. [\[CrossRef\]](#)
137. Chen, X.-M.; Zhang, Z.-J.; Liu, A.-L.; Lv, F.-J.; Li, S.-F. Cholinesterase and Human Lung Cancer Cells (A-549) Inhibitory Activity of the Cassava Peel of Euphorbiaceae In Vitro. In *International Conference on Food Science and Engineering (ICFSE 2016)*; Trans Tech Publications Ltd.: Stafa-Zurich, Switzerland, 2016; ISBN 978-1-60595-390-8.
138. Al-Douri, N.; Al-Jaidi, B.A.; Hammad, H.M.; Shakya, A.K.; Alkhalilah, T.J.M.; Shilbayeh, S.A.R.; Ibrahim, A.A.; Venugopala, K.N.; Kamal, Y.T. Biological Evaluation of *Mercurialis annua* Extracts for Possible Antioxidant, Antiproliferative and Cytotoxic Activity. *Indian. J. Pharm. Educ. Res.* **2022**, *56*, S479–S486. [\[CrossRef\]](#)
139. Nascimento, A.K.L.; Melo-Silveira, R.F.; Dantas-Santos, N.; Fernandes, J.M.; Zucolotto, S.M.; Rocha, H.A.O.; Scortecchi, K.C. Antioxidant and Antiproliferative Activities of Leaf Extracts from *Plukenetia volubilis* Linneo (Euphorbiaceae). *Evid.-Based Complement. Altern. Med.* **2013**, *2013*, 950272. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Islam Shah, T.; Sharma, E.; Shah, G.A. Inhibitory Property of Aqueous Extract of *Ricinus communis* Leaves on Proliferation of Melanoma Treated against A375 Cell Lines. *World J. Pharm. Sci.* **2015**, *3*, 758–761.
141. Prakash, E.; Gupta, D.K. In Vitro Study of Extracts of *Ricinus communis* Linn on Human Cancer Cell Lines. *J. Med. Sci. Public Health* **2014**, *2*, 15–20.
142. Albino, R.C.; Antoniassi, R.; de Faria-Machado, A.F.; Ferraris, F.K.; Amendoeira, F.C.; Ramos, D.F.; Silva, P.E.A.; Leitão, S.G.; Oliveira, D.R. Traditional Detoxification of *Jatropha curcas* L. Seeds. *J. Ethnopharmacol.* **2019**, *241*, 111970. [\[CrossRef\]](#)

143. Florence, D. An Investigation into The Antineoplastic Properties of *Schinziophyton rautanenii* and *Colophospermum mopane*. Ph.D. Thesis, The University of Namibia, Windhuk, Namibia, 2008.
144. Menon, M.; Varghese, L. Evaluation of the Phytochemical Constituents and Tumor Reduction Potentials of *Tragia involucrata* Linn. *J. Appl. Biol. Biotechnol.* **2023**, *11*, 84–91. [\[CrossRef\]](#)
145. Euphorbia Species-Derived Diterpenes and Coumarins as Multidrug Resistance Modulators in Human Colon Carcinoma Cells. *Anticancer. Res.* **2016**, *36*, 2259–2264.
146. Li, Y.; Liu, Y.; Li, Y.; Liu, F.; Zhao, Y.; Xu, J.; Guo, Y. Trigothysoid, N. Inhibits Tumor Proliferation and Migration by Targeting Mitochondria and the STAT3/FAK Pathway. *Arab. J. Chem.* **2023**, *16*, 104930. [\[CrossRef\]](#)
147. Lin, M.; Tang, S.; Zhang, C.; Chen, H.; Huang, W.; Liu, Y.; Zhang, J. Euphorbia Factor L2 Induces Apoptosis in A549 Cells through the Mitochondrial Pathway. *Acta Pharm. Sin. B* **2017**, *7*, 59–64. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Fan, R.Z.; Chen, L.; Su, T.; Li, W.; Huang, J.L.; Sang, J.; Tang, G.H.; Yin, S. Discovery of 8,9-Seco-Ent-Kaurane Diterpenoids as Potential Leads for the Treatment of Triple-Negative Breast Cancer. *J. Med. Chem.* **2021**, *64*, 9926–9942. [\[CrossRef\]](#) [\[PubMed\]](#)
149. Wongprayoon, P.; Leelasart, S.; Jantam, J.; Pootaeng-on, Y.; Oekchuae, S.; Limpachayaporn, P.; Rayanil, K.O.; Charoensuksai, P. A Triterpenoid Friedelan-3 β -Ol Isolated from *Euphorbia lactea* Exhibited Cytotoxic Activity against HN22 Cells by Inducing an S-Phase Cell Cycle Arrest. *J. Appl. Pharm. Sci.* **2022**, *12*, 31–48. [\[CrossRef\]](#)
150. Li, H.H.; Qi, F.M.; Dong, L.L.; Fan, G.X.; Che, J.M.; Guo, D.D.; Zhang, Z.X.; Fei, D.Q. Cytotoxic and Antibacterial Pyran-2-One Derivatives from *Croton crassifolius*. *Phytochem. Lett.* **2014**, *10*, 304–308. [\[CrossRef\]](#)
151. Cui, J.J.; Ji, K.L.; Liu, H.C.; Zhou, B.; Liu, Q.F.; Xu, C.H.; Ding, J.; Zhao, J.X.; Yue, J.M. Cytotoxic Tiglane Diterpenoids from *Croton damayeshu*. *J. Nat. Prod.* **2019**, *82*, 1550–1557. [\[CrossRef\]](#)
152. Karina, P.; Uchôa, S.; Nunes Da Silva, J.; Silveira, R.; Anne, M.; Lima, S.; Braz-Filho, R. Trachylobane and kaurane diterpenes from *Croton floribundus* Spreng. *Quim. Nova* **2013**, *36*, 778–782.
153. Liu, C.-P.; Xu, J.-B.; Zhao, J.-X.; Xu, C.-H.; Dong, L.; Ding, J.; Yue, J.-M. Diterpenoids from *Croton laui* and Their Cytotoxic and Antimicrobial Activities. *J. Nat. Prod.* **2014**, *77*, 1013–1020. [\[CrossRef\]](#)
154. Zhao, B.Q.; Peng, S.; He, W.J.; Liu, Y.H.; Wang, J.F.; Zhou, X.J. Antitubercular and Cytotoxic Tiglane-Type Diterpenoids from *Croton tiglium*. *Bioorg Med. Chem. Lett.* **2016**, *26*, 4996–4999. [\[CrossRef\]](#)
155. Zhang, X.L.; Wang, L.; Li, F.; Yu, K.; Wang, M.K. Cytotoxic Phorbol Esters of *Croton tiglium*. *J. Nat. Prod.* **2013**, *76*, 858–864. [\[CrossRef\]](#)
156. Wang, J.F.; Yang, S.H.; Liu, Y.Q.; Li, D.X.; He, W.J.; Zhang, X.X.; Liu, Y.H.; Zhou, X.J. Five New Phorbol Esters with Cytotoxic and Selective Anti-Inflammatory Activities from *Croton tiglium*. *Bioorg Med. Chem. Lett.* **2015**, *25*, 1986–1989. [\[CrossRef\]](#) [\[PubMed\]](#)
157. Du, Q.; Zhao, Y.; Liu, H.; Tang, C.; Zhang, M.; Ke, C.; Ye, Y. Isolation and Structure Characterization of Cytotoxic Phorbol Esters from the Seeds of *Croton tiglium*. *Planta Med.* **2017**, *83*, 1361–1367. [\[CrossRef\]](#) [\[PubMed\]](#)
158. Cândido-Bacani, P.D.M.; Figueiredo, P.D.O.; Matos, M.D.F.C.; Garcez, F.R.; Garcez, W.S. Cytotoxic Orbitide from the Latex of *Croton urucurana*. *J. Nat. Prod.* **2015**, *78*, 2754–2760. [\[CrossRef\]](#) [\[PubMed\]](#)
159. Abreu, L.S.; do Nascimento, Y.M.; do Espírito-Santo, R.F.; Meira, C.S.; Santos, I.P.; Brandão, R.B.; Souto, A.L.; Guedes, M.L.S.; Soares, M.B.P.; Villarreal, C.F.; et al. Phenylpropanoids from *Croton velutinus* with Cytotoxic, Trypanocidal and Anti-Inflammatory Activities. *Fitoterapia* **2020**, *145*, 104632. [\[CrossRef\]](#) [\[PubMed\]](#)
160. Chen, D.; Cheng, X.; Sun, Y.; Liu, P. A new friedelane triterpenoid possessing cytotoxicity from the leaves and stems of *Drypetes hainanensis*. *Chem. Nat. Compd.* **2014**, *50*, 93–96. [\[CrossRef\]](#)
161. Abdel-Sattar, E.; Abou-Hussein, D.; Petereit, F. Chemical Constituents from the Leaves of Euphorbia Ammak Growing in Saudi Arabia. *Pharmacogn. Res.* **2015**, *7*, 14–17. [\[CrossRef\]](#)
162. Aljubiri, S.M.; Mahgoub, S.A.; Almansour, A.I.; Shaaban, M.; Shaker, K.H. Isolation of Diverse Bioactive Compounds from *Euphorbia balsamifera*: Cytotoxicity and Antibacterial Activity Studies. *Saudi J. Biol. Sci.* **2021**, *28*, 417–426. [\[CrossRef\]](#)
163. Shadi, S.; Saeidi, H.; Ghanadian, M.; Rahimnejad, M.R.; Aghaei, M.; Ayatollahi, S.M.; Iqbal Choudhary, M. New Macrocyclic Diterpenes from *Euphorbia connata* Boiss. with Cytotoxic Activities on Human Breast Cancer Cell Lines. *Nat. Prod. Res.* **2015**, *29*, 607–614. [\[CrossRef\]](#)
164. Foda, S.M.; Zaghloul, M.G.; Marzouk, A.M.; Shaker Mansour, E.-S. Phytochemical Investigation and Biological Studies of *Euphorbia tithymaloides* L. Family Euphorbiaceae. *Delta Univ. Sci. J.* **2022**, *5*, 260–271. [\[CrossRef\]](#)
165. Shamsabadipour, S.; Zarei, S.M.; Ghanadian, M.; Ayatollahi, S.A.; Rahimnejad, M.R.; Saeedi, H.; Aghaei, M. A New Taraxastane Triterpene from *Euphorbia denticulata* with Cytotoxic Activity against Prostate Cancer Cells. *Iran. J. Pharm. Res. IJPR* **2018**, *17*, 336.
166. Yuan, W.J.; Yang, G.P.; Zhang, J.H.; Zhang, Y.; Chen, D.Z.; Li, S.L.; Di, Y.T.; Hao, X.J. Three New Diterpenes with Cytotoxic Activity from the Roots of *Euphorbia ebracteolata* Hayata. *Phytochem. Lett.* **2016**, *18*, 176–179. [\[CrossRef\]](#)
167. Chen, G.; Ma, T.; Ma, Y.; Han, C.; Zhang, J.; Sun, Y. Chemical Composition, Anti-Breast Cancer Activity and Extraction Techniques of Ent-Abietane Diterpenoids from *Euphorbia fischeriana* Steud. *Molecules* **2022**, *27*, 4282. [\[CrossRef\]](#) [\[PubMed\]](#)
168. Zhong, N.F.; Huang, H.H.; Wei, J.C.; Yang, Y.C.; Gao, X.X.; Wei, X.Y.; Wang, A.H.; Jia, J.M. Euphorbiatnoids A–I: Diterpenoids from the Roots of *Euphorbia fischeriana* with Cytotoxic Effects. *Phytochemistry* **2022**, *203*, 113372. [\[CrossRef\]](#) [\[PubMed\]](#)
169. Meng, J.; Li, B.T.; Sheng, G.; Zhang, A.L.; Li, X.C.; Tian, J.M. Cytotoxic Diterpenoids from *Euphorbia fischeriana*. *Chem. Biodivers.* **2021**, *18*, e2000919. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Fu, X.; Yu, D.; Zhu, G.; Xu, J. Three New Abietane Diterpenoids from the Aerial Parts of *Euphorbia fischeriana* and Their Cytotoxic Effects. *Phytochem. Lett.* **2023**, *55*, 56–60. [\[CrossRef\]](#)

171. Zhou, M.; Ma, Q.; He, L.; Chen, Y.H.; Zhu, B.Y.; Wang, J.H.; Yang, Q.; Liu, S.; Ma, L.M. Cytotoxic Jatrophane Diterpenoids from the Aerial Parts of *Euphorbia helioscopia*. *J. Asian Nat. Prod. Res.* **2021**, *23*, 731–737. [\[CrossRef\]](#)
172. Lu, Y.B.; Luo, S.; Wang, Y.X.; Feng, Z.Y.; Gao, K.; Chen, J.J. Jatrophane Diterpenoids with Cytotoxic Activity from the Whole Plant of *Euphorbia helioscopia* L. *Phytochemistry* **2022**, *203*, 113420. [\[CrossRef\]](#)
173. Adedoyin, B.A. In Vitro Cytotoxicity of *Euphorbia heterophylla* against Human Cancer Cell Lines. *Afr. J. Eng. Environ. Res.* **2022**, *3*, 1.
174. Hu, R.; Sang, J.; Li, W.; Tian, Y.; Zou, M.F.; Tang, G.H.; Yin, S. Structurally Diverse Triterpenoids with Cytotoxicity from *Euphorbia hypericifolia*. *Fitoterapia* **2021**, *151*, 104888. [\[CrossRef\]](#)
175. Hegazy, M.E.F.; Hamed, A.R.; Ibrahim, M.A.A.; Talat, Z.; Reda, E.H.; Abdel-Azim, N.S.; Hammouda, F.M.; Nakamura, S.; Matsuda, H.; Haggag, E.G.; et al. Euphosantianane A–D: Antiproliferative Premyrsinane Diterpenoids from the Endemic Egyptian Plant *Euphorbia sanctae-catharinae*. *Molecules* **2018**, *23*, 2221. [\[CrossRef\]](#)
176. Jia, H.Y.; Liao, Z.X.; Liu, F.Y.; Wu, L.; Xu, C.; Zuo, B. A New Phenylpropanoid from the Roots of *Euphorbia nematocypha*. *Nat. Prod. Res.* **2015**, *29*, 650–655. [\[CrossRef\]](#) [\[PubMed\]](#)
177. Xu, C.; Jia, H.-Y.; Zuo, B.; Liao, Z.-X.; Ji, L.-J.; Sun, H.-F.; Wang, Q. Chemical Constituents of the Aerial Parts of *Euphorbia nematocypha*. *Nat. Prod. Commun.* **2016**, *11*, 1934578X1601100210. [\[CrossRef\]](#)
178. Song, N.; Zheng, X.; Wang, J.; Zhu, L.; Wang, C.; Cai, L.; Ding, Z. Cytotoxicity and Molecular-Docking Approach of a New Rosane-Type Diterpenoid from the Roots of *Euphorbia nematocypha*. *Front. Chem.* **2022**, *10*, 912738. [\[CrossRef\]](#) [\[PubMed\]](#)
179. Gao, Y.; Zhou, J.S.; Liu, H.C.; Zhang, Y.; Yin, W.H.; Liu, Q.F.; Wang, G.W.; Zhao, J.X.; Yue, J.M. Phonerilins A–K, Cytotoxic Ingenane and Ingol Diterpenoids from *Euphorbia neriifolia*. *Tetrahedron* **2022**, *123*, 132955. [\[CrossRef\]](#)
180. Ghanadian, M.; Saeidi, H.; Aghaei, M.; Rahiminejad, M.R.; Ahmadi, E.; Ayatollahi, S.M.; Choudhary, M.I.; Bahmani, B. New Jatrophane Diterpenes from *Euphorbia osyridea* with Proapoptotic Effects on Ovarian Cancer Cells. *Phytochem. Lett.* **2015**, *12*, 302–307. [\[CrossRef\]](#)
181. Wang, K.; Yu, H.; Wu, H.; Wang, X.; Pan, Y.; Chen, Y.; Liu, L.; Jin, Y.; Zhang, C. A New Casbane Diterpene from *Euphorbia pekinensis*. *Nat. Prod. Res.* **2015**, *29*, 1456–1460. [\[CrossRef\]](#) [\[PubMed\]](#)
182. Hou, P.; Zeng, Y.; Ma, B.; Bi, K.; Chen, X. A New Cytotoxic Cembrane Diterpene from the Roots of *Euphorbia pekinensis* Rupr. *Fitoterapia* **2013**, *90*, 10–13. [\[CrossRef\]](#) [\[PubMed\]](#)
183. Chen, Y.Y.; Zeng, X.T.; Xu, D.Q.; Yue, S.J.; Fu, R.J.; Yang, X.; Liu, Z.X.; Tang, Y.P. Pimarane, Abietane, and Labdane Diterpenoids from *Euphorbia pekinensis* Rupr. and Their Anti-Tumor Activities. *Phytochemistry* **2022**, *197*, 113113. [\[CrossRef\]](#)
184. Ismail, M.; Ahmed, M.H.; Zaki, M.; Moawad, A.; Mohammed, R. Colorectal Cytotoxic Activities of Phenolic Constituents Isolated from *Euphorbia pseudocactus* Cultivated in Egypt. *J. Appl. Pharm. Sci.* **2022**, *12*, 97–102. [\[CrossRef\]](#)
185. Wu, S.; Gan, L.; Su, T.; Wei, X.; Yin, S. New Ingenane and Ingol Diterpenoids from *Euphorbia royleana*. *Nat. Prod. Res.* **2023**, *37*, 1130–1137. [\[CrossRef\]](#)
186. Banjar, M.F.S.; Mohamed, G.A.; Shehata, I.A.; Abdallah, H.M.; Shati, A.A.; Alfaifi, M.Y.; Elbehairi, S.E.I.; Koshak, A.E.; Ibrahim, S.R.M. Cycloschimperols A and B, New Cytotoxic Cycloartane Triterpenoids from *Euphorbia schimperii*. *Phytochem. Lett.* **2019**, *32*, 90–95. [\[CrossRef\]](#)
187. Aljubiri, S.M.; Mahmoud, K.; Mahgoub, S.A.; Almansour, A.I.; Shaker, K.H. Bioactive Compounds from *Euphorbia schimperiana* with Cytotoxic and Antibacterial Activities. *S. Afr. J. Bot.* **2021**, *141*, 357–366. [\[CrossRef\]](#)
188. Zolfaghari, B.; Yazdiniapour, Z.; Ghanadian, M.; Lanzotti, V. Cyclomyrsinane and Premyrsinane Diterpenes from *Euphorbia sogdiana* Popov. *Tetrahedron* **2016**, *72*, 5394–5401. [\[CrossRef\]](#)
189. Zhu, H.; Ren, X.; Huang, Y.; Su, T.; Yang, L. Chemical Constituents of *Euphorbia stracheyi* Boiss (Euphorbiaceae). *Metabolites* **2023**, *13*, 852. [\[CrossRef\]](#) [\[PubMed\]](#)
190. Ye, Y.; Liu, G.H.; Dawa, D.; Ding, L.S.; Cao, Z.X.; Zhou, Y. Cytotoxic Diterpenoids from the Roots of *Euphorbia stracheyi*. *Phytochem. Lett.* **2020**, *36*, 183–187. [\[CrossRef\]](#)
191. Rédei, D.; Kúsz, N.; Sători, G.; Kincses, A.; Spengler, G.; Burián, K.; Barina, Z.; Hohmann, J. Bioactive Segetane, Ingenane, and Jatrophane Diterpenes from *Euphorbia taur.* *Planta Med.* **2018**, *84*, 729–735. [\[CrossRef\]](#)
192. Silva, V.A.O.; Rosa, M.N.; Tansini, A.; Oliveira, R.J.S.; Martinho, O.; Lima, J.P.; Pianowski, L.F.; Reis, R.M. In Vitro Screening of Cytotoxic Activity of Euphol from *Euphorbia tirucalli* on a Large Panel of Human Cancer-Derived Cell Lines. *Exp. Ther. Med.* **2018**, *16*, 557–566. [\[CrossRef\]](#)
193. Liu, G.; Zhang, Z.; Wang, Y.; Li, X. Highly Oxygenated Ent-Atisane and Podocarpane Diterpenoids from *Excoecaria agallocha*. *Nat. Prod. Res.* **2022**, *36*, 3924–3930. [\[CrossRef\]](#)
194. Asep, S.; Hening, H.; Gema, S.P.; Gigih, S.; Widya, M.C. Sahidin Anticancer Activity of Jatrophane an Isolated Compound from *Jatropha gossypifolia* Plant against Hepatocellular Cancer Cell Hep G2 1886. *Biomed. Pharmacol. J.* **2017**, *10*, 667–673. [\[CrossRef\]](#)
195. Byrappa, M.; Viswanathan, G.; Doss, J.; Ananthi, J.; Rajan, N.; Raja, L.; Venkateshan, N. Cytotoxic Pyrrolidinone from Leaves of *Jatropha tanjorensis*. *Int. J. Res. Sci. Innov.* **2018**, *5*, 45–50.
196. Segun, P.A.; Ogbole, O.O.; Ismail, F.M.D.; Nahar, L.; Evans, A.R.; Ajaiyeoba, E.O.; Sarker, S.D. Bioassay-Guided Isolation and Structure Elucidation of Cytotoxic Stilbenes and Flavonols from the Leaves of *Macaranga barteri*. *Fitoterapia* **2019**, *134*, 151–157. [\[CrossRef\]](#) [\[PubMed\]](#)
197. Wanandri, R.; Rewa, S.; Saputri, R.D.; Tjahjandarie, S.; Tanjung, M. Cytotoxic Activity of Flavonols from *Macaranga gigantea*. *Pharm. J. Farm. Indones.* **2020**, *17*, 360–367.

198. Yang, D.S.; Wei, J.G.; Peng, W.B.; Wang, S.M.; Sun, C.; Yang, Y.P.; Liu, K.C.; Li, X.L. Cytotoxic Prenylated Bibenzyls and Flavonoids from *Macaranga Kurzii*. *Fitoterapia* **2014**, *99*, 261–266. [[CrossRef](#)] [[PubMed](#)]
199. Tanjung, M.; Tjahjandarie, T.S. Dihydroflavonols from the Leaves of *Macaranga recurvata* and Their Cytotoxic and Antioxidant Activities. *J. Chem. Pharm. Res.* **2014**, *6*, 90–95.
200. Zhang, Y.; Zhou, D.; Liu, W.; Li, C.; Hao, L.; Zhang, G.; Deng, S.; Yang, R.; Qin, J.; Li, J.; et al. Cytotoxic Activity and Related Mechanisms of Prenylflavonoids Isolated from *Mallotus conspurcatus* Croizat. *Chem. Biodivers.* **2019**, *16*, e1800465. [[CrossRef](#)]
201. Mbeunkeu, A.B.D.; Azebaze, A.G.B.; Tala, M.F.; Teinkela, J.E.M.; Noundou, X.S.; Krause, R.W.M.; Vardamides, J.C.; Laatsch, H. Three New Pentacyclic Triterpenoids from Twigs of *Manniophyton fulvum* (Euphorbiaceae). *Phytochem. Lett.* **2018**, *27*, 1–8. [[CrossRef](#)]
202. Toussaint-Douhoré, G.Y.; Soro, Y.; Ouédraogo, N.; Vaca-Garcia, C.; Koffi-Attoua, B.; Carraz, M. Liver Cancer Antiproliferative Activity of a New Nor-Cucurbitacin from *Mareya micrantha* Müll. Arg. *Fitoterapia* **2023**, *166*, 105471. [[CrossRef](#)]
203. Johnson-Ajinwo, O.R.; Richardson, A.; Li, W.W. Cytotoxic Effects of Stem Bark Extracts and Pure Compounds from *Margaritaria discoidea* on Human Ovarian Cancer Cell Lines. *Phytomedicine* **2015**, *22*, 1–4. [[CrossRef](#)]
204. Yakubu, O.F.; Adebayo, A.H.; Dokunmu, T.M.; Zhang, Y.J.; Iweala, E.E.J. Cytotoxic Effects of Compounds Isolated from *Ricinodendron heudelotii*. *Molecules* **2019**, *24*, 145. [[CrossRef](#)]
205. Mangisa, M.; Tembu, V.J.; Fouche, G.; Nthambeleni, R.; Peter, X.; Langat, M.K. Ent-Abietane Diterpenoids from *Suregada zanzibariensis* Baill. (Euphorbiaceae), Their Cytotoxic and Anticancer Properties. *Nat. Prod. Res.* **2019**, *33*, 3240–3247. [[CrossRef](#)]
206. Lei, C.; Li, Y.N.; Li, J.N.; Zhou, Y.B.; Cui, M.J.; Fu, K.C.; Li, J.; Hou, A.J. Two New Cytotoxic Maytansinoids Targeting Tubulin from *Trewia nudiflora*. *Planta Med.* **2021**, *88*, 678–684. [[CrossRef](#)] [[PubMed](#)]
207. Liu, Y.P.; Hu, S.; Wen, Q.; Ma, Y.L.; Jiang, Z.H.; Tang, J.Y.; Fu, Y.H. Novel γ -Lactone Derivatives from *Trigonostemon heterophyllus* with Their Potential Antiproliferative Activities. *Bioorg. Chem.* **2018**, *79*, 107–110. [[CrossRef](#)] [[PubMed](#)]
208. Yang, B.; Chen, G.Y.; Song, X.P.; Yang, L.Q.; Han, C.R.; Wu, X.Y.; Zheng, C.J.; Ran, X.; Tang, R.F. Trigoxypins O-T, Six New Heterodimers from *Trigonostemon xyphophylloides*. *Tetrahedron Lett.* **2013**, *54*, 6434–6438. [[CrossRef](#)]
209. de Abrandes, R.A.; Batista, T.M.; Mangueira, V.M.; de Sousa, T.K.G.; Ferreira, R.C.; Moura, A.P.G.; Abreu, L.S.; Alves, A.F.; Velozo, E.S.; Batista, L.M.; et al. Antitumor and Antiangiogenic Effects of Tonantzitlone B, an Uncommon Diterpene from *Stillingia loranthea*. *Naunyn Schmiedeberg's Arch. Pharmacol.* **2022**, *395*, 267–274. [[CrossRef](#)] [[PubMed](#)]
210. Adiga, P.G.; Shridhar, N.B.; Sanganal, J.S.; Rao, S.; Shilpa, N.; Aparna, V.; Santhosh, I.M.; Tirumala, M.; Sindhu, B.M. In Vivo Anti-Tumor Efficacy of *Croton oblongifolius* in DMBA Induced Mammary Tumor in Rats. *Indian J. Anim. Res.* **2018**, *52*, 144–147. [[CrossRef](#)]
211. Majumder, M.; Debnath, S.; Gajbhiye, R.L.; Saikia, R.; Gogoi, B.; Samanta, S.K.; Das, D.K.; Biswas, K.; Jaisankar, P.; Mukhopadhyay, R. *Ricinus communis* L. Fruit Extract Inhibits Migration/Invasion, Induces Apoptosis in Breast Cancer Cells and Arrests Tumor Progression in Vivo. *Sci. Rep.* **2019**, *9*, 14493. [[CrossRef](#)]
212. De Castro, J.B.R.; Wemmenson, F.; Moura, G.; Solheiro Bezerra, A.; De, P.; Rodrigues, A.; Da Costa, H.P.S.; Do, N.; Canabrava, V.; Matheus, C.; et al. Antitumoral activity and in vivo toxicity of *Cnidioscolus quercifolius* pohl (euphorbiaceae). *Int. J. Dev. Res.* **2020**, *10*, 20359. [[CrossRef](#)]
213. Liang, Y.; Zhang, Y.; Wang, G.; Li, Y.; Huang, W. Penduliflaworosin, a Diterpenoid from *Croton crassifolius*, Exerts Anti-Angiogenic Effect via VEGF Receptor-2 Signaling Pathway. *Molecules* **2017**, *22*, 126. [[CrossRef](#)]
214. Ma, L.; Chen, Z.; Wang, W.; Zhang, J.; Zhang, H.; Liu, J. The Molecular Mechanism of ESZM Extract on Treatment ER(–) Breast Cancer in Vivo: Application of “Combat Poison with Poison” Theory. *Preprint* **2021**. [[CrossRef](#)]
215. Cheng, J.; Han, W.; Wang, Z.; Shao, Y.; Wang, Y.; Zhang, Y.; Li, Z.; Xu, X.; Zhang, Y. Hepatocellular Carcinoma Growth Is Inhibited by *Euphorbia helioscopia* L. Extract in Nude Mice Xenografts. *Biomed. Res. Int.* **2015**, *2015*, 601015. [[CrossRef](#)]
216. Huang, Y.Y.; Chen, C.H.; Hsu, C.H.; Kuo, T.Y.; Liu, C.C.; Liao, A.T.C.; Lin, C.S. Inhibiting Autophagy Potentiates the Antitumor Efficacy of *Euphorbia royleana* for Canine Mammary Gland Tumors. *BMC Vet. Res.* **2020**, *16*, 193. [[CrossRef](#)] [[PubMed](#)]
217. Sultana, S. Evaluation of the Antioxidant and In Vivo Anticancer Activity of *Excoecaria agallocha* Bark Extrac. Ph.D. Thesis, Brac University, Dhaka, Bangladesh, 2019.
218. Ghramh, H.A.; Khan, K.A.; Ibrahim, E.H.; Setzer, W.N. Synthesis of Gold Nanoparticles (AuNPs) Using *Ricinus communis* Leaf Ethanol Extract, Their Characterization, and Biological Applications. *Nanomaterials* **2019**, *9*, 765. [[CrossRef](#)] [[PubMed](#)]
219. Alqahtani, M.S.; Al-Yousef, H.M.; Alqahtani, A.S.; Tabish Rehman, M.; AlAjmi, M.F.; Almarfidi, O.; Amina, M.; Alshememry, A.; Syed, R. Preparation, Characterization, and in Vitro-in Silico Biological Activities of *Jatropha pelargoniifolia* Extract Loaded Chitosan Nanoparticles. *Int. J. Pharm.* **2021**, *606*, 120867. [[CrossRef](#)] [[PubMed](#)]
220. Alqahtani, A.A.; Attia, G.H.; Elgamal, A.; Aleraky, M.; Youns, M.; Ibrahim, A.M.; Abdou, R.; Shaikh, I.A.; El Raey, M.A. Cytotoxic Activity of Zinc Oxide Nanoparticles Mediated by *Euphorbia retusa*. *Crystals* **2022**, *12*, 903. [[CrossRef](#)]
221. Aboulthana, W.M.; Omar, N.I.; El-Feky, A.M.; Hasan, E.A.; Ibrahim, N.E.S.; Youssef, A.M. In Vitro Study on Effect of Zinc Oxide Nanoparticles on the Biological Activities of *Croton tiglium* L. Seeds Extracts. *Asian Pac. J. Cancer Prev.* **2022**, *23*, 2671–2686. [[CrossRef](#)] [[PubMed](#)]
222. Banerjee, K.; Das, S.; Choudhury, P.; Ghosh, S.; Baral, R.; Choudhuri, S.K. A Novel Approach of Synthesizing and Evaluating the Anticancer Potential of Silver Oxide Nanoparticles in Vitro. *Chemotherapy* **2017**, *62*, 279–289. [[CrossRef](#)] [[PubMed](#)]

223. El Raey, M.A.; El-Hagrassi, A.M.; Osman, A.F.; Darwish, K.M.; Emam, M. *Acalypha wilkesiana* Flowers: Phenolic Profiling, Cytotoxic Activity of Their Biosynthesized Silver Nanoparticles and Molecular Docking Study for Its Constituents as Topoisomerase-I Inhibitors. *Biocatal. Agric. Biotechnol.* **2019**, *20*, 101243. [\[CrossRef\]](#)
224. Elemike, E.E.; Onwudiwe, D.C.; Singh, M. Eco-Friendly Synthesis of Copper Oxide, Zinc Oxide and Copper Oxide–Zinc Oxide Nanocomposites, and Their Anticancer Applications. *J. Inorg. Organomet. Polym. Mater.* **2020**, *30*, 400–409. [\[CrossRef\]](#)
225. Cherian, A.M.; Snima, K.S.; Kamath, C.R.; Nair, S.V.; Lakshmanan, V.K. Effect of *Baliospermum montanum* Nanomedicine Apoptosis Induction and Anti-Migration of Prostate Cancer Cells. *Biomed. Pharmacother.* **2015**, *71*, 201–209. [\[CrossRef\]](#)
226. Boomi, P.; Poorani, G.P.; Selvam, S.; Palanisamy, S.; Jegatheeswaran, S.; Anand, K.; Balakumar, C.; Premkumar, K.; Prabu, H.G. Green Biosynthesis of Gold Nanoparticles Using *Croton sparsiflorus* Leaves Extract and Evaluation of UV Protection, Antibacterial and Anticancer Applications. *Appl. Organomet. Chem.* **2020**, *34*, e5574. [\[CrossRef\]](#)
227. Abd El-Moaty, H.I.; Ismail, E.H.; Abu-Khudir, R.; Soliman, N.A.; Sabry, D.Y.; Al Abdulsalam, N.K.; Sorour, W.A.; Khalil, M.M.H. Characterization and Evaluation of Multiple Biological Activities of Phytosynthesized Gold Nanoparticles Using Aqueous Extract of *Euphorbia dendroides*. *Nanomater. Nanotechnol.* **2022**, *12*, 18479804221141266. [\[CrossRef\]](#)
228. Lingaraju, K.; Naika, H.R.; Nagaraju, G.; Nagabhushana, H. Biocompatible Synthesis of Reduced Graphene Oxide from *Euphorbia heterophylla* (L.) and Their in-Vitro Cytotoxicity against Human Cancer Cell Lines. *Biotechnol. Rep.* **2019**, *24*, e00376. [\[CrossRef\]](#) [\[PubMed\]](#)
229. Ghramh, H.A.; Khan, K.A.; Ibrahim, E.H. Biological Activities of *Euphorbia peplus* Leaves Ethanolic Extract and the Extract Fabricated Gold Nanoparticles (AuNPs). *Molecules* **2019**, *24*, 1431. [\[CrossRef\]](#) [\[PubMed\]](#)
230. Kumari, A.; Naveen; Dhatwalia, J.; Thakur, S.; Radhakrishnan, A.; Chauhan, A.; Chandan, G.; Choi, B.H.; Neetika; Nidhi. Antioxidant, Antimicrobial, and Cytotoxic Potential of *Euphorbia royleana* Extract-Mediated Silver and Copper Oxide Nanoparticles. *Chem. Pap.* **2023**, *77*, 4643–4657. [\[CrossRef\]](#)
231. Bhuvaneswari, R.; Xavier, R.J.; Arumugam, M. Facile Synthesis of Multifunctional Silver Nanoparticles Using Mangrove Plant *Excoecaria agallocha* L. for Its Antibacterial, Antioxidant and Cytotoxic Effects. *J. Parasit. Dis.* **2017**, *41*, 180–187. [\[CrossRef\]](#) [\[PubMed\]](#)
232. Zhou, J.B.; Peng, G.; Wang, J.; Nie, S.; Li, J.; Jia, Y.C.; Zhang, Q.Y. Ebracteolatin A and Ebracteolatin B Induce Apoptosis of Human Hepatoma Cell Line (HepG2). *Trop. J. Pharm. Res.* **2015**, *14*, 1821–1828. [\[CrossRef\]](#)
233. Chaitanya, G.V.; Alexander, J.S.; Babu, P.P. PARP-1 Cleavage Fragments: Signatures of Cell-Death Proteases in Neurodegeneration. *Cell Commun. Signal.* **2010**, *8*, 31. [\[CrossRef\]](#) [\[PubMed\]](#)
234. Shi, Y. Mechanisms of Caspase Activation and Inhibition during Apoptosis. *Mol. Cell* **2002**, *9*, 459–470. [\[CrossRef\]](#)
235. Hao, Q.; Chen, J.; Lu, H.; Zhou, X. The ARTS of P53-Dependent Mitochondrial Apoptosis. *J. Mol. Cell Biol.* **2023**, *14*, mjac074. [\[CrossRef\]](#)
236. Jaiswal, P.L.; Goel, A.; Mittal, R.D. Survivin: A Molecular Biomarker in Cancer. *Indian J. Med. Res.* **2015**, *141*, 389.
237. Shi, S.-Q.; Fan, Y.-Y.; Xu, C.-H.; Ding, J.; Wang, G.-W.; Yue, J.-M. Supporting Information Cytotoxic 8,9-Seco-Ent-Kaurane Diterpenoids from *Croton kongensis*. *J. Asian Nat. Prod. Res.* **2018**, *20*, 920–927. [\[CrossRef\]](#) [\[PubMed\]](#)

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.