toxins

Communication

Occurrence of Deoxynivalenol and Zearalenone in Commercial Fish Feed: An Initial Study

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Abstract: The control of mycotoxins is a global challenge not only in human consumption but also in nutrition of farm animals including aquatic species. *Fusarium* toxins, such as deoxynivalenol (DON) and zearalenone (ZEN), are common contaminants of animal feed but no study reported the occurrence of both mycotoxins in fish feed so far. Here, we report for the first time the occurrence of DON and ZEN in samples of commercial fish feed designed for nutrition of cyprinids collected from central Europe. A maximal DON concentration of 825 μg kg^{-1} feed was found in one feed whereas average values of 289 μg kg^{-1} feed were noted. ZEN was the more prevalent mycotoxin but the concentrations were lower showing an average level of 67.9 μg kg^{-1} feed.

Keywords: *Fusarium* toxins; aquaculture; feedstuffs

1. Introduction

The mycotoxins deoxynivalenol (DON) and zearalenone (ZEN) appear due to cultivation of cereals world-wide due to their production as secondary metabolites by naturally occurring fungi of the genus

Fusarium [1]. Introduction of these mycotoxins into soil takes places by contaminated parts of the cereals, and subsequently these substances can be transferred to aquatic environments [2]. Due to their relative high stability in the aquatic environment, they can be found in relevant concentrations in surface waters [2,3]. Even more importantly, fish in aquaculture are commonly exposed to feed-borne mycotoxins. Commercial feed for aquaculture often contain high amounts of fishmeal [4], but the growing aquaculture decreased the fish meal stock world-wide and the decreasing availability of fish resources led to the need for alternative protein resources to replace fishmeal in these feed. Thus, cereals are increasingly used for production of fish feed [5].

DON contamination in animal feed components and finished feedstuff is very prevalent in Europe and North America [6]. However, it has up to now only rarely been shown that DON and ZEN are contaminants of fish feed components [7,8], but wheat and less frequently corn, barley, and rye are used for feed production and might contain these mycotoxins in relevant concentrations with high prevalency [9–11]. Especially, wheat is often used for fish feed production due to its high protein content and its benefits concerning the preservation of the pellet shape during the production process of pelleted feed. The amount of wheat in fish feed varies considerably, ranging from approximately 15 to 27 percent for carnivorous fish whereas feed for cyprinids usually contain 20 to 70 percent [11,12]. In addition, whole cereals can also be used as supplementary feed in semi-intensive culture of fish species such as carp [13].

The undesired introduction of cereal-borne mycotoxins into fish feeds leads to so far largely unknown consequences. In mammalian cell lines, it has been found that ZEN is hepatotoxic [14,15], immunotoxic [16,17], and genotoxic [18]. The toxicity of DON is also well recognized in mammalian cell lines including mechanisms such as mitochondrial impairment and apoptosis [19,20], but few studies have assessed its toxicity on cell lines of aquatic organisms [21,22].

Recent investigations on salmonids showed that alterations in the intestinal tract of fish occur upon feeding with DON-contaminated diets [23,24]. A study on zebrafish (*Danio rerio*) showed effects of DON on fecundity and offspring larvae swimming activity [25]. ZEN and its metabolites, namely α-zearalenol (α-ZEL) and β-zearalenol (β-ZEL), have been shown to act as typical estrogenic compounds with partially high estrogenic potencies via affinity to estrogen receptors of fish depending on the fish species [26,27]. Thus, investigations on effects of ZEN in fish focussed on reproduction of zebrafish and early life stages of fathead minnow (*Pimephales promelas*) [28,29]. Only one study investigated the effects of ZEN on growth performance of Atlantic salmon (*Salmo salar*) showing no evidence for an adverse influence of ZEN on this species at concentrations ranging from 60 to 770 µg kg^{-1} feed [23].

Since the sensitivity of farm animals to DON differs remarkably, different guideline values of DON in feed have been proposed [30]. A general recommendation on guidance values of 5 mg kg^{-1} DON in complete feedingstuff was established by the European Commission [31] while specific recommendations concerning ZEN contamination in animal feed are only referring to some animal species such as pigs, cattle, sheep and goats. Nevertheless, DON concentrations below its guidance value have already been shown to affect fish negatively [23–25].

Still the occurrence of *Fusarium* toxins in aquaculture remains mostly unknown and research is needed. Here, we present DON and ZEN concentrations in commercial feed mainly designed for

cultivation of cyprinids in order to assess their possible health risks for aquatic animals and to provide a first basis for deriving acceptable guidance values for DON and ZEN in fish feed.

2. Results and Discussion

2.1. Occurrence of DON

The study of Måge *et al*. [7] reported a contamination prevalence with DON in fish feed components of 33%, whereas others reported 50% DON contamination in samples from cereals, straw, silage and finished feed from all over the world [32]. In contrast, we were able to show that DON could be found in more than 80% of the samples from commercial fish feed showing average values of 289 µg kg^{-1} feed (Table 1). A maximum value for DON of 825 µg kg^{-1} was noted for feed #6. However, its metabolite DOM-1 was not found in the samples. Thus, the DON values in fish feed do not exceed the guidance values for complete feedingstuff [31].

2.2. Occurrence of ZEN

Again Måge *et al*. [7] noted that 33% of the samples taken from fish feed components were contaminated with ZEN, whereas another study reported that in 56% of the samples from finished animal feed ZEN was found with average levels of 99 µg kg^{-1} feed [6]. In Asian countries ZEN was found in 9 fish feed samples showing average concentrations of 76.2 µg kg^{-1} feed [8]. Similarly, in our study ZEN was found in all samples (Table 1), although the concentrations varied considerably (average 67.9 µg kg^{-1} feed, ranging from 3 to 511 µg kg^{-1} feed). Consequently, the ZEN values found in our study do not exceed the values currently recommended by the European Commission [31].

Table 1. Deoxynivalenol (DON) and zearalenone (ZEN) concentrations in commercial fish feed in central Europe designated for feeding of cyprinids.

Feed	Pellet size (mm)	Dry matter (%)	Crude protein [1] (%)	Crude fat [1] (%)	DON (µg kg^{-1})	ZEN (µg kg^{-1})
#1	2.0	90.9	20.0	4.8	768	80
#2	4.5	86.9	42.2	23.8	81	10
#3	1.6	90.7	48.6	13.1	284	15
#4	4.5	92.0	48.6	13.1	117	27
#5	2.8	91.0	48.6	13.1	66	9
#6	3.0	92.2	33.0	6.0	825	511
#7	3.0	94.5	30.0	5.0	150	8
#8	3.0	92.3	34.0	15.0	0	6
#9	3.0	92.5	45.0	12.0	0	3
#10	2.5	91.9	41.0	12.0	176	12
#11	3.0	91.6	35.0	6.0	131	21

[1] According to the manufacturer.

2.3. Possible Consequences of DON and ZEN Contamination in Fish Feed

According to the recommendation of the European Commission [31] ZEN should not exeed levels of 2 mg kg^{-1} feed material (with the exception of maize by-products) and DON should be lower than

5 mg kg^{-1} complete feedstuff. According to our findings for the samples from fish feed these values are not exeeded. However, recent studies showed that DON can be harmful to fishes at levels below these recommendations [23–25]. For ZEN the consequences of food-borne exposure remain mostly unknown [23]. Still, the frequent exposure of fish to single mycotoxins and combinations of different mycotoxins in aquaculture represents a continuous health risk [29,33,34]. Unfortunately, the actual risks remain more or less unknown due to a lack of data which provide evidence for effects in aquatic animals and which clarify the mechanism(s) of action of these substances. In particular, the effects of DON on certain aspects of fish health and cell function *in vivo* have rarely been investigated in fish [24]. However, this is an important issue for future research since economic consequences for the animal industry can be minimized.

2.4. Possible Strategies to Prevent DON and ZEN Contamination in Fish Feed

Chemical substances and natural toxins in the food chain received increasing attention recently which led to risk assessments and the development of different prevention strategies [35,36]. One strategy would include the selection of proper ingredients for feed production. Not surprisingly, even within one production firm the batches of ingredients seem to influence the mycotoxin content of the final feed enormously. This can be seen from feed #3 to #5 which were produced by the same company by using similar ingredients and the same production processes but differed in final pellet size. Nevertheless, the contents of DON and ZEN differ considerably between these three samples (Table 1). Moreover, the contamination of the tested fish feed samples in our study was more prevalent than previously expected from other studies [6,7]. This is certainly due to the combination of ingredients that were chosen for feed production for cyprinids. A quite compatible protein source in fish feed would be reached by inclusion of fish meal at high percentages. However, the availability of fish meal for production of aquaculture feeds is decreasing globally over the last decades and the fish meal prices are increasing accordingly [5]. Thus, alternative protein sources need to be used for fish feed production. A higher percentage of cereals is commonly used for feed production for cyprinids compared to carnivorous fishes [11,12]. But natural contamination of commercial fish feed with DON and ZEN seems to be higher when corn and cereal components are used at higher percentages whereas fish meal-based feed mostly contained less of these mycotoxins (Tables 1 and 2). This assumption is supported by the finding that the most prevalent mycotoxin in central Europe was DON with maximum values of 14.1 mg kg^{-1} barley and 49.0 mg kg^{-1} wheat (found in Austria) and average levels of ZEN and DON of 1.8 mg kg^{-1} and 1.7 mg kg^{-1} in corn gluten meal world-wide, respectively [6]. Soybean and soybean meal are considered to be less contaminated with DON and ZEN [6,37] and the usage of these plant components instead of cereals probably leads to less contamination of animal feedstuffs with *Fusarium* toxins. However, soybean and soybean by-products can be used for fish feed production only at relatively low percentages (depending on the fish species) because they exert estrogenic effects and negative impacts on fish growth at higher levels [38,39]. Thus, the contribution of certain ingredients to the mycotoxin concentrations in fish feed and their possible consequences on fish health should be the subject of future research.

Table 2. Approximate declaration of ingredients of the commercial fish feed from central Europe; all components (C1 to C10) are sorted by decreasing percentage in the final feed; wheat, wheat by-products and rye are shaded in blue; fish components are displayed in green, blood meal is shaded in red, ingredients containing soybean are displayed in orange, and corn and corn by-products are shaded in yellow.

Feed	C1	C2	C3	C4	C5	C6	C7	C8	C9	C10
#1	W	WB	MS	CGF	SFEM	SEM	B	SBM	VO	M
#2	FM	FO	BM	SEM	WDB	W	M	Y	WB	VO
#3	FM	BM	W	SM	FO	M	-	-	-	-
#4	FM	BM	W	SM	FO	M	-	-	-	-
#5	FM	BM	W	SM	FO	M	-	-	-	-
#6	SEM	W	FM	WB	C	CGF	SFEM	FO	WGF	-
#7	SEM	WB	W	SFEM	FO	FM	-	-	-	-
#8	FM	W	SEM	FO	BM	VO	-	-	-	-
#9	FM	W	SEM	WGF	FO	VO	-	-	-	-
#10	W (31%)	SEM (27%)	FM (19%)	BM (6%)	FO (3%)	VO (3%)	-	-	-	-
#11	SEM	W	SM	FM	FO	M	-	-	-	-

Abbreviations: B = barley; BM = blood meal; C = corn; CGF = Corn gluten feed; FM = fish meal; FO = fish oil; M = minerals; MS = malt sprouts; SBM = sugar beet molasses; SEM = soybean extraction meal; SM = soybean meal; SFEM = sunflower feed extraction meal; VO = vegetable oils and fats; W = wheat; WB = wheat bran, WDB = wheat distillery by-product; WGF = wheat gluten feed; Y = yeast.

However, a general problem during our collection of samples was that although the producers of fish feed have to give an ingredient declaration of compounds in feedingstuffs by (percentage) weight of inclusion, only the producer of feed #10 displayed percentages on the packaging. All others refused to declare the percentages even on repeated enquiry. Therefore, an improved labeling policy would help to identify and prevent sources of mycotoxin inclusion in animal feed.

It is also known that the storage of ingredients and the production processes influence mycotoxin contents in fish feed. Pelleted feed are increasingly replaced by extruded diets. Extrusion of feeds increases the digestibility of nutrients. The process of extrusion comprises an increase of temperatures to 110–160 °C. In our study feed #7 and feed #9 are extruded but, both of them still contain ZEN and feed #7 still contains DON. Studies on effects of high temperatures and high pressure on DON concentrations during production of food and feed have yielded contradicting results [40–42]. However, extrusion cooking clearly led to the reduction of DON concentrations in wheat grits depending on the adjustment of physicochemical parameters [43]. Moreover, extrusion can influence the content of ZEN [44]. We have no proof whether the ZEN content in samples from feed #7 and feed #9 was influenced by the production process. Nevertheless, according to the literature the selection of the appropriate course of production processes in addition to optimized storage conditions of ingredients and finished feed should lead to less mycotoxins in feedstuffs to some extent.

In addition, some research group focus on strategies of biological detoxification of mycotoxins, the use of different sorbents or the use of genetically-modified crops for feed production in order to achieve less mycotoxin contamination in feed [45–47].

3. Experimental Section

3.1. Fish Feed Samples

For analyses of DON and ZEN 11 samples from fish feed from central Europe were collected. The samples that were chosen were mainly designed for use of carp feeding, since cyprinids display a major group of species in freshwater aquaculture [5]. All samples were taken from complete feedstuffs with the exception of feed #1 which is designated as a complementary feed for carp. Samples were stored at −20 °C before extraction of mycotoxins.

3.2. Mycotoxin Analyses

DON and DOM-1 in fish feed were analyzed by HPLC-DAD (high performance liquid chromatography (consisting of a pump (LC-10ADVP), an autoinjector (SIL-10ADVP), and a column oven (CTO-10ACVP)) from Shimadzu (Duisburg, Germany)) with diode array detection using the detector SPD-M10AVP (Shimadzu, Duisburg, Germany) after a clean-up with IAC (immuno-affinity columns, DONprep^{TM}, R-Biopharm, Darmstadt, Germany) according to manufacturer's procedure with slight modifications as described previously [48]. The detection limit was 30 µg kg^{-1}, the mean recovery was approximately 90%.

ZEN in feed was determined by HPLC with fluorescence detection after a clean-up with IAC (ZearalaTest, Vicam, Klaus Ruttmann, Hamburg, Germany) according to a slightly modified method of the "Verband Deutscher Landwirtschaftlicher Untersuchungs- und Forschungsanstalten" [49]. The detection limit was 2 µg kg^{-1} and mean recovery was approximately 79%.

4. Conclusions

The values of DON and ZEN in commercial fish feed appear to be lower than the currently recommended values [31]. Still, it can be concluded that some of the commercial feed that were investigated within the present study may be harmful to fish due to their DON content. Unfortunately, we were only able to analyse 11 different fish feed so far. A much broader survey should be conducted to increase knowledge on the occurrence of cereal-borne toxins in feed designated for aquatic species. In this respect it can also be assumed that fumonisins and ergot alkaloids certainly also occur in fish feed but no survey has as yet assessed their concentrations in feed produced for aquatic species.

Acknowledgments

Conflict of Interest

The authors declare no conflict of interest.

References

1. Yazar, S.; Omurtag, G.Z. Fumonisins, trichothecenes and zearalenone in cereals. *Int. J. Mol. Sci.* **2008**, *9*, 2062–2090.

2. Bucheli, T.D.; Wettstein, F.E.; Hartmann, N.; Erbs, M.; Vogelsang, S.; Forrer, H.-R.; Schwarzenbach, R.P. *Fusarium* mycotoxins: Overlooked aquatic micropollutants. *J. Agric. Food Chem.* **2008**, *56*, 1029–1034.

3. Kolpin, D.W.; Hoerger, C.C.; Meyer, M.T.; Wettstein, F.E.; Hubbard, L.E.; Bucheli, T.D. Phytoestrogens and mycotoxins in Iowa streams: An examination of underinvestigated compounds in agricultural basins. *J. Environ. Qual.* **2010**, *39*, 2089–2099.

4. Tacon, A.G.J.; Metian, M. Global overview on the use of fish meal and fish oil in industrially compounded aquafeeds: Trends and future prospects. *Aquaculture* **2008**, *285*, 146–158.

5. FAO. *The State of World Fisheries and Aquaculture*; Food and Agriculture Organization of the United Nations: Rome, Italy, 2012.

6. Rodrigues, I.; Naehrer, K. Prevalence of mycotoxins in feedstuffs and feed surveyed worldwide in 2009 and 2010. *Phytopathol. Mediterr.* **2012**, *51*, 175–192.

7. Måge, A.; Julshamn, K.; Lunestad, B.T. *Overvåkningsprogram for Fôrvarer til Fisk og Andre Akvatiske dyr—Årsrapport 2008 og 2009*; NIFES: Bergen, Norway, 2009.

8. Fegan, D.F.; Spring, P. Recognizing the Reality of the Aquaculture Mycotoxin Problem: Searching for a Common and Effective Solution. In *Nutritional Biotechnology in the Feed and Food Industries*, Proceedings of Alltech's 23rd Annual Symposium. The New Energy Crisis: Food, Feed or Fuel? Lexington, KY, USA, 20–23 May 2007; pp. 343–354.

9. Placinta, C.M.; D'Mello, J.P.F.; Macdonald, A.M.C. A review of worldwide contamination of cereal grains and animal feed with Fusarium mycotoxins. *Anim. Feed Sci. Technol.* **1999**, *78*, 21–37.

10. Schollenberger, M.; Müller, H.M.; Drochner, W. Fusarium toxins in different food samples. *Mycotox. Res.* **2002**, *18*, 39–42.

11. Matz, S.A. *The Chemistry and Technology of Cereals as Food and Feed*; Kluwer Academic Publishers Group: Dordrecht, The Netherlands, 1991; p. 319.

12. Berntssen, M.H.G.; Julshamn, K.; Lundebye, A.K. Chemical contaminants in aquafeeds and Atlantic salmon (*Salmo salar*) following the use of traditional versus alternative feed ingredients. *Chemosphere* **2010**, *78*, 637–646.

13. Urbanek, M.; Hartvich, P.; Vacha, F.; Rost, M. Investigation of fat content in market common carp (*Cyprinus carpio*) flesh during the growing season. *Aquac. Nutr.* **2010**, *16*, 511–519.

14. Maaroufi, K.; Chekir, L.; Creppy, E.E.; Ellouz, F.; Bacha, H. Zearalenone induces modifications of haematological and biochemical parameters in rats. *Toxicon* **1996**, *34*, 535–540.

15. Conková, E.; Laciaková, A.; Pástorová, B.; Seidel, H.; Kovác, G. The effect of zearalenone on some enzymatic parameters in rabbits. *Toxicol. Lett.* **2001**, *121*, 145–149.

16. Marin, L.; Murtha, J.; Dong, W.; Pestka, J.J. Effects of mycotoxins on cytokine production and proliferation in EL-4 thymoma cells. *J. Toxicol. Environ. Health* **1996**, *48*, 379–396.

17. Berek, L.; Petri, I.B.; Mesterhazy, A.; Teren, J.; Molnar, J. Effects of mycotoxins on human immune functions *in vitro*. *Toxicol. in Vitro* **2001**, *15*, 25–30.

18. Abid-Essefi, S.; Ouanes, Z.; Hassen, W.; Baudrimont, I.; Creppy, E.E.; Bacha, H. Cytotoxicity, inhibition of DNA and protein syntheses and oxidative damage in cultured cells exposed to zearalenone. *Toxicol. in Vitro* **2004**, *18*, 467–474.

19. Gutleb, A.C.; Morrison, E.; Murk, A.J. Cytotoxicity assays for mycotoxins produced by *Fusarium* strains: A review. *Environ. Toxicol. Pharmacol.* **2002**, *11*, 309–320.

20. Bensassi, F.; el Golli-Bennour, E.; Abid-Essefi, S.; Bouaziz, C.; Hajlaoui, M.R.; Bacha, H. Pathway of deoxynivalenol-induced apoptosis in human colon carcinoma cells. *Toxicology* **2009**, *264*, 104–109.

21. Pietsch, C.; Bucheli, T.; Wettstein, F.; Burkhardt-Holm, P. Frequent biphasic cellular responses of permanent fish cell cultures to deoxynivalenol (DON). *Toxicol. Appl. Pharmacol.* **2011**, *256*, 24–34.

22. Pietsch, C.; Crivelli, G.; Noser, J.; Wettstein, F.E.; Burkhardt-Holm, P. The role of oxidative stress in zearalenone-mediated toxicity in permanent fish cell cultures. *Toxic. Sci.* **2013**, submitted for publication.

23. Döll, S.; Valenta, H.; Baardsen, G.; Möller, P.; Koppe, W.; Stubhaug, I.; Dänicke, S. Effects of Increasing Concentrations of Deoxynivalenol, Zearalenone and Ochratoxin A in Diets for Atlantic Salmon *(Salmo salar)* on Performance, Health and Toxin Residues. In *Proceedings of 33rd Mycotoxin Workshop*, Freising, Germany, 30 May–1 June 2011.

24. Hooft, J.M.; Elmor, A.E.H.I.; Encarnação, P.; Bureau, D.P. Rainbow trout (*Oncorhynchus mykiss*) is extremely sensitive to the feed-borne *Fusarium* mycotoxin deoxynivalenol (DON). *Aquaculture* **2011**, *311*, 224–232.

25. Sanden, M.; Jorgensen, S.; Hemre, G.-I.; Ornrud, R.; Sissener, N.H. Zebrafish (*Danio rerio*) as a model for investigating dietary toxic effects of deoxynivalenol contamination in aquaculture feeds. *Food Chem. Toxicol.* **2012**, *50*, 4441–4448.

26. Bucheli, T.D.; Erbs, M.; Hartmann, N.; Vogelsang, S.; Wettstein, F.E.; Forrer, H.R. Estrogenic mycotoxins in the environment. *Mitt. Lebensm. Hyg.* **2005**, *96*, 386–403.

27. Cosnefroy, A.; Brion, F.; Maillot-Marechal, E.; Porcher, J.M.; Pakdel, F.; Balaguer, P.; Ait-Aissa, S. Selective activation of zebrafish estrogen receptor subtypes by chemicals by using stable reporter gene assay developed in a zebrafish liver cell line. *Toxicol. Sci.* **2012**, *125*, 439–449.

28. Johns, S.M.; Denslow, N.D.; Kane, M.D.; Watanabe, K.H.; Orlando, E.F.; Sepulveda, M.S. Effects of estrogens and antiestrogens on gene expression of fathead minnow (*Pimephales promelas*) early life stages. *Environ. Toxicol.* **2009**, *26*, 195–206.

29. Schwartz, P.; Thorpe, K.L.; Bucheli, T.D.; Wettstein, F.E.; Burkhardt-Holm, P. Short-term exposure to the environmentally relevant estrogenic mycotoxin zearalenone impairs reproduction in fish. *Sci. Total Environ.* **2010**, *409*, 326–333.

30. Eriksen, G.; Pettersson, H. Toxicological evaluation of trichothecenes in animal feed. *Anim. Feed Sci. Technol.* **2004**, *114*, 205–239.

31. European Commission, Commission Recommendation (EC) of 17 August 2006 on the presence of deoxynivalenol, zearalenone, ochratoxin A, T-2 and HT-2 and fumonisins in products intended for animal feeding. *Off. J. Eur. Un.* **2006**, L229/7–L229/9.

32. Santos, G.A.; Rodrigues, I.; Naehrer, K.; Encarnacao, P. Mycotoxins in aquaculture: Occurrence in feed components and impact on animal performance. *Aquac. Eur.* **2010**, 35, 6–10.

33. Yildirim, M.; Manning, B.B.; Lovell, R.T.; Grizzle, J.M.; Rottinghaus, G.E. Toxicity of moniliformin and fumonisin B1 fed singly and in combination in diets for young Channel Catfish *Ictalurus punctatus*. *J. World Aquac. Soc.* **2000**, *31*, 599–608.

34. Santacroce, M.; Conversano, M.; Casalino, E.; Lai, O.; Zizzadoro, C.; Centoducati, G.; Crescenzo, G. Aflatoxins in aquatic species: Metabolism, toxicity and perspectives. *Rev. Fish Biol. Fish.* **2008**, *18*, 99–130.

35. Mantovani, A.; Frazzoli, C.; la Rocca, C. Risk assessment of endocrine-active compounds in feeds. *Vet. J.* **2009**, *182*, 392–401.

36. Dorne, J.L.C.M.; Fink-Gremmels, J. Human and animal health risk assessment of chemicals in the food chain: Comparative aspects and future perspectives. *Toxicol. Appl. Pharmacol.* in press.

37. Valenta, H.; Dänicke, S.; Blüthgen, A. Mycotoxins in soybean feedstuffs used in Germany. *Mycotox. Res.* **2002**, *18*, 208–211.

38. Pelissero, C.; le Menn, F.; Kaushik, S. Estrogenic effects of dietary soya bean meal on vitellogenesis in cultured Siberian sturgeon *Acipenser baeri. Gen. Comp. Endocrinol.* **1991**, *83*, 447–457.

39. Kaushik, S.J.; Cravedi, J.P.; Lalles, J.P.; Sumpter, J.; Fauconneau, B.; Laroche, M. Partial or total replacement of fish meal by soybean protein on growth, protein utilization, potential estrogenic or antigenic effects, cholesterolemia and flesh quality in rainbow trout, *Oncorhynchus mykiss. Aquaculture* **1995**, *133*, 257–274.

40. Wolf-Hall, C.E.; Hanna, M.A.; Bullerman, L.B. Stability of deoxynivalenol in heat-treated foods. *J. Food Protect.* **1999**, *62*, 962–964.

41. Bretz, M.; Beyer, M.; Cramer, B.; Knecht, A.; Humpf, H.-U. Thermal degradation of the Fusarium mycotoxin deoxynivalenol. *J. Agric. Food Chem.* **2006**, *54*, 6445–6451.

42. Kushiro, M. Effects of milling and cooking processes on the deoxynivalenol content in wheat. *Int. J. Mol. Sci.* **2008**, *9*, 2127–2145.

43. Wu, Q.; Lohrey, L.; Cramer, B.; Yuan, Z.; Humpf, H.-U. Impact of physicochemical parameters on the decomposition of deoxynivalenol during extrusion cooking of wheat grits. *J. Agric. Food Chem.* **2011**, *59*, 12480–12485.

44. Ryu, D.; Hanna, M.A.; Bullerman, L.B. Stability of zearalenone during extrusion of corn grits. *J. Food Protect.* **1999**, *62*, 1482–1484.

45. Huwig, A.; Freimund, S.; Käppeli, O.; Dutler, H. Mycotoxin detoxication of animal feed by different adsorbents. *Toxicol. Lett.* **2001**, *122*, 179–188.

46. Okubara, P.A.; Blechl A.E.; McCormick, S.P.; Alexander, N.J.; Dill-Macky, R.; Hohn, T.M. Engineering deoxynivalenol metabolism in wheat through the expression of a fungal trichothecene acetyltransferase gene. *Theor. Appl. Genet.* **2002**, *106*, 74–83.

47. Karlovsky, P. Biological detoxification of the mycotoxin deoxynivalenol and its use in genetically engineered crops and feed additives. *Appl. Microbiol. Biotechnol.* **2011**, *91*, 491–504.

48. Oldenburg, E.; Bramm, A.; Valenta, H. Influence of nitrogen fertilization on deoxynivalenol contamination of winter wheat—Experimental field trials and evaluation of analytical methods. *Mycotox. Res.* **2007**, *23*, 7–12.

49. *VDLUFA-Methodenbuch III, 6. Ergänzung 2006, Zearalenon 16.9.2*; VDLUFA-Verlag: Darmstadt, Germany, 2006.