

## Supplementary Material

1. Take 1 g of a homogenized tissue (cut into pieces) sample and combine it with 200  $\mu\text{L}$  of isotopic standard working solution;
2. Add 10 mL of acetonitrile/water/acetic acid (79:20:1, v/v/v);
3. Add 4.0 g anhydrous sodium sulfate and 1.5 g of anhydrous sodium acetate in a plastic tube (50 mL), and blend in a in ultra-turrax blender (IKA, T10) until completely dissolved;
4. Centrifuge at 5,000 rpm for 8 min;
5. Separate the supernatant and transfer it to another tube (50 mL);
6. Add 5 mL of hexane and vortex the mixture for 2 min;
7. Keep the mixture under dark for 5 min BEFORE hexane phase (upper phase) removal & discard;
8. Separate and transfer the acetonitrile phase into a glass tube following evaporation under flowing nitrogen gas ( $50 \pm 5$  °C) in a MultiVap 54 (LabTech) concentrator;
9. Redissolve the remaining dry residue in 0.5 mL of acetonitrile/water (10:90, v/v);
10. Vortex (30 s) the extract, filter through a 0.22  $\mu\text{m}$  syringe filter and transfer the volume into an autosampler vial with an insert;
11. Submit a 5  $\mu\text{L}$  aliquot to LC-MS/MS analysis.

**Figure S1.** Analytical steps for extraction of mycotoxin residues from samples of liver and kidneys.

**Table S1.** Analytical parameters of the analytical method for determination of residual mycotoxins in liver and kidneys.

Mycotoxin	RT (min.)	Mass (g/mol)	Molecular ion	Transition ( <i>m/z</i> )	Calibration range (µg/kg)	LOD (µg/kg)	LOQ (µg/kg)
AFM <sub>1</sub>	1.13	328.3	[M+H] <sup>+</sup>	329.0 > 273.1 <sup>a</sup> 329.0 > 229.0 <sup>b</sup>	0.1-100	0.2	0.7
AFB <sub>1</sub>	1.25	312.3	[M+H] <sup>+</sup>	312.7 > 284.9 <sup>a</sup> 312.7 > 241.1 <sup>b</sup>	0.1-100	0.1	0.3
[ <sup>13</sup> C <sub>17</sub> ]-AFB <sub>1</sub>	1.25	329.1	[M+H] <sup>+</sup>	330.3 > 301.5	0.1-100	0.1	0.4
AFB <sub>2</sub>	1.15	314.3	[M+H] <sup>+</sup>	314.7 > 259.0 <sup>a</sup> 314.7 > 287.0 <sup>b</sup>			
AFG <sub>1</sub>	1.18	328.3	[M+H] <sup>+</sup>	328.9 > 243.0 <sup>a</sup> 328.9 > 199.5 <sup>b</sup>	0.1-100	0.2	0.6
AFG <sub>2</sub>	1.16	330.3	[M+H] <sup>+</sup>	330.9 > 245.0 <sup>a</sup> 330.9 > 188.9 <sup>b</sup>	0.1-100	0.2	0.8
[ <sup>13</sup> C <sub>34</sub> ]-FB <sub>1</sub>	5.40	755.6	[M+H] <sup>+</sup>	756.6 > 374.4	0.5-100	0.3	1.0
FB <sub>1</sub>	5.40	721.8	[M+H] <sup>+</sup>	722.5 > 334.0 <sup>a</sup> 722.5 > 352.1 <sup>b</sup>			
FB <sub>2</sub>	5.76	705.8	[M+H] <sup>+</sup>	706.5 > 336.2 <sup>a</sup> 706.5 > 318.3 <sup>b</sup>	0.5-100	0.4	1.0
[ <sup>13</sup> C <sub>18</sub> ]-ZEN	6.03	336.2	[M-H] <sup>-</sup>	335.1 > 185.1	1.0-100	1.0	3.0
ZEN	6.03	318.1	[M-H] <sup>-</sup>	317.1 > 175.1 <sup>a</sup> 317.1 > 130.9 <sup>b</sup>			
α-ZEL	5.81	320.2	[M-H] <sup>-</sup>	319.1 > 275.2 <sup>a</sup> 319.1 > 160.2 <sup>b</sup>	1.0-100	0.8	2.6
β-ZEL	5.83	320.2	[M-H] <sup>-</sup>	319.1 > 275.2 <sup>a</sup> 319.1 > 160.2 <sup>b</sup>	1.0-100	0.6	2.0

RT: Retention time; LOD: Limit of detection; LOQ: Limit of quantification; AF: aflatoxin; FB: fumonisin; ZEN: zearalenone; α-ZEL: α-zearalenol; β-ZEL: β-zearalenol.

<sup>a</sup> Transitions used in the quantification.

<sup>b</sup> Transitions used in the confirmation.