

Supplementary Materials: Domain Shuffling between Vip3Aa and Vip3Ca: Chimera Stability and Insecticidal Activity against European, American, African, and Asian Pests

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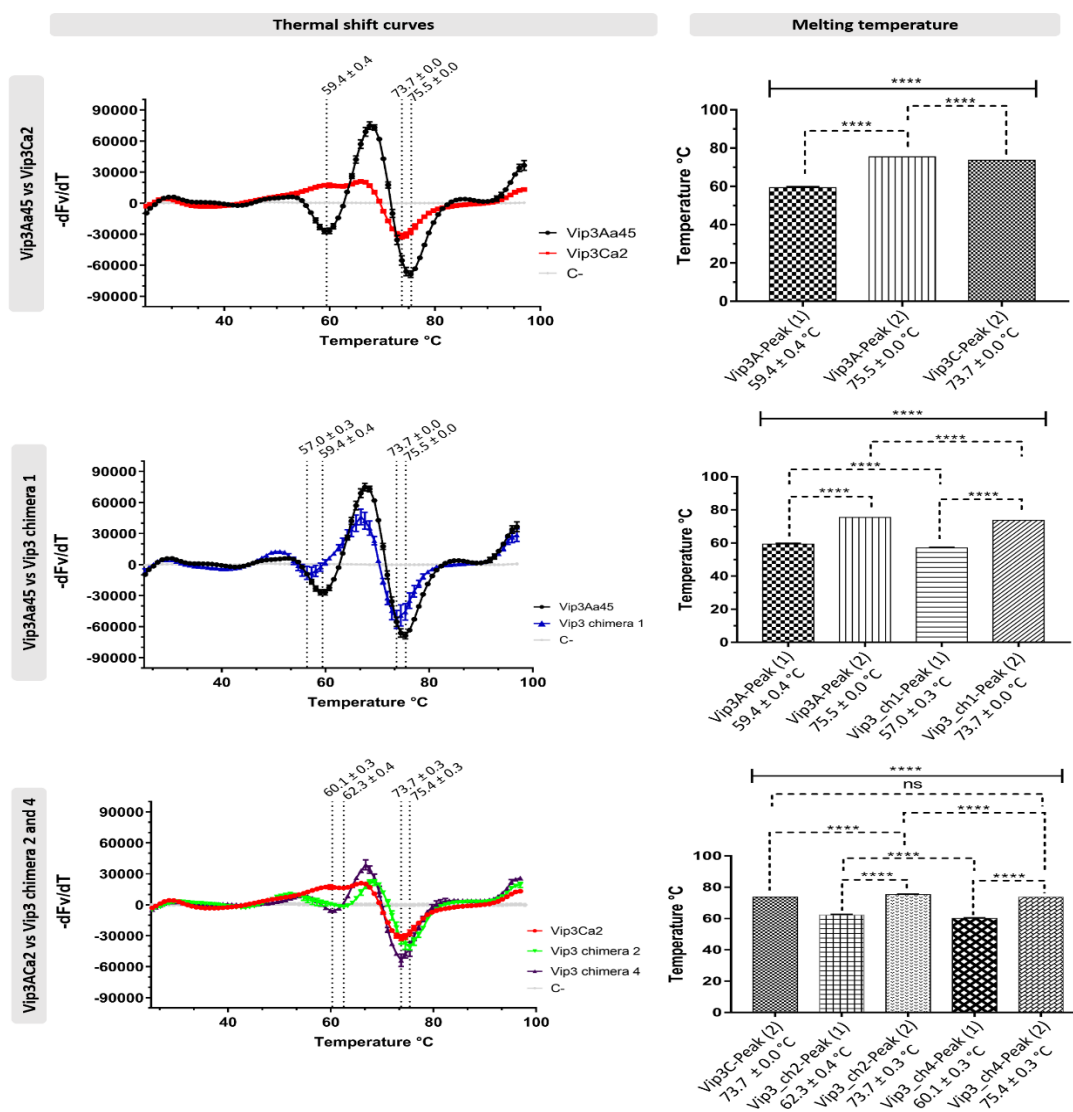


Figure S1. Thermal shift assays and multiple comparison of the thermal transitions of the parental proteins and chimeric proteins. The dashed vertical lines in the thermal shift assays curves indicate the T_m (measured in Celsius degrees) of respective thermal transitions. C- indicate the fluorescence intensity due to the SPYRO-Orange 15X in 20 mM Tris 500 mM NaCl pH 8.6. Thick line indicate the comparison of the T_m by One-way Anova (α 0.05). Dashed line indicate the multiple comparison analyzed by Tukey's range test (α 0.05). "****" indicate a p value less than 0.0001 and "ns" indicate a p value greater than 0.05

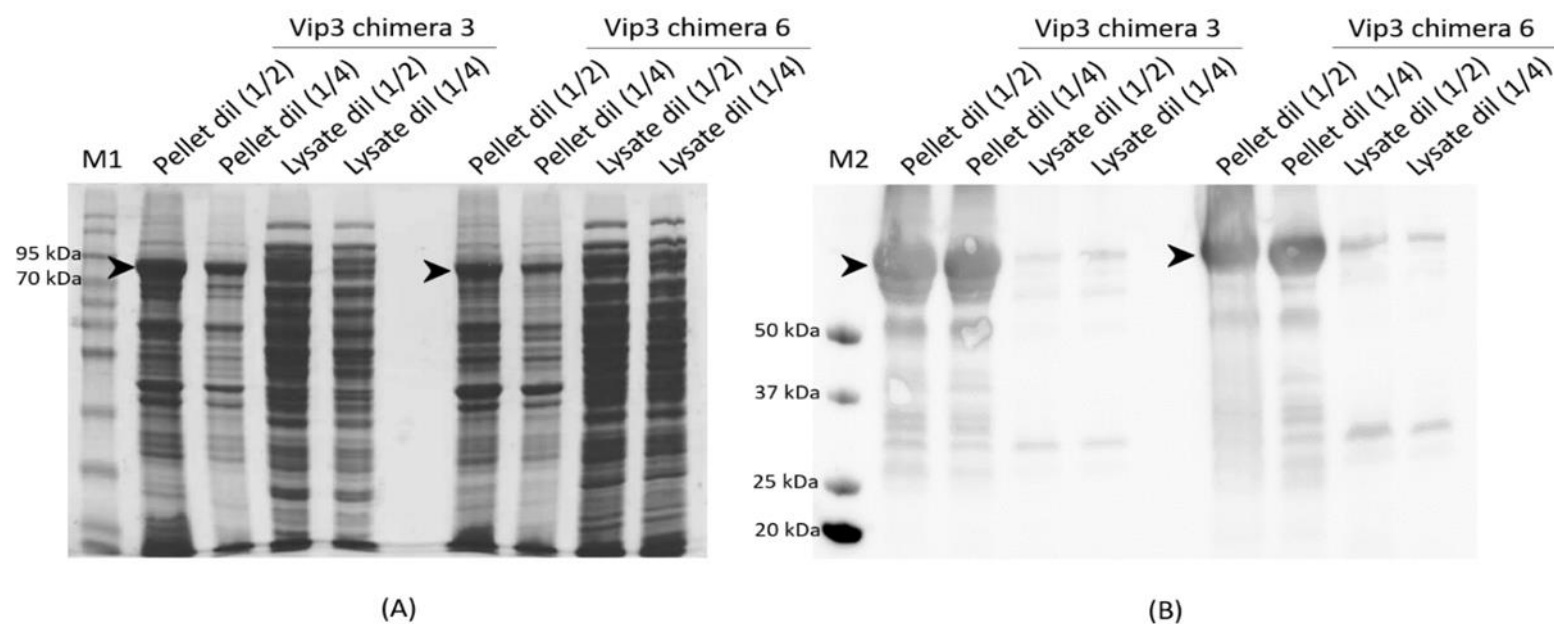


Figure S2. Expression of the chimeric Vip3 proteins (Vip3_ch3 and Vip3_ch6). **(A)** SDS-PAGE gel of different dilutions of the pellet and supernatant of the Vip3_ch3 and Vip3_ch6 proteins. **(B)** Western blot analysis different dilutions of the pellet and supernatant of the respective chimeric Vip3 proteins. The dilutions of the lysates were made with 50 mM phosphate buffer, 500 mM NaCl pH 8.0 while the pellet were dissolved in the same volume of the supernatant and the dilutions were made with 50 mM phosphate buffer, 500 mM NaCl pH 8.0. The arrowhead indicate the protein band corresponding to the chimeric Vip3 proteins. M1: Molecular Mass Marker “PINK Plus Prestained Protein Ladder” (Genedirex). M2: Molecular Mass Marker “Precision Plus Protein™ Dual Color Standards” (Biorad) developed with “Precision Protein™ Strep Tactin-HRP conjugate.

Table S1. Comparison analyses of the respective dose-response assays (LC values) of the parental and Vip3 chimeric proteins in *S. frugiperda* and *O. furnacalis*.

Statistical variables	<i>Spodoptera frugiperda</i>		<i>Ostrinia furnacalis</i>	
	Vip3Aa45 vs Vip3ch2	Vip3Ca2 vs Vip3ch2	Vip3Ca2 vs Vip3ch4	Vip3ch2 vs Vip3ch4
Null hypothesis	LogEC50 same for all data sets	LogEC50 same for all data sets	LogEC50 same for all data sets	LogEC50 same for all data sets
Alternative hypothesis	LogEC50 different from each sets	LogEC50 different from each sets	LogEC50 different from each sets	LogEC50 different from each sets
P value	0.22	< 0.0001	< 0.0001	< 0.0001
F (DFn, DFd)	1.561 (1, 35)	110.4 (1, 35)	294.2 (1, 36)	49.1 (1, 36)
Conclusion ($\alpha=0.05$)	Do not reject null hypothesis	Reject null hypothesis	Reject null hypothesis	Reject null hypothesis
Preferred model	LogEC50 same for all data sets	LogEC50 different from each sets	LogEC50 different from each sets	LogEC50 different from each sets

Table S2. Construction of the chimeric Vip3 proteins from the Vip3Aa and Vip3Ca proteins.

Chimeric Genes	Individual DNA regions								Full ORF of <i>vip3</i> Chimeric Genes		
	DNA Amplicon A				DNA Amplicon B				DNA Amplicon C		
	DNA source	Size (bp)	Forward primer	Reverse primer	DNA source	Size (bp)	Forward primer	Reverse primer	DNA	Forward primer	Reverse primer
<i>vip3 chimera 1</i>	<i>vip3Ca2</i>	610	3	6	<i>vip3Aa45</i>	1836	5	2	A+B	3	2
<i>vip3 chimera 2</i>	<i>vip3Aa45</i>	610	1	8	<i>vip3Ca2</i>	1880	7	4	A+B	1	4
<i>vip3 chimera 5</i>	<i>vip3Aa45</i>	1573	1	10	<i>vip3Ca2</i>	895	9	4	A+B	1	4
<i>vip3 chimera 6</i>	<i>vip3Ca2</i>	1594	3	12	<i>vip3Aa45</i>	874	11	2	A+B	3	2
<i>vip3 chimera 3</i>	<i>vip3 chimera 5</i>	610	1	8	<i>vip3 chimera 6</i>	1836	7	2	A+B	1	2
<i>vip3 chimera 4</i>	<i>vip3 chimera 6</i>	610	3	6	<i>vip3 chimera 5</i>	1880	5	2	A+B	3	4

Table S3. Primers used in construction and sequencing of the genes encoding the chimeric Vip3 proteins.

Primers	Sequences* (5'→3')	Source
<i>Overlapping PCR</i>		
<i>End primers</i>		
[1] Vip3Aa45 FXA/BamHI	<i>cgcgatccatcgaaggtcg</i> atgaacaagaataataactaaat	Designed in this study
[2] Vip3Aa45 FXA/NotI	<i>aaggaaaaagcgccgctt</i> acttaataagagacatcgtaa	Designed in this study
[3] Vip3Ca2 FXA/BamHI	<i>cgcgatccatcgaaggtcg</i> atgaacatgaataataactaaat	Designed in this study
[4] Vip3Ca2 Fxa/NotI	<i>aaggaaaaagcgccgctt</i> atcaatctttccttaata	Designed in this study
<i>Annealing primers</i>		
[5] ch1 20Ca-66Aa_F	<i>gaaatttgataaattaacatttgc</i> tacagaaactagttaaag	Designed in this study
[6] ch1 20Ca-66Aa_R	ctttgaactagtttctgtg <i>caaatgttaatttatcaaatttc</i>	Designed in this study
[7] ch2 20Aa-66Ca_F	gaaaaatttgaggaattaactttg <i>ccacagaaagcactctaagag</i>	Designed in this study
[8] ch2 20Aa-66Ca_R	<i>ctcttagagtgctttctgtg</i> cacaaagttaattcctcaaattttc	Designed in this study
[9] ch5 20-30Aa-33Ca_F	gattaattactttaacatgtaaa <i>tcttacctgcgagaatatttattag</i>	Designed in this study
[10] ch5 20-36Aa-33Ca_R	<i>ctaataaatattctgcgaggttaaga</i> ttacatgttaaagtaattaatc	Designed in this study
[11] ch6 20-33Ca-33Aa_F	<i>cattaaccttaaatgtaaat</i> catatttaagagaactactgc	Designed in this study
[12] ch6 20-33Ca-33Aa_R	gcagtagttcttctaataatga <i>tttacattttaagggttaatg</i>	Designed in this study
<i>Sequencing</i>		
T7 promoter	taatacgactcactatag	pet system manual
T7 terminator	gctagttattgctcagcgg	pet system manual
Sp6 promoter	atttagtgacactatag	pgem-t easy manual
M13 Forward (−20)	gtaaaacgacggccag	pcrtopo2.1 manual
M13 reverse	caggaaacagctatgac	pcrtopo2.1 manual
Vip3 internal 1	gatgtaatgaacaaaattatgc	Designed in this study
Vip3 internal 2	ctaaaacaaaattatcaagtcg	Designed in this study

* The italic and underlined nucleotides indicate the extra base pairs need it to cut the NotI and BamHI close to the edge of the fragment. The italic nucleotides shown the NotI and BamHI restriction sites. The underlined nucleotides are the recognition sequence for the FXA protease. The nucleotides in red are the part of the primer in the overlapping PCR corresponding to the Vip3Ca protein.