



Review

The Potential Impact of Probiotics on the Gut Microbiome of Athletes

Laura Wosinska 1,2,3, Paul D. Cotter 2,3, Orla O'Sullivan 2,3,* and Caitriona Guinane 1,*

- Department of Biological Sciences, Cork Institute of Technology, Bishopstown, T12 P928 Cork, Ireland; Laura.Wosinska@teagasc.ie
- ² Teagasc Food Research Centre, Moorepark, Fermoy, P61 C996 Cork, Ireland; paul.cotter@teagasc.ie
- ³ APC Microbiome Ireland, T12 YT20 Cork, Ireland
- * Correspondence: caitriona.guinane@cit.ie (C.G.); orla.osullivan@teagasc.ie (O.O.); Tel.: +353-21-433-5598 (C.G.); +353-0-254-2222 (O.O.)

Received: 15 August 2019; Accepted: 19 September 2019; Published: 21 September 2019



Abstract: There is accumulating evidence that physical fitness influences the gut microbiome and as a result, promotes health. Indeed, exercise-induced alterations in the gut microbiome can influence health parameters crucial to athletic performance, specifically, immune function, lower susceptibility to infection, inflammatory response and tissue repair. Consequently, maintenance of a healthy gut microbiome is essential for an athlete's health, training and performance. This review explores the effect of exercise on the microbiome while also investigating the effect of probiotics on various potential consequences associated with over-training in athletes, as well as their associated health benefits.

Keywords: athletes; probiotic; microbiome; overtraining; fitness; exercise

1. A Brief Introduction to Exercise and the Microbiome

Our gut microbiome, an elusive and recently designated organ, is under ever greater scrutiny. Uncovering the significance of the microbiome in human health and disease has become possible with the advent of various genomic tools accompanied by improvements in culture-based approaches (culture-omics). The gut contains a rich and diverse microbial ecosystem whose activities can influence the health of the host [1]. More specifically the gut is home to trillions of microbial species [2] that co-exist with human cells. The dominant microbial populations in the gut are bacteria from the Bacteroidetes and Firmicutes phyla, followed by members of the phyla Actinobacteria, Proteobacteria and Verrucomicrobia. The microbiome also consists of a fungal community, including species of Candida and Saccharomyces [3], viruses (primarily bacteriophage) [4], as well as members from the archaeal domain [2]. Despite the microbiome being stable in adults, certain factors can alter its structure, including, diet, antibiotics, probiotics and indeed exercise/fitness (for review see [5,6]). The microbiome appears to have an effect on most aspects of human health. In addition to a role for specific microbes, or groups of microbes, the importance of microbial diversity has been recognized in many studies, with a decreased diversity associated with a number of gastrointestinal (GIT) conditions such as Crohn's disease [7], certain cancers [8] and Type 1 diabetes [9]. An altered microbiome can affect energy metabolism, immune function and oxidative stress, all of which are vital for athlete's performance and overall health (for review see [10]). A potential, non-medical, route to maintaining athlete's health is through probiotics. Probiotics are defined as live bacteria that, when ingested in adequate amounts, confer a benefit to the host [11]. Although such health benefits have frequently been credited to specific bacterial strains from the genera Bifidobacterium and Lactobacillus, the potential of next-generation probiotic (NGP) candidates and designer probiotics is also being recognized.

Nutrients **2019**, 11, 2270 2 of 23

Although it is generally accepted that microbial diversity is an indicator of human health, this hypothesis has been challenged recently by Shade, who postulated that "mechanisms maintaining or changing microbial diversity are many and complex" and assumptions that an increased diversity is better may oversimplify complex mechanisms of health and disease [12]. As previously stated, certain factors including an individual's lifestyle can alter the structure of the microbiome. It has been noted across a number of studies that athletes have increased gut microbial diversity compared with more sedentary controls [13–15]. Clarke et al. [14] demonstrated significant gut microbiome differences between male professional elite rugby players and a cohort of non-athlete male controls. Notably, the athletes had significantly higher gut microbial diversity accompanied by elevated levels of the Akkermansia genus. A subsequent study of the same cohort [13] concluded that athletes had a higher abundance of short-chain fatty acids (SCFA) metabolic pathways. In another instance, Petersen et al. [15], investigated the gut microbiome of cyclists and concluded that 30 out of 33 cyclists also had an increased abundance of Akkermansia. It was also observed that exercise regime is proportional to the abundance of Prevotella, which in turn has been associated with increased branched-chain amino acid (BCAA) pathways, important in muscle recovery. There was also an increase in Methanobrevibacter smithii within the professional cyclist group compared to amateur cyclists, a microbe that utilises H₂ in the colon to make SCFA and Adenosine triphosphate (ATP) [15]. More recently, Cronin et al. investigated the impact of exercise and/or whey protein supplementation over an eight-week period on a cohort of sedentary adults. In this instance, the differences between the microbial communities of the three test populations were more subtle, a fact that could be attributed to the shorter period over which time these interventions took place relative to the potentially life-long practices of some athletes, i.e., microbial architecture is not easily changed and it may take more time to notice significant impacts [16]. Recently, Jang et al. compared the gut microbiome of bodybuilders, distance runners and controls (n = 45), each of the groups were ingesting a different sport specific diet. The results show that bodybuilders ingesting a high-protein and fat diet had increased relative abundance of Faecalibacterium, Sutterella, Clostridium, Haemophilus and Eisenbergiella, but decreased abundances of Bifidobacteria, Parasutterella and Eubacterium. The study also concluded that individuals practicing anaerobic sports show similar gut microbiome patterns to bodybuilders [17]. A pioneering study by Scheiman et al. discovered that the genus Veillonella was enriched in the gut microbiome of marathon runners, compared to non-runners. Furthermore, using a mouse model they demonstrated that Veillonella atypica increased endurance, reduced inflammatory cytokines, and converted lactate to acetate/propionate [18]. The present studies suggest that exercise can alter the microbiome in a beneficial manner, where the alteration in the composition of the microbiome can be associated with increased health parameters and thus possibly affect athletic performance in a beneficial way. Reported gut microbiome changes/differences associated with exercise and fitness are summarized in Table 1.

Whilst exercise confers numerous physiological effects on the host including mood regulation [19], improving cardiovascular symptoms [20], alleviation of fatigue [21] and anti-inflammatory effects, a healthy balance between training load and recovery needs to be maintained to prevent the condition of overtraining occurring [22]. Studies in the past have observed the effect of overtraining on the microbiome; Allen et al. concluded that voluntary and forced exercise altered the mice microbiome in different ways. Of particular interest were the phyla *Tenericutes* and *Proteobacteria*, both elevated in forced exercised mice, compared to mice that exercised voluntarily. *Tenericutes* spp. have been linked to intestinal inflammation in human subjects and *Proteobacteria* bacteria are known for their lipopolysaccharide production [23]. Similarly, Karl et al. observed that there was a large increase in pathogenic taxa post-exercise in soldiers including, *Peptostreptococcus*, *Staphylococcus*, *Peptoniphilus*, *Acidaminococcus*, and *Fusobacterium* and a decrease in more beneficial taxa *Bacteroides*, *Faecalibacterium*, *Collinsellaa* and *Roseburia*. The authors postulated that the increase in more toxic species may possibly explain the observation of increased intestinal permeability within the soldier cohort [24]. More recently, Yuan et al. concluded that excessive exercise had a negative impact on microbial diversity in murine models [25].

Nutrients **2019**, 11, 2270 3 of 23

Table 1.	Studios	of o	vorcico	and the	micro	hiomo
Table L	SHIGHES	αP	XPICISE .	and ine	11116 163	DIOTHE

Subject Group	Microbiome Change	Key Findings	Study Reference
Rugby players	↑Akkermansia, Prevotella, ↓Ruminococcaceae Bacteroides Lactobacillus	Akkermanisa was associated with better immunity and gut barrier function while Prevotella was correlated to biosynthesis of branched-chain amino acid (BCAA) pathways which help with muscle recovery	[14]
Professional male athletes	<i>†Akkermansia</i> metabolic pathways/higher short-chain fatty acids (SCFA) metabolic pathways	Rugby players had a higher abundance of health-promoting <i>Akkermansia</i> genus, which has been associated with an improved gut barrier function and immune function stimulation.	[13]
Cyclists	†Prevotella, Methanobrevibacter Smithii	Prevotella was correlated to biosynthesis of BCAA pathways which help with muscle recovery, M. smithii has been associated with degradation of H ₂ which is used to make ATP/SCFA resulting in a more energetically efficient body	[15]
Sedentary adults challenged to eight week exercise regime	↓in <i>Archaea</i> species and an ↑ in microbial diversity	Microbial diversity has been linked to an overall better health	[16]
Marathon runners	↑Veillonella	Veillonella has been shown to metabolize lactate to SCFA, lower inflammation and increase performance in murine models	[18]
Bodybuilders and Distance runners	↑Faecalibacterium, Sutterella, Clostridium, Haemophilus, Eisenbergiella ↓Bifidobacterium, Parasutterella and Eubacterium	Different sports and their sport specific diets can affect the gut microbiome in different ways	[17]

Various studies throughout the years have investigated overtraining and its effect on human health particularly host immunity [26,27] and susceptibility to infection [22,28,29]. The potential side effects of overtraining are illustrated in Figure 1. Furthermore, overtraining has been associated with an increased incidence of mental illnesses in athletes [30–33] and oxidative stress [34,35], where studies in animal models have shown the relationship between the microbiome and oxidative stress [36,37]. One of the most common side effects of overtraining is gut dysfunction and it has been frequently noted in athletes [38,39]. Gastrointestinal (GIT) symptoms are frequently observed with athletes who tend to travel for training or competition, purposes [40,41]. Antibiotics can be used to overcome the GIT infections, however, certain complications may arise when using antibiotics. Firstly, studies have shown that the microbiome takes about six weeks to resemble its original state post antibiotic therapy [42], it increases possible tendon rupture [43] and can cause antibiotic associated diarrhea [44].



Figure 1. The effects of overtraining on the wellbeing of an athlete.

Nutrients **2019**, 11, 2270 4 of 23

The review aims to provide a panoramic view of what is currently known about probiotics and their effect on gut microbiome of athletes. A recent systematic review by Moller et al. [45] has provided an overview of the effects of traditional probiotics on physically active individuals and athletes, with a particular focus on the clinical trials performed to date. Here we explore the relationship between fitness and the gut microbiome focusing particularly on overtraining. We also explore the available literature on how, possibly, probiotics can help to alleviate or prevent the different side effects of overtraining. We also discuss next generation and designer probiotics, as well as the safety and toxicity considerations associated with novel strains.

2. Traditional Probiotics

Traditionally, the majority of probiotic strains were representatives of the lactic acid bacteria group (LAB) i.e., *Bifidobacterium*, *Lactobacillus* [46], however other bacterial and yeast strains are also commonly used such as *Escherichia coli* and *Saccharomyces* respectively. Although probiotics have existed for quite some time, it is only recently that their commercial potential has been realized, with the market expected to exceed \$67 billion by 2024 [47]. Probiotics can consist of one or a combination of a few strains, either as capsules, powders or as a component of a food e.g., yoghurt [48]. One of the main concerns relating to the probiotic market is that some strains do not have substantial proof of efficacy. It is also important for consumers to be aware that the benefits of one probiotic strain cannot be inferred to another [48,49]. Nonetheless, there is increasing evidence of the ability of specific probiotics to, for example, modulate the immune system, impact on tight-junction proteins and inhibit pathogen colonization; this list is not exhaustive, and the benefits of probiotics are strain-dependent. Focusing on athletes as noted above, extreme exercise can be associated with undesirable symptoms and specific probiotics can help to ease or prevent certain gastrointestinal disorders, help with the brief immunosuppression period, and reduce susceptibility to infections. Ways in which existing and novel probiotics could contribute to athlete health are provided in Table 2.

2.1. Escherichia Coli

Escherichia coli, is the most diversely studied prokaryotic model organism in science. *E. coli* belongs to the *Enterobacteriaceae*, a large family of Gram-negative bacteria consisting of commensal and pathogenic bacteria, *E. coli* is a facultative anaerobe, meaning it can either respire or ferment, depending on the environment, which maximises its growth in the gut. *E. coli* Nissle 1917 (EcN) possesses antagonistic properties against *Salmonella*, *Yersinia*, *Shigella and Listeria* [50]. A study by Wehkamp found that EcN induces the production of defensin in epithelial cells [51]. Various studies have also demonstrated that *E. coli* affects the intestinal epithelial barrier [52,53], additionally EcN can be used for irritable bowel disorder [53], constipation (for review see [54]), and has pro-inflammatory potential [55]. EcN has been shown to repair the gut barrier function in vitro [53] and in murine models [52]. Henker et al, investigated whether EcN is an effective therapeutic agent against acute diarrhea in infants and toddlers. The randomized double blind placebo controlled trial, including 113 subjects concluded that, EcN successfully reduced the duration and incidence of diarrhea compared to placebo, and improved the general health of the subject [56].

EcN has shown great promise in mitigating effects of gut barrier dysfunction, diarrhea and impairment immune system, experienced often with the condition of overtraining. In order to prove its efficacy more research is needed, especially involving athlete cohorts.

2.2. Lactobacillus

Lactobacillus species are Gram-positive bacteria belonging to the lactic acid bacteria group, capable of lactic acid fermentation metabolism. Lactobacilli are frequently resistant to bile salts; an important probiotic trait as it allows for the probiotic to survive in the hostile acidic environment of the gastrointestinal tract [57]. Lactobacilli are frequently found in various fermented foodstuffs, silage, human gut and the vagina [49–58]. Lactobacilli are amongst the most widely used and characterized

Nutrients **2019**, 11, 2270 5 of 23

probiotics to date. Various strains from the species: *Lactobacillus rhamnosus*, *Lactobacillus plantarum* and *Lactobacillus acidophillus* have been considered important probiotics, each strain exhibiting individual functions. A small selection of very many examples are provided here, *Lactobacillus casei* GG has been found to shorten diarrheal distress in subjects suffering from viral gastroenteritis [59], *Lactobacillus johnsonii* BFE 6128 and *Lb. plantarum* BFE 1685 have been shown to aid in modulating the immune system by inducing the secretion of the cytokine IL-8 in vitro [60], *Lb. rhamnosus* 4B15 and *Lactobacillus gasseri* 4M13 have been suggested to inhibit the expression of inflammatory cytokines at transcriptional level in vitro thus showing anti-inflammatory potential [61]. Among the aforementioned benefits, *lactobacilli* are capable of inhibiting enteric pathogens through the production of lactic acid, bacteriocins and hydrogen peroxide [62]. It is worth to mention that *lactobacilli* are biofilm matrix formers, which makes the bacteria more resistant to antibiotics, and could potentially allow for longevity of the bacteria in the gut [63].

Most studies to date have investigated the potential impact of *lactobacilli* in either animal models or non-active human populations and, as of today, little has been done in athletes. Cox et al investigated whether Lb. fermentum VRI-003 had any effect on mucosal immunity in elite male distance runners. They observed a significant decrease in the duration and severity of respiratory illness, and a two-fold increase in interferon gamma (INF $^{\circ}$) [64]. Another study examined Lb. fermentum (PCC $^{\otimes}$) and its effects on gastrointestinal and respiratory health in competitive cyclists. In this study West and colleagues deduced that Lb. fermentum (PCC®) was successful in reducing the severity of gastrointestinal symptoms in male cyclists, in addition, it significantly reduced (20%-60%) cytokine imbalance caused by acute exercise. Moreover, the study also concluded that the severity and duration of lower respiratory illness decreased in male cyclists but increased in female cyclists, taking the probiotic compared to placebo [65]. It is also worth noting that a recent study by Shing et al, reported that a multi-strain probiotic formulation consisting of Lb. acidophilus, Lb. rhamnosus, Lb. casei, Lb. plantarum, Lb. fermentum, B. lactis, B. breve, B. bifidum and Streptococcus thermophilus increased run to fatigue time in male runners, and showed small to moderate improvement in gut permeability [66]. Overtraining can result in a compromised immune system and gut barrier function; in particular lactobacilli are capable of reducing the severity and duration of upper respiratory tract infections (URTI) and gastrointestinal problems, as well as modulating the immune system and increasing performance and thus a good candidate probiotic for athletes.

2.3. Bifidobacterium

Bifidobacteria are Gram-positive anaerobic bacteria from the phylum Actinobacteria. Associated strains are typically among the first colonisers of the infant gut and contribute to the infants' immune system maturation and the utilisation of certain dietary components [67]. They are generally bile-acid resistant [68] and represent a large proportion of the bacterial microbiome obtained from infant faecal samples, on average about 60%–91% present in breast fed infants, but proportions decrease greatly later in life [69]. Numerous strains from the species: *Bifidobacterium breve*, *Bifidobacterium longum*, Bifidobacterium bifidum exhibit probiotic properties, several examples are listed below. The Bifidobacteria genus is represented by many different strains that have been assigned probiotic properties, shown here is a small selection of strains, although many more have been noted in literature A mixture of B. breve, B. longum and Bifidobacterium infantis has been associated with the treatment of antibiotic associated diarrhea [70], while two Bifidobacterium strains: B. breve strain Yakult and B. bifidum strain Yakult have been shown by in vitro studies to have a possible role in modulating the immune system [71]. Even though bifidobacteria are frequently used in probiotic preparations and many studies have elucidated their benefits in vivo, little has been studied in athlete populations. West et al investigated whether a single strain of a probiotic (B. animlais subsp. lactis BI-04) compared against a multi-strain probiotic (Lb. acidophilus NCFM and B. animalis subsp. lactis BI-04) was effective in reducing the risk of upper respiratory tract infection in physically active individuals. The study concluded that B. animalis subsp. lactis BI-04 reduced the risk of upper respiratory illness by 27%. The multi-strain probiotic

Nutrients **2019**, 11, 2270 6 of 23

did not show a significant decrease in risk of URTI [72]. Another worthwhile study investigated a multi-strain probiotic composed of *B. bifidum*, *B. longum* and *Lb. gasseri* and its effect on the duration and incidence of infection in elite rugby players. The findings were that the multi-strain probiotic formulation reduced both the incidence and duration of URTI and gastrointestinal symptoms [73]. In addition to the aforementioned studies, Jäger and colleagues concluded that a probiotic composed of *B. breve* BR03 and *S. thermophilus* FP4 has a positive effect on the reduced performance and range of motion followed by intense muscle damaging exercise [74]. More recently, Pugh et al. demonstrated that a four week supplementation of *Lb. acidophilus* (CUL 60/ CUL 21), *B. bifidum* (CUL20) and *B. animalis* subsp. *Lactis* (CUL34) successfully lowered the incidence and severity of GIT symptoms both during training and a marathon race [75]. *Bifidobacteria* supplementation can positively help with the burden of gastrointestinal distress found often in overtrained athletes.

2.4. Saccharomyces

Saccharomyces is another commonly used probiotic; it is a non-pathogenic yeast, and is one of the most common fungal species found in the microbiome [3]. The gut mycobiome represents about 0.1% of the total microbiome [76]. Saccharomyces boulardii is most widely used for the treatment of traveler's diarrhea [77]; it can prevent antibiotic-associated diarrhoea and treat recurrent Clostridium difficile infections [78]. Additionally, research suggests that taking S. boulardii benefits irritable bowel syndrome sufferers [79]. It is not to be overlooked that immunocompromised patients are at risk of fungemia, if supplementing with Saccharomyces [78]. The effect of S. boulardii on diarhoea was elucidated by Kurugol and Koturoglu, where they enrolled 200 young children in a randomized placebo study. The study concluded that S. boulardii successfully reduced the duration of diarrhea and hospital stay associated with it, compared to placebo [80]. In a similar study, Billoo et al. investigated S. boulardii and its possible impact on diarrheal episodes in 100 children, the study concluded that the probiotic reduced the number of diarrhea episodes by 50%, lowered stool frequency and the duration of the illness [81].

It can be postulated that *S. boulardii* could possibly be an effective treatment for athletes and their reoccurring gastrointestinal problems.

Table 2. Traditional probiotics and next-generation probiotics and their benefits.

Colon, oral cavity, breast milk and vagina Yoghurt, kombucha, sauerkraut, kefir etc. Bile resistant, easing lactose intolerance, antibiotic-associated diarrhoea, eczema, immune system modulation, cholesterol lowering abilities Travellers' diarrhoea, antibiotic-associated diarrhoea, preventing recurring Clostridium difficile infections, irritable bowel syndrome Antagonistic properties against a variety of pathogens, production of defensin, tight-junction protein modification, irritable bowel disorder, constipation, pro-inflammatory properties and colon cancer Immune system modulation, intestinal homeostasis [92–94] Akkermansia Colon Co	Probiotic Genus *	Found in the Body	Dietary Source:	Potential Benefits Attributed to Specific Strains	References
Colon, oral cavity, breast milk and vagina Saucharomyces Colon, decaying fruit, plants, soil, insects Wine, yoghurt, kombucha, sauerkraut etc. Wine, yoghurt, kombucha, sauerkraut etc. Wine, yoghurt, kombucha, soil, insects Wine, yoghurt, kombucha, soil, insects Wine, yoghurt, kombucha, sauerkraut etc. Wine, yoghurt, kombucha, sauerkraut etc. Travellers' diarrhoea, antibiotic-associated diarrhoea, preventing recurring Clostridium difficile infections, irritable bowel syndrome Antagonistic properties against a variety of pathogens, production of defensin, tight-junction protein modification, irritable bowel disorder, constipation, pro-inflammatory properties and colon cancer Immune system modulation, intestinal homeostasis [92–94] Akkermansia Colon - Golon	Lactobacillus	Colon, gut and vagina		modulation, alleviating inflammation, lowering cholesterol, cancer protection, modulating brain activity, preventing	[60–62,68,82–87]
Saccharomyces Colon, decaying fruit, plants, soil, insects Soil, insects Soil, insects Colon Colon Capsules Colon Colon Capsules Colon Col	Bifidobacterium			antibiotic-associated diarrhoea, eczema, immune system	[68,71,88–90]
Antagonistic properties against a variety of pathogens, production of defensin, tight-junction protein modification, irritable bowel disorder, constipation, pro-inflammatory properties and colon cancer Bacteroidetes Colon - Immune system modulation, intestinal homeostasis Colon - Gut barrier function, fat mass storage, glucose homeostasis, immune system stimulation, production of Vitamin B12 Faecalibacterium Colon - Immune system modulation, ease inflammation [102,103] Improve insulin sensitivity, increase energy production, production, produce Vitamin B12, maintain intestinal homoestasis, colon [104–107]	Saccharomyces	, ,	, 0	Travellers' diarrhoea, antibiotic-associated diarrhoea, preventing recurring <i>Clostridium difficile</i> infections, irritable	[77–79]
Bacteroidetes Colon - Immune system modulation, intestinal homeostasis [92–94] Akkermansia Colon - Gut barrier function, fat mass storage, glucose homeostasis, immune system stimulation, production of Vitamin B12 [95–101] Faecalibacterium Colon - Immune system modulation, ease inflammation [102,103] Improve insulin sensitivity, increase energy production, production produce Vitamin B12, maintain intestinal homoestasis, colon [104–107]	Escherichia coli	Colon	Capsules	Antagonistic properties against a variety of pathogens, production of defensin, tight-junction protein modification, irritable bowel disorder, constipation, pro-inflammatory	[50–54,91]
Akkermansia Colon - Gut barrier function, fat mass storage, glucose homeostasis, immune system stimulation, production of Vitamin B12 Faecalibacterium Colon - Immune system modulation, ease inflammation [102,103] Improve insulin sensitivity, increase energy production, produce Vitamin B12, maintain intestinal homoestasis, colon [104–107]	Bacteroidetes	Colon	-		[92–94]
Faecalibacterium Colon - Immune system modulation, ease inflammation [102,103] Improve insulin sensitivity, increase energy production, Eubacterium Colon - produce Vitamin B12, maintain intestinal homoestasis, colon [104–107]	Akkermansia	Colon	-	Gut barrier function, fat mass storage, glucose homeostasis,	[95–101]
Eubacterium Colon - produce Vitamin B12, maintain intestinal homoestasis, colon [104–107]	Faecalibacterium	Colon	-	Immune system modulation, ease inflammation	[102,103]
Ü , ,	Eubacterium	Colon	-		[104–107]

^{*} Some of the mentioned genera are regarded as potential probiotics.

Nutrients **2019**, 11, 2270 8 of 23

3. Next-Generation Probiotics

Next-generation probiotics still merit the definition of a probiotic and although they have not been used as health modulators to date, have potential to be of importance to the probiotic market. Next-generation probiotics are outside of the commonly used probiotic spectrum (*Lactobacilli*, *Bifidobacteria* etc.), however large-scale genomic initiatives have identified putative probiotic strains with potential health benefit, mainly from the genera *Bacteroides*, *Akkermasia*, *Faecalibacterium* and *Eubacterium*.

One promising candidate is a strain of *Bacteroides fragilis* which has been proposed to stimulate T-cell immune responses in vitro [108], this is of interest as athletes have decreased numbers of T cells following intensive anaerobic exercise [109]. Similarly, strains of, Bacteroides acidifaciens have been suggested to induce IgA production in murine models and as a consequence elevating the production of IgA+ B cells and B cells; this is important because IgA plays a pivotal role in upholding intestinal homeostasis namely preventing the adherence of pathogens in the intestine [94], studies in the past have demonstrated the decrease in salivary IgA in endurance athletes [110,111] as well as when decreased it has been linked to a higher incidence of URTI [112]. Studies have illustrated the potential role of Akkermanisa in obesity, diabetes and inflammation (for review see [96]). Akkermansia muciniphilia has been associated with maintaining gut barrier function, which can be compromised in endurance athletes [113,114], and glucose homeostasis [97,98] and its direct correlation to athletic performance [115] and additionally it has the potential to stimulate the immune system [97,99–101]. The most recent study has concluded that A. muciniphilia is capable of synthesising Vitamin B12 de novo, however whether humans can benefit from the product remains unclear [95]. Considering that recent studies demonstrated an increase in Akkermansia in athletes this is a species that definitely warrants further investigation for use in athletes [13–15]. Another potential probiotic is the Faecalibacterium prausnitzii. F. prausnitzii A2-165 has been proposed to have immunodulatory capabilities, through induction of IL-10 and T-cell responses in human and murine dendric cells [102,103,116]. Another worthy strain belongs to the genus Eubacterium. Eubacterium hallii L2-7 has been demonstrated to improve insulin sensitivity and increase energy metabolism in obese and diabetic murine models [104], E. hallii DMS 3353 and E. hallii DSM 17630 have been indicated to produce Vitamin B12 and maintain intestinal homeostasis through the utilisation of glucose and various fermentation intermediates like acetate and lactate in vitro studies [105], while, Eubacterium limosum JCM 6421 has been suggested to produce SCFAs that have been shown to decrease the levels of the pro-inflammatory cytokine IL-6 and increase mucosal integrity [107].

Preliminary research has shown the benefit of the various new generation probiotics however more proof is required to ensure their efficacy, as well as testing for their safety in humans. The potential benefits of next-generation probiotics are also summarized in Table 2.

4. Designer Probiotics

In recent years, research has uncovered numerous functions provided by probiotics, including antagonistic activities, anti-inflammatory and tight-junction modification all of which are of particular benefit to athlete health. Designer probiotics are simply commensal strains of bacteria that have been engineered or modified in a way that they are able to resist the countless stresses that they meet both outside and inside our bodies (lyophilisation/manufacturing/acids/temperature) or simply just to improve the functions most beneficial to the host. The advent of synthetic biology has allowed for engineering of both probiotic strains and commensal strains to acquire and execute new functions (for review see [117]). The most recent concept in the probiotic field is the idea of a "biodrug", which essentially allows for oral administration of a live recombinant probiotic strain of bacteria for the treatment/prevention of diseases. In the past 10 years, recombinant probiotics have been engineered, for the delivery of therapeutic molecules (usually proteins, fragile in nature, easily denatured by changes in the environment) such as DNA, peptides, single-chain variable fragments, various enzymes or cytokines. Probiotics are excellent vectors of transmission for such therapeutics, due to; a) their

Nutrients **2019**, 11, 2270 9 of 23

ease of colonization and direct delivery to the mucosa; b) resistance to gastric acid and bile salts; c) continued colonization and longevity of protection against pathogens; d) cost of delivery being relatively non-expensive; e) prolonged shelf-life and stability (for review see [118]). Designer probiotics could potentially be an attractive proposition to athletes due to their amenability to being tailored to the athletes specific needs.

Lb. plantarum NC8 which has been engineered to possess angiotensin-converting enzyme inhibitory peptides (ACEIPs), showed reduction of high blood pressure in rats [117]. It has also been shown that engineering Lactococcus lactis NZ29000, B. longum and Lb. gasseri ATCC 33323, managed the symptoms of diabetes and induced insulin secretion. More probiotics are currently under investigation for pathogen infection, cancer or human immunodeficiency virus (HIV), however certain issues arise with engineering probiotics [117]. Tackling the problem of biocontainment should be addressed to prevent the spread of genetically modified microorganisms (GMO) organisms into the natural environment [118] and, of course, the consumer acceptance of GMO products is still a major issue. With that being said, designer probiotics are a great alternative to todays, often impermanent medication.

5. Safety Considerations

When a novel probiotic strain is being considered, evidence needs to be presented to prove its efficacy and safety for human consumption. Even though probiotics are generally considered safe for use, certain side effects may arise. In a statement released by the World Health Organisation (WHO) and the Food and Agriculture Organisation of United Nations (FAO) in 2002, probiotics in theory can have four possible side effects: 1) they can be responsible for lateral gene transfer of antibiotic resistant genes i.e. Lactobacillus spp. 2) They can be responsible for systematic infections i.e. Fungemia. 3) They can stimulate the immune system excessively due to their immune system modulation potential, possibly causing inflammation; and 4) they are detrimental to metabolic activities, as some strains produce D-lactate which can be linked to d-acidosis (for review see [119]). However, with the advent of whole genome sequencing some of these issues can be identified or at least indicated using bioinformatics tools. Genus, species and strain level need to be provided, the nomenclature must agree with the International Committee of Systemics of Prokaryotes, and covered by the International Code of Nomenclature (ICN) of Prokaryotes for bacteria, for fungi, the nomenclature is covered by the ICN for algae, fungi and plants, the organism under consideration should be deposited in a recognized culture collection and identified by the means of current methodologies such as whole-genome sequencing (WGS). As well as using WGS to identify the organism, sequencing should be used to fully analyse the genetic content of the probiotic (both chromosomal and extra-chromosomal) to determine the presence of any transferable antimicrobial resistance loci, virulence factors or production of adverse metabolites that might deem the strain unsafe for human consumption [120]. The European Food Safety Authority (EFSA) guidelines also require full antimicrobial resistance (AMR) testing. AMR testing requires two sets of data, firstly the phenotypic testing determining the minimum inhibitory concentration (MIC) of the antimicrobial, using internationally recognized standards (ISO or similar), serial two-fold dilutions should be applied using relevant controls. The results should be compared to established and published MIC cut off values. Secondly, WGS is required to test for the presence of known AMR genes. Probiotic supplements are sold in many different doses depending on the product and strain. Most commercially available probiotics contain about 1-10 billion colony forming units (CFU) per serving. Generally, the CFU of the product should be equivalent to human studies, showing a positive effect. As of now no strict guidelines exist on the dosage of probiotics [121,122]. Consumers should also be aware that for the most part multi-strain probiotics are more effective than monostrain preparations [123]. It is to be assumed that, new probiotic strains with the intent of consumption, should be of human origin (although Saccharamyces spp., is an exception), their safety, benefits, and dose ratio, should be clearly stated and assessed (for review see [124]) Before the consumer purchases a probiotic it is recommended to research the probiotic strain best suited for their needs paying particular attention to studies performed in human subjects. Customers should ensure the commercial probiotic dosage is consistent with

the dosage used in human studies. An important consideration to keep in mind is that the use of probiotics in sport is still a novel area; therefore, most studies to date are preliminary pilot studies, where the dosage and exact benefit of the strain are not fully optimized, and therefore should be reasoned with caution.

6. Probiotics in Treating the Overtraining Syndrome

Overtraining can put the athlete at risk of developing asthma [125], infection such as upper respiratory illness [126,127], gastrointestinal complaints [128] as well as depression and anxiety [129]. Various other symptoms such as immunity suppression and chronic fatigue are also very common [26]. The symptoms can have negative effects for the athlete; impeding progress and decreasing performance. Various studies have explored the positive relationship between athletes and probiotics, and how they can possibly be used to ease or prevent symptoms associated with overtraining. The effects of probiotics on athletes and physically active individuals are summarized in Table 3.

Table 3. Summary of studies using probiotics on physically active and athlete cohorts.

Subject Group	Intervention	Result	Limitations of the Study	References
20 male elite distance runners. Randomized double-blinded, placebo controlled trial,	Lb. fermentum VRI-003	\downarrow risk and severity of respiratory systems \uparrow INF $^{\checkmark}$	A small sample size	[64]
99 male and female competitive cyclists. Randomized, double-blinded, placebo trial	Lb. fermentum (PCC®)	↓ severity of GIT symptoms↓ severity/duration of lowerrespiratory illness↓ cytokine imbalance	Inclusion criteria for antibiotics was only four weeks, study relied on self-reported illness, reported a higher rate of lower respiratory illness in females.	[65]
10 male runners. Randomized, double-blinded, placebo trial	Lb. acidophilus, Lb. rhamnosus, Lb. casei, Lb. plantarum, Lb. fermentum, B. lactis, B. breve, B.bifidum and Streptococcus thermophilus	↑ increased run time to fatigue, small to moderate improvement in gut permeability	Study only investigated males, sample size was too small, short study duration of $4\ weeks$.	[66]
465 physically active males and females. Randomized double blind placebo controlled trial	B. animalis subsp. lactis BI-04 (BI-04), Lb. acidophilus NCFM and B.animalis subsp. lactis BI-04 (NCFM and BI-04)	↓ the risk of URTIs by 27%	No separation between recreational and professional athletes, study relied on self-reported illness data, inclusion criteria for antibiotic use was only four weeks.	[72]
30 male elite male rugby players. Randomized, double-blinded, Placebo trial.	B. bifidum B. longum Lb. gasseri	↓ in the incidence of URTI/GIT	Relatively small sample size, study only looked at males, short study duration of 4 weeks, relied on self-reported illness data as opposed to measuring immune system markers	[73]
15 resistance-trained men. Randomized, double-blinded, placebo trial.	B. breve BR03 and S. thermophilus FP4	positive effect on the reduced performance and range of motion followed by intense muscle damaging exercise	Small sample size, looked at males only, didn't include antibiotic use in inclusion criteria, short study duration of 3 weeks	[74]
24 amateur athletes	Lb. acidophilus (CUL60/CUL21), B. bifidum (CUL20), B. animalis subsp. Lactis (CUL34)	↓ incidence and severity of GIT symptoms, both during training and a marathon race	Small sample size, ratio of males to females was skewed, athletic levels weren't standardized, not double-blinded.	[75]
24 amateur male athletes	Lb. rhamnosus IMC 501 and Lb. paracasei IMC 502	↓antioxidant levels followed by exercise	Small sample size, short study duration of 4 weeks, studied males only, study did not include placebo in control group	[130]
23 endurance male athletes Randomized, double-blind, placebo controlled trial.	EcologicWPerformance or OMNi-BiOTiCWPOWER,	↓ zonulin ↓TNF-alpha	Small sample size, only looked at men, looked at only one marker or impaired intestinal permeability	[131]
27 trained amateur athletes	Lb. acidophilus LAFTI®	↓fatigue	Small sample size, sample size of control group was significantly lower than test group, the ratio of male to females was skewed, not randomized or double blinded.	[132]
44 university student athletes. Randomized, double-blind, placebo controlled trial.	Lb. gasseri OLL2809 combined with alpha-lactalbumin	prevents the exercise induced drops in Natural Killer cells positive effect on minor fatigue ↑mood from a depressive state	Study only looked at males, all participants were at a university level, relied on self-reported illness, short study duration of 4 weeks	[133]
33 highly trained individuals. Randomized, double-blinded, placebo controlled trial.	B. bifidum W23, B. W51, Enterococcus faecium W54, Lb. acidophilus W22, Lb. brevis W63, and Lactococcus lactis W58	↓drops of tryptophan levels caused by intense exercise ↓incidence of URTI's	Relatively small sample size, women were overrepresented; the severity of illness could not be calculated due to lack of replies from participants.	[134]

6.1. Antioxidant Boosting

Oxidative stress is the imbalance between oxidant and antioxidant levels in the body; it results in the creation of reactive oxygen species (ROS). Our bodies counteract them with special enzymes that neutralize the reactive species and if the species are not neutralized then they can compromise the cell [135]. Diseases such as rheumatoid arthritis, heart disease, Parkinson's disease and aging can all be linked to oxidative stress. Intense physical regime coupled to the increased oxygen consumptions results in athletes having a higher abundance of reactive oxygen species [130]. Various probiotic species have been reported to have antioxidant activities; Lactobacillus delbrueckii ssp. bulgaricus, Lactobacillus delbrueckii ssp. lactis, Lb. acidophilus, Lb. casei [136]. Studies have also demonstrated the antioxidant activity of B. animalis, which was found to absorb hydroxyl radicals and superoxidase anions in vivo, as well as, increasing the general antioxidase activity in murine species [135]. Certain probiotic strains have also been found to manufacture metabolites that have antioxidant abilities such as; Lb. fermentum E3 and E18 have been found to produce glutathionine (GSH), and Clostridium butyricum has been detected to produce the SCFA butyrate [135]. SCFAs have health-aiding properties and are thought to help with the immune system [137,138] as well as insulin resistance [139]. Martarelli et al has investigated if two probiotic strains (Lb. rhamnosus IMC 501 and Lb. paracasei IMC 502) have any effect on the oxidative stress of athletes during a four-week time frame of intense training [130]. The study came to two findings; firstly, intense exercise significantly increased the oxidative stress in athletes, and secondly, the probiotic course successfully lowered the oxidative stress caused by the intense exercise.

6.2. Tight-Junction Protein Modification

The mucosal layer of the intestine is made up of several constituents namely; enterocytes and epithelial cells, which are interconnected by tight junctions. This intestinal barrier is pivotal in nutrient and water absorption, while preventing harmful substances entering the bloodstream [140,141]. During intense continuous exercise, cardiovascular and thermoregulatory responses augment the blood flow, increasing the phosphorylation state of the tight-junction proteins consequently increasing their permeability (Figure 2). Increased permeability can cause; vulnerability to allergies and infections, along with various gastrointestinal complaints [142]. A randomized, double-blinded, placebo-controlled trial has investigated two probiotic brands in relation to markers of intestinal barrier integrity on 23 endurance male athletes. The probiotics of choice were Ecologic®Performance (*B. bifidum* W23, *B. lactis* W51, *Enterococcus faecium* W54, *Lb. acidophilus* W22, *Lb. brevis* W63, *L. lactis* W58) and OMNi-BiOTiC®POWER (*E. faecium* W54, *Lb. acidophilus* W22, *Lb. brevis* W63, *L. lactis* W58, *B. bifidum* W23, *B. lactis* W51). The study investigated the concentration of Zonulin in faeces, a marker of impaired gut barrier. The study concluded that the probiotic treatment effectively decreased Zonulin concentrations in faeces from above normal to below normal, during the 14 days of the study, when compared to a placebo [142].

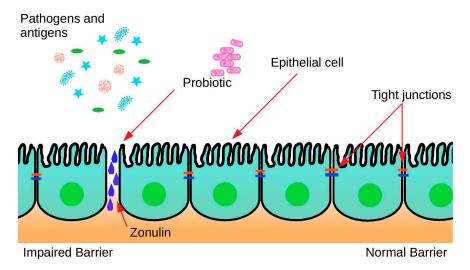


Figure 2. Decreased intestinal permeability.

6.3. Immune System Modulation

It has been documented that overtraining can lead to immunosuppression within the individual [26]. The tissue trauma associated with excessive training results in overproduction of cytokines (IL6, TNF-alpha) which subsequently lead to chronic fatigue-like behaviour in athletes, followed by the induction of a humoral response, resulting in the suppression of cell-mediated immunity resulting in a higher chance of infections (for review see [143]).

Much research has focused on potential probiotics and their strain specific effect on the immune system. Throughout the years several different probiotics have been found to influence the host immune system. Studies have shown that Lb. plantarum 299v is capable of inducing mucin, MUC2 and MUC3, which is important in hindering pathogen colonization [144,145]. Mucin is a large glycosylated protein which protects the intestines by a minimum of three mechanisms: a) it creates a gel-like structure to trap various molecules; and b) it can bind to various molecules, pathogens or proteins through very specific binding sites and c) through disposal of trapped organisms or proteins [146]. Escherichia coli have also been assigned some immunomodulatory functions in recent years, EcN has been found to induce beta-defensins, defensins are small peptides which are the innate immune systems first line of defence, they are produced by epithelial cells of the intestinal tract, skin and lung. Defensins have a broad antagonistic spectrum against various pathogens and are active against both Gram-positive and negative bacteria as well as fungi [147]. A cheese product containing live a culture of Lb. rhamnosus HN001 and Lb. acidophillus NCFM has been proven to improve the ability of natural killer cells (NK Cells) to kill cancerous cells and phagocytosis in elderly subjects. Clancy et al. [132] investigated if a daily dose of Lb. acidophilus could reverse a defect in INF-y secretion in fatigued athletes. The study has concluded that treatment with Lb. acidophillus has significantly increased IFN-y secretion in fatigued athletes to comparable levels of the control cohort. Another worthy study has investigated the effect of probiotic supplementation on respiratory infections and gastrointestinal symptoms of 141 marathon runners. The probiotic of choice was *Lb. rhamnosus GG*, compared with a placebo pill. The findings of the study were as follows: there was no significant improvement in the URTI, however, Lb. rhamnosus GG decreased the duration of the gastrointestinal symptoms by 33% during the training period, compared to the placebo [148]. Various studies have elucidated immune modulation induced by Akkermansia. A study by Derrien et al. determined that A. muciniphilia is capable of up regulating genes involved in antigen presentation, as well as aiding in B and T cell maturation [100]. Ottman also investigated A. muciniphilia and its immune modulation activity, and concluded that Akkermansia could activate Toll like receptor 2 (TLR) and TLR4 and induce IL8 and IL10 [101].

Nutrients **2019**, 11, 2270 14 of 23

6.4. Infection

Overtraining can lower the immune system and in turn result in an "open window" period, where sensitivity to infection is increased [126,149]. Athletes are vulnerable to GI, URTI, and skin infections [150].

To benefit the residential flora, probiotic strains need to show antagonistic activities against pathogens. There are two types of antimicrobials produced by certain probiotic strains: high molecular weight compounds called bacteriocins, and low molecular weight compounds including, but not restricted to: hydrogen peroxide, lactic acid, acetic acid and reuterin [151]. Lactic acid, acetic acid and hydrogen peroxide are the most common antimicrobial compounds secreted by probiotic species. LMW compound production is specific to its species, and the effectiveness of microbial inhibition is directly proportional to the amount of organic acid produced [152]. Bacteriocins are small peptides that have been ribosomally-synthesized which possess antagonistic properties against other bacteria, to which the producing strain is immune (for review see [153]). Bacteriocins can either be narrow spectrum (bactericidal against closely related bacteria) or broad spectrum (bactericidal against a wide variety of strains (for review see [151,154]). They can be classified into three groups, Class 1 bacteriocins are lantibiotics which possess lanthionine or beta-metyllanthionine residues, like nisin, Class 2 can be classified as non-lanthionine containing bacteriocins such as lactacin F, and finally bacteriolysins which are non-bacteriocin lytic proteins like lysostaphin [153]. Bacteriocins are an important trait for a probiotic to possess, they can aid in the survival of the producer within a gut [155], help inhibit the proliferation of other bacteria and function as sensing molecules (for review see [155,156]). The production of bacteriocins is dependent on a variety of factors including, the bacterial species producing the antimicrobial compound, and the environmental conditions present. Different bacteria produce various types of bacteriocins, for example, Lb. reuteri is known to produce reuterin, L. lactis produces Nisin A, E. faecalis DS16 makes Cytolisin, Lb. plantarum manufactures Plantaricin S and Lb. acidophillus makes acidophilicin. The production of antimicrobial entities is an important probiotic trait. It allows for longer survival of the producer in the intestine, as well as having the potential to outcompete pathogenic bacteria maintaining a healthy bacterial balance in the gut. Athletes tend to be prone to gastroenteritis and other infections, and the antimicrobial compounds produced by the probiotics can help in easing or preventing the symptoms.

6.5. Mental Health

The ubiquity of depressive symptoms in elite athletes ranges from 4%–68%, with the consensus that female athletes are more likely to develop depressive episodes than male athletes. Athletes, who take part in individual sports, rather than team-based sports, tend to be more susceptible to depressive symptoms. Elite athletes face risk factors that contribute to depression including, genetic factors, environmental factors, injury, competitive failure, retirement from sport, pain, concussion and of course overtraining [157].

A study by *Sashihara* [133] and colleges investigated if *Lb. gasseri* OLL2809 combined with alpha-lactalbumin decreases symptoms of depression. The study has found that *Lb. gasseri* OLL2809 has successfully elevated symptoms of depression in college athletes. Similarly, Strasser et al. investigated the effect of a multi-species probiotic formulation (*B. bifidum* W23, *B. lactis* W51, *E. faecium* W54, *Lb. acidophilus* W22, *Lb. brevis* W63, *L. lactis* W58) and its possible effect on tryptophan levels, often implicated in the etiology of depression [158]. The study concluded that the multispecies probiotic formulation had a positive effect on the tryptophan-kynurenine pathway by successfully reducing the exercise-induced depletion of tryptophan levels that occur after excessive exercise [134].

7. Conclusion and Future Prospects

In conclusion, the relationship between physical fitness and the microbiome is a complex one. Exercise and physical fitness can positively alter the composition of the microbiome and benefit host

Nutrients **2019**, 11, 2270 15 of 23

health, consequently maintaining athletic potential. Studies suggest that different forms of exercise may influence the abundance of different bacterial populations and, hence, alter the microbiome in a different way and, therefore, studies in the future should consider this. As addressed in this review, strenuous exercise can put an athlete in a predicament where they are more prone to infection, inflammation, depression and gut permeability dysfunction. Probiotics offer an effective strategy to prevent or improve those symptoms and allow for continued athletic success without the drawback of the aforementioned symptoms. Future research should aim to clarify the optimal dosage for each individual strain with proven efficacy, as well as establishing whether the probiotic strains colonize our intestine or are simply transient microorganisms with beneficial effects. The impact of probiotics on the athletic population is a new and exciting area, where a limited amount of research has been conducted to date, and although research has shown great promise, little is known about the benefits of probiotics in highly active individuals and ultimately if they benefit from them.

Funding: Research in the group is funded by Science Foundation Ireland in the form of a centre grant (APC Microbiome Ireland grant number SFI/12/RC/2273).

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Janssens, Y.; Nielandt, J.; Bronselaer, A.; Debunne, N.; Verbeke, F.; Wynendaele, E.; Van Immerseel, F.; Vandewynckel, Y.-P.; De Tré, G.; De Spiegeleer, B. Disbiome database: Linking the microbiome to disease. BMC Microbiol. 2018, 18, 50. [CrossRef]
- 2. Gill, S.R.; Pop, M.; DeBoy, R.T.; Eckburg, P.B.; Turnbaugh, P.J.; Samuel, B.S.; Gordon, J.I.; Relman, D.A.; Fraser-Liggett, C.M.; Nelson, K.E. Metagenomic analysis of the human distal gut microbiome. *Science* 2006, 312, 1355–1359. [CrossRef]
- 3. Nash, A.K.; Auchtung, T.A.; Wong, M.C.; Smith, D.P.; Gesell, J.R.; Ross, M.C.; Stewart, C.J.; Metcalf, G.A.; Muzny, D.M.; Gibbs, R.A.; et al. The gut mycobiome of the Human Microbiome Project healthy cohort. *Microbiome* **2017**, *5*, 153. [CrossRef] [PubMed]
- 4. Lozupone, C.A.; Stombaugh, J.I.; Gordon, J.I.; Jansson, J.K.; Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature* **2012**, *489*, 220–230. [CrossRef]
- 5. O'Donovan, C.M.; O'Sullivan, O.; Cotter, P.D. *Gut Microbiol—A Relatively Unexplored Domain*; Elsevier: Amsterdam, The Netherlands, 2018.
- 6. Jandhyala, S.M. Role of the normal gut microbiota. World J. Gastroenterol. 2015, 21, 8787. [CrossRef] [PubMed]
- 7. Manichanh, C.; Rigottier-Gois, L.; Bonnaud, E.; Gloux, K.; Pelletier, E.; Frangeul, L.; Nalin, R.; Jarrin, C.; Chardon, P.; Marteau, P. Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. *Gut* 2006, 55, 205–211. [CrossRef] [PubMed]
- 8. Arthur, J.C.; Perez-Chanona, E.; Mühlbauer, M.; Tomkovich, S.; Uronis, J.M.; Fan, T.-J.; Campbell, B.J.; Abujamel, T.; Dogan, B.; Rogers, A.B. Intestinal inflammation targets cancer-inducing activity of the microbiota. *Science* **2012**, *338*, 120–123. [CrossRef]
- 9. De Goffau, M.C.; Luopajarvi, K.; Knip, M.; Ilonen, J.; Ruohtula, T.; Harkonen, T.; Orivuori, L.; Hakala, S.; Welling, G.W.; Harmsen, H.J.; et al. Fecal microbiota composition differs between children with beta-cell autoimmunity and those without. *Diabetes* **2013**, *62*, 1238–1244. [CrossRef]
- 10. Mach, N.; Fuster-Botella, D. Endurance exercise and gut microbiota: A review. *J. Sport Health Sci.* **2017**, *6*, 179–197. [CrossRef]
- 11. Hill, C.; Guarner, F.; Reid, G.; Gibson, G.R.; Merenstein, D.J.; Pot, B.; Morelli, L.; Canani, R.B.; Flint, H.J.; Salminen, S.; et al. Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic. *Nat. Rev. Gastroenterol. Hepatol.* **2014**, *11*, 506–514. [CrossRef]
- 12. Shade, A. Diversity is the question, not the answer. ISME J. 2017, 11, 1. [CrossRef] [PubMed]
- 13. Barton, W.; Penney, N.C.; Cronin, O.; Garcia-Perez, I.; Molloy, M.G.; Holmes, E.; Shanahan, F.; Cotter, P.D.; O'Sullivan, O. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. *Gut* 2017, 67, 625–633. [CrossRef] [PubMed]

Nutrients **2019**, 11, 2270 16 of 23

14. Clarke, S.F.; Murphy, E.F.; O'Sullivan, O.; Lucey, A.J.; Humphreys, M.; Hogan, A.; Hayes, P.; O'Reilly, M.; Jeffery, I.B.; Wood-Martin, R.; et al. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut* 2014, 63, 1913–1920. [CrossRef] [PubMed]

- 15. Petersen, L.M.; Bautista, E.J.; Nguyen, H.; Hanson, B.M.; Chen, L.; Lek, S.H.; Sodergren, E.; Weinstock, G.M. Community characteristics of the gut microbiomes of competitive cyclists. *Microbiome* **2017**, *5*, 98. [CrossRef] [PubMed]
- 16. Cronin, O.; Barton, W.; Skuse, P.; Penney, N.C.; Garcia-Perez, I.; Murphy, E.F.; Woods, T.; Nugent, H.; Fanning, A.; Melgar, S.; et al. A Prospective Metagenomic and Metabolomic Analysis of the Impact of Exercise and/or Whey Protein Supplementation on the Gut Microbiome of Sedentary Adults. *MSystems* **2018**, 3, e00044. [CrossRef] [PubMed]
- 17. Jang, L.-G.; Choi, G.; Kim, S.-W.; Kim, B.-Y.; Lee, S.; Park, H. The combination of sport and sport-specific diet is associated with characteristics of gut microbiota: An observational study. *J. Int. Soc. Sports Nutr.* **2019**, *16*, 21. [CrossRef] [PubMed]
- 18. Scheiman, J.; Luber, J.M.; Chavkin, T.A.; MacDonald, T.; Tung, A.; Pham, L.-D.; Wibowo, M.C.; Wurth, R.C.; Punthambaker, S.; Tierney, B.T. Meta-omics analysis of elite athletes identifies a performance-enhancing microbe that functions via lactate metabolism. *Nat. Med.* **2019**, 25, 1104–1109. [CrossRef] [PubMed]
- 19. Meyer, J.D.; Koltyn, K.F.; Stegner, A.J.; Kim, J.-S.; Cook, D.B. Influence of exercise intensity for improving depressed mood in depression: A dose-response study. *Behav. Ther.* **2016**, *47*, 527–537. [CrossRef]
- Duscha, B.D.; Slentz, C.A.; Johnson, J.L.; Houmard, J.A.; Bensimhon, D.R.; Knetzger, K.J.; Kraus, W.E. Effects of exercise training amount and intensity on peak oxygen consumption in middle-age men and women at risk for cardiovascular disease. *Chest* 2005, 128, 2788–2793. [CrossRef]
- 21. Puetz, T.W.; Flowers, S.S.; O'Connor, P.J. A randomized controlled trial of the effect of aerobic exercise training on feelings of energy and fatigue in sedentary young adults with persistent fatigue. *Psychother. Psychosom.* **2008**, *77*, 167–174. [CrossRef]
- 22. Meeusen, R.; Duclos, M.; Foster, C.; Fry, A.; Gleeson, M.; Nieman, D.; Raglin, J.; Rietjens, G.; Steinacker, J.; Urhausen, A. Prevention, diagnosis and treatment of the overtraining syndrome: Joint consensus statement of the European College of Sport Science (ECSS) and the American College of Sports Medicine (ACSM). *Eur. J. Sport Sci.* 2013, 13, 1–24. [CrossRef]
- 23. Allen, J.M.; Berg Miller, M.E.; Pence, B.D.; Whitlock, K.; Nehra, V.; Gaskins, H.R.; White, B.A.; Fryer, J.D.; Woods, J.A. Voluntary and forced exercise differentially alters the gut microbiome in C57BL/6J mice. *J. Appl. Physiol.* 2015, 118, 1059–1066. [CrossRef] [PubMed]
- 24. Karl, J.P.; Margolis, L.M.; Madslien, E.H.; Murphy, N.E.; Castellani, J.W.; Gundersen, Y.; Hoke, A.V.; Levangie, M.W.; Kumar, R.; Chakraborty, N. Changes in intestinal microbiota composition and metabolism coincide with increased intestinal permeability in young adults under prolonged physiological stress. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2017**, *312*, G559–G571. [CrossRef] [PubMed]
- 25. Yuan, X.; Xu, S.; Huang, H.; Liang, J.; Wu, Y.; Li, C.; Yuan, H.; Zhao, X.; Lai, X.; Hou, S. Influence of excessive exercise on immunity, metabolism, and gut microbial diversity in an overtraining mice model. *Scand. J. Med. Sci. Sports* **2018**, *28*, 1541–1551. [CrossRef] [PubMed]
- 26. Walsh, N.P.; Gleeson, M.; Shephard, R.J.; Gleeson, M.; Woods, J.A.; Bishop, N.; Fleshner, M.; Green, C.; Pedersen, B.K.; Hoffman-Goete, L. *Position Statement Part One: Immune Function and Exercise*; Loughborough University: Loughborough, UK, 2011.
- 27. Sina, C.; Gavrilova, O.; Förster, M.; Till, A.; Derer, S.; Hildebrand, F.; Raabe, B.; Chalaris, A.; Scheller, J.; Rehmann, A.G. Protein-coupled receptor 43 is essential for neutrophil recruitment during intestinal inflammation. *J. Immunol.* **2009**, *183*, 7514–7522. [CrossRef]
- 28. Shouval, D.S.; Biswas, A.; Goettel, J.A.; McCann, K.; Conaway, E.; Redhu, N.S.; Mascanfroni, I.D.; Al Adham, Z.; Lavoie, S.; Ibourk, M. Interleukin-10 receptor signaling in innate immune cells regulates mucosal immune tolerance and anti-inflammatory macrophage function. *Immunity* **2014**, *40*, 706–719. [CrossRef] [PubMed]
- 29. Sutterwala, F.S.; Noel, G.J.; Clynes, R.; Mosser, D.M. Selective suppression of interleukin-12 induction after macrophage receptor ligation. *J. Exp. Med.* **1997**, *185*, 1977–1985. [CrossRef]

Nutrients **2019**, 11, 2270 17 of 23

30. Yano, J.M.; Yu, K.; Donaldson, G.P.; Shastri, G.G.; Ann, P.; Ma, L.; Nagler, C.R.; Ismagilov, R.F.; Mazmanian, S.K.; Hsiao, E.Y. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. *Cell* **2015**, *161*, 264–276. [CrossRef]

- 31. Valles-Colomer, M.; Falony, G.; Darzi, Y.; Tigchelaar, E.F.; Wang, J.; Tito, R.Y.; Schiweck, C.; Kurilshikov, A.; Joossens, M.; Wijmenga, C. The neuroactive potential of the human gut microbiota in quality of life and depression. *Nat. Microbiol.* **2019**, *4*, 623. [CrossRef]
- 32. Naseribafrouei, A.; Hestad, K.; Avershina, E.; Sekelja, M.; Linløkken, A.; Wilson, R.; Rudi, K. Correlation between the human fecal microbiota and depression. *Neurogastroenterol. Motil.* **2014**, *26*, 1155–1162. [CrossRef]
- 33. Maes, M.; Meltzer, H.Y.; Bosmans, E.; Bergmans, R.; Vandoolaeghe, E.; Ranjan, R.; Desnyder, R. Increased plasma concentrations of interleukin-6, soluble interleukin-6, soluble interleukin-2 and transferrin receptor in major depression. *J. Affect. Disord.* 1995, 34, 301–309. [CrossRef]
- 34. Margonis, K.; Fatouros, I.G.; Jamurtas, A.Z.; Nikolaidis, M.G.; Douroudos, I.; Chatzinikolaou, A.; Mitrakou, A.; Mastorakos, G.; Papassotiriou, I.; Taxildaris, K. Oxidative stress biomarkers responses to physical overtraining: Implications for diagnosis. *Free Radic. Biol. Med.* **2007**, *43*, 901–910. [CrossRef] [PubMed]
- 35. Tanskanen, M.; Atalay, M.; Uusitalo, A. Altered oxidative stress in overtrained athletes. *J. Sports Sci.* **2010**, *28*, 309–317. [CrossRef] [PubMed]
- 36. Hsu, Y.J.; Chiu, C.C.; Li, Y.P.; Huang, W.C.; Te Huang, Y.; Huang, C.C.; Chuang, H.L. Effect of intestinal microbiota on exercise performance in mice. *J. Strength Cond. Res.* **2015**, *29*, 552–558. [CrossRef] [PubMed]
- 37. Xu, J.; Xu, C.; Chen, X.; Cai, X.; Yang, S.; Sheng, Y.; Wang, T. Regulation of an antioxidant blend on intestinal redox status and major microbiota in early weaned piglets. *Nutr.* **2014**, *30*, 584–589. [CrossRef] [PubMed]
- 38. Jeukendrup, A.; Vet-Joop, K.; Sturk, A.; Stegen, J.; Senden, J.; Saris, W.; Wagenmakers, A. Relationship between gastro-intestinal complaints and endotoxaemia, cytokine release and the acute-phase reaction during and after a long-distance triathlon in highly trained men. *Clin. Sci.* **2000**, *98*, 47–55. [CrossRef] [PubMed]
- 39. Holland, A.M.; Hyatt, H.W.; Smuder, A.J.; Sollanek, K.J.; Morton, A.B.; Roberts, M.D.; Kavazis, A.N. Influence of endurance exercise training on antioxidant enzymes, tight junction proteins, and inflammatory markers in the rat ileum. *BMC Res. Notes* **2015**, *8*, 514. [CrossRef]
- 40. Schwellnus, M.P.; Derman, W.E.; Jordaan, E.; Page, T.; Lambert, M.I.; Readhead, C.; Roberts, C.; Kohler, R.; Collins, R.; Kara, S. Elite athletes travelling to international destinations > 5 time zone differences from their home country have a 2–3-fold increased risk of illness. *Br. J. Sports Med.* **2012**, *46*, 816–821. [CrossRef]
- 41. Svendsen, I.S.; Taylor, I.M.; Tønnessen, E.; Bahr, R.; Gleeson, M. Training-related and competition-related risk factors for respiratory tract and gastrointestinal infections in elite cross-country skiers. *Br. J. Sports Med.* **2016**, *50*, 809–815. [CrossRef]
- 42. Palleja, A.; Mikkelsen, K.H.; Forslund, S.K.; Kashani, A.; Allin, K.H.; Nielsen, T.; Hansen, T.H.; Liang, S.; Feng, Q.; Zhang, C. Recovery of gut microbiota of healthy adults following antibiotic exposure. *Nat. Microbiol.* **2018**, *3*, 1255. [CrossRef]
- 43. Sode, J.; Obel, N.; Hallas, J.; Lassen, A. Use of fluroquinolone and risk of Achilles tendon rupture: A population-based cohort study. *Eur. J. Clin. Pharmacol.* **2007**, *63*, 499–503. [CrossRef] [PubMed]
- 44. Wiström, J.; Norrby, S.R.; Myhre, E.B.; Eriksson, S.; Granström, G.; Lagergren, L.; Englund, G.; Nord, C.E.; Svenungsson, B. Frequency of antibiotic-associated diarrhoea in 2462 antibiotic-treated hospitalized patients: A prospective study. *J. Antimicrob. Chemother.* **2001**, *47*, 43–50. [CrossRef] [PubMed]
- 45. Möller, G.B.; da Cunha Goulart, M.J.V.; Nicoletto, B.B.; Alves, F.D.; Schneider, C.D. Supplementation of Probiotics and Its Effects on Physically Active Individuals and Athletes: Systematic Review. *Int. J. Sport Nutr. Exerc. Metab.* **2019**, *29*, 481–492. [CrossRef]
- 46. Parvez, S.; Malik, K.A.; Ah Kang, S.; Kim, H.Y. Probiotics and their fermented food products are beneficial for health. *J. Appl. Microbiol.* **2006**, *100*, 1171–1185. [CrossRef] [PubMed]
- 47. PRNewswire Probiotics Market Will Register 7.5% CAGR to Exceed \$67 Billion by 2024: Global Market Insights, Inc. Available online: https://www.prnewswire.com/news-releases/probiotics-market-will-register-75-cagr-to-exceed-67-billion-by-2024-global-market-insights-inc-681334981.html (accessed on 20 November 2018).
- 48. Santosa, S.; Farnworth, E.; Jones, P.J. Probiotics and their potential health claims. *Nutr. Rev.* **2006**, *64*, 265–274. [CrossRef] [PubMed]

Nutrients **2019**, 11, 2270 18 of 23

49. Ramos, C.L.; Thorsen, L.; Schwan, R.F.; Jespersen, L. Strain-specific probiotics properties of Lactobacillus fermentum, Lactobacillus plantarum and Lactobacillus brevis isolates from Brazilian food products. *Food Microbiol.* **2013**, *36*, 22–29. [CrossRef]

- 50. Altenhoefer, A.; Oswald, S.; Sonnenborn, U.; Enders, C.; Schulze, J.; Hacker, J.; Oelschlaeger, T.A. The probioticEscherichia colistrain Nissle 1917 interferes with invasion of human intestinal epithelial cells by different enteroinvasive bacterial pathogens. *FEMS Immunol. Med Microbiol.* **2004**, 40, 223–229. [CrossRef]
- 51. Wehkamp, J.; Harder, J.; Wehkamp, K.; Wehkamp-von Meissner, B.; Schlee, M.; Enders, C.; Sonnenborn, U.; Nuding, S.; Bengmark, S.; Fellermann, K. NF-κB-and AP-1-mediated induction of human beta defensin-2 in intestinal epithelial cells by Escherichia coli Nissle 1917: A novel effect of a probiotic bacterium. *Infect. Immun.* 2004, 72, 5750–5758. [CrossRef]
- 52. Ukena, S.N.; Singh, A.; Dringenberg, U.; Engelhardt, R.; Seidler, U.; Hansen, W.; Bleich, A.; Bruder, D.; Franzke, A.; Rogler, G.; et al. Probiotic Escherichia coli Nissle 1917 inhibits leaky gut by enhancing mucosal integrity. *PLoS ONE* **2007**, *2*, e1308. [CrossRef]
- 53. Zyrek, A.A.; Cichon, C.; Helms, S.; Enders, C.; Sonnenborn, U.; Schmidt, M.A. Molecular mechanisms underlying the probiotic effects of Escherichia coli Nissle 1917 involve ZO-2 and PKCzeta redistribution resulting in tight junction and epithelial barrier repair. *Cell. Microbiol.* **2007**, *9*, 804–816. [CrossRef]
- 54. Chmielewska, A.; Szajewska, H. Systematic review of randomised controlled trials: Probiotics for functional constipation. *World J. Gastroenterol.* **2010**, *16*, 69. [PubMed]
- 55. Ukena, S.N.; Westendorf, A.M.; Hansen, W.; Rohde, M.; Geffers, R.; Coldewey, S.; Suerbaum, S.; Buer, J.; Gunzer, F. The host response to the probiotic Escherichia coli strain Nissle 1917: Specific up-regulation of the proinflammatory chemokine MCP-1. *BMC Med Genet*. **2005**, *6*, 43. [CrossRef] [PubMed]
- 56. Henker, J.; Laass, M.; Blokhin, B.M.; Bolbot, Y.K.; Maydannik, V.G.; Elze, M.; Wolff, C.; Schulze, J. The probiotic Escherichia coli strain Nissle 1917 (EcN) stops acute diarrhoea in infants and toddlers. *Eur. J. Pediatr.* **2007**, 166, 311–318. [CrossRef] [PubMed]
- 57. De Smet, I.; Van Hoorde, L.; Vande Woestyne, M.; Christiaens, H.; Verstraete, W. Significance of bile salt hydrolytic activities of lactobacilli. *J. Appl. Bacteriol.* **1995**, *79*, 292–301. [CrossRef] [PubMed]
- 58. Pavlova, S.; Kilic, A.; Kilic, S.; So, J.S.; Nader-Macias, M.; Simoes, J.; Tao, L. Genetic diversity of vaginal lactobacilli from women in different countries based on 16S rRNA gene sequences. *J. Appl. Microbiol.* **2002**, 92, 451–459. [CrossRef] [PubMed]
- 59. Isolauri, E.; Kaila, M.; Mykkänen, H.; Ling, W.H.; Salminen, S. Oral bacteriotherapy for viral gastroenteritis. *Dig. Dis. Sci.* **1994**, *39*, 2595–2600. [CrossRef]
- 60. Vizoso Pinto, M.G.; Schuster, T.; Briviba, K.; Watzl, B.; Holzapfel, W.H.; Franz, C.M.A.P. Adhesive and Chemokine Stimulatory Properties of Potentially Probiotic Lactobacillus Strains. *J. Food Prot.* **2007**, *70*, 125–134. [CrossRef]
- 61. Oh, N.S.; Joung, J.Y.; Lee, J.Y.; Kim, Y. Probiotic and anti-inflammatory potential of Lactobacillus rhamnosus 4B15 and Lactobacillus gasseri 4M13 isolated from infant feces. *PLoS ONE* **2018**, *13*, e0192021. [CrossRef]
- 62. Reid, G. The scientific basis for probiotic strains of Lactobacillus. Appl. Environ. Microbiol. 1999, 65, 3763–3766.
- 63. Giwercman, B.; Jensen, E.; Høiby, N.; Kharazmi, A.; Costerton, J. Induction of beta-lactamase production in Pseudomonas aeruginosa biofilm. *Antimicrob. Agents Chemother.* **1991**, *35*, 1008–1010. [CrossRef]
- 64. Cox, A.J.; Pyne, D.B.; Saunders, P.U.; Fricker, P.A. Oral administration of the probiotic Lactobacillus fermentum VRI-003 and mucosal immunity in endurance athletes. *Br. J. Sports Med.* **2010**, *44*, 222–226. [CrossRef] [PubMed]
- 65. West, N.P.; Pyne, D.B.; Cripps, A.W.; Hopkins, W.G.; Eskesen, D.C.; Jairath, A.; Christophersen, C.T.; Conlon, M.A.; Fricker, P.A. Lactobacillus fermentum (PCC®) supplementation and gastrointestinal and respiratory-tract illness symptoms: A randomised control trial in athletes. *Nutr. J.* 2011, 10, 30. [CrossRef] [PubMed]
- 66. Shing, C.M.; Peake, J.M.; Lim, C.L.; Briskey, D.; Walsh, N.P.; Fortes, M.B.; Ahuja, K.D.K.; Vitetta, L. Effects of probiotics supplementation on gastrointestinal permeability, inflammation and exercise performance in the heat. *Eur. J. Appl. Physiol.* **2013**, *114*, 93–103. [CrossRef] [PubMed]
- 67. Hidalgo-Cantabrana, C.; Delgado, S.; Ruiz, L.; Ruas-Madiedo, P.; Sánchez, B.; Margolles, A. Bifidobacteria and Their Health-Promoting Effects. *Microbiol. Spectr.* **2017**, *5*.

68. Ruiz, L.; Margolles, A.; Sanchez, B. Bile resistance mechanisms in Lactobacillus and Bifidobacterium. *Front. Microbiol.* **2013**, *4*, 396. [CrossRef]

- 69. Junick, J.; Blaut, M. Quantification of Human Fecal Bifidobacterium Species by Use of Quantitative Real-Time PCR Analysis Targeting thegroELGene. *Appl. Environ. Microbiol.* **2012**, *78*, 2613–2622. [CrossRef] [PubMed]
- 70. Selinger, C.P.; Bell, A.; Cairns, A.; Lockett, M.; Sebastian, S.; Haslam, N. Probiotic VSL#3 prevents antibiotic-associated diarrhoea in a double-blind, randomized, placebo-controlled clinical trial. *J. Hosp. Infect.* **2013**, *84*, 159–165.
- 71. Imaoka, A.; Shima, T.; Kato, K.; Mizuno, S.; Uehara, T.; Matsumoto, S.; Setoyama, H.; Hara, T.; Umesaki, Y. Anti-inflammatory activity of probiotic Bifidobacterium: Enhancement of IL-10 production in peripheral blood mononuclear cells from ulcerative colitis patients and inhibition of IL-8 secretion in HT-29 cells. *World J. Gastroenterol.* **2008**, *14*, 2511–2516. [CrossRef]
- 72. West, N.P.; Horn, P.L.; Pyne, D.B.; Gebski, V.J.; Lahtinen, S.J.; Fricker, P.A.; Cripps, A.W. Probiotic supplementation for respiratory and gastrointestinal illness symptoms in healthy physically active individuals. *Clin. Nutr.* **2014**, *33*, 581–587. [CrossRef]
- 73. Haywood, B.A.; Black, K.E.; Baker, D.; McGarvey, J.; Healey, P.; Brown, R.C. Probiotic supplementation reduces the duration and incidence of infections but not severity in elite rugby union players. *J. Sci. Med. Sport* **2014**, *17*, 356–360. [CrossRef]
- 74. Jäger, R.; Purpura, M.; Stone, J.; Turner, S.; Anzalone, A.; Eimerbrink, M.; Pane, M.; Amoruso, A.; Rowlands, D.; Oliver, J. Probiotic Streptococcus thermophilus FP4 and Bifidobacterium breve BR03 supplementation attenuates performance and range-of-motion decrements following muscle damaging exercise. *Nutrients* 2016, *8*, 642. [CrossRef] [PubMed]
- 75. Pugh, J.N.; Sparks, A.S.; Doran, D.A.; Fleming, S.C.; Langan-Evans, C.; Kirk, B.; Fearn, R.; Morton, J.P.; Close, G.L. Four weeks of probiotic supplementation reduces GI symptoms during a marathon race. *Eur. J. Appl. Physiol.* **2019**, *119*, 1491–1501. [CrossRef] [PubMed]
- 76. Qin, J.; Li, R.; Raes, J.; Arumugam, M.; Burgdorf, K.S.; Manichanh, C.; Nielsen, T.; Pons, N.; Levenez, F.; Yamada, T.; et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Naturer* **2010**, *464*, 59–65. [CrossRef] [PubMed]
- 77. McFarland, L.V. Meta-analysis of probiotics for the prevention of traveler's diarrhea. *Travel Med. Infect. Dis.* **2007**, *5*, 97–105. [CrossRef] [PubMed]
- 78. Thygesen, J.B.; Glerup, H.; Tarp, B. Saccharomyces boulardii fungemia caused by treatment with a probioticum. *BMJ Case Rep.* **2012**. [CrossRef] [PubMed]
- 79. Choi, C.H.; Jo, S.Y.; Park, H.J.; Chang, S.K.; Byeon, J.-S.; Myung, S.-J. A randomized, double-blind, placebo-controlled multicenter trial of Saccharomyces boulardii in irritable bowel syndrome: Effect on quality of life. *J. Clin. Gastroenterol.* **2011**, 45, 679–683. [CrossRef] [PubMed]
- 80. Kurugöl, Z.; Koturoğlu, G. Effects of Saccharomyces boulardii in children with acute diarrhoea. *Acta Paediatr.* **2005**, *94*, 44–47. [CrossRef]
- 81. Billoo, A.; Memon, M.; Khaskheli, S.; Murtaza, G.; Iqbal, K.; Shekhani, M.S.; Siddiqi, A.Q. Role of a probiotic (Saccharomyces boulardii) in management and prevention of diarrhoea. *World J. Gastroenterol.* **2006**, *12*, 4557. [CrossRef]
- 82. Walter, J. Ecological Role of Lactobacilli in the Gastrointestinal Tract: Implications for Fundamental and Biomedical Research. *Appl. Environ. Microbiol.* **2008**, *74*, 4985–4996. [CrossRef]
- 83. Dugas, B.; Mercenier, A.; Lenoir-Wijnkoop, I.; Arnaud, C.; Dugas, N.; Postaire, E. Immunity and probiotics. *Immunol. Today* **1999**, 20, 387–390. [CrossRef]
- 84. Bezirtzoglou, E.; Stavropoulou, E. Immunology and probiotic impact of the newborn and young children intestinal microflora. *Anaerobe* **2011**, *17*, 369–374. [CrossRef] [PubMed]
- 85. Tulumoglu, S.; Yuksekdag, Z.N.; Beyatli, Y.; Simsek, O.; Cinar, B.; Yaşar, E. Probiotic properties of lactobacilli species isolated from children's feces. *Anaerobe* **2013**, *24*, 36–42. [CrossRef] [PubMed]
- 86. Montes, R.; Bayless, T.M.; Saavedra, J.; Perman, J. Effect of milks inoculated with Lactobacillus acidophilus or a yogurt starter culture in lactose-maldigesting children. *J. Dairy Sci.* **1995**, 78, 1657–1664. [CrossRef]
- 87. Kim, Y.; Whang, J.Y.; Whang, K.Y.; Oh, S.; Kim, S.H. Characterization of the Cholesterol-Reducing Activity in a Cell-Free Supernatant of Lactobacillus acidophilus ATCC 43121. *Biosci. Biotechnol. Biochem.* **2014**, 72, 1483–1490. [CrossRef] [PubMed]

Nutrients **2019**, 11, 2270 20 of 23

88. Almeida, C.C.; Lorena, S.L.; Pavan, C.R.; Akasaka, H.M.; Mesquita, M.A. Beneficial effects of long-term consumption of a probiotic combination of Lactobacillus casei Shirota and Bifidobacterium breve Yakult may persist after suspension of therapy in lactose-intolerant patients. *Nutr. Clin. Pract.* **2012**, 27, 247–251. [CrossRef] [PubMed]

- 89. Kim, J.Y.; Kwon, J.H.; Ahn, S.H.; Lee, S.I.; Han, Y.S.; Choi, Y.O.; Lee, S.Y.; Ahn, K.M.; Ji, G.E. Effect of probiotic mix (*Bifidobacterium bifidum*, *Bifidobacterium lactis*, *Lactobacillus acidophilus*) in the primary prevention of eczema: A double-blind, randomized, placebo-controlled trial. *Pediatric Allergy Immunol*. **2010**, 21, e386–e393. [CrossRef] [PubMed]
- 90. Zanotti, I.; Turroni, F.; Piemontese, A.; Mancabelli, L.; Milani, C.; Viappiani, A.; Prevedini, G.; Sanchez, B.; Margolles, A.; Elviri, L.; et al. Evidence for cholesterol-lowering activity by Bifidobacterium bifidum PRL2010 through gut microbiota modulation. *Appl. Microbiol. Biotechnol.* **2015**, *99*, 6813–6829. [CrossRef] [PubMed]
- 91. Behnsen, J.; Deriu, E.; Sassone-Corsi, M.; Raffatellu, M. Probiotics: Properties, examples, and specific applications. *Cold Spring Harb. Perspect. Med.* **2013**, *3*, a010074. [CrossRef]
- 92. Troy, E.B.; Kasper, D.L. Beneficial effects of Bacteroides fragilis polysaccharides on the immune system. *Front. Biosci. A J. Virtual Library* **2010**, *15*, 25. [CrossRef]
- 93. Curtis, M.M.; Hu, Z.; Klimko, C.; Narayanan, S.; Deberardinis, R.; Sperandio, V. The Gut Commensal Bacteroides thetaiotaomicron Exacerbates Enteric Infection through Modification of the Metabolic Landscape. *Cell Host Microbe* **2014**, *16*, 759–769. [CrossRef]
- 94. Yanagibashi, T.; Hosono, A.; Oyama, A.; Tsuda, M.; Suzuki, A.; Hachimura, S.; Takahashi, Y.; Momose, Y.; Itoh, K.; Hirayama, K.; et al. IgA production in the large intestine is modulated by a different mechanism than in the small intestine: Bacteroides acidifaciens promotes IgA production in the large intestine by inducing germinal center formation and increasing the number of IgA + B cells. *Immunobiology* **2013**, *218*, 645–651. [PubMed]
- 95. Kirmiz, N.; Galindo, K.; Cross, K.L.; Luna, E.; Rhoades, N.; Podar, M.; Flores, G.E. Comparative genomics guides elucidation of vitamin B12 biosynthesis in novel human associated Akkermansia. *bioRxiv* **2019**. [CrossRef]
- 96. Cani, P.D.; de Vos, W.M. Next-Generation Beneficial Microbes: The Case of Akkermansia muciniphila. *Front. Microbiol.* **2017**, *8*, 1765. [CrossRef] [PubMed]
- 97. Everard, A.; Belzer, C.; Geurts, L.; Ouwerkerk, J.P.; Druart, C.; Bindels, L.B.; Guiot, Y.; Derrien, M.; Muccioli, G.G.; Delzenne, N.M.; et al. Cross-talk between Akkermansia muciniphila and intestinal epithelium controls diet-induced obesity. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 9066–9071. [CrossRef] [PubMed]
- 98. Shin, N.-R.; Lee, J.-C.; Lee, H.-Y.; Kim, M.-S.; Whon, T.W.; Lee, M.-S.; Bae, J.-W. An increase in the Akkermansia spp. population induced by metformin treatment improves glucose homeostasis in diet-induced obese mice. *Gut* 2014, 63, 727–735. [CrossRef] [PubMed]
- 99. Plovier, H.; Everard, A.; Druart, C.; Depommier, C.; Van Hul, M.; Geurts, L.; Chilloux, J.; Ottman, N.; Duparc, T.; Lichtenstein, L.; et al. A purified membrane protein from Akkermansia muciniphila or the pasteurized bacterium improves metabolism in obese and diabetic mice. *Nat. Med.* **2016**, 23, 107. [CrossRef] [PubMed]
- 100. De Vos, W.M.; Müller, M.; Norin, E.; Hooiveld, G.; Van Baarlen, P.; Derrien, M. Modulation of Mucosal Immune Response, Tolerance, and Proliferation in Mice Colonized by the Mucin-Degrader Akkermansia muciniphila. *Front. Microbiol.* **2011**, *2*, 166.
- 101. Sanz, Y.; Ottman, N.; Reunanen, J.; Meijerink, M.; Pietilä, T.E.; Kainulainen, V.; Klievink, J.; Huuskonen, L.; Aalvink, S.; Skurnik, M.; et al. Pili-like proteins of Akkermansia muciniphila modulate host immune responses and gut barrier function. *PLoS ONE* **2017**, *12*, e0173004.
- 102. Rossi, O.; van Berkel, L.A.; Chain, F.; Tanweer Khan, M.; Taverne, N.; Sokol, H.; Duncan, S.H.; Flint, H.J.; Harmsen, H.J.M.; Langella, P.; et al. Faecalibacterium prausnitzii A2-165 has a high capacity to induce IL-10 in human and murine dendritic cells and modulates T cell responses. *Sci. Rep.* **2016**, *6*, 18507. [CrossRef] [PubMed]
- 103. Martín, R.; Miquel, S.; Chain, F.; Natividad, J.M.; Jury, J.; Lu, J.; Sokol, H.; Theodorou, V.; Bercik, P.; Verdu, E.F.; et al. Faecalibacterium prausnitzii prevents physiological damages in a chronic low-grade inflammation murine model. *BMC Microbiol.* **2015**, *15*, *67*. [CrossRef]

Nutrients **2019**, 11, 2270 21 of 23

104. Udayappan, S.; Manneras-Holm, L.; Chaplin-Scott, A.; Belzer, C.; Herrema, H.; Dallinga-Thie, G.M.; Duncan, S.H.; Stroes, E.S.; Groen, A.K.; Flint, H.J. Oral treatment with Eubacterium hallii improves insulin sensitivity in db/db mice. *NPJ Biofilms Microbiomes* **2016**, *2*, 16009. [CrossRef] [PubMed]

- 105. Engels, C.; Ruscheweyh, H.-J.; Beerenwinkel, N.; Lacroix, C.; Schwab, C. The Common Gut Microbe Eubacterium hallii also Contributes to Intestinal Propionate Formation. *Front. Microbiol.* **2016**, 7, 713. [CrossRef] [PubMed]
- 106. Fekry, M.I.; Engels, C.; Zhang, J.; Schwab, C.; Lacroix, C.; Sturla, S.J.; Chassard, C. The strict anaerobic gut microbe Eubacterium hallii transforms the carcinogenic dietary heterocyclic amine 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine (PhIP). *Environ. Microbiol. Rep.* **2016**, *8*, 201–209. [CrossRef] [PubMed]
- 107. Kanauchi, O.; Fukuda, M.; Matsumoto, Y.; Ishii, S.; Ozawa, T.; Shimizu, M.; Mitsuyama, K.; Andoh, A. Eubacterium limosum ameliorates experimental colitis and metabolite of microbe attenuates colonic inflammatory action with increase of mucosal integrity. *World J. Gastroenterol.* **2006**, *12*, 1071. [CrossRef] [PubMed]
- 108. Brubaker, J.O.; Li, Q.; Tzianabos, A.O.; Kasper, D.L.; Finberg, R.W. Mitogenic activity of purified capsular polysaccharide A from Bacteroides fragilis: Differential stimulatory effect on mouse and rat lymphocytes in vitro. *J. Immunol.* **1999**, *162*, 2235–2242.
- 109. Hack, V.; Weiss, C.; Friedmann, B.; Suttner, S.; Schykowski, M.; Erbe, N.; Benner, A.; Bartsch, P.; Droge, W. Decreased plasma glutamine level and CD4⁺ T cell number in response to 8 wk of anaerobic training. *Am. J. Physiol. Endocrinol. Metab.* **1997**, 272, E788–E795. [CrossRef] [PubMed]
- 110. Laing, S.; Gwynne, D.; Blackwell, J.; Williams, M.; Walters, R.; Walsh, N. Salivary IgA response to prolonged exercise in a hot environment in trained cyclists. *Eur. J. Appl. Physiol.* **2005**, *93*, 665–671. [CrossRef]
- 111. Mackinnon, L.T.; Ginn, E.; Seymour, G.J. Decreased salivary immunoglobulin A secretion rate after intense interval exercise in elite kayakers. *Eur. J. Appl. Physiol. Occup. Physiol.* **1993**, *67*, 180–184. [CrossRef]
- 112. Neville, V.; Gleeson, M.; Folland, J.P. Salivary IgA as a risk factor for upper respiratory infections in elite professional athletes. *Med. Sci. Sports Exerc.* **2008**, *40*, 1228–1236. [CrossRef]
- 113. Pals, K.L.; Chang, R.-T.; Ryan, A.J.; Gisolfi, C.V. Effect of running intensity on intestinal permeability. *J. Appl. Physiol.* **1997**, *82*, 571–576. [CrossRef]
- 114. Øktedalen, O.; Lunde, O.; Opstad, P.; Aabakken, L.; Kvernebo, K. Changes in the gastrointestinal mucosa after long-distance running. *Scand. J. Gastroenterol.* **1992**, 27, 270–274. [CrossRef]
- 115. Soman, V.R.; Koivisto, V.A.; Deibert, D.; Felig, P.; DeFronzo, R.A. Increased insulin sensitivity and insulin binding to monocytes after physical training. *N. Engl. J. Med.* **1979**, *301*, 1200–1204. [CrossRef] [PubMed]
- 116. Sokol, H.; Pigneur, B.; Watterlot, L.; Lakhdari, O.; Bermúdez-Humarán, L.G.; Gratadoux, J.-J.; Blugeon, S.; Bridonneau, C.; Furet, J.-P.; Corthier, G. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. *Proc. Natl. Acad. Sci. USA* 2008, 105, 16731–16736. [CrossRef] [PubMed]
- 117. Chua, K.J.; Kwok, W.C.; Aggarwal, N.; Sun, T.; Chang, M.W. Designer probiotics for the prevention and treatment of human diseases. *Curr. Opin. Chem. Biol.* **2017**, *40*, 8–16. [CrossRef]
- 118. Amalaradjou, M.A.; Bhunia, A.K. Bioengineered probiotics, a strategic approach to control enteric infections. *Bioengineered* **2013**, *4*, 379–387. [CrossRef]
- 119. Doron, S.; Snydman, D.R. Risk and safety of probiotics. *Clin. Infect. Dis.* **2015**, *60* (suppl. 2), S129–S134. [CrossRef] [PubMed]
- 120. Panel, E.F. Guidance on the Characterisation Microorganisms 1 Used as Feed Additives or as Production Organisms; EFSA: Parma, Italy, 2017.
- 121. Organisation, W.G. Probiotics and Prebiotic; WGO: Forest City, IA, USA, 2017.
- 122. Health, N.I.O. Probiotics. Available online: https://ods.od.nih.gov/factsheets/Probiotics-HealthProfessional/(accessed on 25 June 2019).
- 123. Timmerman, H.; Koning, C.; Mulder, L.; Rombouts, F.; Beynen, A. Monostrain, multistrain and multispecies probiotics—A comparison of functionality and efficacy. *Int. J. Food Microbiol.* **2004**, *96*, 219–233. [CrossRef]
- 124. Anadón, A.; Martínez-Larrañaga, M.R.; Arés, I.; Martínez, M.A. Prebiotics and Probiotics in Feed and Animal Health. In *Nutraceuticals in Veterinary Medicine*; Springer: Berlin/Heidelberg, Germany, 2016; pp. 3–23.

Nutrients **2019**, 11, 2270 22 of 23

125. Helenius, I.J.; Tikkanen, H.O.; Sarna, S.; Haahtela, T. Asthma and increased bronchial responsiveness in elite athletes: Atopy and sport event as risk factors. *J. Allergy Clin. Immunol.* **1998**, *101*, 646–652. [CrossRef]

- 126. Nieman, D.C.; Johanssen, L.M.; Lee, J.W.; Arabatzis, K. Infectious episodes in runners before and after the Los Angeles Marathon. *J Sports Med. Phys. Fit.* **1990**, *30*, 316–328.
- 127. Spence, L.; Brown, W.J.; Pyne, D.B.; Nissen, M.D.; Sloots, T.P.; McCormack, J.G.; Locke, A.S.; Fricker, P.A. Incidence, etiology, and symptomatology of upper respiratory illness in elite athletes. *Med. Sci. Sports Exerc.* **2007**, *39*, 577–586. [CrossRef]
- 128. Ter Steege, R.W.; Van Der Palen, J.; Kolkman, J.J. Prevalence of gastrointestinal complaints in runners competing in a long-distance run: An internet-based observational study in 1281 subjects. *Scand. J. Gastroenterol.* 2008, 43, 1477–1482. [CrossRef] [PubMed]
- 129. Gouttebarge, V.; Frings-Dresen, M.; Sluiter, J. Mental and psychosocial health among current and former professional footballers. *Occup. Med.* **2015**, *65*, 190–196. [CrossRef] [PubMed]
- 130. Martarelli, D.; Verdenelli, M.C.; Scuri, S.; Cocchioni, M.; Silvi, S.; Cecchini, C.; Pompei, P. Effect of a Probiotic Intake on Oxidant and Antioxidant Parameters in Plasma of Athletes During Intense Exercise Training. *Curr. Microbiol.* **2011**, *62*, 1689–1696. [CrossRef] [PubMed]
- 131. Lamprecht, M.; Frauwallner, A. Exercise, intestinal barrier dysfunction and probiotic supplementation. *Med. Sport Sci.* **2012**, *59*, 47–56. [PubMed]
- 132. Clancy, R.; Gleeson, M.; Cox, A.; Callister, R.; Dorrington, M.; D'este, C.; Pang, G.; Pyne, D.; Fricker, P.; Henriksson, A. Reversal in fatigued athletes of a defect in interferon γ secretion after administration of Lactobacillus acidophilus. *Br. J. Sports Med.* **2006**, *40*, 351–354. [CrossRef]
- 133. Sashihara, T.; Nagata, M.; Mori, T.; Ikegami, S.; Gotoh, M.; Okubo, K.; Uchida, M.; Itoh, H. Effects of Lactobacillus gasseri OLL2809 and α-lactalbumin on university-student athletes: A randomized, double-blind, placebo-controlled clinical trial. *Appl. Physiol. Nutr. Metab.* **2013**, *38*, 1228–1235. [CrossRef] [PubMed]
- 134. Strasser, B.; Geiger, D.; Schauer, M.; Gostner, J.M.; Gatterer, H.; Burtscher, M.; Fuchs, D. Probiotic Supplements Beneficially Affect Tryptophan-Kynurenine Metabolism and Reduce the Incidence of Upper Respiratory Tract Infections in Trained Athletes: A Randomized, Double-Blinded, Placebo-Controlled Trial. *Nutrients* 2016, 8, 752. [CrossRef]
- 135. Wang, Y.; Wu, Y.; Wang, Y.; Xu, H.; Mei, X.; Yu, D.; Wang, Y.; Li, W. Antioxidant Properties of Probiotic Bacteria. *Nutrients* **2017**, *9*, 521. [CrossRef]
- 136. Saide, J.; Gilliland, S. Antioxidative activity of lactobacilli measured by oxygen radical absorbance capacity. *J. Dairy Sci.* **2005**, *88*, 1352–1357. [CrossRef]
- 137. Brown, A.J.; Goldsworthy, S.M.; Barnes, A.A.; Eilert, M.M.; Tcheang, L.; Daniels, D.; Muir, A.I.; Wigglesworth, M.J.; Kinghorn, I.; Fraser, N.J. The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *J. Biol. Chem.* 2003, 278, 11312–11319. [CrossRef]
- 138. Arpaia, N.; Campbell, C.; Fan, X.; Dikiy, S.; van der Veeken, J.; Deroos, P.; Liu, H.; Cross, J.R.; Pfeffer, K.; Coffer, P.J. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* 2013, 504, 451. [CrossRef] [PubMed]
- 139. Gao, Z.; Yin, J.; Zhang, J.; Ward, R.E.; Martin, R.J.; Lefevre, M.; Cefalu, W.T.; Ye, J. Butyrate improves insulin sensitivity and increases energy expenditure in mice. *Diabetes* **2009**, *58*, 1509–1517. [CrossRef] [PubMed]
- 140. Zuhl, M.; Schneider, S.; Lanphere, K.; Conn, C.; Dokladny, K.; Moseley, P. Exercise regulation of intestinal tight junction proteins. *Br. J. Sports Med.* **2014**, *48*, 980–986. [CrossRef] [PubMed]
- 141. Wells, J.M.; Konstaninov, S.; Konings, I.; Karczewski, J. Effects of probiotic and commensals on epithelial barrier function. *Int. J. Probiotics Prebiotics* **2008**, *3*, 127–132.
- 142. Lamprecht, M.; Bogner, S.; Schippinger, G.; Steinbauer, K.; Fankhauser, F.; Hallstroem, S.; Schuetz, B.; Greilberger, J.F. Probiotic supplementation affects markers of intestinal barrier, oxidation, and inflammation in trained men; a randomized, double-blinded, placebo-controlled trial. *J. Int. Soc. Sports Nutr.* **2012**, *9*, 45. [CrossRef] [PubMed]
- 143. Hackney, A.C.; Koltun, K.J. The immune system and overtraining in athletes: Clinical implications. *Acta Clin. Croat.* **2012**, *51*, 633–641.
- 144. West, N.; Pyne, D.; Peake, J.; Cripps, A. Probiotics, immunity and exercise: A review. *Exerc. Immunol. Rev.* **2009**, 15, e26.

Nutrients **2019**, 11, 2270 23 of 23

145. Mack, D.R.; Michail, S.; Wei, S.; McDougall, L.; Hollingsworth, M.A. Probiotics inhibit enteropathogenic E. coli adherence in vitro by inducing intestinal mucin gene expression. *Am. J. Physiol. Gastrointest. Liver Physiol.* **1999**, 276, G941–G950. [CrossRef]

- 146. Snyder, J.D.; Walker, A. Structure and Function of Intestinal Mucin: Developmental Aspects. *Int. Arch. Allergy Immunol.* **1987**, *82*, 351–356. [CrossRef]
- 147. Möndel, M.; Schroeder, B.; Zimmermann, K.; Huber, H.; Nuding, S.; Beisner, J.; Fellermann, K.; Stange, E.; Wehkamp, J. Probiotic E. coli treatment mediates antimicrobial human β-defensin synthesis and fecal excretion in humans. *Mucosal Immunol.* **2009**, *2*, 166. [CrossRef]
- 148. Kekkonen, R.A.; Vasankari, T.J.; Vuorimaa, T.; Haahtela, T.; Julkunen, I.; Korpela, R. The effect of probiotics on respiratory infections and gastrointestinal symptoms during training in marathon runners. *Int. J. Sport Nutr. Exerc. Metab.* **2007**, *17*, 352–363. [CrossRef] [PubMed]
- 149. Peters, E.M.; Bateman, E. Ultramarathon running and upper respiratory tract infections. An epidemiological survey. *S. Afr. Med J.* **1983**, *64*, 582–584. [PubMed]
- 150. Romano, R.; Lu, D.; Holtom, P. Outbreak of community-acquired methicillin-resistant Staphylococcus aureus skin infections among a collegiate football team. *J. Athl. Train.* **2006**, *41*, 141. [PubMed]
- 151. Suskovic, J. Antimicrobial Activity—The Most Important Property of probiotic and starter lactic acid bacteria. *Food Technol. Biotechnol.* **2010**, *48*, 296–307.
- 152. Shokryazdan, P.; Sieo, C.C.; Kalavathy, R.; Liang, J.B.; Alitheen, N.B.; Faseleh Jahromi, M.; Ho, Y.W. Probiotic potential of Lactobacillus strains with antimicrobial activity against some human pathogenic strains. *BioMed Res. Int.* **2014**, 2014, 927268. [CrossRef] [PubMed]
- 153. Cotter, P.D.; Hill, C.; Ross, R.P. Food microbiology: Bacteriocins: Developing innate immunity for food. *Nature Rev. Microbiol.* **2005**, *3*, 777. [CrossRef] [PubMed]
- 154. Hegarty, J.W.; Guinane, C.M.; Ross, R.P.; Hill, C.; Cotter, P.D. Bacteriocin production: A relatively unharnessed probiotic trait? *F1000Research* **2016**, *5*, 2587. [CrossRef]
- 155. Dobson, A.; Cotter, P.D.; Ross, R.P.; Hill, C. Bacteriocin Production: A Probiotic Trait? *Appl. Environ. Microbiol.* **2012**, *78*, 1–6. [CrossRef]
- 156. Hegarty, J.W.; Guinane, C.M.; Ross, R.P.; Hill, C.; Cotter, P.D. Lack of Heterogeneity in Bacteriocin Production Across a Selection of Commercial Probiotic Products. *Probiotics Antimicrob. Proteins* **2017**, *9*, 459–465. [CrossRef]
- 157. Reardon, C.L.; Hainline, B.; Aron, C.M.; Baron, D.; Baum, A.L.; Bindra, A.; Budgett, R.; Campriani, N.; Castaldelli-Maia, J.M.; Currie, A. Mental health in elite athletes: International Olympic Committee consensus statement. *Br. J. Sports Med.* **2019**, *53*, 667–699. [CrossRef]
- 158. Dougherty, D.M.; Marsh-Richard, D.M.; Mathias, C.W.; Hood, A.J.; Addicott, M.A.; Moeller, F.G.; Morgan, C.J.; Badawy, A.A.-B. Comparison of 50-and 100-g L-tryptophan depletion and loading formulations for altering 5-HT synthesis: Pharmacokinetics, side effects, and mood states. *Psychopharmacology* **2008**, *198*, 431. [CrossRef]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).