



Article Disinfection in Wastewater Treatment Plants: Evaluation of Effectiveness and Acute Toxicity Effects

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Abstract: In Italy, urban wastewater disinfection is regulated in the third part of Legislative Decree n. 152/2006, which states that wastewater treatment plants (WWTPs) must include a disinfection unit, with a capacity exceeding 2000 Population Equivalent (PE). This treatment shall ensure microbial quality and health security. The legislation provides the following limits for wastewater: Escherichia coli (E. coli) concentration below 5000 CFU 100 mL⁻¹ (recommended value), active chlorine concentration below 0.2 mg L^{-1} and lack of acute toxicity. The compliance with these conditions is shown by means of the study of correct disinfectant dosage, which also depends on wastewater characteristics. An investigation at the regional level (from 2013 to 2016) shows a correlation between acute toxicity discharge and disinfection treatment through chemical reagents (mainly with the use of chlorine compounds and peracetic acid). The experimental work concerns two active sludge WWTPs in northern Italy with small capacity (10,000-12,000 PE). The activities provide the assessment of microbiological quality and toxicity of WWTPs effluents in relation to the dosage of sodium hypochlorite and peracetic acid, by means of the use of batch tests. The results show that with similar disinfectant dosage and comparable initial E. coli concentration, peracetic acid exhibits the best performance in terms of microbial removal (with removal yields up to 99.99%). Moreover, the acute toxicity was evident at higher doses and therefore with higher residuals of peracetic acid (2.68 mg L^{-1}) compared to the free residual chlorine (0.17 mg L^{-1}) .

Keywords: toxicity; microbial quality; disinfection; urban wastewater

1. Introduction

Many human pathogens can be transmitted by waters contaminated by wastewater effluents, which should be disinfected to prevent the spread of pathogenic microorganisms, particularly when the wastewater is used for watering, drinking or bathing purposes [1].

Wastewater disinfection is applied to provide protection to humans against exposure to waterborne pathogenic microorganisms. Microbial inactivation is achieved in these processes by induced biochemical changes within the pathogenic microbial population. The nature of these biochemical changes is dependent upon the microbial population and the applied disinfectant. All disinfectants have the ability to involve changes in wastewater composition that many persist after the disinfectant-specific. One possible outcome of these changes is an alteration of effluent toxicity [2].

Chlorination is the most widely used method for disinfecting the effluents from wastewater treatment plants (WWTPs) and from drinking water treatment plants (DWTPs), but can cause the

formation of mutagenic/carcinogenic and toxic by-products that are potentially harmful to human and aquatic organisms [3–10].

Recently, peracetic acid (PAA) has also been proposed as a wastewater disinfectant. Peracetic acid (CH₃CO₃H) is a strong oxidant which presents several advantages, including: the treatment being easy to implement (without the need of high investment); the large spectrum of microbial activity even in the presence of heterogeneous organic matter; absence of residual or toxic and/or mutagenic by-products; not requiring dechlorination; presenting low dependency on pH; and short contact time [11]. Major disadvantages associated with PAA disinfection are the increase of organic content in the effluent due to acetic acid, and thus the potential for microbial regrowth (acetic acid is already present in PAA mixtures and is also formed after PAA decomposition) [12].

The disinfection efficiency of PAA in wastewater applications has been demonstrated in the scientific literature [13–16]. The formation of disinfection by-products (DBPs) during PAA disinfection was studied by Dell'Erba [17].

The European Community's (EC) environmental regulations aim to reduce the pollution of surface water caused by municipal wastewater (Council Directive 91/271/EEC 1991 as amended by the Commission Directive 98/15/EEC of 27 February 1998) [18,19]. This requires the European Union member states to ensure that discharge of urban wastewater and its effects are monitored [20].

In order to prevent sanitary hazards related to the uses of recipient water bodies, the current Italian regulations prescribe WWTP effluent emission limits for a wide range of chemical compounds, toxicity, and bacterial discharge, such as *Escherichia coli* [21].

In Italy the legislation provides the following limits for wastewater: active chlorine concentration below 0.2 mg L⁻¹, lack of acute toxicity and *E. coli* concentration below 5000 CFU 100 mL⁻¹ (recommended value). In order to respect this value, many WWTPs apply a wastewater disinfection process, because sometimes the *E. coli* concentration in the effluent is higher than the limit established by the local authorities.

Disinfection techniques should be analyzed case by case, because each effluent shows different chemical and physical characteristics, and, sometimes, the reuse of wastewater (i.e., for agricultural use) is applied [22].

In a different way, depending on the treatment adopted, there are different *E. coli* removal yields for each different stage of WWTPs [23].

It is well known that some disinfectant treatments, despite promoting a reduction in pathogenic organisms and organic contaminants, can also produce toxic and genotoxic compounds, depending on the precursors present in the wastewater and the concentrations of disinfectant used [24].

To clarify this issue, a series of toxicity tests, using *Daphnia magna*, *Vibrio fischeri* and *Pseudokirchneriella subcapitata* [25–29], were performed on wastewater effluent samples taken from two WWTPs.

The toxicity responses were measured in disinfected and undisinfected effluent samples in order to evaluate the changes in toxicity attributable to disinfection processes.

The determination of toxicity in wastewater was carried out using *Daphnia magna*. The test is based on the observation of the number of immobile organisms after 24 h of exposure to the sample. Immobilization is the effect on the organisms caused by the toxic substances present in the sample.

In addition to the test with *Daphnia magna* as indicated in D.Lgs 152/2006 and further modifications, acute toxicity tests were carried out using bioluminescent bacteria (*Vibrio fischeri*) and monocellular green algae (*Pseudokirchneriella subcapitata*).

The test with *Vibrio fischeri* was based on the observation of luminescence inhibition after 15 min and 30 min. In the case of *Pseudokirchneriella subcapitata* the test was based on the observation of inhibition of algal growth after 72 h.

The study of disinfectant agents suitable for microbial population removal without acute toxicity is very important, especially in Italy due to the different enforcement (at a local scale) of *E. coli* limit values, which, according to national legislation, are recommendations only.

The aim of the present study was to compare two different disinfectants (sodium hypochlorite and peracetic acid) in terms of acute toxicity on the water body and microbial inactivation. Moreover, the aim was to find a dosage range which allows the respect of *E. coli* limit values, acute toxicity, disinfectant residual and COD (Chemical Oxygen Demand).

The disinfection tests were applied to the effluent from two active sludge WWTPs in northern Italy with small capacities (10,000 to 12,000 PE), and two disinfectant agents (sodium hypochlorite and PAA) were used. These activities provided the assessment of microbiological quality and toxicity of WWTPs effluents in relation to the dosage of sodium hypochlorite and peracetic acid, by means of the use of batch tests.

2. Materials and Methods

2.1. Description of the WWTPs and Characteristics of the Influent

The experimental work were carried out on two urban WWTPs in northern Italy with small capacities (12,000 PE and 10,000 PE respectively).

The urban WWTPs identified by number 1 and 2 are based on a conventional active sludge (CAS) process with a pre-denitrification scheme. They treat domestic, meteoric and industrial sewage collected in a unit drainage system. Table 1 presents the main features of each WWTP.

Тур	e of Treatment	$\rightarrow PT \rightarrow DEN \rightarrow OX-NIT \rightarrow SII \rightarrow D_{(NaClO)} \rightarrow OX-NIT \rightarrow SII \rightarrow OX-NIT \rightarrow SII \rightarrow OX-NIT \rightarrow SII \rightarrow OX-NIT \rightarrow SII \rightarrow OX-NIT \rightarrow OX-NIT \rightarrow SII \rightarrow OX-NIT \rightarrow OX-OX-NIT \rightarrow OX-OX-OX-OX-OX-OX-OX-OX-OX-OX-OX-OX-OX-O$				
	_	WWTP 1	WWTP 2			
Capacity (P	opulation Equivalent)	12,000 10,000				
	Flow rate (m ³ d ^{-1})	2980	2000			
	COD (mg L^{-1})	260 ± 158	433 ± 508.5			
	$BOD_5 (mg L^{-1})$	$\begin{array}{c} & & & & & \\ & & & & & \\ \hline & & & & & \\ \hline & & & &$	199 ± 167.6			
Influent	TN (mg L^{-1})	35 ± 25	51 ± 43.6			
	TP (mg L^{-1})	4 ± 5.7	6 ± 8.3			
	TSS (mg L^{-1})	170 ± 230.3	243 ± 468.6			
	<i>E. coli</i> (CFU 100 mL ^{-1})	$4800 \div 26,000$	$4500 \div 7600$			

Table 1. Characteristics of the wastewater treatment plants (WWTPs) analyzed.

Legend: PT = pretreatment (screening and grit/grease removal), DEN = biological denitrification, OX-NIT = biological oxidation-nitrification S II = secondary sedimentation, D (NaClO) = disinfection with NaClO (emergency). TN = Total Nitrogen; TP = Total Phosphorus.

Both WWTPs are equipped with an emergency disinfection unit with the use of sodium hypochlorite. The contact tanks are built with a longitudinal baffle serpentine flow basin, with a volume of 90 m³ for WWTP 1 and 35 m³ for WWTP 2 (the contact time depends on the flow rate, usually about 30 min for WWTP 1 and 20 min for WWTP 2). There are no flash-mixing tanks.

2.2. Experimental Tests

Laboratory scale disinfection tests were executed (in the WWTPs analyzed) using 40 L of purified effluent taken with an automatic sampler. Each day, three disinfection tests were carried out with different sodium hypochlorite doses, three tests with different peracetic acid doses (PAA) and a reference sample without the dosage of disinfectant agents. pH was measured at the beginning and end of the tests. The solutions were put into a jar test for mixing (Figure 1) with a rotation speed of 45 rpm.



Figure 1. Jar test.

Disinfection tests were carried out on the effluent of WWTP 1 by testing different disinfectant dosages, while for the WWTP 2 the doses were chosen on the basis of the results obtained for WWTP 1. Table 2 presents the experimental program and the analysis/tests carried out in the work.

Table 2. Experimental program and analysis/tests carried out in the research.

	Type of Dosage o		Contact	Analysis/Tests						
	Disinfectant Agents	(mg min L ⁻¹)	Time (min)	Parameters Measured on Site	Physical-Chemical Microbiolog Analysis Analysis		Ecotoxicological Tests	Respirometric Tests		
WWTP 1	NaClO	15 ÷ 135	30	Free and total residual chlorine; pH	COD, BOD ₅ , TSS, TN, N-NH ₄ ⁺ ,	E. coli	Daphnia magna, Vibrio fischeri, Pseudokirchneriella subcapitata	endogenous OUR; exogenous OUR		
	CH ₃ CO ₃ H	10 ÷ 90	30	residual PAA; pH	$N-NO_3$, $N-NO_2^-$, TP					
WWTP 2	NaClO	15 ÷ 50	30	Free and total residual chlorine; pH	COD, BOD ₅ , TSS, TN, N-NH ₄ ⁺ ,	E. coli	Daphnia magna, Vibrio fischeri, Pseudokirchneriella subcapitata	endogenous OUR; exogenous OUR		
	CH ₃ CO ₃ H	15 ÷ 50	30	residual PAA; pH	N-NO ₃ ⁻ , N-NO ₂ ⁻ , TP					

2.3. Analytical Methods

The concentrations of COD, BOD₅, N-NH₄⁺, N-NO₂⁻, N-NO₃⁻, Total Nitrogen (TN), Total Phosphorus (TP) and Total Suspended Solids (TSS) were measured according to standard methods for water and wastewater [30–33]. BOD₅ was determined at 20 °C by inoculation of activated sludge from the WWTP. The pH was measured with a probe Sentix 940-3 WTW[®].

The concentrations of residual chlorine and residual acid peracetic were measured by means of colorimetric tests.

Free residual chlorine was measured with Hach Lange LCK 410 kit.

Total chlorine and peracetic acid residues were measured with the method "Hach 10070 Pillows powder". In case of PAA the conversion of mg $Cl_2 L^{-1}$ to mg PAA L^{-1} (by multiplying the value of 1.07) is necessary [34].

The respirometric tests of oxygen uptake rate (OUR) were carried out according to ISO 8192:2007 [35].

The *E. coli* concentration was measured using two methods: the number of colonies of *E. coli* (CFU) (Standard Method) [30] and the most probable number of microorganisms per volume (MPN) [36].

The CFU method allows the counting of the number of *E. coli* colonies grown on a membrane on agar soil supplemented with chromogenic substances. The sample is filtered through a cellulose ester membrane of 0.45 μ m nominal porosity. After an incubation period of 18–24 h at 44 ± 1 °C, the results are read under ultraviolet light (366 nm). The 4-methylumbelliferyl- β -D-glucuronide (MUG) compound, embedded in the soil, is hydrolyzed by β -glucuronidase of *E. coli*, releasing

4-methylumbelliferone compound which thus produces ultraviolet light-fluorescent blue-green colonies. The results are expressed as colony forming units per 100 mL sample.

The MPN method expresses results as the most likely number of microorganisms per volume. A dehydrated soil is added to a sample; then the mixture is transferred in boxes that are sealed and then incubated at 36 ± 2 °C for 18–22 h. After the incubation, samples with a yellow color intensity equal to or greater than that of the Quanti-Tray/2000 Comparator are considered positive for coliform bacteria. Yellow samples, which are tested under UV light (365 nm) in a dark room, exhibiting any degree of fluorescence are considered positive for *E. coli*. The most probable amount of *E. coli* in 100 mL of the sample can be determined through statistical tables.

The determination of the acute toxicity of the WWTPs effluents was carried out by multispecific ecotoxicological tests: *Daphnia magna* at 24 h [37], *Vibrio fischeri* at 15 min and 30 min [38] and *Pseudokirkneriella subcapitata* at 72 h [39].

Neonates of *Daphnia magna* were held in a temperature-controlled room at $20 \pm 2 \degree C$ illuminated with fluorescent lamps for 16 h d⁻¹. In the acute toxicity tests, daphnids were exposed to samples of different concentrations of wastewater and the tested agent; the immobile daphnids were counted after 24 h of exposure.

The luminescence of reconstituted liquid dried bacteria *Vibrio fischeri* NRRL-B-11177 was measured on a Microtox mod. 500 luminometer according to the ISO standard. The luminescent bacteria were exposed to wastewater samples for 15 min and 30 min. The percentage inhibition was calculated for each concentration relative to the control.

The green, unicellular algae *Pseudokirchneriella subcapitata* were cultured in a nutrient solution prepared and kept on an orbital shaker at 100 rpm at a constant room temperature of 23 ± 2 °C and under continuous fluorescent illumination (6000–10,000 lx.). After 72 h the growth of algae was determined by algal density, which was measured by counting algal cells in a Burker counting cell or absorbance measurement with the spectrophotometer. The inhibition of specific growth rates for each concentration was calculated in comparison to the control.

3. Results and Discussion

3.1. WWTPs Performance Evaluation

Table 3 shows the concentrations of COD, BOD₅, TSS, TN, nitrogenous forms and TP measured in each WWTP during the disinfection tests.

The analyses were carried out to evaluate the performance of WWTPs. Table 3 shows that disinfection does not involve a significant modification in qualitative characteristics of the effluent.

It can be observed that the effluents of the WWTPs analyzed respect the limit values reported by Italian legislation [18], for COD, BOD_5 and TSS, and by the Lombardy Regulation n. 3/2006 (in Tab. 5 and Tab. 6) [40] for TN and TP. Furthermore, the average removal efficiency of COD, BOD_5 and TSS is higher than 90% for both of WWTPs; as concerns TN and TP, the average removal yields of WWTP 1 (65% and 77% respectively) are lower than the performance obtained in WWTP 2 (90% for both pollutants).

Moreover, the results of respirometric tests of OUR have confirmed the good performance of WWTPs and the good "state of health" of biomass.

3.2. Disinfectant Residual

3.2.1. Sodium Hypochlorite

The results reported in Figure 2 show the free and total residual chlorine for all disinfection tests carried out with the use of different doses of sodium hypochlorite; moreover, the limit value of active chlorine concentration (equal to 0.2 mg L^{-1}) is reported.

				$COD (mg L^{-1})$	$BOD_5 (mg L^{-1})$	SST (mg L^{-1})	TN (mg L^{-1})	$N-NH_4^+ (mg L^{-1})$	$N-NO_3^{-}$ (mg L^{-1})	$N-NO_2^{-1}$ (mg L ⁻¹)	TP (mg L^{-1})
		IN W	WTP	381	203	196	45.7	36.0	1.2	0.75	5.6
	DAV #1	IN	test	13.4	<5	5	10.9	<0.2	11	0.04	1
	DAI #1	0.1.17	NaClO	$13.4 \div 14$	<5	$5 \div 6$	$10.9 \div 11.1$	<0.2	$11 \div 11.3$	$0.04 \div 0.05$	$1 \div 1.1$
		001	PAA	$15.2 \div 19.9$	<5 ÷ 7.6	<5	$11.2 \div 11.6$	$<\!0.1 \div 0.1$	$11.4 \div 12.3$	$0.04 \div 0.09$	$1 \div 1.1$
	IN		WTP	270	142	150	40.9	35.0	1.0	0.67	4.8
	DAY #2	IN	test	13.9	<5	5	10.8	0.1	8.6	< 0.01	1
	2.11 "=	OUT	NaClO	$13.9 \div 14.7$	<5	$5 \div 5.2$	$10.8\div10.9$	$0.1 \div 0.2$	$8.6 \div 8.8$	< 0.01	$1 \div 1.1$
		001	PAA	$15.8 \div 17.5$	<5 ÷ 7.6	$5 \div 5.1$	$10.8 \div 11.2$	$0.1 \div 0.2$	$8.6 \div 9.2$	$<0.01 \div 0.01$	$1 \div 1.2$
		IN W	WTP	290	117	138	21.9	11.8	1.5	0.53	21.9
	DAY #3	IN	test	15.7	11.5	<5	10.5	0.11	9.8	0.06	1.3
	2111 110	OUT	NaClO	$15.7 \div 16.3$	$11.5 \div 12$	<5	$10.5\div10.6$	$0.11 \div 0.13$	$9.8 \div 9.9$	$0.06 \div 0.07$	$1.3 \div 1.4$
		001	PAA	$17.3 \div 22.4$	$12 \div 15.3$	<5	$10.4 \div 10.6$	$0.14 \div 0.15$	$10 \div 10.5$	$0.05 \div 0.07$	$1.3 \div 1.4$
WWTP 1		IN W	WTP	69	29	54	17.3	2.7	1	0.67	2.4
	DAY #4	IN test		12	14	<5	12.7	<0.1	12.6	0.04	0.7
		OUT	NaClO	$12 \div 12.2$	$14 \div 14.6$	<5	$12.7 \div 12.9$	< 0.1	$12.6 \div 12.7$	$0.04 \div 0.05$	$0.7 \div 0.8$
		001	PAA	$11 \div 13$	$15 \div 19$	<5	$12.3 \div 13.5$	< 0.1	$12.6 \div 13.1$	$0.03 \div 0.04$	$0.7 \div 0.9$
		IN W		266	148	216	41.3	34	1.1	0.55	4.4
	DAY #5	IN	test	22.6	13.1	6	15	<0.1	14.9	0.03	1.2
	2111 110	OUT	NaClO	$22.6 \div 22.7$	$13.1 \div 14$	$6 \div 6.1$	$15 \div 15.1$	< 0.1	$14.9 \div 15.1$	$0.03 \div 0.04$	$1.2 \div 1.3$
			PAA	$17.5 \div 18.1$	$18.6 \div 20.8$	<5 ÷ 5	$14.9 \div 15$	<0.1	$14.7 \div 14.9$	$0.02 \div 0.05$	$1.2 \div 1.3$
	IN		WTP	172	79	60	32	29	1.2	0.76	3.2
	DAY #6	IN	test	31.4	14.1	13	13.8	1.8	11.6	0.4	1.3
	2111 #0	OUT	NaClO	$31.4 \div 32$	$14.1\div14.7$	$13 \div 13.1$	$13.8 \div 13.9$	$1.8 \div 1.9$	$11.6 \div 11.8$	$0.40 \div 0.41$	$1.3 \div 1.5$
		001	PAA	$30.4 \div 33$	$15.8 \div 18$	$12 \div 13$	$13.9 \div 14.7$	$1.8 \div 2.1$	$11.3 \div 11.5$	$0.35 \div 0.36$	$1.3 \div 1.6$
		INW		304	153	156	33.4	32.8	0.98	0.15	3.6
	DAY #1	IN test		9.4	<5	<5	2	<0.2	1.8	n.a.	0.3
	2111 #1	OUT	NaClO	$9.4 \div 9.6$	<5	<5	$2 \div 2.3$	<0.2	$1.8 \div 2$	n.a.	$0.3 \div 0.5$
		001	PAA	$9.6 \div 15.6$	<5	<5	$2.3 \div 3.8$	<0.1	$2.2 \div 2.3$	n.a.	$0.3 \div 0.6$
		IN W	WTP	413	155	152	39.3	37.5	1.09	0.18	4.5
	DAY #2	IN	test	25.2	8.2	12	5.4	< 0.1	4.2	0.08	0.7
WWTP 2	2.11 "=	NaClO		$25.2 \div 25.4$	$8.2 \div 8.3$	$12 \div 12.3$	$5.4 \div 5.5$	< 0.1	$4.2 \div 4.3$	$0.08 \div 0.09$	$0.7 \div 0.8$
		001	PAA	$25 \div 35$	$6 \div 6.5$	$8 \div 10$	$5.3 \div 5.4$	$< 0.1 \div 0.1$	$4.4 \div 4.5$	$0.08 \div 0.09$	$0.6 \div 0.7$
	IN WWTP		452	251	280	44.6	38.2	1.1	0.21	5.9	
	DAY #3	IN	test	20.1	<5	5	3.9	<0.1	2.7	0.01	0.3
	DAI 113	NaCl		$20.1 \div 20.4$	<5	$5 \div 5.6$	$3.9 \div 4.1$	<0.1	$2.7 \div 2.8$	$0.01 \div 0.02$	$0.3 \div 0.4$
		001	PAA	$19.5 \div 24$	<5	$5 \div 6$	$3.3 \div 3.8$	< 0.1	$2.8 \div 2.9$	$0.01 \div 0.03$	$0.3 \div 0.4$
	Limit values (mg L^{-1})			125	25	35	15	-	-	-	2

Table 3. Concentration of pollutants in the influent and effluent of WWTPs.

IN WWTP: concentrations of pollutants measured in the influent wastewater to WWTP. IN test: concentrations of pollutants measured in the effluent of WWTP before the disinfection unit. OUT NaClO: concentrations of pollutants measured in the samples after the disinfection tests with NaClO. OUT PAA: concentrations of pollutants measured in the samples after the disinfection tests with NaClO. OUT PAA: concentrations of pollutants measured in the samples after the disinfection tests with NaClO. OUT PAA: concentrations of pollutants measured in the samples after the disinfection tests with PAA.



Figure 2. Free and total residual chlorine.

In Figure 2 the acute toxicity is also shown for different content of free chlorine. It can be noted that, according to several studies [1,41–44] there is a good correlation between the concentration of chloramines (indirectly calculated as the difference between total chlorine and free chlorine) and the acute toxicity observed with *Daphnia magna*, *Pseudokirchneriella subcapitata* and *Vibrio fischeri*. Moreover, high content of free chlorine involves acute toxicity, although the chloramine concentrations persist in the aquatic environment for a long time (and only sunlight/ventilation lead to their removal) and the risks due to their presence could be high.

3.2.2. Peracetic Acid

Figure 3 shows the residual PAA and COD concentration after disinfection tests with peracetic acid. Unlike the free residual chlorine, for the residual PAA, in Italy, there are no limit values.



Figure 3. Residual peracetic acid (PAA).

COD concentration was monitored in the case of disinfection with peracetic acid due to the increase in the concentration of organic matter; in the tests, this increase was approximately equal to 50% for WWTP 1 and equal to 60% for WWTP 2, but the COD concentration was always below the limit of 125 mg L^{-1} (Decree Italian Law 152/2006).

In disinfection tests the decay of PAA was not evaluated, because the aim of the present study was not to find an optimal disinfectant dosage but a range of dosages that would respect the limit values of *E. coli* and toxicity. It was not considered decay because Rossi et al., 2007 [14] have shown that it becomes significant with doses and contact times far higher than those applied.

3.3. Acute Toxicity Assessment

The results of the acute toxicity of experimental disinfection tests using both disinfectants are shown below.

The sample is considered toxic when at least one of these three conditions occurs:

- in the case of *Daphnia magna*, the percentage of immobilisation is greater or equal to 50%;
- in the case of *Vibrio fischeri*, the inhibition of luminescence is greater or equal to 50%;
- in the case of *Pseudokirchneriella subcapitata*, inhibition of algal growth is greater or equal to 50%.

There is conformity when the limit on free active chlorine is 0.2 mg L⁻¹, the *E. coli* limit of 5000 CFU 100 mL⁻¹ and the absence of acute toxicity are observed at the same time.

3.3.1. Sodium Hypochlorite

In Table 4 the results of the acute toxicity assessments, using three species from different trophic levels of aquatic ecosystems, with an active chlorine dosage are shown. Grey cells show the values of the limits that are not observed.

	Active Chlorine	ctive Chlorine Free Residual		Acute Toxicity					
	Dosage (mg $min I^{-1}$)	Chlorine (mg	Daphnia	Vibrio	fischeri	P. subcapitata	IN	OUT	Conformity
	min L)	L)	<i>magna</i> 24 h	15 min	30 min	72 h	11N	001	
	60	0.26	100/100	81/100	81/100	27/100	4800	5	NO
	105	0.47	0/100	88/100	88/100	37/100	4800	<1	NO
	135	0.998	0/100	87/100	87/100	59/100	4800	<1	NO
	15	0.088	0/100	0/100	0/100	0/100	7000	5400	NO
	30	0.175	90/100	83/100	82/100	90/100	7000	<1	NO
	60	0.224	100/100	82/100	82/100	100/100	7000	<1	NO
WWTP 1	15	0.109	0/100	0/100	1/100	5/100	26,000	21,000	NO
	25	0.114	0/100	23/100	31/100	1/100	26,000	9500	NO
	30	0.096	0/100	18/100	25/100	8/100	26,000	9300	NO
	20	0.097	5/100	0/100	5/100	1/100	8100	3800	YES
	30	0.169	0/100	0/100	5/100	3/100	8100	320	YES
	40	0.129	45/100	9/100	13/100	16/100	8100	74	YES
	30	0.058	0/100	11/100	16/100	2/100	6100	3700	YES
	35	0.1	0/100	14/100	20/100	1/100	6100	290	YES
	50	0.13	0/100	40/100	42/100	3/100	6100	17	YES
	35	0.057	0/100	80/100	78/100	70/100	7100	58	NO
	40	0.05	0/100	86/100	88/100	47/100	7100	37	NO
	50	0.203	100/100	91/100	93/100	65/100	7100	21	NO
	25	n.a.	100/100	91/100	93/100	65/100	4500	38	NO
WWTP 2	30	n.a.	100/100	0/100	5/100	8/100	4500	33	NO
	35	n.a.	100/100	8/100	6/100	1/100	4500	36	NO
	15	0.05	5/100	0/100	0/100	4/100	7600	5000	YES
	20	0.069	0/100	31/100	25/100	0/100	7600	100	YES
	25	0.07	0/100	6/100	6/100	0/100	7600	230	YES

Table 4. Acute toxicity assessment with active chlorine.

As regards the WWTP 1, the acute toxicity was increased as the free residual chlorine value increases. Acute toxicity has been reported for values above 0.17 mg L^{-1} . An active chlorine dosage

between 30 and 50 mg min L^{-1} allowed the contemporary respect of the *E. coli* limit value and a non-toxicity of disinfected effluent.

Concerning the WWTP 2, acute toxicity has been reported for values above 0.20 mg L⁻¹. An active chlorine dosage between 20 and 25 mg min L⁻¹ allowed the contemporary respect of the *E. coli* limit and a non-toxicity of disinfected effluent.

The use of low dosages of active chlorine has involved a scarce disinfection efficiency, especially in case of high *E. coli* concentrations, that, for WWTP 1, has reached 26,000 CFU 100 mL⁻¹. A proper solution for disinfection efficiency increasing could be two or three-step chlorination, also in order to reduce the DBPs formation and the toxicity effects [45,46].

3.3.2. Peracetic Acid

In Table 5 the results of the acute toxicity assessment, using three species from different trophic levels of aquatic ecosystems with a PAA dosage, are reported. Grey cells show the values of the limits that are not observed.

Concerning the WWTP 1, the acute toxicity was increased as the residual PAA value increases. Acute toxicity has been reported for values above 2.68 mg L⁻¹. A PAA dosage between 20 and 40 mg min L⁻¹ allowed the contemporary respect of the *E. coli* limit and a non-toxicity of disinfected effluent.

The results obtained for WWTP 2 effluent is similar to WWTP 1: in fact, a PAA dosage between 20 and 35 mg min L^{-1} allowed the contemporary respect of the *E. coli* limit value and a non-toxicity of disinfected effluent.

	PAA Dosage Residual			toxicity	<i>E. coli</i> (CFU 100 mL ⁻¹)				
	(mg min	PAA (mg L ⁻¹)	Daphnia	Vibrio	fischeri	<i>P. subcapitata</i> 72 h	IN	OUT	Conformity
	L ¹)		magna 24 h	15 min	30 min		110	001	
	30	1.50	0/100	13/100	5/100	5/100	4800	1	YES
	60	3.32	100/100	100/100	96/100	100/100	4800	<1	NO
	90	5.67	100/100	100/100	100/100	54/100	4800	<1	NO
	40	4.49	100/100	99/100	99/100	100/100	7000	<1	NO
	50	2.68	20/100	71/100	37/100	20/100	7000	<1	NO
	60	3.53	100/100	100/100	96/100	100/100	7000	<1	NO
WWTP 1	30	1.61	0/100	13/100	20/100	6/100	26,000	38	YES
	40	2.14	0/100	5/100	10/100	4/100	26,000	29	YES
	50	6.42	100/100	86/100	51/100	73/100	26,000	8	NO
	20	0.43	0/100	0/100	0/100	0/100	8100	5000	YES
	30	0.96	0/100	0/100	3/100	0/100	8100	180	YES
	40	1.39	0/100	23/100	16/100	3/100	8100	15	YES
	30	1.30	0/100	19/100	13/100	0/100	6100	16	YES
	35	1.50	0/100	33/100	22/100	1/100	6100	45	YES
	40	1.80	0/100	45/100	32/100	3/100	6100	110	YES
	35	1.71	5/100	82/100	44/100	1/100	7100	50	NO
	40	2.25	5/100	100/100	80/100	1/100	7100	22	NO
	50	2.57	0/100	100/100	94/100	1/100	7100	37	NO
WWTP 2	25	4.00	0/100	0/100	0/100	0/100	4500	75	YES
	30	4.00	0/100	0/100	0/100	0/100	4500	54	YES
	35	4.00	0/100	0/100	0/100	0/100	4500	21	YES
	15	0.64	0/100	0/100	0/100	0/100	7600	7500	NO
	20	0.07	0/100	0/100	0/100	0/100	7600	3800	YES
	25	0.08	0/100	3/100	0/100	1/100	7600	2600	YES

Table 5. Acute toxicity assessment with PAA dosage.

3.4. OUR Tests

The OUR values shown in Figure 4 consider only the part of oxygen demand to degrade the organic matter (the values reported are obtained by the difference between the OUR of each effluent and the endogenous values). When the free residual chlorine limit value is higher than 0.2 mg L^{-1} , a slowing down of oxygen consumption, to degrade the organic matter present in the effluent, is observed. The effect of chlorination is generally a reduction in carbonaceous oxygen uptake rate (OUR) [47].



Figure 4. Results of oxygen uptake rate (OUR) tests: effects of free residual chlorine (**a**) and residual PAA (**b**).

Moreover, Figure 4 shows that there is a correlation between the reduction of OUR and the presence of toxicity (especially for *Vibrio fischeri*) in case of sodium hypochlorite use. Concerning disinfection with PAA there is not a correlation with toxicity; indeed an increase in organic content, due to acetic acid that is readily biodegradable, and OUR enhancement were observed.

4. Conclusions

The aim of the present study was to evaluate the ecotoxicological behavior of secondary effluents disinfected with sodium hypochlorite and PAA using three species from different trophic levels of aquatic ecosystems. Moreover, the work is aimed to find a dosage range, which allows the respect of *E. coli* limit values, acute toxicity, disinfectant residual and COD.

The experimental activities provided assessment of microbiological quality and toxicity of WWTPs effluents in relation to the dosage of sodium hypochlorite and peracetic acid, by means of the use of batch tests.

The results showed that the acute toxicity was increased as the disinfectant residual value increases. Moreover, residual acute toxicity of peracetic acid was higher (2.68 mg L^{-1}) than residual chlorine (0.17 mg L^{-1}). As concerns the chlorination, a good correlation between the content of chloramines and the acute toxicity was observed.

As concerns the WWTP 1, an active chlorine dosage between 30 and 50 mg min L^{-1} allowed the contemporary respect of the limit of *E. coli* and a non-toxicity of disinfected effluent and a PAA dosage between 20 and 40 mg min L^{-1} allowed the contemporary respect of the *E. coli* limit and a non-toxicity of disinfected effluent. Concerning the WWTP 2, an active chlorine dosage between 20 and

25 mg min L^{-1} allowed the contemporary respect of the *E. coli* limit and a non-toxicity of disinfected effluent and a PAA dosage between 20 and 35 mg min L^{-1} allowed the contemporary respect of the *E. coli* limit value and a non-toxicity of disinfected effluent.

The results show that with similar disinfectant dosage and comparable initial *E. coli* concentration, peracetic acid displayed the best performance in terms of microbial removal (with removal yields up to 99.99%).

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