



Article Onchocerciasis Drug Discovery: In Vitro Evaluation of FDA-Approved Drugs against *Onchocerca gutturosa* in Gambia

Suzanne Gokool^{1,†}, Simon Townson^{1,*,†}, Andrew Freeman¹, Jadzia Siemienski-Kleyn¹, Jakub Zubrzycki¹, Senyo Tagboto¹, Marc P. Hübner^{2,3} and Ivan Scandale⁴

- ¹ Tropical Parasitic Diseases Unit, Northwick Park Institute for Medical Research, Watford Road, Harrow, London HA1 3UJ, UK; suzanne.gokool@bristol.ac.uk (S.G.); stagboto@uhas.edu.gh (S.T.)
- ² Institute for Medical Microbiology, Immunology and Parasitology, University Hospital Bonn, Venusberg-Campus 1, 53127 Bonn, Germany; huebner@uni-bonn.de
- ³ German Center for Infection Research (DZIF), Partner Site Bonn-Cologne, 53127 Bonn, Germany
- ⁴ Drugs for Neglected Diseases Initiative, 1202 Geneva, Switzerland; iscandale@dndi.org
- * Correspondence: s.townson@imperial.ac.uk
- ⁺ These authors contributed equally to this work.

Abstract: Onchocerciasis treatment and control relies mainly on the use of ivermectin which has high activity against the microfilarial stage of *Onchocerca volvulus* but limited activity against the long-lived, tissue dwelling adult nematodes. As this neglected tropical disease has now been targeted for elimination, there is an urgent need for new drugs to combat these parasites, ideally with macrofilaricidal activity. In this study, we have examined the anti-*Onchocerca* activity of a range of existing FDA-approved drugs with a view to repurposing, which can lead to rapid and relatively inexpensive development. From the Pharmakon-1600 library, 106 drugs were selected and tested against *O. gutturosa* adult male parasites using a concentration of 1.25×10^{-5} M in an in vitro 5-day standard assay to assess motility and viability (using MTT/formazan colorimetry). The findings revealed that 44 drugs produced marginal/moderate activity (50–99% motility and/or MTT reductions) including cefuroxime sodium, methenamine, primaquine phosphate and rivastigmine tartrate, while 23 drugs produced good activity (100% motility reductions and significant MTT reductions), including atovaquone, isradipine, losartan, rifaximin, cefaclor and pyrantel pamoate. Although this study represents only a first step, some of the identified hits indicate there are potential anti-*Onchocerca* drug candidates worthy of further investigation.

Keywords: onchocerciasis; drug discovery; anthelmintics; *O. gutturosa*; motility and MTT inhibition; FDA-approved drugs

1. Introduction

Onchocerciasis (river blindness) is caused by the tissue-dwelling filarial worm *Onchocerca volvulus*. The infection is transmitted between humans, by the bite of the blackfly vector of the *Simulium* genus. Circulating microfilariae (mf) accumulate in the skin but in high-intensity infections, the mf can also enter the tissues of the eye. Death of the mf causes the pathologies of pruritus, skin atrophy, skin de-pigmentation, papular rash, eye lesions and blindness in humans [1]. Related to high morbidity is reduced work productivity, which can then lead to social stigmatisation; this poverty promoting nematode infection has been included within the group of neglected tropical diseases (NTDs) [2].

Without an available vaccine, current preventive chemotherapy and elimination of onchocerciasis rely on mass drug administration (MDA) programs (large scale distribution without diagnosis and supervision of health-care staff) that distribute ivermectin (Mectizan[®], Merck, Rahway, NJ, USA) either on an annual or biannual basis [2]. This drug, belonging to the family of macrocyclic lactones, targets only the mf stage of the parasite by killing mf and exerting embryostatic effects on the adult female worm; that is, mf release



Citation: Gokool, S.; Townson, S.; Freeman, A.; Siemienski-Kleyn, J.; Zubrzycki, J.; Tagboto, S.; Hübner, M.P.; Scandale, I. Onchocerciasis Drug Discovery: In Vitro Evaluation of FDA-Approved Drugs against *Onchocerca gutturosa* in Gambia. *Pharmaceutics* **2024**, *16*, 210. https://doi.org/10.3390/ pharmaceutics16020210

Academic Editors: Tihomir Tomašič and Tao Sun

Received: 16 November 2023 Revised: 18 January 2024 Accepted: 25 January 2024 Published: 31 January 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). from the uterus is temporarily suppressed [3]. Both of these mechanisms have the effect of reducing the skin mf by up to 99% two months after treatment [4]. In order to have an impact on filarial transmission reduction, treatment needs to be administered for the duration of the reproductive life span of the long-lived (up to 15 years [5]), tissue-dwelling adult worms, so program success is constrained by the absence of drugs with macrofilaricidal activity. There are now concerns of developing ivermectin resistance in Onchocerca parasites, as has already been reported in nematodes of veterinary importance [6-8], and construed in human onchocerciasis [9]. A study in endemic communities exposed to frequent rounds of treatment in Ghana demonstrated that although ivermectin retained microfilarial activity, sub-optimal responses to treatment could be due to the development of resistance by *O. volvulus*, resulting in a decreased effect on the inhibition of mf release [9]. Similarly, in Cameroon, studies on communities given multiple rounds of ivermectin therapy compared to those that were ivermectin naive indicated that continuous exposure to this drug had a reduced embryostatic effect on the female adult worms [10]. The findings from these studies highlight the possible emerging problems of ivermectin resistance and the urgent need for alternative methods of treatment. Recent studies demonstrated that the closely related drug moxidectin (registered for use in humans by the United States Food and Drug Administration (US FDA) in 2018) may offer improved treatment over ivermectin, although no macrofilaricidal effect has been observed so far [11,12].

The Pharmakon 1600 Collection, a US FDA-approved library, consists of 1600 drugs that have reached clinical evaluation and demonstrated biological activity against known targets. By screening a selection of this drug set, the identification of any potential antifilarial candidates could be rapidly repurposed and prove useful in onchocerciasis elimination programs, as many of the drugs represented in the library are available on the market. Humans are the only viable hosts of *O. volvulus* and there are no laboratory models that support the complete life cycle of this parasite; as such, drug discovery for onchocerciasis has to rely in part, on the use of surrogate parasites and animal hosts. Several in vitro and in vivo standard operating protocols for testing drugs against the adult stage of *Onchocerca* have been developed and optimized [13] and references therein. Using a World Health Organisation approved 5-day motility/MTT-based assay with the cattle filarial nematode *Onchocerca gutturosa*, in this study we have assessed the activity of 106 selected drugs with a range of biological activities (see Table 1).

Table 1. Selected Pharmakon-1600 drugs (106) for in vitro evaluation against *O. gutturosa* adult male parasites. Bioactivity was categorised by MicroSource Discovery Systems, Inc. (Gaylordsville, CT, USA).

Bioactivity	Drug
Antibacterial	Carbadox; Cefaclor; Cefamandole nafate; Cefoperazone; Cefoxitin sodium; Cefsulodin sodium; Ceftibuten; Cefuroxime sodium; Chlorhexidine dihydrochloride; Chloroxylenol; Demeclocycline hydrochloride; Doxycycline hydrochloride; Furazolidone; Gramicidin; Lasalocid sodium; Merbromin; Methacycline hydrochloride; Methenamine; Minocycline hydrochloride; Nitrofurantoin; Nitroxoline; Ofloxacin; Oxytetracycline; Rifampcin; Rifaximin; Sulfaquinoxaline sodium; Teicoplanin
Anticancer	Azaserine; Bleomycin; Daunorubicin; Doxorubicin; Epirubicin hydrochloride; Isotretinon; Lomustine; Mitoxantrone hydrochloride; Tretinoin
Antihypertensive/ vasodilator	Dipyridamole; Guanethidine; Losartan; Nicardipine hydrochloride; Nicotinyl alcohol tartrate; Nifedipine
Anti-infective	Benzethonium chloride; Broxyquinoline; Dequalinium chloride; Methylbenzethonium chloride; Nitrofurazone; Oxyquinoline hemisulfate; Phenylethyl alcohol; Resorcinol monoacetate
Anti-inflammatory/ antihistamine	Dexamethasone acetate; Doxylamine succinate; Meloxicam sodium; Prednisolone tebutate; Sulfasalazine
Antiparasitic	Amitraz; Atovaquone; Candicidin; Clorsulon; Diethylcarbamazine citrate; Flubendazole; Hexetidine; Homidium bromide; Iodoquinol; Levamisole hydrochloride; Moxidectin; Primaquine phosphate; Pyrantel pamoate;
Antiviral	Öseltamivir phosphate; Valganciclovir hydrochloride

Bioactivity	Drug
Neurological	Acepromazine maleate; Almotriptan; Ampyzine sulfate; Apomorphine hydrochloride; Armodafinil; Bupropion; Chlorpromazine; Danazol; Desipramine hydrochloride; Dopamine hydrochloride; Isradipine (also antihypertensive/vasodilator); Methsuximide; Methylphenidate hydrochloride; Olanzapine; Oxidopamine hydrochloride; Penfluridol; Rivastigmine tartrate; Selegiline hydrochloride; Zaleplon
Various	Alendronate sodium; Anisindione; Ascorbyl palmitate; Bromhexine hydrochloride; Butacaine; β-Carotene; Clopidogrel sulfate; Dienestrol; Dioxybenzone; Docusate sodium; Fluorescein; Mangafodipir trisodium; Methylergonovine maleate; Propoxycaine hydrochloride; Riboflavin; Sennoside A; Tetrahydrozoline hydrochloride

2. Materials and Methods

A workflow diagram for the experimental procedure used in this study is depicted in Figure 1.

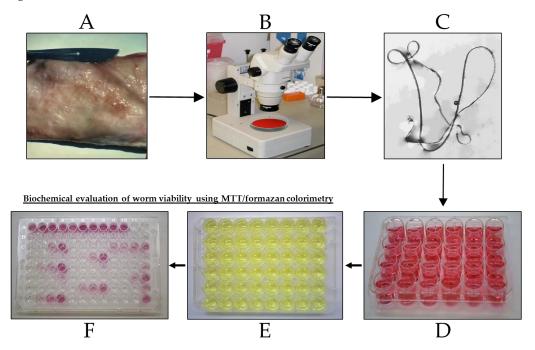


Figure 1. Workflow diagram for the *Onchocerca gutturosa* adult worm in vitro 5-day motility/MTT assay. (**A**) Nuchal ligament connective tissue from naturally infected cattle; (**B**) Tissue in culture medium placed under a dissecting microscope ($40 \times$ magnification) to isolate worms using fine forceps; (**C**) Isolated worm floating on tissue culture medium, worm length approximately 2 cm; (**D**) Worms were maintained individually in each well of a 24-well plate containing 2 mL culture medium, at 37 °C under an atmosphere of 5% CO₂ in air; this was replaced after 24 h with culture medium containing the test drug at a final concentration of 1.25×10^{-5} M. Worm motility was determined microscopically every 24 h up to 120 h; (**E**) Worm viability was then assessed by the transfer of each worm to the well of a 48-well plate containing 0.5 mg/mL MTT, incubation at 37 °C for 30 min; (**F**) Each worm was transferred to a well in a 96-well plate containing 200 µL dimethylsulfoxide to solubilize the formazan. After 1 h at 23–26 °C, the plate was gently agitated and formazan formation measured using absorbance (490 nm) on an ELISA reader (Dynatech, Willenhall, UK).

2.1. Parasites—Onchocerca gutturosa

Adult male worms were obtained postmortem from naturally infected, freshly slaughtered cattle in Gambia, West Africa. The material used in this study was purchased from local butchers by the West Africa Livestock Innovation Centre (WALIC), Banjul, Gambia. The adult male worms were dissected from the cattle nuchal ligament connective tissue and transferred to each well of a sterile 24-well (2 mL) plate (Fisher Scientific, Loughborough, UK), and maintained for at least 24 h in culture before use. The culture medium used was Minimum Essential Medium (MEM) with Earl's Salts and L-Glutamine (Life Technologies Ltd., Loughborough, UK) supplemented with 10% heat inactivated new-born calf serum (Life Technologies Ltd., UK) and 200 units/mL penicillin, 200 μ g/mL streptomycin and 0.5 μ g/mL amphotericin B (Life Technologies Ltd., UK). Only normally active worms were used for the test and all assays were conducted at 37 °C under an atmosphere of 5% CO₂ in air.

2.2. Origin of Drugs Tested

The drugs of the Pharmakon 1600 Collection were supplied by MicroSource Discovery Systems Inc. (Gaylordsville, CT, USA) as 10 mM DMSO stock solutions in microtubes in a 96-well plate format and stored at -20 °C. The positive control drug, Immiticide[®] (melarsomine dihydrochloride, Merial, Duluth, GA, USA) was supplied as a dry solid, known amounts of this drug were solubilized in 1 mL DMSO.

2.3. In Vitro Drug Activity against O. gutturosa Adult Worms as Described by Townson et al. [13]

The selected Pharmakon drugs with the positive control Immiticide® were screened using a final concentration of 1.25×10^{-5} M. At this concentration, the drugs were not toxic to mammalian cells (LLMCK2 monkey kidney cells, evaluated by microscopy). Two trials were performed using different numbers of worms (according to availability) and drugs. Trial 1: Four worms per group were used for the test and positive control drugs, and eight worms for the untreated control group. Only 100 of the selected 106 drugs were tested due to an insufficient number of available worms. Trial 2: This was performed using three separate assays; 23 drugs were tested, of which 17 were retested from Trial 1 (confirmation tests), and the 6 remaining drugs that could not be tested in Trial 1; two worms per group were used for the test and positive control drugs and six worms for the untreated control group. Worm viability was assessed using the following parameters: measurement of mean worm motility scores on a scale of 0 (immotile) to 10 (maximum) every 24 h, terminating at 120 h, using an Olympus inverted microscope. Biochemical evaluation of worm viability using MTT/formazan colorimetry was carried out after the last motility reading (120 h), outlined in Figure 1. Inhibition of formazan formation is correlated with worm damage or death (viability). The results of the test drugs were compared to the untreated controls. The results were calculated as motility reduction (%) and MTT reduction (%) compared to the untreated controls (Microsoft Office Excel, 2010) and designated: good activity, 100% motility and/or MTT reduction; moderate/marginal activity, 50-99% motility and/or MTT reduction; inactive, <50% motility reduction and MTT reduction. The test drug is considered active when the motility and/or MTT reductions of \geq 50% is observed by comparison to the untreated controls. Statistical analysis was performed on only the Trial 1 data, using the two-tailed t test for the comparison between the motility and optical density means of the test drug and untreated control (Microsoft Office Excel, 2010) with a significance level of p < 0.05.

3. Results

Of the 106 selected drugs tested, 39 were inactive (<50% motility reduction and MTT reduction), 44 showed marginal/moderate activity (50–99% motility and/or MTT reduction) and 23 showed good activity (100% motility and/or MTT reduction) after 120 h drug exposure.

The results for the 44 drugs that showed moderate or marginal activity in Trial 1 are shown in Table 2. For the majority of these drugs, the MTT reductions correlated with the motility reductions, indicating that the worms did not only have reduced motility but were permanently damaged. Examination of the data shows that for 13 drugs with the greatest motility reductions of >80%, the MTT reduction was \geq 70% (*p*-values < 0.001), with the highest number of hits in the antibacterial (cefuroxime sodium, demeclocycline)

hydrochloride, methenamine), antiparasitic (amitraz, primaquine phosphate) and neurological (armodafinil, chlorpromazine, dopamine hydrochloride, rivastigmine tartrate) bioactivity groups.

Table 2. Results of the 44 identified drugs with moderate or marginal activity after 120 h drug exposure. Mot Redn—motility reduction; MTT Redn—MTT reduction.

Drug	Mo	ot Red	MTT Red		
Drug	%	<i>p</i> -Value	%	<i>p</i> -Value	
IMMITICIDE (positive control)	100.00	< 0.0001	91.09	< 0.0001	
Acepromazine Maleate	53.57	< 0.0001	57.67	< 0.001	
Amitraz	89.29	< 0.0001	77.23	< 0.0001	
Ampyzine Sulfate	78.57	< 0.0001	76.98	< 0.0001	
Apomorphine Hydrochloride	78.57	< 0.0001	76.49	< 0.0001	
Armodafinil	82.14	< 0.0001	84.65	< 0.0001	
Ascorbyl Palmitate	57.14	< 0.0001	67.08	< 0.001	
Bleomycin	53.57	< 0.0001	53.22	< 0.01	
Bromhexine Hydrochloride	57.14	< 0.0001	56.93	< 0.01	
Broxyquinoline	82.14	< 0.0001	82.18	< 0.0001	
Candicidin	67.86	< 0.0001	72.03	< 0.001	
Carbadox	60.71	< 0.0001	66.83	< 0.001	
Cefsulodin Sodium	50.00	< 0.0001	21.04	0.18	
Ceftibuten	75.00	< 0.0001	71.53	< 0.001	
Cefuroxime Sodium	92.86	< 0.0001	79.70	< 0.0001	
Chlorpromazine	82.14	< 0.0001	79.46	< 0.0001	
Clopidogrel Sulfate	28.57	< 0.01	53.47	< 0.01	
Demeclocycline Hydrochloride	82.14	< 0.0001	76.24	< 0.001	
Dexamethasone Acetate	78.57	< 0.0001	61.14	< 0.001	
Dienestrol	78.57	< 0.0001	75.74	< 0.0001	
Docusate Sodium	75.00	< 0.0001	66.58	< 0.001	
Dopamine Hydrochloride	82.14	< 0.0001	69.55	< 0.001	
Doxorubicin	60.71	< 0.0001	64.36	< 0.001	
Doxycycline Hydrochloride	53.57	< 0.0001	37.13	< 0.05	
Epirubicin Hydrochloride	75.00	< 0.0001	77.48	< 0.0001	
Fluorescein	50.00	< 0.0001	45.54	< 0.01	
Guanethidine	78.57	< 0.0001	79.95	< 0.0001	
Mangafodipir Trisodium	89.29	< 0.0001	76.49	< 0.0001	
Methenamine	92.86	< 0.0001	78.47	< 0.0001	
Methsuximide	71.43	< 0.0001	66.34	< 0.001	
Methylergonovine Maleate	57.14	< 0.0001	55.45	< 0.001	
Methylphenidate Hydrochloride	75.00	< 0.0001	88.12	< 0.0001	
Minocycline Hydrochloride	78.57	< 0.0001	72.52	< 0.001	
Moxidectin	78.57	< 0.0001	71.78	< 0.001	
Nicotinyl Alcohol Tartrate	67.86	< 0.0001	70.30	< 0.001	
Nifedipine	92.86	< 0.0001	82.92	< 0.0001	
Prednisolone Tebutate	96.43	< 0.0001	82.18	< 0.0001	
Primaquine Phosphate	92.86	< 0.0001	84.16	< 0.0001	
Propoxycaine Hydrochloride	60.71	< 0.0001	60.15	< 0.001	
Riboflavin	60.71	<0.0001	56.19	<0.01	
Rifampin	57.14	<0.0001	56.44	<0.01	
Rivastigmine Tartrate	92.86	<0.0001	81.93	< 0.001	
Sennoside A	53.57	<0.0001	52.23	<0.01	
Tetrahydrozoline Hydrochloride	53.57	<0.0001	53.22	<0.01	
Valganciclovir Hydrochloride	50.00	<0.0001	53.71	<0.01	

The 23 drugs which demonstrated good activity are shown in Table 3 (includes the chemical structures); 17 of these drugs were retests from Trial 1, and 6 drugs were new tests in Trial 2. All of the drugs produced 100% motility reduction and high levels of MTT reduction, and for most drugs there was a good level of concordance between the results of Trial 1 when compared to Trial 2. The highest number of hits was found in the antibacterial (cefaclor, chlorhexidine dihydrochloride, gramicidin, lasalocid sodium, nitrofurantoin, nitrofurazone, nitroxoline, rifaximin), anti-infective (benzethonium chloride, dequalinium chloride, methylbenzethonium chloride, oxyquinoline hemisulfate) and antiparasitic (atovaquone, hexetidine, homidium bromide, iodoquinol, levamisole hydrochloride, pyrantel pamoate) bioactivity groups.

Drug (Molecular	Molecular Structure		Trial 1		Tr	Trial 2	
Weight/Bioactivity)	Molecular Structure	Mot Redn (%)	MTT Redn (%)	<i>p</i> -Value	Mot Redn (%)	MTT Redn (%)	
IMMITICIDE, positive control (501.34/Anthelmintic)	$H_{2}N \xrightarrow{N} S^{As} S^{As} S^{NH_{2}}$	100.00	91.09	<0.0001	100.00	Range 77.19–98.88	
ATOVAQUONE (366.85/Antimalarial)	CI OH OH	100.00	90.10	<0.0001	100.00	74.42	
BENZETHONIUM CHLORIDE (448.09/Antiinfective)	H_3C CH_{BH_3} $CI^ CI^ CH_3$ $CI^ CH_3$ $CI^ CH_3$ $CI^ CH_3$ $CI^ CH_3$ CH_3 $CI^ CH_3$ CH_3 CH	100.00	88.12	<0.0001	100.00	71.91	
CHLORHEXIDINE DIHYDROCHLORIDE (578.38/Antibacterial)		100.00	90.10	<0.0001	100.00	100.00	

Table 3. Results of the 23 drugs identified with good activity after 120 h drug exposure. Mot Redn—motility reduction; MTT Redn—MTT reduction; nd—not determined; *p*-value applies to both motility and MTT reductions.

Drug (Molecular	Molecular Structure Trial 1		Trial 1			ial 2
Weight/Bioactivity)	Molecular Structure	Mot Redn (%)	MTT Redn (%)	<i>p</i> -Value	Mot Redn (%)	MTT Redn (%)
GRAMICIDIN, gramicidin A shown (1882.34/Antibacterial)		100.00	76.24	<0.0001	100.00	84.27
IODOQUINOL (396.96/Antiamoebic)	OH I I	100.00	80.20	<0.0001	100.00	87.64
ISRADIPINE (371.40/Calcium channel blocker)	H_3C_0 H_3C_0 H_3C_0 H_3C_0 CH_3	100.00	87.13	<0.0001	100.00	78.95
LASALOCID SODIUM (612.79/Antibacterial)	$HO \rightarrow f \rightarrow $	100.00	89.11	<0.0001	73.33	90.70

Trial 1 Trial 2 Drug (Molecular **Molecular Structure** Weight/Bioactivity) Mot Redn (%) MTT Redn (%) *p*-Value Mot Redn (%) MTT Redn (%) LEVAMISOLE CIH HYDROCHLORIDE 100.00 92.08 < 0.0001 55.81 100.00 (240.76/Anthelmintic) OH N=NCI LOSARTAN 100.00 < 0.0001 100.00 88.12 82.56 (422.92/Antihypertensive) H₃Ć 0_0 S CH. MELOXICAM SODIUM 47.67 100.00 89.11 < 0.0001 100.00 (373.39/Antiinflammatory) Ó 0 Na⁺ H₃C (CH, METHYLBENZETHONIUM CHLORIDE 100.00 91.09 < 0.0001 67.44 100.00 H₃C Cl (462.12/Antiinfective) H₃C⁷H₃C `CH₃

Drug (Molecular	Molecular Structure	Trial 1			Tr	ial 2
Weight/Bioactivity)	Molecular Structure	Mot Redn (%)	MTT Redn (%)	<i>p</i> -Value	Mot Redn (%)	MTT Redn (%)
MITOXANTRONE HYDROCHLORIDE (517.41/Antineoplastic)		100.00	91.09	<0.0001	100.00	70.93
NITROFURANTOIN (238.16/Antibacterial)		100.00	75.25	<0.0001	100.00	92.70
NITROFURAZONE (198.14/Antibacterial)	$ \begin{array}{c} $	100.00	85.15	<0.0001	100.00	95.51
OXYQUINOLINE HEMISULFATE (243.24/Antiinfective)	OH N O=S-OH OH	100.00	86.14	<0.0001	100.00	91.57

Drug (Molecular	Molecular Structure		Trial 1			Trial 2	
Weight/Bioactivity)	Molecular Structure	Mot Redn (%)	MTT Redn (%)	<i>p</i> -Value	Mot Redn (%)	MTT Redn (%)	
PYRANTEL PAMOATE (594.69/Anthelmintic)	CH3 CH3 OH OH OH OH OH OH OH OH	100.00	85.15	<0.0001	100.00	90.45	
RIFAXIMIN (785.90/Antibacterial)	$\begin{array}{c} \begin{array}{c} CH_{3} \\ HO \\ \hline \\ HO \\ CH_{3} \\ H_{3} \\ CH_{3} \\ CH_{3$	100.00	91.09	<0.0001	100.00	68.60	
CEFACLOR (367.81/Antibacterial)		nd	nd	nd	100.00	96.51	
DEQUALINIUM CHLORIDE (527.59/Antiinfective)		nd	nd	nd	100.00	61.40	

Drug (Molecular	Molecular Structure	Trial 1			Tr	ial 2
Weight/Bioactivity)	Molecular Structure	Mot Redn (%)	MTT Redn (%)	<i>p</i> -Value	Mot Redn (%)	MTT Redn (%)
HEXETIDINE (339.61/Antifungal)		nd	nd	nd	100.00	88.60
HOMIDIUM BROMIDE (394.32/Antiprotozoal)	H ₂ N CH ₃ Br ⁻	nd	nd	nd	100.00	81.58
NITROXOLINE (190.16/Antibacterial)		nd	nd	nd	100.00	82.46
PENFLURIDOL (523.98/Antipsychotic)		nd	nd	nd	100.00	64.04

4. Discussion

With the urgent need to identify drugs with potential macrofilaricidal activity against Onchocerca parasites, using the strategy of drug repurposing to identify new drugs for the prompt development of therapeutics for the treatment of filariasis is not a new concept; indeed, the drugs currently in use to treat filarial infections, ivermectin, diethylcarbamazine, moxidectin and doxycycline, have all been repurposed from the veterinary or medical fields [14]. Previous studies have screened libraries and drugs for activity against filarial parasites [15] and the Pharmakon 1600 library itself has also been screened for antischistosomal activity [16]. All human-infecting filarial nematodes, with the exception of L. loa, carry the endosymbiotic Wolbachia bacteria which has been shown to be essential for O. volvulus fertility and viability [17,18]. Studies using drugs from diverse libraries in anti-Wolbachia screens have revealed promising candidates for further development [19]. Rifampicin (also known as rifampin), an antibiotic used for the treatment of tuberculosis, has been developed for testing in clinical trials based on the effect of high dose, long term exposure using in vitro assays against O. gutturosa adult male worms [20] and Wolbachia [21]. In addition, in vivo studies have demonstrated more than 90% Wolbachia depletion using Brugia malayi and Onchocerca ochengi models [22]. To test whether the treatment time for onchocerciasis could be reduced, rifampicin in combination with albendazole is currently being investigated in a clinical trial in Cameroon [23]. Emodepside, a veterinary drug licensed for the oral treatment of gastrointestinal nematodes, exhibited high activity in vitro and in vivo against various filarial parasites [24]. A Phase I clinical trial in healthy humans has been completed [25] and recruitment for a Phase II trial in onchocerciasis patients is currently underway [26]. Similarly, oxfendazole is a drug that is used against intestinal helminths in the veterinary field over several decades. In vitro and in vivo animal studies showed that oxfendazole is also active against filarial nematodes [27,28]. Oxfendazole was tested in Phase 1 clinical trials [29] and will be tested in onchocerciasis, loiasis, mansonellosis and *Trichuris trichiura* patients in a Phase 2 clinical basket trial [30]. In this study, we aimed to identify selected existing drugs contained within the FDA-approved Pharmakon 1600 library with the potential to be rapidly developed as macrofilaricides against onchocerciasis.

The results of the standard anthelmintics, contained within the antiparasitic bioactivity group, were as expected and demonstrate that screening of diverse libraries using this 5-day motility/MTT in vitro assay is suitable for identifying new drug candidates with activity against Onchocerca parasites. Nevertheless, these types of in vitro assays do not tell us all we need to know about the activity of drugs, since host factors may play an important role. Diethylcarbamazine citrate and flubendazole were inactive in this study and this result corresponds to previous in vitro findings [27,31,32]. However, clinical studies in Mexico demonstrated that flubendazole has high macrofilaricidal activity [33,34] and in vivo laboratory studies confirmed this activity [35]. Recently, the important role of the immune system in supporting the macrofilaricidal efficacy for the related drug oxfendazole was demonstrated in the *Litomosoides sigmodontis* filarial mouse model [28]. Despite the limitation that some candidates may require an intact immune system for efficacy, several promising candidates were identified. In this study, levamisole hydrochloride displayed good activity (100% motility reduction), the parasites were completely immotile after 5 days of exposure (Table 3), and this result is in accordance with previous studies [31], although the MTT result indicated paralysis rather than worm death. Unsurprisingly, pyrantel pamoate (anthelmintic), which is used for roundworm and pinworm infections, showed good activity against O. gutturosa parasites in this study; this drug is a depolarising neuromuscular blocking agent which causes paralysis of the worms [36]. Treatment of onchocerciasis patients with pyrantel pamoate showed no notable activity against adult worms of O. volvulus [37], possibly due to poor oral uptake or suboptimal dosage/treatment length. Moxidectin showed moderate activity; the motility of the parasites was reduced by 78.6% with a comparable reduction in viability (71.8%) indicating slow killing of the worms in vitro (Table 2). This drug was licensed to treat human onchocerciasis in 2018 [38]. Of the drugs that displayed moderate/marginal activity, cefuroxime sodium, methenamine, primaquine phosphate and rivastigmine tartrate had significant effects on the parasites with motility reductions of >90% and MTT reductions of >70% (*p*-values < 0.001, Table 2). Oral formulations are available for these drugs and therefore they should be considered "drugs of interest" to be further investigated for use against *Onchocerca* infections. Cefuroxime sodium and methenamine are used for the treatment of bacterial infections, primaquine phosphate (antimalarial) is used for the treatment of hypnozoites, the dormant form of *Plasmodium* parasites during malaria tertiana and rivastigmine tartrate has neurological activity in the treatment of dementia associated with Alzheimer's or Parkinson's diseases. Rifampin (internationally known as rifampicin) also showed moderate activity (Table 2) and is currently in development for the treatment of onchocerciasis [15]. Such a slow macrofilaricidal efficacy is known for antibiotics that target and eliminate the *Wolbachia* endosymbionts of filariae, such as doxycycline [15]. The semi-synthetic derivative of rifampin, rifaximin, completely reduced the motility of the parasites (Table 3); however, due to poor absorption, rifaximin is only used to treat gastrointestinal infections.

Drugs belonging to novel classes, with available oral formulations, rendered O. gutturosa male worms completely immotile (good activity, 100% motility reduction) but the parasites were not dead as indicated by the MTT reductions (Table 3); longer exposure to these drugs, or a longer period in culture following exposure to the drug, may result in parasite death. Isradipine (calcium channel blocker) and losartan (angiotensin receptor blocker) are both used to treat hypertension and cause vasodilation by blocking different receptors. Penfluridol with neurological activity is commonly used as an antipsychotic drug. The anticancer drug, mitoxantrone hydrochloride, is used in the treatment of prostate cancer and leukaemia. Of interest is the activity of atovaquone (antimalarial) against the O. gutturosa worms. In addition to its use as prophylaxis and treatment against malaria parasites, it is also used for the treatment of pneumonia caused by fungal infection and some other microbial infections [39]. Several studies have investigated this drug for the treatment of different types of cancers [40]. Also of interest is the antibacterial cefaclor which belongs to the large cephalosporin family of antibiotics; this drug class is structurally related to penicillin and used to treat a wide range of bacterial infections. Cefuroxime sodium, which was moderately/marginally active against the parasites, together with cefaclor, are both second-generation drugs and oral formulations are available for both drugs. We speculate that the active antibacterial drugs tested in this study had a direct effect on worms, with the possibility of an indirect effect by killing Wolbachia in the longer term.

In this study we have taken the first step to identify FDA-approved drugs with potential anti-*Onchocerca* activity; some of the identified hits should be further investigated for repurposing. Of lower priority are the drugs designed for topical use and those that can only be administered parenterally. Further investigation of the candidate drugs that displayed promising in vitro activity against *O. gutturosa* adult male parasites was curtailed due to the unexpected, imposed travel and work restrictions with regard to studies in Gambia during the COVID-19 pandemic. Further in vitro trials are required to retest the hits of interest to estimate activity endpoints and EC₅₀ values. This data together with the available pharmacokinetic and toxicity profiles can be used to rapidly inform the development of the drugs for further evaluation against *Onchocerca* and other filarial species of medical and veterinary importance. In addition, these hits may provide a good starting point to assess related compounds of interest and for the synthesis of new drugs.

Author Contributions: Conceptualization, S.T. (Simon Townson) and I.S.; methodology, S.G., S.T. (Simon Townson) and A.F.; software, S.G.; formal analysis, S.G. and S.T. (Simon Townson); investigation, S.G., S.T. (Simon Townson), A.F., J.S.-K. and J.Z.; data curation, S.G. and A.F.; writing—original draft preparation, S.G. and S.T. (Simon Townson); writing—review and editing, M.P.H., S.G., S.T. (Simon Townson), S.T. (Senyo Tagboto) and I.S.; supervision, S.G., S.T. (Simon Townson) and I.S.; project administration, S.T. (Simon Townson) and I.S.; funding acquisition, S.T. (Simon Townson). All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by funding from the Drugs for Neglected Diseases initiative, Geneva. M.P.H. is funded under Germany's Excellence Strategy—EXC2151 390873048. M.P.H. is a member of the German Center for Infection Research (DZIF). M.P.H. received funding from the German Center for Infection Research (TTU 09.701).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are available upon reasonable request.

Conflicts of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

- Brattig, N.W.; Cheke, R.A.; Garms, R. Onchocerciasis (river blindness)—More than a century of research and control. *Acta Trop.* 2021, 218, 105677. [CrossRef]
- 2. WHO. Available online: https://www.who.int/health-topics/neglected-tropical-diseases#tab=tab_1 (accessed on 15 October 2023).
- 3. Duke, B.O.; Zea-Flores, G.; Muñoz, B. The embryogenesis of *Onchocerca volvulus* over the first year after a single dose of ivermectin. *Trop. Med. Parasitol.* **1991**, 42, 175–180.
- 4. Basáñez, M.G.; Pion, S.D.; Boakes, E.; Filipe, J.A.; Churcher, T.S.; Boussinesq, M. Effect of single-dose ivermectin on *Onchocerca volvulus*: A systematic review and meta-analysis. *Lancet Infect. Dis.* **2008**, *8*, 310–322. [CrossRef]
- 5. Plaisier, A.P.; van Oortmarssen, G.J.; Remme, J.; Habbema, J.D. The reproductive lifespan of *Onchocerca volvulus* in West African savanna. *Acta Trop.* **1991**, *48*, 271–284. [CrossRef]
- Geerts, S.; Gryseels, B. Drug resistance in human helminths: Current situation and lessons from livestock. *Clin. Microbiol. Rev.* 2000, 13, 207–222. [CrossRef]
- 7. von Samson-Himmelstjerna, G.; Harder, A.; Sangster, N.C.; Coles, G.C. Efficacy of two cyclooctadepsipeptides, PF1022A and emodepside, against anthelmintic-resistant nematodes in sheep and cattle. *Parasitology* **2005**, *130*, 343–347. [CrossRef]
- 8. Prichard, R.K.; Geary, T.G. Perspectives on the utility of moxidectin for the control of parasitic nematodes in the face of developing anthelmintic resistance. *Int. J. Parasitol. Drugs Drug Resist.* **2019**, *10*, 69–83. [CrossRef]
- 9. Osei-Atweneboana, M.Y.; Awadzi, K.; Attah, S.K.; Boakye, D.A.; Gyapong, J.O.; Prichard, R.K. Phenotypic evidence of emerging ivermectin resistance in Onchocerca volvulus. *PLoS Negl. Trop. Dis.* **2011**, *5*, e998. [CrossRef]
- Nana-Djeunga, H.C.; Bourguinat, C.; Pion, S.D.; Bopda, J.; Kengne-Ouafo, J.A.; Njiokou, F.; Prichard, R.K.; Wanji, S.; Kamgno, J.; Boussinesq, M. Reproductive status of Onchocerca volvulus after ivermectin treatment in an ivermectin-naïve and a frequently treated population from Cameroon. *PLoS Negl. Trop. Dis.* 2014, *8*, e2824. [CrossRef]
- 11. Milton, P.; Hamley, J.I.D.; Walker, M.; Basáñez, M.G. Moxidectin: An oral treatment for human onchocerciasis. *Expert Rev. Anti Infect. Ther.* **2020**, *18*, 1067–1081. [CrossRef]
- 12. Pfarr, K.M.; Krome, A.K.; Al-Obaidi, I.; Batchelor, H.; Vaillant, M.; Hoerauf, A.; Opoku, N.O.; Kuesel, A.C. The pipeline for drugs for control and elimination of neglected tropical diseases: 2. Oral anti-infective drugs and drug combinations for off-label use. *Parasit. Vectors* **2023**, *16*, 394. [CrossRef]
- 13. Townson, S.; Ramirez, B.; Fakorede, F.; Mouries, M.A.; Nwaka, S. Challenges in drug discovery for novel antifilarials. *Expert Opin. Drug Discov.* **2007**, *2*, S63–S73. [CrossRef]
- 14. Tagboto, S.; Orish, V. Drug development for onchocerciasis-the past, the present and the future. *Front. Trop. Dis.* **2022**, *3*, 953061. [CrossRef]
- 15. Ehrens, A.; Hoerauf, A.; Hübner, M.P. Current perspective of new anti-Wolbachial and direct-acting macrofilaricidal drugs as treatment strategies for human filariasis. *GMS Infect. Dis.* **2022**, *10*, Doc02. [CrossRef]
- 16. Panic, G.; Vargas, M.; Scandale, I.; Keiser, J. Activity Profile of an FDA-Approved Compound Library against *Schistosoma mansoni*. *PLoS Negl. Trop. Dis.* **2015**, *9*, e0003962. [CrossRef]
- 17. Tamarozzi, F.; Halliday, A.; Gentil, K.; Hoerauf, A.; Pearlman, E.; Taylor, M.J. Onchocerciasis: The role of Wolbachia bacterial endosymbionts in parasite biology, disease pathogenesis, and treatment. *Clin. Microbiol. Rev.* **2011**, *24*, 459–468. [CrossRef]
- 18. Hoerauf, A. Filariasis: New drugs and new opportunities for lymphatic filariasis and onchocerciasis. *Curr. Opin. Infect. Dis.* 2008, 21, 673–681. [CrossRef]
- Johnston, K.L.; Cook, D.A.N.; Berry, N.G.; David Hong, W.; Clare, R.H.; Goddard, M.; Ford, L.; Nixon, G.L.; O'Neill, P.M.; Ward, S.A.; et al. Identification and prioritization of novel anti-Wolbachia chemotypes from screening a 10,000-compound diversity library. *Sci. Adv.* 2017, *3*, eaao1551. [CrossRef]
- 20. Townson, S.; Tagboto, S.; McGarry, H.F.; Egerton, G.L.; Taylor, M.J. *Onchocerca parasites* and Wolbachia endosymbionts: Evaluation of a spectrum of antibiotic types for activity against *Onchocerca gutturosa* in vitro. *Filaria J.* **2006**, *5*, 4. [CrossRef]
- 21. Fenollar, F.; Maurin, M.; Raoult, D. Wolbachia pipientis growth kinetics and susceptibilities to 13 antibiotics determined by immunofluorescence staining and real-time PCR. *Antimicrob. Agents Chemother.* **2003**, *47*, 1665–1671. [CrossRef]

- 22. Aljayyoussi, G.; Tyrer, H.E.; Ford, L.; Sjoberg, H.; Pionnier, N.; Waterhouse, D.; Davies, J.; Gamble, J.; Metuge, H.; Cook, D.A.N.; et al. Short-Course, High-Dose Rifampicin Achieves Wolbachia Depletion Predictive of Curative Outcomes in Preclinical Models of Lymphatic Filariasis and Onchocerciasis. *Sci. Rep.* **2017**, *7*, 210. [CrossRef]
- Wanji, S.; Hoerauf, A.; Klarmann-Schulz, U. ISRCTN38954299—The Efficacy of Rifampicin Plus Albendazole against River Blindness (Onchocerciasis) in Cameroon. Available online: https://www.isrctn.com/ISRCTN38954299 (accessed on 5 November 2023).
- Hübner, M.P.; Townson, S.; Gokool, S.; Tagboto, S.; Maclean, M.J.; Verocai, G.G.; Wolstenholme, A.J.; Frohberger, S.J.; Hoerauf, A.; Specht, S.; et al. Evaluation of the in vitro susceptibility of various filarial nematodes to emodepside. *Int. J. Parasitol. Drugs Drug Resist.* 2021, 17, 27–35. [CrossRef]
- Gillon, J.Y.; Dennison, J.; van den Berg, F.; Delhomme, S.; Dequatre Cheeseman, K.; Peña Rossi, C.; Strub Wourgaft, N.; Specht, S.; Pedrique, B.; Monnot, F.; et al. Safety, tolerability and pharmacokinetics of emodepside, a potential novel treatment for onchocerciasis (river blindness), in healthy male subjects. *Br. J. Clin. Pharmacol.* 2021, *87*, 3949–3960. [CrossRef]
- 26. DNDi. Available online: https://dndi.org/research-development/portfolio/emodepside/ (accessed on 15 October 2023).
- Hübner, M.P.; Martin, C.; Specht, S.; Koschel, M.; Dubben, B.; Frohberger, S.J.; Ehrens, A.; Fendler, M.; Struever, D.; Mitre, E.; et al. Oxfendazole mediates macrofilaricidal efficacy against the filarial nematode Litomosoides sigmodontis in vivo and inhibits Onchocerca spec. motility in vitro. *PLoS Negl. Trop. Dis.* 2020, 14, e0008427. [CrossRef]
- 28. Risch, F.; Scheunemann, J.F.; Reichwald, J.J.; Lenz, B.; Ehrens, A.; Gal, J.; Fercoq, F.; Koschel, M.; Fendler, M.; Hoerauf, A.; et al. The efficacy of the benzimidazoles oxfendazole and flubendazole against Litomosoides sigmodontis is dependent on the adaptive and innate immune system. *Front. Microbiol.* **2023**, *14*, 1213143. [CrossRef]
- 29. Bach, T.; Galbiati, S.; Kennedy, J.K.; Deye, G.; Nomicos, E.Y.H.; Codd, E.E.; Garcia, H.H.; Horton, J.; Gilman, R.H.; Gonzalez, A.E.; et al. Pharmacokinetics, Safety, and Tolerability of Oxfendazole in Healthy Adults in an Open-Label Phase 1 Multiple Ascending Dose and Food Effect Study. *Antimicrob. Agents Chemother.* **2020**, *64*, e01018. [CrossRef]
- 30. eWHORM. Available online: https://ewhorm.org/ (accessed on 5 November 2023).
- 31. Townson, S.; Connelly, C.; Dobinson, A.; Muller, R. Drug activity against Onchocerca gutturosa males in vitro: A model for chemotherapeutic research on onchocerciasis. *J. Helminthol.* **1987**, *61*, 271–281. [CrossRef]
- Strote, G.; Wieland, S.; Darge, K.; Comley, J.C. In vitro assessment of the activity of anthelmintic compounds on adults of Onchocerca volvulus. *Acta Leiden*. 1990, 59, 285–296. [PubMed]
- Dominguez-Vazquez, A.; Taylor, H.R.; Greene, B.M.; Ruvalcaba-Macias, A.M.; Rivas-Alcala, A.R.; Murphy, R.P.; Beltran-Hernandez, F. Comparison of flubendazole and diethylcarbamazine in treatment of onchocerciasis. *Lancet* 1983, 1, 139–143. [CrossRef]
- 34. Mackenzie, C.D.; Geary, T.G. Flubendazole: A candidate macrofilaricide for lymphatic filariasis and onchocerciasis field programs. *Expert Rev. Anti Infect. Ther.* **2011**, *9*, 497–501. [CrossRef] [PubMed]
- Hübner, M.P.; Ehrens, A.; Koschel, M.; Dubben, B.; Lenz, F.; Frohberger, S.J.; Specht, S.; Quirynen, L.; Lachau-Durand, S.; Tekle, F.; et al. Macrofilaricidal efficacy of single and repeated oral and subcutaneous doses of flubendazole in Litomosoides sigmodontis infected jirds. *PLoS Negl. Trop. Dis.* 2019, 13, e0006320. [CrossRef] [PubMed]
- Fissiha, W.; Kinde, M.Z. Anthelmintic Resistance and Its Mechanism: A Review. *Infect. Drug Resist.* 2021, 14, 5403–5410. [CrossRef]
 [PubMed]
- 37. Kale, O. Small-scale trials of six drugs against Onchocerca volvulus. Tropenmed. Parasitol. 1978, 29, 163–167.
- 38. Kura, K.; Milton, P.; Hamley, J.I.D.; Walker, M.; Bakajika, D.K.; Kanza, E.M.; Opoku, N.O.; Howard, H.; Nigo, M.M.; Asare, S.; et al. Can mass drug administration of moxidectin accelerate onchocerciasis elimination in Africa? *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 2023, 378, 20220277. [CrossRef]
- 39. Baggish, A.L.; Hill, D.R. Antiparasitic agent atovaquone. Antimicrob. Agents Chemother. 2002, 46, 1163–1173. [CrossRef]
- Cheng, G.; Hardy, M.; Topchyan, P.; Zander, R.; Volberding, P.; Cui, W.; Kalyanaraman, B. Potent inhibition of tumour cell proliferation and immunoregulatory function by mitochondria-targeted atovaquone. *Sci. Rep.* 2020, *10*, 17872. [CrossRef] [PubMed]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.