Human Immunodeficiency Virus-1 (HIV-1)-Mediated Apoptosis: New Therapeutic Targets

Zukile Mbita, Rodney Hull and Zodwa Dlamini*

College of Agriculture and Environmental Sciences, University of South Africa, Florida Science Campus, C/o Christiaan de Wet and Pioneer Avenue P/Bag X6, Johannesburg 1710, South Africa; E-Mails: mbitaz@unisa.ac.za (Z.M.); hullr@unisa.ac.za (R.H.)

* Author to whom correspondence should be addressed; E-Mail: dlamizl@unisa.ac.za; Tel.: +27-11-471-2272.

Received: 13 March 2014; in revised form: 12 June 2014 / Accepted: 8 July 2014 / Published: 19 August 2014

Abstract: HIV has posed a significant challenge due to the ability of the virus to both impair and evade the host’s immune system. One of the most important mechanisms it has employed to do so is the modulation of the host’s native apoptotic pathways and mechanisms. Viral proteins alter normal apoptotic signaling resulting in increased viral load and the formation of viral reservoirs which ultimately increase infectivity. Both the host’s pro- and anti-apoptotic responses are regulated by the interactions of viral proteins with cell surface receptors or apoptotic pathway components. This dynamic has led to the development of therapies aimed at altering the ability of the virus to modulate apoptotic pathways. These therapies are aimed at preventing or inhibiting viral infection, or treating viral associated pathologies. These drugs target both the viral proteins and the apoptotic pathways of the host. This review will examine the cell types targeted by HIV, the surface receptors exploited by the virus and the mechanisms whereby HIV encoded proteins influence the apoptotic pathways. The viral manipulation of the hosts’ cell type to evade the immune system, establish viral reservoirs and enhance viral proliferation will be reviewed. The pathologies associated with the ability of HIV to alter apoptotic signaling and the drugs and therapies currently under development that target the ability of apoptotic signaling within HIV infection will also be discussed.

Keywords: apoptosis; HIV; HIV encoded proteins; antiviral; HIVAN; HAND
1. Introduction

Human immunodeficiency virus (HIV) is a retrovirus that causes Acquired Immunodeficiency Syndrome (AIDS) due to immune system compromise that leads to the body’s inability to fight off opportunistic infections. HIV infection results in the catastrophic loss of cells through apoptosis and chief among these are uninfected T cells, where the activation of programmed cell death pathways aid in the suppression of the immune system [1–3]. Apoptosis directed at the immune cells leaves the body helpless with no defense mechanisms. The devastating effects of HIV-1 infections are known, but how the HIV retrovirus manipulates the immune system in its favor remains elusive and unclear. Understanding the mechanisms that HIV-1 influences can lead to new therapeutic intervention and innovations.

The primary site of HIV infection is the lining of mucosal surfaces; a site that is abundantly inhabited by immature dendritic cells [4,5]. HIV infects all the CD4\(^+\) cells, including lymphocytes, monocytes and macrophages. These cells form the first line of defense, the innate immune system. In addition, HIV-1 persistently replicates in macrophages, compromising the integrity and function of the immune system, resulting in new HI viruses that further infect CD4 T cells, leading to severe immunosuppression [6]. However, while the percentage of lymphocytes declines, the percentage of macrophages does not. Since macrophages are resistant to the cytopathic effects of the virus, they survive for longer periods and contribute to further viral replication, serving as an important reservoir for HIV-1 persistence and replication [7,8]. CD8\(^+\) cytotoxic T lymphocytes kill the infected CD4\(^+\) cells, resulting in an immune-compromised body susceptible to opportunistic infections that can result in other pathologies. For new therapeutic interventions, the mechanisms that dictate the resistance or susceptibility of cells to HIV-induced apoptosis must be elucidated.

Proteins coded for by HIV have been implicated in the increase in cell death. These include gp120 [3,9] and Tat-proteins [10]. HIV induced apoptotic cell death was shown to occur in many different cell types [1,10]. These tissue specific increases in HIV induced apoptosis provide explanations for some of the diverse pathologies observed in HIV infected individuals. The apoptotic pathways induced by HIV are components of the intrinsic as well as the extrinsic apoptotic pathways and include the MAPK pathway [10], caspase 8 and 9 [11,12], destruction of host structural proteins [11], down regulation of BCL-2 [13], mitochondrial cytochrome c release [14], shortening of telomeres [15], HIV-1 long terminal repeats (LTR) and the transcription and activation of NF-\(\kappa\)B [16].

Currently, there is no cure for HIV infection. However, antiretroviral therapy against HIV has been very successful, but now research is focusing on achieving the ultimate goal, a cure. Consequently, understanding the host factors that control disease progression will be fundamental to the development of new therapeutic strategies [17]. In this review, the apoptotic pathways that result in the demise of the immune system and possible apoptotic-mediated interventions for therapeutic purposes will be covered. Finally, this paper will focus on the drugs currently under development.

2. Mediators of Apoptosis in HIV

Apoptosis occurs via two distinct pathways, an intrinsic (Figure 1) and an extrinsic pathway (Figure 2). The extrinsic (or external) pathway is initiated by the binding of ligands such as Fas ligand (FasL),
TNF, and TRAIL/Apo-2 ligands to their death receptors FAS/CD95, TNFR1, DR4, and DR5 [18]. The pathway relies on the activation of caspases 8 and 10 which in turn activate the effector caspases 3 and 7. The intrinsic (or internal) pathway is initiated by the disruption of the mitochondrial membrane, resulting in the release of cytochrome c, regulated by the bcl-2 family of proteins [18]. The bcl-2 family contains pro- and anti-apoptotic proteins, with the pro-apoptotic proteins binding the anti-apoptotic proteins to initiate an apoptotic signal by promoting the dimerization of Bax and Bak [19]. These proteins function by creating pores within the mitochondrial membrane. Once released cytochrome c is responsible for the assembly of a caspase-activating complex, which in turn activates caspase-9 [19]. While some of the HIV proteins such as Vpr can induce apoptosis in infected CD4 T cells via the mitochondrial pathway, (Figure 1), other HIV proteins (e.g., Nef) can induce apoptosis in CD4 T cells by the death receptor pathway (Figure 2).

2.1. Viral Entry and the Cell Surface Receptors

The primary and secondary cellular receptors CD4 and CCR5/CXCR4 are recognized by the viral envelope proteins. These envelope proteins then insert into the lipid membrane, and this is followed by the fusion of the viral and cellular membranes allowing entry of the viral particles. This fusion is facilitated by the fact that the viral envelope is formed when the viral particle, which is released through budding, consists of a portion of the plasma membrane of the host cell [20]. HIV-1 infection of CD4 T cells is favored by cell-to-cell contact, through the formation of the virological synapse [21]. It is important to note that both infectious and non-infectious HIV particles are able to induce selective apoptosis of CD4 T cells, suggesting that noninfectious HIV particles, which make up a large portion of plasma virus, contribute to the decline in CD4 counts in patients [22,23].

2.2. Death Receptors

The Tumor Necrosis Factor Receptor (TNFR) gene family contains the death receptor proteins. These molecules all contain a cysteine rich extracellular domain and a homologous cytoplasmic death domain. However, they do have diverse primary sequences allowing them to recognize specific ligands. The cytoplasmic death domains are responsible for the transmission of death signals through adapter proteins that contain death domains such as FADD, TRADD or DAXX [24].

2.2.1. Fas/FasL

The Fas antigen is a member of the TNF-α family. Cross linking of this receptor led to apoptosis that is similar to that induced by TNF [25]. The Fas/Fas ligand (FasL) apoptotic pathway has been studied extensively, and it plays a major role in the induction of apoptosis in peripheral blood T cells [26]. Consequently, this pathway has been identified as a possible mechanism that contributes to apoptosis of CD4 T cells in AIDS [27]. Both soluble and membrane bound Fas and FasL are present at higher levels in HIV infected patients, and other studies showed that CD4 cross-linking and Fas ligation resulted in apoptosis of both CD4 and CD8 T cells [28,29]. The expression of the Tumor necrosis factor receptors Fas/FasL increases in human macrophages infected with HIV. HIV infection also up-regulates FasL in CD4 T cells after they are exposed to soluble Tat, gp120 or Nef.
proteins [30]. Treatment of HIV infected cells with Anti-Fas antibodies did not completely block apoptosis, which points to the involvement of multiple death pathways [22].

**Figure 1.** The involvement of HIV proteins in intrinsic apoptotic pathways: The viral proteins are marked in yellow. Bcl-2 is a central molecule in these mitochondrial-mediated cell death pathways. This protein is a target of Tat, Nef and HIV protease. Depending on the pathway activated the virus either shifts the balance of pro-apoptotic Bcl-2 family proteins to anti-apoptotic counterparts or vice versa. HIV proteins also affect the activation of caspases, and control their inhibitors as well, thereby, influencing the p53 signaling pathway.
**Figure 2.** The influence of HIV proteins on the extrinsic apoptotic pathways: HIV proteins influence the expression of death receptors Fas or Tumor necrosis factor (TNF)–related apoptosis-inducing ligand (TRAIL)-R1/R1 (DR4/DR5). Up-regulation of these induces receptor—mediated cell death. HIV proteins also act on caspases, the p53 pathway and influence ubiquitin associated protein degradation.

2.2.2. TNF

Tumor Necrosis Factor α (TNFα) binds to either the 55 kDa TNFR1 or the 75 kDa TNFR2. Binding leads to the activation of transcription factors NF-κB, AP-1, c-Jun N-terminal kinase, and p38. HIV
proteins are known to bind to the Tumor Necrosis Factor Receptor (TNFR) which results in apoptosis in surrounding bystander T cells. Additionally, HIV-1 proteins target the TNFR pathway, altering gene expression and leading to increased HIV replication in infected cells [31]. The HIV-1 proteins Nef and Vpr both mimic TNFR signaling, resulting in the positive regulation of viral particle production (LTR activation) through the activation of NF-κB and JNK pathways [31]. In the later stages of infection higher levels of TNF are produced leading to an increase in the death of uninfected bystander T cells, which gives rise to the subsequent immune suppression. TNFR stimulation leads to a decrease in the levels of the anti-apoptotic proteins Bcl-XL and activation of caspase 8 [32]. A model has been proposed explaining the interactions observed between TNF signaling and disease progression. In the initial stages of infection viral proteins activate the TNFR pathway to mimic the effects of this pathway. These include a G2 cell cycle arrest, the increase in changes in transcription leading to cell death, interleukin 1 secretion, cell proliferation and cell differentiation [33]. As the disease progresses the onset of AIDS is caused by viral proteins enhancing TNF signaling leading to an increase in apoptosis of bystander T cells. Pro-inflammatory cytokine production is also increased attracting more T cell and macrophages to the infected cell, further increasing the number of bystander cells that are induced to undergo apoptosis [31].

2.2.3. TRAIL

Tumor necrosis factor (TNF)–related apoptosis-inducing ligand (TRAIL), a TNF super family member, induces the apoptosis of virus-infected and tumor cells. When exposed to HIV, uninfected CD4 T cells express TRAIL, DR5, and activated caspase-3 resulting in their eventual apoptosis [22]. Several other studies suggested a role for TRAIL in the apoptosis of CD4 T cells in HIV infection. For example, CD4 and CD8 T cells from HIV—infected patients were observed to be more susceptible to TRAIL-induced apoptosis in vitro than T cells from healthy donors [34]. TRAIL was also shown to induce selective apoptosis of uninfected CD4 T cells in HIV–infected human peripheral-blood leukocyte–non-obese diabetic–severe combined immunodeficient (hu-PBLNOD-SCID) mice [35]. TRAIL produced by monocytes exposed to the HIV Tat protein also resulted in the apoptosis of uninfected CD4 T cells [36].

The TRAIL protein is expressed on the cell membrane or secreted, and both the soluble and membrane-bound forms induce the apoptosis of cells expressing death receptors [37]. TRAIL has 2 death receptors capable of inducing apoptosis (DR4 and DR5), and 3 other receptors that engage ligands without initiating apoptosis [38,39]. The TRAIL gene is regulated by type 1 interferon (IFN)-α/β, which is mainly produced by plasmacytoid dendritic cells (pDCs) and has been shown to have a broad antiviral activity, including activity against HIV [40]. This apoptosis is partially prevented by anti-TRAIL antibodies, a situation similar to that observed for Fas/FasL and points to the involvement of multiple death mechanisms or receptors [22].

2.2.4. Co-Receptors CCR5/CXCR4

In order for the virus to enter the host cells, the viral surface protein Env must first bind to the host receptor CD4 and consequently, to either the CCR5 or CXCR4 co-receptor, (Figure 1). CCR5 has three known natural ligands the presence of which reduces HIV infection by directly competing
with Env for binding sites. These ligands: RANTES, MIP-1α and MIP-1β are produced by CD8+ T cells while CCR5 is expressed on the surface of macrophages, microglia and central and effector memory T cells [41]. CXCR4 is expressed on the cell surface lymphocytes [42], however, CXCR4 is more broadly expressed than CCR5 being found on the surface of most hematopoietic and parenchymal cells [41]. The physiological ligand for CXCR4 is the chemokine stromal cell-derived factor-1 (SDF-1) [42].

The T cell infecting strains preferentially induce apoptosis through interaction with CXCR4. Dual trophic strains have no preference for the co-receptor bound to induce apoptosis [43]. A change in HIV-1 bias for binding to CXCR4 over CCR5 precedes AIDS development and the decline in CD4 cell number. However, this co-receptor switch is not a requirement for disease progression [44], but CCR5 dependent apoptosis is an absolute requirement for the HIV-1 R5 trophic mediated killing of uninfected bystander cells [45]. Irrespective of the co-receptor used HIV is still able to induce TRAIL and DR5 expression and preferential apoptosis of CD4 T cells [22].

2.3. HIV Proteins and Apoptosis

HIV-1 encodes only 15 proteins [46] (Table 1) and thus must exploit multiple host cell functions for successful infection [47]. These include three structural proteins Gag, Pol and Env. These polyproteins are proteolysed to give rise to smaller individual proteins; Gag gives rise to four proteins MA (matrix), CA (capsid), NC (nucleocapsid) and p6. Pol gives rise to three proteins PR (protease), RT (reverse transcriptase) and IN (integrase). Finally, Env gives rise to two proteins SU (surface or gp120) and TM (trans-membrane or gp41). The remaining six proteins encoded by HIV include the two gene regulatory proteins Tat and Rev as well as the four accessory proteins Vif, Vpr, Nef and Vpu [48].

2.3.1. HIV Protease

HIV protease is crucial for the life cycle of HIV, as this protease is responsible for cleaving the Gag/Pol polyprotein into functional subunits. Mutation or inhibition of this enzyme results in non-infectious viral particles [49]. Furthermore, HIV protease can also cleave proteins such as actin, laminin-B, desmon, vimentin; cleavage of vimentin results in changes in the nuclear morphology and chromatin organization [11,50,51]. The cleavage of these cytoskeletal proteins can result in apoptosis [11] (Figure 1). Over expression of HIV protease in yeast and mammalian cells results in cell lysis with both cell types displaying a loss of plasma membrane integrity and changes in membrane permeability. In yeast cells the cell walls breakdown following HIV protease over expression [52].

The viral protease can also act on components of the apoptotic machinery, killing infected and uninfected lymphocytes through the action of several host molecules, by the extrinsic pathway through members of the tumor necrosis factor family (Figure 2), or via the mitochondrial apoptotic pathway [49] (Figure 1). Over expression of the viral protease in cultured cells results in increased apoptosis which can be visualized as increased DNA fragmentation. The increase in apoptosis observed here was due to the proteolytic degradation of Bcl-2 an anti-apoptotic protein by the HIV protease [53]. HIV protease is also able to cleave procaspase-8 into a unique form known as casp8p41 which is able to induce the mitochondrial-dependent pathway of apoptosis [54]. HIV protease can also
lead to an increase in the levels of active NF-κβ through the proteolytic processing of the precursor molecule [55]. Additionally, NF-κβ can also be up-regulated through the actions of casp8p41 [56].

**Table 1.** Pro and Anti-apoptotic functions of HIV proteins.

<table>
<thead>
<tr>
<th>Pro/anti Apoptotic</th>
<th>Mechanism</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>HIV protease</td>
<td>Pro-apoptotic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Cytoskeletal damage</td>
<td>[51]</td>
</tr>
<tr>
<td></td>
<td>Damage to Plasma membrane</td>
<td>[52]</td>
</tr>
<tr>
<td></td>
<td>Proteolytic cleavage of Bcl-2</td>
<td>[53]</td>
</tr>
<tr>
<td></td>
<td>Cleavage of procaspase 8</td>
<td>[54]</td>
</tr>
<tr>
<td>Anti-apoptotic</td>
<td>Increased NF-κβ signaling</td>
<td>[55,56]</td>
</tr>
<tr>
<td>Tat</td>
<td>Pro-apoptotic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Up-regulate Caspase 3 and 8</td>
<td>[57]</td>
</tr>
<tr>
<td></td>
<td>Up-regulation of FasL and RCAS</td>
<td>[58]</td>
</tr>
<tr>
<td></td>
<td>Up-regulation of Bax</td>
<td>[58]</td>
</tr>
<tr>
<td></td>
<td>Decrease in FOXO3a signaling - FLIP decrease</td>
<td>[59,60]</td>
</tr>
<tr>
<td></td>
<td>Altered microtubule stability resulting in Bcl2 inhibition</td>
<td>[61,62]</td>
</tr>
<tr>
<td></td>
<td>Increased ROS production</td>
<td>[63]</td>
</tr>
<tr>
<td>Anti-apoptotic</td>
<td>Increased resistance of cells to TNF, Fas and TRAIL</td>
<td>[58]</td>
</tr>
<tr>
<td></td>
<td>Decrease in Caspase 10</td>
<td>[64]</td>
</tr>
<tr>
<td></td>
<td>Increase in FLIP transcription</td>
<td>[64]</td>
</tr>
<tr>
<td></td>
<td>Increase in Bcl2 activity</td>
<td>[65]</td>
</tr>
<tr>
<td></td>
<td>Decrease in FOXO3 leading to a decrease in Bim and Puma transcription</td>
<td>[66]</td>
</tr>
<tr>
<td>Nef</td>
<td>Pro-apoptotic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Up-regulation of FasL</td>
<td>[67]</td>
</tr>
<tr>
<td></td>
<td>Increase in JNK signaling leading to increase in p53 transcription</td>
<td>[68]</td>
</tr>
<tr>
<td></td>
<td>Decrease in Bcl-2 and Bcl-xL activity</td>
<td>[69]</td>
</tr>
<tr>
<td></td>
<td>Binding to CXCR4</td>
<td>[70]</td>
</tr>
<tr>
<td>Anti-apoptotic</td>
<td>Inhibition of caspase 3 and 8</td>
<td>[71]</td>
</tr>
<tr>
<td></td>
<td>Inhibition of Ask1</td>
<td>[72]</td>
</tr>
<tr>
<td></td>
<td>Phosphorylation of BAD</td>
<td>[73,74]</td>
</tr>
<tr>
<td></td>
<td>Up-regulate MAPK and JNK</td>
<td>[75]</td>
</tr>
<tr>
<td></td>
<td>Bind p53 and prevent p53 mediated apoptosis</td>
<td>[76]</td>
</tr>
<tr>
<td></td>
<td>Down modulate the expression of molecules of the MHC class I</td>
<td>[77]</td>
</tr>
<tr>
<td></td>
<td>PAK activation</td>
<td>[78]</td>
</tr>
<tr>
<td></td>
<td>Inhibition of caspases 9 via increased nuclear export of TRNA via eEF1A and Exp-t</td>
<td>[79]</td>
</tr>
<tr>
<td>Vpr</td>
<td>Pro-apoptotic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>ANT mitochondrial membrane permeability</td>
<td>[80]</td>
</tr>
<tr>
<td></td>
<td>Bax activation</td>
<td>[81]</td>
</tr>
<tr>
<td>Anti-apoptotic</td>
<td>Survivin</td>
<td>[82]</td>
</tr>
<tr>
<td>Vpu</td>
<td>Pro-apoptotic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Decrease in NF-κβ signaling</td>
<td>[83]</td>
</tr>
<tr>
<td></td>
<td>Increase in p53 protein levels</td>
<td>[84]</td>
</tr>
<tr>
<td></td>
<td>Increases the sensitivity of cells to Fas associated apoptosis</td>
<td>[85]</td>
</tr>
<tr>
<td></td>
<td>Increase in JNK signaling</td>
<td>[86]</td>
</tr>
<tr>
<td>Anti-apoptotic</td>
<td>Unknown role but cell type dependent decrease has been observed</td>
<td>[87]</td>
</tr>
</tbody>
</table>
Table 1. Cont.

<table>
<thead>
<tr>
<th>Pro/anti Apoptotic</th>
<th>Mechanism</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Env</td>
<td>Pro-apoptotic</td>
<td>Up-regulation of Fas and Fas/L, and an increase in Fas mediated apoptosis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Decrease in the transcription of the FLICE-like inhibitory protein (FLIP)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Up-regulating Bax. Intrinsic apoptosis pathway</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Activation of the p38 but not AKT or ERK</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Induces membrane expression of TNF-α</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Hemifusion cell killing associated with caspase 3 and high ROS</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Syncitia formation leading fused cells to undergo apoptosis through the intrinsic pathway, involving the activation of Cdk1/cyclinB, Nk-κβ, mTOR, MAPK, p53 and PUMA</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Molecular mimicry of Fas</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Up-regulation of caspase 1</td>
</tr>
<tr>
<td></td>
<td></td>
<td>gp-160-CD4 complex blocks nuclear pores</td>
</tr>
<tr>
<td></td>
<td></td>
<td>contagious apoptosis through caspase activation and alterations in mitochondrial trans-membrane potential</td>
</tr>
<tr>
<td>Anti-apoptotic</td>
<td></td>
<td>High levels of CD4 expression lead to the retention of gp160-CD4 complexes within the Endoplasmic reticulum.</td>
</tr>
</tbody>
</table>

A list of HIV proteins and their pro and anti-apoptotic functions.

2.3.2. Nef

The Nef accessory protein was originally thought to be a negative regulator of HIV replication, hence the name Negative factor (Nef). One of the primary functions of Nef and another accessory protein Vpu (discussed below) is the down regulation of CD4. Despite the fact that the CD4 receptor plays a critical role during viral entry, as continued expression of CD4 inhibits viral replication [103].

Nef is found as a soluble as well as a membrane bound protein on the surface of HIV infected cells, and facilitates its own secretion from infected cells by increasing the formation of exosomes [103]. Both versions are able to bind to CD4+ T lymphocytes and upon cross-linking by anti-Nef antibodies, the CD4+ T cells undergo apoptosis [104–106]. When Nef is added extra-cellularly it is also able to induce apoptosis in a variety of cell types by interacting with the chemokine receptor CXCR4 [70]. It has been reported that this death pathway relies on a protein kinase involved in apoptosis signaling that did not require Fas [69,107] or the activity of caspases [69]. Despite this, the over-expression of Nef in a T lymphocyte cell line led to an increase in the number of Fas/FasL molecules on the cell surface. This led to an increase in apoptosis which could be blocked using Fas/FasL blocking antibodies [67].

The induction of Fas ligand expression by Nef occurs following the direct binding of Nef to the T Cell Receptor Complex (TCR). This results in a TCR-Nef complex which is able to up-regulate Fas/L expression without the requirement for any specific antigen [77] (Figure 2). Transgenic Drosophila that over-express Nef revealed that Nef stimulated JNK dependent apoptosis and down-regulated
the innate immune pathway mediated by Relish and NF-κB [68]. Nef is also able to down-regulate the expression of anti-apoptotic proteins Bcl-2 and Bcl-XL [69].

Nef plays an anti-apoptotic role in HIV infected cells giving time for viral particles to mature (Figures 1 and 2). Nef was able to prevent apoptosis and up-regulate MAPK and JNK activities in the presence of TNF-α stimulation [75]. This characteristic of Nef to increase the resistance of cells to TNF-α induced apoptosis increases the pathogenesis of Mycobacterium tuberculosis infections, as cells infected with M. tuberculosis that would normally undergo apoptosis survive in an HIV positive background. Although evidence suggests that Nef and M. tuberculosis both activate TNF-α production individually, a combination of both leads to a larger decrease in TNF-α production. This implies that they activate TNF-α via different pathways [108]. Nef is also able to protect infected cells from Fas-mediated apoptosis by inhibiting the activities of caspase 3 and caspase 8 [71]. Nef associates with and inhibits apoptosis signal-regulating kinase 1 (ASK1), a serine/threonine kinase [72] and Nef is also able to bind p53 preventing p53 mediated apoptosis [76]. Serine kinase activity was found to be associated with Nef in T cell lines expressing hybrid CD8-Nef, as well as in lymphocytes infected with HIV [109]. This association and downstream effects of Nef signaling could be suppressed using serine/threonine kinase inhibitors [78]. The identity of this serine threonine kinase may be Nef-mediated p21-activated kinase (PAK) [110] or src like tyrosine kinases Hck and Lck, both of which are required for T cell activation [111]. It is thought that Nef associates with and activates PAK by forming a complex containing itself, PAK, phosphoinositide 3-kinase (PI3K), the small Rho-GTPases, Cdc42, Rac1, as well as a GEF providing factor. By activating PAK, Nef is also able to block the intrinsic apoptotic pathway by phosphorylating the pro-apoptotic Bad, leading to its dissociation from anti-apoptotic members of the Bcl2 family, decreasing apoptotic stimuli [73,74,78]. The Nef-Pak 2 interaction also plays a role in T cell development and motility as well as an increase in TCR signaling [78]. Nef is also able to decrease apoptosis by enhancing the nuclear transport of tRNA by translation elongation factor EF1A. The nuclear export of tRNA is performed via the Exportin pathway (Exp-1) through the direct interaction of aminoacylated tRNAs with eEF1A [112]. This elongation factor also plays a role in the regulation of the cytoskeleton by associating with actin and microtubules, and also plays a role in apoptosis. However, its role in apoptosis is conflicting, with some studies suggesting it increases the rate of apoptosis when expressed at higher levels, while other studies show eEF1A prevents cell death [112]. Nef was found to interact with eEF1A and increase its transport from the nucleus in an Exp-t dependent manner. Nef was able to rescue cells treated with the pro-apoptotic agent Brefeldin A (BFA) which acts by disrupting the cytoskeleton and increasing ER and Golgi apparatus stress. This is achieved by Nef increasing the nuclear export of NEF/eEF1A/tRNA complexes from the nucleus. The tRNA is then able to prevent mitochondrial cytochrome c release by binding to cytochrome c, inhibiting caspase-9 activation [79].

2.3.3. Tat

The Trans-Activator of Transcription protein (Tat) is a small highly conserved protein that varies between 86 and 101 amino acids depending on the viral subtype [113]. Fas/FasL has also been shown to be up-regulated by Tat, which is secreted from infected cells, and can act on uninfected cells and enhance susceptibility to CD95-induced apoptosis [114]. The primary function of the HIV-1 Tat
regulatory protein is the elongation of viral transcripts by facilitating the interaction of cellular factors with the long terminal repeat (LTR) viral promoter. Tat defective virus does not replicate efficiently in tissue culture. Cells infected with viruses lacking Tat show defects in elongation despite transcription beginning normally and results in few productive transcripts being generated [16].

Tat can modify the homeostasis of the immune system, by stimulating secretion of interleukin (IL)-1β, IL-4, IL-6, IL-8, IL-10, transforming growth factor (TGF)-β1, tumor necrosis factor (TNF)-α, and TNF-β [115,116]. In addition, Tat also induces the expression of the enzyme cyclooxygenase (COX)-2 and the synthesis of prostaglandin E2 (PGE2) [6]. Expression studies in both primary CD4 T cells and T cell lines highlighted the ability of Tat to induce apoptosis or to increase the sensitivity of the cells to pro-apoptotic signals [57]. Mutation studies also indicated that the ability of Tat to induce an apoptotic response was independent of its trans-activation and FasL functions, while expression studies revealed that Tat was able to induce apoptosis by up-regulating the expression of caspase 8 [57] and decreasing the phosphorylation of the forkhead promoter FOXO3a [59] (Figure 2). This will lead to a decrease in the expression of the anti-apoptotic caspase 8 inhibitor, FLIP [60]. Furthermore, the acetylation of Tat enhances its ability to bind and stimulate microtubule assembly which results in the formation of abnormally stable microtubules [61] leading to Bcl-2 dependent apoptosis [62]. Finally, Tat may be able to induce apoptosis through oxidative damage. Tat is known to increase the generation of reactive oxygen species, and Tat dependent apoptosis is inhibited by antioxidants such as N acetylcysteine (NAC). Tat also has the ability to decrease antioxidant defense systems, such as reducing catalase activity and reducing the ratio of GSH:GSSG [63].

The ability of Tat to inhibit the phosphorylation of Foxo3a may also play an anti-apoptotic role as this would inhibit the transcription of pro-apoptotic genes such as Puma and Bim [66]. Tat is also able to up-regulate the expression of the anti-apoptotic proteins Bcl-2 [65] and FLIP while down regulating the expression of caspase 10 [64] (Figure 2).

These conflicting roles of Tat in some pathways such as in the case of FOXO3a and FLIP may be due to the fact that the effect that Tat has on apoptosis seems to be reliant on the current levels of the protein. At low levels Tat decreases the sensitivity of cells to Fas, TNFα and TRAIL signaling [58]. At high levels Tat leads to an increase in the expression of FasL [117], caspase 8, Bax [58] and Receptor-binding Cancer Antigen expressed on SiSo cells (RCAS1). This receptor is similar to TRAIL, having a soluble and membrane bound form. Similarly, both forms can initiate cell cycle arrest and apoptosis in cells that express the receptor [118].

2.3.4. Vpu

The Vpu protein is a 77–86 amino acid protein. Vpu enhances the release of mature virus from an infected cell [119], and is also able to down-regulate the expression of CD4 to allow for efficient viral replication and assembly. Phosphorylated Vpu is thought to bind to CD4 and to the SCF- βTrCP E3 ligase complex, resulting in polyubiquitination of CD4 leading to its degradation by the proteasome [120]. This allows Vpu to regulate the expression of NF-κβ by interfering with the ability of the E3 ligase to transfer ubiquitin to phosphorylated Iκβ-α [83]. In a similar manner Vpu is able to interact with the E3 ligase complex and prevent the ubiquitination and degradation of p53. Consequently, this results in higher levels of active p53 and therefore, higher levels of apoptosis [84]. The ability of
Vpu to interact with the SCF- βTrCP E3 ligase complex is conserved in *Drosophila* where Vpu binds to the homologous SLIMB/b-TrCP complex. As with mammals this results in an increase in apoptosis in the fly [86]. Vpu also increases the sensitivity of cells to Fas associated apoptosis [85], counteracts Tetherin, the host restriction factor active against HIV, through the ubiquitination of Tetherin in a manner similar to CD4 degradation [121].

Vpu has also been indicated in the E3 ligase independent up-regulation of apoptosis. Deletion of Vpu from HIV resulted in a decrease in the sensitivity of infected cells to Fas-induced death [85]. In the *Drosophila* model the expression of Vpu resulted in SCF- βTrCP dependent and SCF- βTrCP independent apoptosis. However, all forms of apoptosis relied on the activation of the JNK pathway by activating the upstream JNKKks, DTAK1 and SLPR. This resulted in the expression of Reaper, resulting in the inactivation of *Drosophila* Inhibitor of Apoptosis1 [86].

Like other HIV proteins Vpu protein has also been shown to play a protective role against apoptosis. Two HIV isolates that demonstrated an increase in the levels of apoptosis they caused in uninfected peripheral blood mononuclear cells (PBMC), contained mutations in the *Vpu* gene. At the same time the *vpu* mutants induced a decreased apoptotic response in an infected CD4-T cell line. This demonstrated that the apoptotic response of cells to *vpu* mutants depended largely on the type of cell used [87].

2.3.5. Vpr

HIV-1 Viral protein regulatory is a small 96 amino acid protein, the truncation of which results in a decrease in the replication rate of the virus. Vpr is responsible for the trans-activation of the HIV-1 LTR as well as for the import of pre-integration complexes. It is also responsible for the initiation of cell cycle arrest in the G2 phase [122], through ATR via the activation of γ-H2AX and BRCA1, and not through ATM or p53 [123]. Vpr also activates Wee the negative regulator of Cdk1 resulting in cell cycle arrest [124]. Vpr was also observed to associate directly with chromatin and may function by interacting with chromatin or with the replication machinery [122]. Alternatively Vpr could be interacting with the proteosome pathway as inhibition of this pathway inhibits the Vpr induced G2 cell cycle arrest [125].

Vpr is able to induce apoptosis through caspase 8 and 9 by causing sustained ERK activation [126] which is able to activate these caspases in a pathway that is FADD and Fas independent [127]. However, most Vpr mediated apoptosis occurs through mitochondrial injury, resulting in the release of cytochrome c and the activation of caspase 9. This occurs without the activity of caspase 8, the presence of Fas/L [128] or active p53 [129]. As inhibition of ATR in cells expressing Vpr resulted in the inhibition of cell cycle arrest and apoptosis, it was concluded that there was a relationship between the ability of Vpr to induce cell cycle arrest and its ability to induce apoptosis [130]. Some studies have concluded that the mitochondrial membrane permeabilisation that occurs during Vpr mediated apoptosis is due to the direct formation of large conductive pores due to the binding of Vpr to the Adenine Nuclear Transporter (ANT) [131]. It has been reported that Vpr acts directly on the mitochondria and its pro-apoptotic functions are not inhibited by the knockout of caspase activators Therefore, the ability of Vpr to induce apoptosis does not rely on caspase activation during the apoptotic response [132]. However, other studies used knockdown of ANT to show that the presence or absence
of ANT did not affect Vpr mediated apoptosis. Rather Vpr induced apoptosis requires the activity of Bax. Vpr activates Bax through the upstream signaling pathway consisting of ATR and GADD45a, as the RNAi mediated silencing of either of these genes prevented Vpr induced apoptosis [133]. Vpr is also able to initiate mitochondrial permeabilisation by inducing the cleavage of BID to tBID [126].

Vpr can also function as an anti-apoptotic protein by activating the transcription of the inhibitor of apoptosis (IAP) Survivin. Survivin inhibits caspase activity and stops apoptotic signaling by Bax, TNF-α and Fas. The expression of Survivin is cell cycle dependent, with the highest expression occurring in the G2 phase. The establishment of a G2 cell cycle arrest by Vpr is the most likely mechanism behind the increase in Survivin expression [134]. It has also been reported that low level expression of Vpr protects cells from apoptotic stimuli. At these low levels there is no G2 cell cycle arrest. Therefore, like Vpu, Vpr plays different roles in apoptosis depending on its expression levels. With high levels leading to apoptosis and lower levels leading to an anti-apoptotic signal [135].

Vpr can reduce neuronal death by reducing the expression of pro-inflammatory cytokines such as IL-1β, IL-8 and TNF-α [136]. A large 1507 amino acid protein interacts with Vpr and was given the name Vpr Binding protein (VprBP). This protein is responsible for the regulation of p53 by binding to p53 promoter elements. This interaction between Vpr and VprBP allows Vpr to down-regulate p53 and therefore inhibit apoptosis [137].

2.3.6. Env

The HIV Envelope protein is initially synthesized as a large precursor molecule known as gp160. This protein is then cleaved to give rise to the secreted gp120 and the trans-membrane gp40, Gp120 remains loosely associated with gp40 at the cell envelope [138]. Env contains a long signaling peptide that is located upstream of the region that forms gp120 following proteolytic cleavage. Replacing or removing this signal peptide resulted in decreased levels of apoptosis. This increase in apoptosis induced by the signaling peptide of Env was found to depend on the up-regulation of caspase 1 [99]. T cell apoptosis mediated by Env is due to impaired cytokine production, with Th1 cytokines such as IFN-γ, IL-2 and IL-12 blocking apoptosis. The full length Env precursor gp-160 may associate with CD4 to form a complex that is capable of blocking the nuclear pores [100]. This leads to calcium ion build up within the nucleus and activates calcium dependent endonucleases, which cleave DNA resulting in apoptosis [138]. However, other studies indicate that this binding inhibits apoptosis as, high levels of CD4 expression lead to the retention of gp160-CD4 complexes within the Endoplasmic reticulum. At lower levels of CD4 expression the complex is transported to the Golgi apparatus where the gp160 is cleaved into gp120. The gp120-CD4 complex is then able to create pores in the membrane of the Golgi apparatus, migrate to the cell membrane and then to act in a similar way on the plasma membrane resulting in necrosis [102] (Figure 3).
**Figure 3.** Mechanisms of apoptosis induction by the HIV-1 envelope protein at the cell surface. This can result from a hemifusion event through the transient interaction between Env- and CD4/CXCR4. This hemifusion event results in the exchange of membrane lipids and proteins. Alternately, Env-expressing cells can fuse with Env-negative cells resulting in the formation of a syncytium and death for the new fused cell. Here cell death relies on the p53 pathway via p38 and Cdk1 signaling.

The binding of gp120 to CD4 and CCR5/CXCR4 results in the transmission of death signals through these receptors leading to apoptosis. Alternatively, apoptosis may be achieved through membrane fusions. This may be either, a hemifusion event, or the formation of a syncytium [20]. Cross-linking of CD4 T cells with the envelope protein gp120 prior to T cell receptor (TCR) stimulation results in the up-regulation of Fas and Fas/L, activation the Fas/FasL (CD95/CD178) pathway and an increase in Fas mediated apoptosis [88]. Infected T cells co-express CD178 thus becoming potential killers of uninfected CD95-expressing T cells [67]. The binding of gp120 to CD4 separately from the TCR also results in a decrease in the transcription of the FLICE-like inhibitory protein (FLIP),
which inhibits the proteolytic activation of caspase 8. In addition to this Gp120 and CD4 binding also activates intrinsic apoptotic pathways by up-regulating the pro-apoptotic Bax independently of Bcl-2. This then results in an increase in mitochondria-dependent apoptosis [89]. The activation of the p38 signaling pathway but not AKT or ERK is another reported outcome for the binding of gp120 to either CD4, or CXCR4. In the absence of AKT or ERK activation, p38 signaling leads to cell death, while the presence of AKT or ERK leads to cell survival [90]. Gp120 from HIV-1 and Fas, share a seven amino acid motif VEINCTR. This sequence is proximal to the immunogenic V3 loop of gp120 and antibodies found in the sera from HIV positive patients react with peptides containing this motif [97]. Due to molecular mimicry antibodies against gp120 bind Fas activating the Fas pathway and leading to apoptosis [98].

The association of Gp120 with CXCR4, results in a Fas independent death signal that leads to the depolarization of the mitochondrial cell membrane, cytochrome c release and the activation of caspase 9 [9]. Bystander T-cell killing can be mediated by macrophages since ligation of the chemokine receptor CXCR4 by gp120 or its ligand on macrophages induces membrane expression of TNF-α which triggers apoptosis on CD8+ T cells that express the receptor TNF-RII [91].

The association of Env with the CXCR4 receptor can result in cell-cell fusion between infected and uninfected cells [20] (Figure 3). Mutation studies revealed that the fusogenic activity mediated by gp41 is a requirement for the depletion of uninfected bystander T cells [139]. In order for this to occur, gp41 must mediate a close cell to cell contact. This then results in the death of the single uninfected cell through gp41 mediated transfer of lipids from the infected Env expressing cell to the uninfected cell [92]. This hemifusion cell killing can be inhibited by the over-expression of Bcl-2 [93] (Figure 3) and seems to be dependent on caspase 3 but not caspase 8.

Hemifusion apoptotic pathways also do not require p38 or p53 signaling and result in the generation of high levels of reactive oxygen species [94]. Membrane bound gp40 with associated gp120 binding to CXCR4 more commonly leads to the formation of short lived syncitia. These fusions between infected and uninfected cells lead to the cell undergoing apoptosis through the intrinsic pathway. It is unknown whether caspases are required for this apoptotic process with different studies reporting conflicting results [20,140]. The apoptotic cascade following cell fusion is initiated by the activation of Cdk1/cyclinB and NF-κβ. This is followed by an aborted entry into the prophase of mitosis where the nuclei of the cells fuse. At this stage p53 is phosphorylated by mTOR (Mammalian Target of Rapamycin) and cytoplasmic p38 mitogen-activated protein kinase (MAPK). Activated p53 then activates the transcription of Bax and Puma, activating mitochondrial apoptosis (95,96) (Figure 3). When apoptosis is initiated in cells expressing Env they can transfer this apoptotic signal to surrounding healthy cells. Different methods of inducing apoptosis in the donor cell still result in apoptosis in the target cell. In order for this contagious apoptosis to occur the donor cells have to be undergoing pre-apoptotic chromatin condensation. Target cells undergo apoptosis through caspase activation and alterations in mitochondrial trans-membrane potential [101].

3. HIV Related Apoptotic Pathways in Immune Related Cells

HIV-1 infects a variety of human cells with immune cells being the most commonly infected. These include CD4+ T cells, dendritic cells and macrophages. The first cells that HIV-1 encounters are
dendritic cells, the potent antigen presenting cells that play a role in rapid infection of the T cells. Macrophages are known to be the major reservoir of persistent replication competent HIV-1 for extended periods, while Dendritic cells (DC) play a pivotal role in HIV-1 pathogenesis [141].

Dendritic cells (DC) are clustered in groups according to their location and are either blood DCs or are found in and around the skin and mucosal membranes. Blood DCs are subdivided into Myeloid DCs (myDCs) and plasmacytoid DCs (pcDCs) [142]. A normal immature DC cell serves to capture an antigen and migrate to the T cell areas of secondary lymphoid organs where it matures [143]. Blood DCs numbers are depleted upon infection which reduces IFN-alpha, resulting in a high viral load [142].

3.1. CD4⁺ T Cells

HIV infection is associated with a steady decline in the numbers of CD4⁺ T lymphocytes [144]. Under normal circumstances the pool of CD4⁺ cells is maintained through thymopoeisis and apoptosis [58]. Following viral infection CD4⁺ T cell numbers are depleted through increased apoptosis via multiple viral induced pathways. This increase in cell death is observed in both infected and uninfected cells. The levels of the bystander un-infected T cells that are lost due to apoptosis exceeds that of the infected T cells [58,144]. Additionally, both infectious and non-infectious HIV particles are able to induce selective apoptosis of CD4 T cells, suggesting that noninfectious HIV particles, which make up a large portion of plasma virus, contribute to the decline in CD4 counts in patients [22,23].

The primary and secondary cellular receptors CD4 and CCR5/CXCR4 are recognized by the viral envelope proteins. These envelope proteins then insert into the lipid membrane allowing the entry of the virus into the cell [20]. HIV-1 infection of CD4 T cells is favored by cell-to-cell contact, through the formation of the virological synapse [21]. Once the virus has infected the cell apoptosis can be initiated in a variety of means. These include the action of HIV protease which cleaves Bcl-2 and procaspase-8, with cytotoxic affects produced by other HIV proteins [145]. The cleavage of procaspase 8 performed by HIV-1 protease gives rise to a unique 41 kDa protein fragment known as Casp8p41. This fragment is observed to co-localise with infected and apoptotic cells. The levels of Casp8p41 correlates with CD4 cell count, with an increase in the level of Casp8p41 being associated with a decrease in the level of CD4 cells [12,146]. The generation of this HIV specific cleavage product seems to be necessary for the initiation of cell death in infected cells [12] as well as bystander T cells [147], and initiates cell death through the intrinsic apoptotic pathway involving the downstream activation of caspase 9 through the Bax/Bak facilitated release of cytochrome c [148].

The killing of uninfected bystander T cells occurs through the release of viral proteins gp120, Tat and Nef from infected. Gp120 triggers death in uninfected T cells by binding the receptors CD4, CXCR4 and CCR5 which leads to Fas up-regulation and a decrease in FLIP. This in turn leads to cell death through the intrinsic apoptotic pathway [145]. Tat enters the bystander cells and up-regulates TRAIL. TRAIL is induced by TLR on the surface of pcDCs which were shown to be able to kill CD4⁺ T cells. pcDCs presenting TRAIL were found in sites of high CD4⁺ T cell depletion [149]. HIV gp120 released from HIV infected DCs was implicated in T cell dysfunction [145,150]. Finally, Nef appears to kill uninfected T cells through the extrinsic and intrinsic pathways [145].
However, the main trigger for the death of these cells appears to be HIV-1 DNA products that arise due to the activity of the sterile alpha motif (SAM) domain and histidine—aspartate (HD) domain—containing protein 1 (SAMHD1). This enzyme acts to deplete dNTP stores and thus prevents complete DNA synthesis [151]. The presence of these viral particles is sensed by the interferon-γ—inducible protein 16 (IFI16) [152]. Unfortunately, although this enzyme protects these bystander cells from infection the resulting viral cDNA particles initiate apoptosis within these cells through the activation of caspases 3 and caspases 1. While caspase 3 initiates apoptosis through classical pathways, caspases 1 is involved in the innate immune response. Here the inflammasome initiates apoptosis through a process known as pyroptosis [153]. The levels of the detector IFI16 directly correlate with viral load as well as with the levels of cell death [152].

At the same time the failure of the virus to properly integrate into the host can also lead to apoptosis. The integration of viral cDNA requires DNA double strand break repair enzymes to repair single strand gaps between viral DNA and the target DNA as well as to circularise unintegrated viral cDNA. DNA damage is sensed by the molecular sensor ataxia-telangiectasia-mutated (ATM) which in turn activates DNA-dependent protein kinase (DNA-PK). The lack of either ATM or DNA-PK leads to errors in the integration of viral cDNA into host DNA [154,155]. The inhibition of the activity of DNA-PK leads to a decrease in the levels of HIV induced apoptosis. This implies that inhibition of DNA-PK by the virus would lead to a population of latently infected cells which do not produce new viral particles but still have the ability to do so [155]. This also implicates DNA-PK as a useful drug target [155].

3.2. The Contribution of HIV Proteins to Apoptosis in Macrophages

Macrophages are part of the cells of the innate immune system and along with CD4$^+$ cells are the principal targets of HIV infection [74,145,156]. Viral entry seems to be modulated through CCR5, CD4 and CXCR4 surface proteins which also affects strain susceptibility [156]. In contrast to macrophages, peripheral blood monocytes are less frequently infected with HIV-1 in vivo [157]. Macrophages do not undergo cytotoxic events after infection and thus survive [158]. This is followed by the formation of vacuoles harboring viral particles [158]. These vacuoles known as virus containing compartments (VCCs) are present in uninfected macrophages but are more prominent following infection [145]. These surviving cells harboring viral particles act as reservoirs evading treatments such as HAART [74]. The ability of Macrophages to infect other cell types has a great effect on HIV pathogenesis. These cells include brain cells and T cells [159]. Direct transmission of virions to other cell types is through a virological synapse. This synapse is formed between an infected macrophage and uninfected cells expressing a receptor [159].

HIV encoded proteins play an important role in regulating apoptosis in macrophages. For instance, Nef elevated anti-apoptotic effects in the human macrophage precursor cell line. This enhances survival. Monocyte derived macrophages (MDMs) are involved in formation of HIV reservoirs affecting AIDS pathogenesis [74]. Tat can also enhance infectivity of HIV by recruiting monocytes, macrophages and dendritic cells. The expression of HIV co-receptors is also induced by the presence of Tat [145]. Vpu functions to degrade CD4 in the endoplasmic reticulum and counteracts host restriction, such as tetherin, to enable viral release. Degradation of CD4 follows its ubiquitination.
A similar fate is dealt to tetherin. Vpu mutation hinders infection of macrophages [121]. Vpr is an absolute requirement for viral replication in macrophages through its ability to increase the activity of various transcription factors and to stimulate HIV replication [145].

4. HIV-Associated Pathologies: The Role of Apoptosis

Over 33 million people worldwide suffer from HIV/AIDS [160]. Highly active antiretroviral therapy (HAART) has extended the life expectancy of HIV-1 patients and has significantly reduced the viral burden [161]. Despite treatment serious HIV-1 related complications and pathologies have been shown to develop over time. These pathologies may include HIV-1 associated neurocognitive disorders (HAND), HIV-associated nephropathy (HIVAN), HIV-associated vascular complications and HIV-associated tumors such as Kaposi sarcoma [136] (Table 2). Although productive HIV-1 infection of primary neurons has not been demonstrated, HIV is able to affect neurons through indirect mechanisms. HIVAN is caused by infection of renal epithelial cells, where the kidney serves as a reservoir for HIV-1 [161,162]. It has been documented that 1% of HIV patients suffer from HIV-associated vascular complications [163]. Cardiovascular diseases progression is associated with the increased production of reactive oxygen species (ROS), such as superoxide. This leads to aberrant cell signaling, vascular smooth muscle cell hypertrophy and migration [164], endothelial dysfunction, and, potentially, apoptosis [165]. In people with HIV, cancer is a significant cause of mortality and morbidity, with 30%–40% of HIV patients developing a malignancy during their life time. Most of these HIV-related cancers (“AIDS-associated malignancies” or “opportunistic” cancer) are established as AIDS-defining and include Kaposi’s sarcoma (KS), non-Hodgkin’s lymphoma (NHL), and invasive cervical cancer (ICC) to name but a few examples.

Table 2. HIV related pathologies.

<table>
<thead>
<tr>
<th>HIV-Associated Neurocognitive Disorders (HAND)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Prevalence</strong></td>
</tr>
<tr>
<td>40%–50% of HIV-1 positive patients.</td>
</tr>
<tr>
<td><strong>Symptoms</strong></td>
</tr>
<tr>
<td>Impaired cognitive activity, memory, learning, attention, problem solving, decision making, confusion, forgetfulness, behavioral changes, and nerve pain.</td>
</tr>
<tr>
<td><strong>Histological patterns</strong></td>
</tr>
<tr>
<td>Macrophage infiltration, activated microglia, reduced synaptic/dendritic density and selective neuronal loss.</td>
</tr>
<tr>
<td><strong>Underlying causes</strong></td>
</tr>
<tr>
<td>• Neuro-inflammation characterized by pro-inflammatory events</td>
</tr>
<tr>
<td>• Release of pro-inflammatory cytokines such as IL-1β, -6, TNF-α, and chemokines</td>
</tr>
<tr>
<td>• Higher levels of TNF-α, IL-1β, IL-6, IL-8, monocyte chemo attractant protein-1, macrophage inflammatory protein-1 and CXCL10 are observed in vivo and in vitro.</td>
</tr>
<tr>
<td>• Levels of these neuro-inflammatory factors are associated with higher viral load in cerebrospinal fluid (CSF).</td>
</tr>
</tbody>
</table>
Table 2. Cont.

### HIV-Associated Neurocognitive Disorders (HAND)

**Role of HIV proteins**
- HIV-1 gene products are also known to modulate the levels of cytokines in macrophages.
- Tat stimulates cytokine/chemokine networks in monocytes and macrophages. Tat is also implicated in apoptosis using an excitotoxic mechanism to cause neurotoxicity. This excitotoxic mechanism involves the use of indirect and direct oxidative stress coupled with increased intra-cellular calcium and caspase3 activation. [166,175]
- Further apoptosis is due to the mitochondrial release of cytochrome c and microtubule damage. This mechanism is similar to that induced by protease in lymphocytes with apoptosis as the end result [14,166,176].
- The surface of glial cells and neurons display CCR5 and CXCR4 which are targeted by gp120. This attachment, as in CD4 cells, causes apoptosis. An increase in gp120 concentration was shown to cause an increase in programmed cell death. Gp120 also inhibits the size and quantity of neurite growth and is known to activate caspases 3 [166,167].
- Vpr is the main viral protein responsible for neuropathology through pro-inflammatory cytokines. [177]
- The same proteins use similar mechanisms to cause apoptosis in both the nervous system and immune system. HIV-1 invades the central nervous system (CNS) during early infection via infiltrating monocytes and lymphocytes that are infected in the periphery [178].

### HIV-Associated Nephropathy (HIVAN)

**Prevalence**
Many patients with HIVAN ultimately progress to end stage renal disease (ESRD). 90% of all ESRD cases attributed to HIVAN occur in African Americans [179,180,181,182,183,184,185,186,187,188,189,190,191].

**Symptoms**
Inflammation is the major pathology. [181,182,183,184,185,186,187,188,189,190,191].

**Histological patterns**
- Collapse of the glomerulus, cystic tubular dilatation and ultra-structurally actin cytoskeletal effacement. [183,184,185,186,187,188,189,190,191].
- Histologic and molecular evidence of injury to glomerular podocytes. [183,184,185,186,187,188,189,190,191].
- Normally terminally differentiated podocytes lose podocyte-specific proteins such as podocin and synaptopodin, and undergo proliferation and apoptosis [183,184,185,186,187,188,189,190,191].

**Underlying causes**
- HIVAN is caused by infection of renal epithelial cells, where the kidney serves as a reservoir for HIV-1. Unlike neurons, HIV-1 can directly infect the renal tubular epithelium cells [RTEC] leading to HIVAN [161,162,163,164,165,166,167,168,169,170,171,172,173,174,175,176,177,178,179,180,181,182,183,184,185,186,187,188,189,190,191].
- HIV-1 proteins interfere with signaling pathways that maintain cellular quiescence. The deregulated podocytes cannot re-differentiate into the quiescent state and are gradually depleted [188,189,190,191].
- Remaining podocytes undergo hypertrophy to cover a larger surface area resulting in denuded segments of the basement membrane that promote the development of sclerotic lesions [185,186,187,188,189,190,191].

**Role of HIV proteins**
- Vpr induces ERK, caspases-8 dependent apoptosis and hyperploidy in RTECs [190,191,192].
- Nef protein activates ERK in podocytes. [190,191,192].
- The ubiquitin-like protein FAT10 is up-regulated by HIV infection [190,191,192].
Table 2. Cont.

**HIV-Associated Vascular Complications**

<table>
<thead>
<tr>
<th>Prevalence</th>
<th>1% of HIV patients.</th>
</tr>
</thead>
</table>

**Symptoms**
Coronary heart disease, pulmonary hypertension (PH), and atherosclerosis
Increase the risk for noninfectious pulmonary conditions, including
- chronic obstructive pulmonary disease
- lung cancer
- pulmonary hypertension (PH).

**Underlying causes**
Increased production of reactive oxygen species (ROS), such as superoxide, leading to
- aberrant cell signaling
- vascular smooth muscle cell hypertrophy and migration
- Endothelial dysfunction, and potentially apoptosis.

**Role of HIV proteins**
Documented, but not yet fully understood. Suspected to play a role through promoting apoptosis, growth, and proliferation of a variety of cells in vitro by interacting with molecular partners in the infected host

<table>
<thead>
<tr>
<th>HIV-Associated Tumors, Kaposi Sarcoma (KS)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Prevalence</strong></td>
</tr>
<tr>
<td>For some time, Kaposi sarcoma was seen in 30%–40% of patients with AIDS, often as the presenting sign. The incidence of Kaposi sarcoma has fallen markedly in recent times, although its prevalence has not.</td>
</tr>
<tr>
<td><strong>Symptoms</strong></td>
</tr>
<tr>
<td>Raised red, purple, brown, or black blotches found on the skin, mouth gastrointestinal tract and respiratory tract.</td>
</tr>
</tbody>
</table>

**Histological patterns**
Initially inflammatory dermatosis with signs of vasoformation. Later abnormal elongated spindle cells are present and are arranged in haphazard clusters. Dense vascularization with Hyaline globules

**Underlying causes**
- Up regulation of Bcl-2 expression associated with reduced endothelial cell apoptosis.
- Bcl-2 favors the angiogenic process which is switched off in healthy tissues
- Decreased expression of anti-apoptotic molecules occurs through the inhibition of endothelial cell adhesion onto the ECM or decreased expression of antigenic growth factors.
- The increase in Bcl-2 levels in late-stage KS lesions is accompanied with an increase in vascular cell apoptosis.

**Role of HIV proteins**
The de-regulation of apoptosis by HIV proteins has been shown to play a significant role in tumor development. Tat effects are cell-type dependent selectively promoting apoptosis in various cell systems. Tat increases Bcl-2 expression

This table lists the major HIV related pathologies that are linked to the ability of the virus to alter the patterns of apoptosis. It shows details of the prevalence of these pathologies, their symptoms, histology, underlying causes and the role played by the viral encoded protein products in contributing to the disorder.
5. Therapeutic Targets: Targeting Apoptotic Pathways

Various modulators of apoptosis have been identified over the years and strategies that target these regulators have been approved and applied in clinical trials. Both death receptors and classic apoptosis markers, such as Bcl-2 proteins, caspases and Bax, have been used as apoptotic therapeutic targets [215,216]. Caspases have been shown as early as 1999 to be effective in targeting HIV-infected cells [216,217]. Recently a caspase 3 (CASP3) has been engineered that is only activated by the HIV-1 protease. This protease was able to limit HIV replication in tissue culture by inducing apoptosis, implying that this mutant synthetic form of CASP3 could be used to treat resistant strains of HIV-1. Due to the mechanisms involved it was postulated that the possibilities of the HI virus developing resistance to CASP3 is unlikely [218].

The mTORC1 pathway was shown to be a therapeutic target for preventing the progression of HIVAN [219]. As previously stated HIV related renal failure is a result of tubular microcysts in cells leading to apoptosis, HIV-1 promotes renal tubular epithelial cell protein synthesis. The mTORC1 pathway plays an important role in mRNA translation and has been shown to be the cause of many renal diseases, and this pathway is activated in tubular cells upon HIV-1 infection [219]. Rehman et al. proceeded to inhibiting the pathway using rapamycin which resulted in diminished protein synthesis and thus no hypertrophy of the tubular cells [219]. Rapamycin is also commonly used to suppress the immune system as the mammalian target of rapamycin (mTOR) controls the activation and proliferation of T cells. Rapamycin can therefore reduce the number of T cells available for infection [220].

5.1. Therapies and Their Shortcomings

The ability of HIV to influence apoptotic pathways has allowed the virus to evade the immune system by blocking apoptosis of infected cells, by inducing apoptosis in other immune related cells or by evading the detection of infected cells. Two distinct types of therapies in relation to apoptosis have been approached [221]. Therapies based on the reactivation of latent virus aim to increase apoptosis in infected cells At the same time therapies aimed at blocking the apoptosis of immune related cells prevents the development of immunodeficiency [221,222].

Some potential reactivation therapies aimed at inducing apoptosis in infected cells include PI3K/Akt inhibitors that block HIV replication and favor apoptosis leading to the clearance of apoptotic cells [145]. Another strategy involves the use of IAP inhibitors in infected macrophages. Here HIV-1 infection of macrophages leads to an increased resistance of the macrophages to apoptosis through the expression of macrophage colony stimulating factor (M-CSF) which increases the expression of anti-apoptotic proteins while inhibiting the expression of death receptors. Conversely, the viral protein Vpr is also unable to induce apoptosis in macrophages. This can be reversed through the inhibition of IAP1 and IAP2 [145]. The use of Caspase 1 inhibitors such as VX-765, inhibits cell death due to the initiation of the IFI16 pyroptosis pathway and provides numerous advantages. Firstly they act on host pathways and would therefore avoid pathogen resistance. Secondly they are predicted to be clinically safe and finally, they are effective pyroptosis inhibitors. However, the value of pyroptosis inhibition in halting disease progression still has to be evaluated [152]. Another example
of reactivation therapy involves the experimental use of Histone deacetylase inhibitors (HDACi) such as vorinostat, panobinostat and rhomempsin. These inhibitors lead to increased DNA transcription and result in an increase in the levels of HIV RNA in resting memory CD4+ T cells. This will presumably increase the innate immune response leading to increased apoptosis in the infected cells. However, the predicted increase in apoptosis does not occur without the presence of an effective HIV-specific cytotoxic T-lymphocyte (CTL) response [222]. This is a common observation regarding reactivation therapy, where the inhibition of anti-apoptotic viral signals does not always result in the increase of apoptosis in infected cells and some evidence suggest it may actually increase viral load [221]. One method that could be used to overcome this is to sensitize the cells to apoptotic signals “priming” them for apoptosis [222].

Most conventional therapies, such as antiretroviral and protease inhibitors, generally decrease viral replication and the depletion of immune cells due to increased apoptosis, leading to a decrease in immunodeficiency. However, failures in these therapies due to the development of mutant viral strains, does not always lead to an increase in apoptosis [221]. This is observed in the use of enfuvirtide to inhibit gp41 fusion activity. Mutations develop that interfere with the ability of the drug to bind to gp41. These mutants also demonstrate a reduced ability to induce apoptosis in bystander cells due to reduced fusogenic activity [223]. Antiviral treatment also leads to HIV-1 protease mutations which inhibit the ability of the enzyme to cleave procaspase 8 into the pro-apoptotic Casp8p41. This results in a decrease in apoptosis [221]. Memory CD4 T cells also down-regulate procaspase 8 and demonstrate an intrinsic resistance to apoptosis [222]. Impaired pro-apoptotic ability as an evolutionary response of the virus to antiviral therapy may allow for increased viral replication in infected cells to compensate for decreased replication fitness [221].

5.2. Drugs Targeting HIV Proteins

The ability of HIV to cause pathologies as mentioned above led to many studies on elucidating the role that HIV proteins play in regulating apoptosis through molecular mechanisms, and signaling pathways. The practical application of all the information gathered is the development of a new generation of drugs that are more effective and have fewer side effects on patients. The different drugs that have been developed or are in the process of development include drugs targeting the effects of gp120, Tat-protein and the action of HIV protease (Table 3).

One strategy includes revisiting plant drug sources which are currently under-utilized with 36 plant families containing 46 plant species that have documented anti-HIV activity [224]. Many of these plant derived drugs were active against HIV protease, viral replication and syncitia formation by acting on gp120 and Tat-protein [224]. Since HIV protease, Tat-protein and gp120 are all implicated in apoptosis, these extracts potentially inhibit apoptosis.
Table 3. Drugs targeting apoptosis in HIV.

<table>
<thead>
<tr>
<th>Drug</th>
<th>Target</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Drugs targeting Tat</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bovine Dialyzable Leukocyte Extract (bDLE)</td>
<td>Down regulation of Tat-protein lowers the expression of anti-apoptotic protein BCL-2 in infected cells</td>
<td>[81]</td>
</tr>
<tr>
<td>PI3K inhibitors and Akt inhibitors</td>
<td>Counter Tat-protein induced protection on infected cells</td>
<td>[82]</td>
</tr>
<tr>
<td>picolinic acid (PA) and fusaric acid (FA)</td>
<td>Target the conserved RING finger on Tat, inhibiting trans-activation</td>
<td>[225,226]</td>
</tr>
<tr>
<td>β-arrestin 2</td>
<td>Reduces apoptosis</td>
<td>[3]</td>
</tr>
<tr>
<td><strong>Drugs targeting HIV protease</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Polyoxometalates</td>
<td>Act against HIV protease</td>
<td>[228]</td>
</tr>
<tr>
<td>Single-chain Fv (scFv)</td>
<td>An artificial derivative of mAb1696</td>
<td></td>
</tr>
<tr>
<td>P27 peptide</td>
<td>Peptide derivative of the C- and N-terminal domains of HIV protease which inhibit dimerisation Uncouples the protease dimer and induces inhibition</td>
<td>[11]</td>
</tr>
<tr>
<td>mAb1696 antibody</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12-aminododecanoic acid (12-Ado)</td>
<td>Template for HIV protease dimerisation inhibition</td>
<td>[230]</td>
</tr>
<tr>
<td>C3-substituted cyclopentyltetrahydrofuranyl GRL-02031</td>
<td>Allosteric inhibitors Bind the flap region of HIV protease</td>
<td>[11,231]</td>
</tr>
<tr>
<td></td>
<td>Another derivative of Cp-THF</td>
<td>[232]</td>
</tr>
<tr>
<td><strong>Drugs targeting gp120</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sifuvirtide</td>
<td>Fusion Inhibitor</td>
<td>[233]</td>
</tr>
<tr>
<td>Enfuvirtide</td>
<td>Fusion inhibitor</td>
<td>[41]</td>
</tr>
<tr>
<td>Maraviroc</td>
<td>CCR5 antagonist</td>
<td>[41]</td>
</tr>
<tr>
<td>4-phenyl-1-4-phenylbutyl piperidine (PPBP)</td>
<td>sigma-1 receptor agonist acting against gp120</td>
<td>[167]</td>
</tr>
<tr>
<td>Selective serotonin reuptake inhibitors (SSRIs)</td>
<td>Reduces cytokine receptor expression in the nervous system, reducing gp120 binding targets</td>
<td>[234]</td>
</tr>
<tr>
<td><strong>Other drugs</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Double stranded RNA Activated Caspase Oligomerizer (DRACO), NAPVSIPQ (NAP)</td>
<td>Selects for viral infected cells only based on the length of RNA transcription helices. Increases apoptosis by caspase activation protects against mitochondrial release of cytochrome c.</td>
<td>[235]</td>
</tr>
<tr>
<td></td>
<td></td>
<td>[176]</td>
</tr>
</tbody>
</table>

This table lists some of the drugs currently under development that target the ability of HIV encoded proteins to manipulate the apoptotic machinery of the host cells.
5.2.1. Drugs Targeting Tat

The immune system of HIV-1 patients was partially restored using bovine Dialyzable Leukocyte Extract (bDLE). This was even effective in individuals that were in the advanced stages of the disease. bDLE acts by preventing all stages of viral infection by excluding the HIV envelope on infected CD4 cells. This seems to be accomplished through the down regulation of Tat expression, resulting in lower levels of BCL-2 in infected cells [81]. PI3K inhibitors and Akt inhibitors are another class of drugs that target Tat by interfering with the ability of Tat to protect infected cells from apoptosis. The associated increase in apoptosis is not observed in uninfected cells [82]. Picolinic acid (PA) and Fusaric acid (FA) are two transition metals that have anti-HIV activity due to their ion chelating properties. This inhibits Tat trans-activation, most likely by targeting the conserved zinc finger of Tat [225,226], as chelation of the zinc finger results in a denatured, non-functional protein [226].

Investigations into the relationship between Tat protein and SIRT1, which plays a role in the activation of the p53 pathway leading to apoptosis in T-cells, [236–242] showed that Tat is able to inactivate SIRT1, thereby, activating the p53 pathway resulting in T-cell depletion [80,236]. This implies that SIRT1 could be used as a potential therapeutic target for inhibiting apoptosis [236].

5.2.2. Drugs Targeting gp120

The inhibition or blocking of gp120 results in a decrease in syncitia formation or hemifusion events and therefore a decrease in gp120 dependent apoptosis. The entry of HIV into cells can be prevented by blocking gp120. This can be achieved through the over-expression of β-arrestin 2 which reduces apoptosis by down regulating µOR which in turn is up-regulated by gp120 [3]. Two drugs target gp120 in the nervous system 4-phenyl-1-4-phenylbutyl piperidine (PPBP) and Tianeptine. PPBP is a sigma-1 receptor agonist that partially inhibits gp120, significantly decreasing the apoptotic effects of the protein [167]. Tianeptine is a Selective serotonin reuptake inhibitor (SSRIs) that reduced cytokine receptor expression in the nervous system. With less receptors for gp120 to bind, caspase 3 activation is decreased resulting in reduced apoptosis [234]. The entry inhibitors Enfuvirtide (a gp41 derivative) and Maravirocis (a CCR5 antagonist) both have limitations. The effects of Enfuviridte in primary macrophages are not fully understood, while resistance to maraviroc has been observed. At the same time, other CCR5 inhibitors show serious side effects and a lack of clinical efficacy [145].

5.2.3. Drugs Targeting HIV Protease

As previously stated the HIV protease initiates apoptosis by physically damaging the host cell’s structural proteins. HIV protease is also involved in viral maturation and assembly. However, so far attempts to accurately establish where this protease cleaves its substrates have been unsuccessful, complicating drug design. Inhibiting HIV protease has been effective in preventing viral maturation and damage to host proteins, resulting in a stall in apoptosis. This was shown using an HIV protease inhibitor saquinavir [243]. Current Protease inhibitors are competitive inhibitors using a structure based design. This increases the risk of resistance emergence, as they target the same active site. If resistance emerges, the danger is that this resistance may be to multiple protease inhibitors at once. One solution is the use of protease inhibitors that target other parts of the protease, namely: allosteric
inhibitors. However, screening using structure-based design strategies has proved tedious due to high levels of toxicity at the cellular level. High throughput screening methods for allosteric inhibitors are required. The solution is the use of a cellular based system, which will allow for the screening of multiple attributes of a drug at the same time, including potency and toxicity. Potential compounds that are candidate allosteric inhibitors of HIV protease include; Polyoxometalates, C3-substituted cyclopentyltetrahydrofuranyl (Cp-THF) and dimerization inhibitors [11].

Polyoxometalates (POM) are oxide compounds from transition metals with a preference for tungsten, molybdenum and vanadium [244]. Early polyoxometalates were tested on HIV but had devastating side effects which resulted in the termination of drug testing [245]. However, less toxic POMs are being sought after. The mechanisms as to how they work are yet to be elucidated [246]. They were first tested for effect on reverse transcriptase which was negative [247]. Later they were shown to act against HIV protease [228], as the activity of POMs is easily interrupted by DMSO (Dimethyl Sulfoxide) which has previously been reported to stabilize HIV protease [246]. At least one POM has been reported to bind the hinge region of HIV protease. This kind of binding does not target the active site making POMs an allosteric inhibitor, which should prevent the development of resistance. Synthesis of POMs is also less laborious and more cost effective relative to organic inhibitors.

A specific class of POMs, Keggin and Dawson POMs are characterized by an organic side chain, and of the total of 28 POMs selected as potential drugs due to their known activity against HIV protease, six Dawson POMs and a single Keggin POM displayed total deactivation of HIV protease, showing that Dawson POMs are more efficient. A butyl side-chain trailed by a propionic acid side-chain enhanced activity among DMSO soluble POMs. NMR studies displayed very little change was caused by POM binding, thus a low-affinity binding is predicted [246].

The dimerization region of HIV protease is highly conserved making it a worthy target in HIV drug development [248]. The design of dimerization inhibitors was accelerated by the use of 12-aminododecanonic acid (12-Ado) as a template. 12-Ado permitted modification including directional changes and amino acid modification. These modifications displayed a cumulative effect with increasing potent modifications [230]. The β-sheet on the N-terminus of the protease monomer is most often targeted, although the C-terminus is also targeted [249,250]. Since partial dissociation of mature protease occurs, resulting in the disruption of the active site, dimerization inhibitors are classed as competitive inhibitors [251,252]. Inhibitors generally have low solubility due to being hydrophobic or containing a substantial hydrophobic constitution.

An antibody dimerization inhibitor known as mAb1696, uncouples the protease dimer and inhibits HIV protease activity [229]. Single-chain Fv (scFv) is an artificial derivative of mAb1696 retaining its potency at higher doses. The proposed mode of action involves detaching the N-terminal residues from the β-sheet resulting in a Hydrogen bonded loop. This allows for antibody binding preventing dimerization and thus disrupting the active site. scFv and mAb1696 both show activity against HIV-1 and HIV-2 Proteases as well as a resistant variant. scFv prevents self-cleavage of HIV protease on the C-terminal but not on the N-terminus. The ability of mAb to bind to HIV protease is optimal on free N-termini, however, even small additions hinder its binding [229].

The C- and N-terminal domains of HIV protease themselves have inhibitory activity by preventing dimerization [250,253]. A synthetic peptide derivative of these termini fused with the Tat-protein cell permeable domain (CPD) is known as P27. P27 was active against protease and its resistant variant,
R1, carrying eight resistance mutations. MTT assays on MT-2 cells showed protection of HIV-induced cell death in a dose dependent manner. Higher doses also led to decreased viral capsid formation, implying lower viral release. A combination of p27 and a potent active site inhibitor showed enhanced activity. Longer gag-pol peptides were observed signifying inhibition of viral maturation [253]. A-seco type triterpenoids are also protease inhibitors suspected to be dimerization inhibitors [254].

C3-substituted cyclopentyltetrahydrofuranyl (Cp-THF)-derived P2-ligands are inhibitors that bind the flap region of HIV protease. A derivative with a 3-(R)-hydroxy group displayed high inhibitory activity even against resistant proteases. Assays were conducted with various C-3 substituted polar group Cp-THFs to select a compound with maximum inhibitory and antiviral activity. By testing each of these modified Cp-THFs derivatives against a panel of proteases, including wild-type HIV1 and HIV2 with their resistant variants, an effective inhibitor can be identified [231,232,255–257]. Using this technique two compounds designated 3c and 3d were identified as being the most potent inhibitors of HIV protease. These two varied due to their stereo chemistry, with the more potent 3c displaying a unique water-mediated interaction with the backbone of HIV protease on the amide bond of Glycine 48 [231].

5.3. Other Drugs of Interest

Since peptides regulate most physiological processes, naturally synthesized peptides are an obvious source for new drugs. NAPVSIPQ (NAP) is a peptide derived from activity-dependent neuroprotective protein (ADNP). This peptide drug protects cells against the mitochondrial release of cytochrome c. Since it also stabilizes the Tau protein, NAP links the protection of microtubule network to the protection against apoptosis. NAP acts upstream of caspases 3 activation, preventing the release of cytochrome c [176].

Double stranded RNA Activated Caspase Oligomerizer (DRACO) is a broad spectrum antiviral drug. It selectively causes the apoptosis of virally infected cells. Healthy and infected cells are differentiated by the length of RNA transcription helices present in the cell. Viruses produce long dsRNA helices while the RNA helices of mammalian cells are much smaller. DRACO itself is a chimeric protein that binds viral dsRNA with one domain and binds pro-caspases with its other domain. If two or more DRACO molecules bind to the same dsRNA cells are then killed in a caspase dependent manner [235].

5.3.1. Aptamers

Aptamers are small nucleic acids characterized by antiviral activity. The variation in aptamers is in what they bind. Targets include gp120, Tat-protein, HIV protease and Reverse transcriptase [258–268]. B40 is an RNA aptamer binding gp120, preventing viral entry. Real time surface plasmon resonance technology demonstrated that this was due to a disruption of CCR5 and gp120 interactions. B40 binds to the gp120 core, mutating gp120 core residues resulted in disruption of B40 binding. This binding also induces conformational changes affecting binding of other compounds [259,261].

The use of RNA aptamers provides for use of small interference RNA (siRNA). A siRNA aptamer against Tat-protein and a gp120 binding aptamer were tested in vivo [263], using RAG-humice which is a humanized mouse model which can sustain long term HIV infection [260]. The first molecule
was an aptamer-siRNA chimera (Ch A-1). Ch A-1 was able to reduce viral load to undetectable quantities which lasted beyond treatment time. In contrast, the siRNA alone had no detectable effect on viral load. Both Ch-A-1 and the second aptamer, aptamer A-1, were able to significantly reduce cell death [263]. A preventative approach involves use of aptamer and siRNA chimeras to knockdown HIV target cells, i.e., CD4 cells and macrophages within the host. This has proved effective in humanized mouse models [266].

The techniques of Nano-delivery as well as DNA aptamers have been formulated to address the instability of RNA aptamers, Targeted delivery would also improve the effectiveness of the drug [267,268]. Nanoparticle, pRNA, was used to deliver a siRNA chimera. This chimera targeted gp120 which is expressed on the surface of infected cells. This allows a gp120 specific aptamer to dock on gp120 expressing cells. The inclusion of Ab’ pRNA–siRNA chimera 2’-Fluoro backbone modifications of pyrimidines, stabilized the moieties from degradation and assisted in the activity of dicer in gene silencing [267]. The technique of systematic evolution of ligands by exponential enrichment (SELEX) allows the generation of siRNA carrying DNA aptamers. The DNA aptamer was generated through direct conversion of a CD4 specific RNA aptamer. These were used to perform siRNA against HIV protease. DNA aptamers with siRNA proved more stable and efficient in protease silencing than the RNA aptamer alone [268].

5.3.2. Nanoformulation of ARVs

Targeted drug delivery promises to increase the effectiveness and safety of drugs. Multiple nano-delivery methods have been tested in the treatment of HIV. These include; polymeric nanoparticles [269], solid lipid nanoparticles (SLNs) [270], liposomes [271], nano-emulsions [272], dendrimers [273] and drug conjugates [274]. The size of Nano-delivery subjects is below one micron. Nano-delivery provides manageable toxicity patterns, adjustable drug release, low costs and high dosage tolerance. Additional advantages include protection from metabolism and a longer retention within the patient [275]. Active targeting allows for specific ligands on cell surfaces to be targeted, resulting in a specific delivery to a specific cell type. Additionally, the option to administer different drugs using one delivery system is available. This option eclipses error in administration and confers the ability to modulate individual drug release. The modes of delivery are specialized enough to allow Intracellular delivery [276], lymphatic system delivery [277] and central nervous system delivery [278]. With intracellular delivery lysosomal destruction can be bypassed allowing for nuclear or cytoplasmic delivery [276].

6. Conclusions

Despite advances in HIV treatments, there is still a high rate of infection and prevalence in sub-Saharan Africa, with the mortality rate due to HIV-infection being devastating throughout the world. In addition to this, the nature of HIV-infection may lead to other pathological condition such as tumourigenesis and HIV associated pathologies continue to affect patients despite the use of current treatments. Although it is currently the most effective treatment towards HIV-infection, discordant patients remain non-responsive to HAART. HAART also has side-effects such as lipodystrophy. The fact that the virus has developed the means to induce or inhibit apoptosis in
ways that will benefit its survival has been a source of great interest and intensive study. By elucidating the mechanisms behind the ability of viral encoded proteins to alter apoptotic pathways, a long list of possible drug targets has been constructed. These vary from cell type specific signaling, to targeting interactions with cell surface receptors and components of the intrinsic and extrinsic apoptotic pathways. There are existing drugs that target the activity of HIV encoded proteins, such as HIV protease inhibitors. However, as mentioned above there is a large and varied list of different drugs using multiple approaches with a wide variety of targets that are currently under development. These drugs all share the common characteristic that they act to inhibit the ability of HIV to manipulate the apoptotic machinery of the host, thus, making it difficult for the virus to evade the immune system by decreasing the number of immune -competent cells by increased apoptosis. These drugs also decrease the ability of the virus to inhibit apoptosis in infected cells, allowing increased viral replication. By understanding and targeting the ability of the virus to manipulate the apoptotic machinery of the host these new therapies can aid to combat HIV/AIDS and improve the quality of life for HIV positive people including discordant patients.

Acknowledgments

We would like to thank Mojakgomo Rahaba, Boitumelo Mofolo and Mahlori Mkhabele for their assistance in writing this review article.

Author Contributions

Z.M., R.H. and Z.D contributed equally to this manuscript.

Conflicts of Interest

The authors declare no conflict of interest.

References and Notes


Virusess 2014, 6


211. Eccles, S.A. Parallels in invasion and angiogenesis provide pivotal points for therapeutic intervention. *Int. J. Dev. Biol.* 2004, 48, 583–598.


© 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).