

Article

The Virome of Babaco (*Vasconcellea* × *heilbornii*) Expands to Include New Members of the *Rhabdoviridae* and *Bromoviridae*

Edison G. Reyes-Proaña^{1,2,†}, Maria G. Cañada-Bautista¹, Juan F. Cornejo-Franco³, Robert A. Alvarez-Quinto^{4,†}, Dimitre Mollov⁵, Eduardo Sanchez-Timm^{1,3} and Diego F. Quito-Avila^{1,3,*}

¹ Facultad de Ciencias de la Vida, Escuela Superior Politécnica del Litoral, ESPOL, Km 30.5 Vía Perimetral Campus Gustavo Galindo, Guayaquil 090902, Ecuador; edison@uidaho.edu (E.G.R.-P.); mcanada@espol.edu.ec (M.G.C.-B.); lesanche@espol.edu.ec (E.S.-T.)

² Department of Entomology and Plant Pathology, University of Idaho, Moscow, ID 83843, USA

³ Centro de Investigaciones Biotecnológicas del Ecuador, Escuela Superior Politécnica del Litoral, CIBE-ESPOL, Km 30.5 Vía Perimetral Campus Gustavo Galindo, Guayaquil 090902, Ecuador; jcornejo@espol.edu.ec

⁴ Department of Botany and Plant Pathology, Oregon State University, Corvallis, OR 97330, USA; alvarero@oregonstate.edu

⁵ USDA-ARS, Corvallis, OR 97330, USA; dimitre.mollov@usda.gov

* Correspondence: dquito@espol.edu.ec

† These authors contributed equally to this work.

Abstract: Babaco (*Vasconcellea* × *heilbornii*) is a subtropical species in the *Caricaceae* family. The plant is native to Ecuador and represents an important crop for hundreds of families. The objective of this study was to characterize, at the genomic level, two new babaco viruses identified by high-throughput sequencing. The viruses, an ilarvirus and a nucleorhabdovirus, were found in a symptomatic babaco plant from a commercial nursery in the Azuay province of Ecuador. The tripartite genome of the new ilarvirus, provisionally named babaco ilarvirus 1 (BabIV-1), is related to subgroup 3 ilarviruses, including apple mosaic virus, apple necrotic mosaic virus, and prunus necrotic ringspot virus as the closest relatives. The genome of the nucleorhabdovirus, provisionally named babaco nucleorhabdovirus 1 (BabRV-1), showed the closest relation with joa yellow blotch-associated virus and potato yellow dwarf nucleorhabdovirus. Molecular-based detection methods found BabIV-1 and BabRV-1 in 21% and 36%, respectively, of plants surveyed in a commercial babaco nursery, highlighting the importance of enforcing virus testing and nursery certification programs for babaco.

Keywords: *Nucleorhabdovirus*; *Iilarvirus*; high-throughput sequencing; plant virus discovery; mountain papaya



Citation: Reyes-Proaña, E.G.; Cañada-Bautista, M.G.; Cornejo-Franco, J.F.; Alvarez-Quinto, R.A.; Mollov, D.; Sanchez-Timm, E.; Quito-Avila, D.F. The Virome of Babaco (*Vasconcellea* × *heilbornii*) Expands to Include New Members of the *Rhabdoviridae* and *Bromoviridae*. *Viruses* **2023**, *15*, 1380. <https://doi.org/10.3390/v15061380>

Academic Editors: Zongtao Sun and Mengji Cao

Received: 3 May 2023

Revised: 13 June 2023

Accepted: 14 June 2023

Published: 16 June 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Babaco (*Vasconcellea* × *heilbornii*) is an herbaceous tree native to the Ecuadorian highlands. It belongs to the *Caricaceae* family and shares morphological and anatomical traits with its tropical relative, papaya (*Carica papaya*) [1]. The babaco fruit has unique and exotic organoleptic properties and has been commercialized fresh or processed in local and international markets. In Ecuador, babaco is produced in small orchards, usually under greenhouse conditions. Nevertheless, the vegetative propagation, the lack of pathogen-free certified material, as well as the unregulated nursery operations have caused an exponential dissemination of viruses in the main producing areas, threatening the profitability and sustainability of this native crop [2,3].

In the last decade, high-throughput sequencing (HTS) has advanced the discovery of pathogenic and non-pathogenic viruses in agricultural and ornamental ecosystems [4]. In babaco, HTS has identified several disease-associated viruses including babaco mosaic virus (BabMV, genus *Potexvirus*), papaya ringspot virus (PRSV, genus *Potyvirus*), babaco virus Q (BabVQ, umbra-like virus), babaco cheravirus 1 (BabChV-1, genus *Cheravirus*), and

babaco nepovirus 1 (BabNV-1, genus *Nepovirus*) [2,3]. In addition, the genomic sequence for putative cryptic viruses, such as babaco cryptic virus-1 (BabCV-1, a partiti-like virus) and babaco endogenous pararetrovirus (BabEPV, a hypothetical integrated pararetrovirus), has been reported [3].

Here, we report two additional babaco-infecting viruses belonging to distinct genera, *Iilarvirus* (family *Bromoviridae*) and *Alphanucleorhabdovirus* (family *Rhabdoviridae*). The genus *Iilarvirus* currently comprises 22 species recognized by the International Committee on Taxonomy of Viruses [5]. *Iilarviruses* have non-enveloped, spherical virions that are 16–36 nm in diameter and single-stranded positive-sense tripartite RNA genome [6]. RNA 1 contains a single open reading frame (ORF) coding for the virus replicase (Protein 1a). Depending on the species, RNA 2 can have one or two ORFs encoding a viral polymerase (ORF 2a) and an RNA silencing suppressor (ORF 2b), expressed by a subgenomic RNA [7,8]. RNA 3 contains two ORFs (ORF 3a and 3b) which code for the movement protein (MP) and the coat protein (CP), respectively [8]. *Iilarviruses* are pollen- and seed-borne but can also be plant-to-plant transmitted by thrips, which act as carriers of virus-contaminated pollen [9].

The family *Rhabdoviridae* is a large and diverse family of viruses with unsegmented or bi-segmented negative-sense single-stranded RNA genomes that infect vertebrates, invertebrates, and plants. *Rhabdovirus* virions typically consist of bullet-shaped or bacilliform particles ranging from 100 to 430 nm in length and 45 to 100 nm in diameter, whereas some consist of non-enveloped filamentous particles [10].

The subfamily *Betarhabdovirinae* groups six genera of plant- and arthropod-infecting rhabdoviruses: *Alphanucleorhabdovirus*, *Betanucleorhabdovirus*, *Gammanucleorhabdovirus*, *Cytorhabdovirus*, *Dichorhavirus*, and *Varicosavirus*. *Alphanucleorhabdoviruses*, having unsegmented genomes of 11–15 kb, have been reported from monocots and dicots and are transmitted in a persistent-replicative manner by leafhoppers, planthoppers, or aphids [11,12]. The genome organization of *alphanucleorhabdoviruses* includes leader (l) and trailer (t) sequences at the 3' and 5' of the genome, respectively, and enclose six canonical ORFs encoding the nucleocapsid (N), phosphoprotein (P), the movement protein (MP), also referred to as Y protein, the matrix (M) protein, the glycoprotein (G), and the polymerase (L) [13,14].

The objective of this study was to characterize, at the genomic level, two new babaco viruses and infer their genetic relationships with other members of the *Bromoviridae* and *Rhabdoviridae* families, respectively.

2. Materials and Methods

2.1. Plant Material and Virus Detection

A single six-month-old babaco plant showing virus-like symptoms, such as leaf yellowing, premature leaf abscission, and mild leaf mosaic (Figure 1), was obtained from a commercial nursery in the Azuay province in Ecuador and was selected for this study. The plant was kept under greenhouse conditions and tested by reverse-transcription PCR (RT-PCR) for a panel of known babaco-infecting viruses described so far [3], including BabMV, PRSV, BabChV-1, BabNV-1, BabVQ, BabCV, and BabEPV, using the primers listed in Supplementary Table S1.

Total RNA was extracted following a modified protocol described by Halgren, A. et al. [15]. Briefly, 100 mg of leaf tissue were ground and homogenized in 1 mL of lithium-chloride based RNA-extraction buffer containing 1% β -mercaptoethanol. The supernatant was mixed with an equal volume of 6 M potassium-acetate (pH 6.5) and centrifuged; then, the resulting supernatant was mixed (1:1) with isopropanol and centrifuged to form a pellet. The pellet was precipitated by centrifugation and rinsed two times with wash buffer (TrisHCl + NaCl + EDTA) + Ethanol (1:1) in the presence of glassmilk. Finally, the glassmilk–RNA complex was vacuum dried, and the RNA was resuspended in 100 μ L of water and stored at -20 °C.

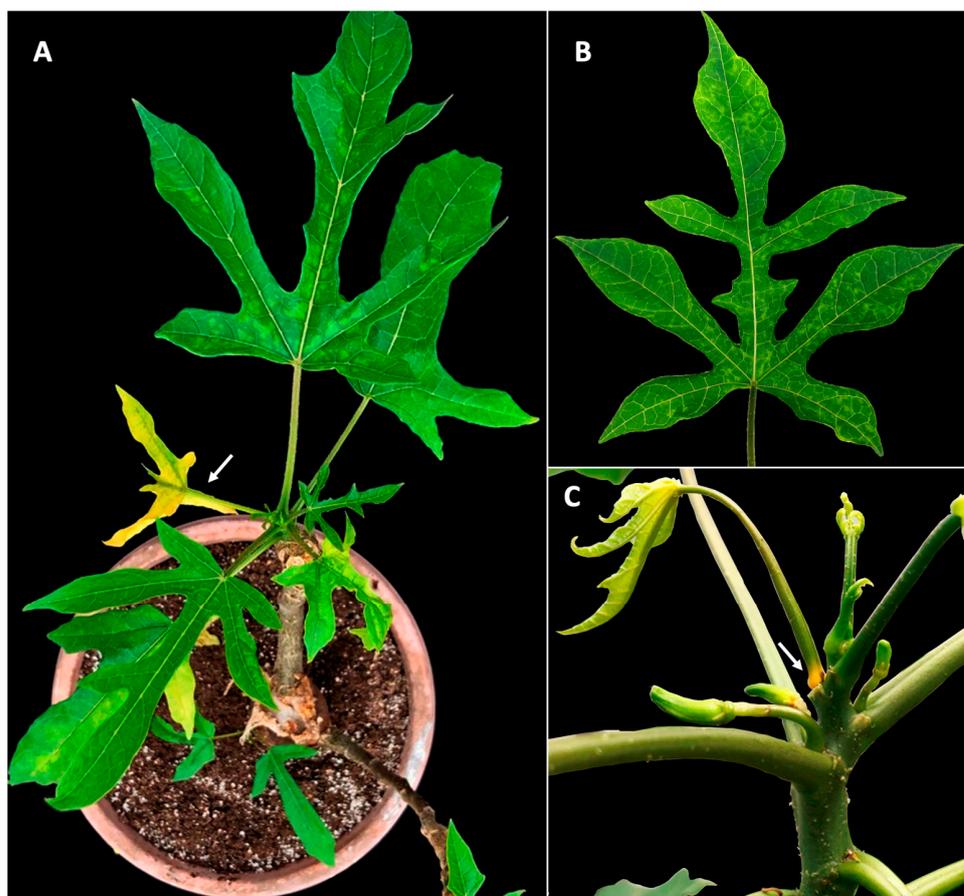


Figure 1. Symptomatic babaco (*Vasconcellea × heilbornii*) plant subjected to this study. (A) Leaf mosaic and yellowing (arrow). (B) Detached leaf with vein banding and mosaic. (C) Yellowing (arrow) at the petiole base of a leaf with premature abscission. Pictures correspond to the same plant at different times.

Reverse transcription (RT) was carried out in a 15 μL reaction containing 0.1 μL of SuperScript III (Thermo Fisher, Waltham, MA, USA), 0.5 μL of random hexamers (40 μM), and 1 μL of RNA template (~45 ng) following manufacturer instructions. PCR was completed using GoTaq[®] Green Master mix (Promega, Madison, WI, USA) in 10 μL reactions with 1 μL of template and 0.5 μL of each primer at 40 μM . To validate RNA quality and synthesized cDNA, a 721 nt region corresponding to NADH dehydrogenase ND2-subunit was amplified as a housekeeping gene by primers F: 5'-GGACTCCTGACGTATACGAAGGATC-3' and R: 5'-AGTAGATGCTATCACACATACAAT-3' [16,17]. To detect the presence of BabIV-1 and BabRV-1, specific primers were designed. For BabIV-1 detection, a 397 nt fragment corresponding to a fragment of the MP and the CP was amplified by F: 5'-GGTTAGAAGCAGGATTGCGTC-3' and R: 5'-CTTGAGCTCCAGTCGGATGAC-3'. On the other hand, primers F: 5'-GGAGTCGGCATTGGTATAGCA-3' and R: 5'-CCGTTGTCCTTTA ACACCCTAG-3' targeted a 923 nt region corresponding to the N and P genes from BabRV-1. PCR conditions were as follows: initial denaturation at 95 $^{\circ}\text{C}$ for 4 min followed by 40 cycles of 95 $^{\circ}\text{C}$ for 40 s, 58 $^{\circ}\text{C}$ for 40 s, and 1 min at 72 $^{\circ}\text{C}$, and a final extension step at 72 $^{\circ}\text{C}$ for 7 min.

2.2. Virus Sequencing, Assembly, and Annotation

The nearly complete genome was obtained by HTS from total RNA extracted from 100 mg of symptomatic leaf tissue using the RNeasy Plant Mini Kit (Qiagen, Hilden, North Rhine-Westphalia, Germany). The total RNA was subjected to ribosomal-RNA removal by using the Ribo-Zero Plant kit. The cDNA library was prepared by Illumina's TruSeq

Stranded total RNA and sequenced on an Illumina HiSeq 4000 platform as pair-ended reads (2×150 bp) at Macrogen Inc. (Teheran-ro, Gangnam-gu, Seoul, Republic of Korea). Raw reads were preprocessed using the BBDuk adapter/quality trimming tool version 38.84 to remove adapters and low-quality reads. Trimmed reads were assembled using SPAdes assembler version 3.15.2 [18]. Since the genome of babaco has not been determined thus far, assembled contigs were mapped to the genome of *Carica papaya* (NCBI PRJNA264084) to filter out common contigs between these two members of the *Caricaceae* family. Non-host contigs were analyzed by BLAST search using a custom viral sequences database (retrieved from the NCBI in November 2021) and the full NCBI non-redundant protein sequences (nr) database.

Putative viral genomes were confirmed by Sanger sequencing of overlapping RT-PCR products using the primers listed in Supplementary Table S2. Amplicons were generated using the proof-reading polymerase Platinum SuperFi II DNA (Thermo Fisher, Waltham, MA, USA). RNA termini for each viral genome were determined from double-stranded RNA (dsRNA) using a 5'/3' RACE Kit, Second Generation System (Roche, Basel, Switzerland), and the primers listed in Supplementary Table S2. DsRNA was extracted from fresh leaf tissue following the protocol described by Dodds et al. [19]. Briefly, 15 g of leaf tissue were ground in liquid nitrogen and homogenized for 30 min in a mixture containing 40 mL of 2X STE (50 mM Tris-Cl, 100 mM NaCl, 1 mM EDTA, pH 7.1), 8 mL of 10% SDS, 23 mL of STE-saturated phenol, and 0.5 mL of B-mercaptoethanol. The homogenate was centrifuged at $10,000 \times g$ for 20 min. The supernatant was transferred to a new flask, measured, and mixed with 0.2 volumes of 100% ethanol and 1.5 g of cellulose fibers medium (Sigma-Aldrich, St. Louis, MO, USA). The cellulose-bound dsRNA was pelleted and washed twice in STE/18%EtOH (18% ethanol, *v/v*, in STE). The dsRNA was eluted in 15 mL of STE. DNA and ssRNA were digested by the addition of 30 U of DNase I (Sigma), 1000 U RNase T1 (Sigma), and 300 μ L of 2 M $MgCl_2$ at 37 °C for 1 h. The digestion was terminated by the addition of 1 mL of 0.5 M EDTA, pH 8.0, and 5 mL of 100% EtOH. The dsRNA was homogenized in the presence of 0.2 g of cellulose fibers medium and was precipitated and washed in 30 mL of STE/18%EtOH. The final elution was completed in 3 mL of STE.

Amplified DNA products were cloned using a pGEM[®]-T easy vector system (Promega, Madison, WI, USA) and sequenced at Macrogen Inc. (Teheran-ro, Gangnam-gu, Seoul, Republic of Korea). The complete viral genomes, including the corresponding terminal sequences, were assembled using Geneious Prime 2022.0.2. For genome annotation, putative ORFs were predicted using Unipro UGENE v40.03 [20] and ORFfinder (NCBI). The hypothetical protein homologs were identified by BLASTp and the non-redundant protein sequences (nr) database (<https://blast.ncbi.nlm.nih.gov/>, accessed on 13 January 2022). Predicted amino acid sequences were analyzed to identify functional conserved domains. This analysis was performed using the Conserved Domains database at NCBI (<https://www.ncbi.nlm.nih.gov/Structure/cdd/wrpsb.cgi>, accessed on 13 January 2022).

2.3. Phylogenetic Analysis

Phylogenetic analyses were performed based on nucleotide and amino acid sequences retrieved from NCBI GenBank and aligned using MUSCLE version 3.8 [21]. The best protein substitution model was inferred using MEGAX [22]. For the new ilarvirus, the analysis was performed for all four predicted proteins encoded by the genome. The first two proteins were related to virus replication (P1a and P2a), movement protein (MP), and coat protein (CP). The best protein models were LG + G + I, rtREV + G + F + I, LG + G, and LG + G + F for P1a, P2a, MP, and CP, respectively. For the rhabdovirus, the amino acid sequence of the viral replicase (L protein) was used, implementing rtREV substitution model. Phylogenetic relationships were estimated using RAxML [23] on CIPRES with 1000 bootstrap replicates.

2.4. Mechanical Transmission and Virus Survey

Leaves of the original virus-infected plant were ground in 0.05 M phosphate buffer (pH 7.0) 1:10 (wt/vol), as described by Hull [24]. The two youngest, fully developed leaves of each test plant were dusted with carborundum (silicon carbide 600 mesh) and rubbed with the infectious sap using a soft sponge. Three plant species belonging to the family *Caricaceae*, babaco (*Vasconcellea* × *heilbornii*), chamburo (*Vestipulata pubescens*), and papaya (*Carica papaya*), were evaluated for mechanical transmission, with five replicates for babaco and papaya, while 10 chamburo plants were used. Inoculated plants were kept in an insect-proof cage within a greenhouse at 22–25 °C and 12 h natural light. Inoculated plants were virus tested at 15-, 30-, and 90-days post inoculation (dpi).

A group of fifty six-month-old babaco nursery plants showing mild leaf mosaic were sampled for determining the prevalence of the new viruses in a single commercial nursery in the Azuay province. These samples were subjected to total RNA extraction, as described above, and RT-PCR for virus detection using the primers listed in Supplementary Table S1.

3. Results

3.1. Virus Detection

The symptomatic babaco sample investigated in this study tested negative by RT-PCR for BabMV, PRSV, BabChV-1, and BabNV-1, which have been shown to be associated with leaf symptoms in babaco [3]. BabVQ, an umbra-like virus considered non-pathogenic [25], along with BabCV and BabEPV, both asymptomatic ‘cryptic’ viruses, were RT-PCR detected in the sample.

3.2. High-Throughput Sequencing

HTS resulted in a raw dataset of 54,785,178 paired-end reads. Post-trimming reads (54,689,746) were de novo assembled, resulting in 94,378 contigs, from which 23,934 contigs mapped to the genome of papaya (*Carica papaya*). BLAST search of the remaining contigs (70,444) identified a 4.7 kb-long contig showing 98% identity (100% coverage) to BabVQ. A total of 11 contigs ranging from 0.4 to 2.6 kb in length were assembled from 0.000006% of the total reads. These contigs showed homology to available partial sequences of both BabCV (98% identity) and BabEPV (89% identity), supporting the results obtained by RT-PCR.

Additional contigs of 3.2, 2.5, and 1.7 kb in length were identified with homology (nt identities in the range 67–71%) to corresponding RNA genomic segments of members of the genus *Iilarvirus* (family *Bromoviridae*), whereas a 12.7 kb contig showed homology to members of the genus *Alphanucleorhabdovirus* (family *Rhabdoviridae*). A summary of the most significant virus-related contigs assembled in this study, assembly details, and their corresponding homologous viruses are presented in Table 1. A significant difference in virus-reads abundance was observed between BabVQ (11.94% of total reads) and other viruses (0.006–0.028% of total reads) (Table 1).

3.3. Genome Organization, Sequence Comparison, and Phylogenetics of a New *Iilarvirus*

The complete genomic sequence of a putative new ilarvirus, provisionally named babaco ilarvirus 1 (BabIV-1), consisted of three RNA segments of 3299 nt (RNA1), 2675 nt (RNA2), and 1802 nt (RNA3). The genomic sequence segments of BabIV-1 were deposited in the GenBank under accession numbers OQ256238, OQ256239, and OQ256240, respectively.

RNA 1 contains a single ORF at nucleotide position 33–3143 coding for the hypothetical protein 1a (P1a) of 1037 aa and a predicted molecular weight of 117 kDa. This protein has two conserved domains: the viral methyltransferase (MET, PF01660, F53-T387) and the helicase (HEL, PF01443, K744-T1001) (Figure 2). At the amino acid level, P1a of BabIV-1 is more closely related to that of lilac leaf chlorosis virus (LLCV, YP_009104367.1; 65% identity; 99% coverage) and prunus necrotic ringspot virus (PNRSV, AEP04410; 63% identity; 99% coverage). The motif (₇₄₇DGVAXCXXXT₇₅₆), a common feature in P1a of ilarviruses [26], was identified in the counterpart from BabIV-1.

Table 1. Summary of the most significant viral genomes assembled from high-throughput sequencing.

Virus	Contig Length (nt)	Genomic RNA Segment	Number of Reads Assembled into Contig (%) ⁽¹⁾	Average Sequencing Depth (per nt)	GenBank Closest Hit (Accession Number)	Nucleotide Identity (%) (Query Coverage%) to Closest Hit
Babaco virus Q	4707	-	6,534,602 (11.940%)	144,256	Babaco virus Q (MN648673)	98% (100%)
New babaco ilarvirus	3201	RNA1	15,806 (0.028%)	482.5	Apple necrotic mosaic virus, RNA1 (KY808376)	68% (60%)
	2508	RNA2	4221 (0.007%)	153	Apple necrotic mosaic virus, RNA2 (LC108994)	71% (53%)
	1719	RNA3	13,788 (0.025%)	760	Prunus necrotic ringspot virus, RNA3 (MZ451064)	67% (36%)
New babaco rhabdovirus	12,770	-	3633 (0.006%)	32.3	Joa yellow blotch-associated virus (MW014292)	68% (91%)
Endogenous pararetrovirus (BabEPV)	632–2682	-	3681 (0.007)	4–108	Citrus endogenous pararetrovirus (ARO38276)	89% (45%)
Babaco cryptic virus (BabCV)	360–382	-	34 (0.000062)	3	Babaco cryptic virus—1 (MN648674)	98% (87%)

⁽¹⁾ From total reads post-trimming (54,689,746).

The single ORF found in RNA 2 is positioned at nucleotides 25–2553 and codes for an 843 aa protein (P2a) of 97 kDa with the conserved viral RNA-dependent RNA polymerase (RdRp) domain (PF00978, T305-R720) (Figure 2). The putative P2a of BabIV-1 shares the highest level of amino acid identity with its counterpart from apple necrotic mosaic virus (ApNMV, AVI05085; 62% identity; 95% coverage). The highly conserved motif ⁶⁰⁹ASGDDSLI₆₁₆, which is present in P2a of other ilarviruses [27], was identified within the corresponding P2a of BabIV-1.

RNA 3 contains two non-overlapping ORFs, ORF3a, and ORF3b. Translation of ORF3a (position 96–947 nt) results in a 284 aa protein of 32 kDa having conserved domains of the bromovirus MP family (PF01573, E16-Q229). The amino acid sequence of BabIV-1, the hypothetical MP, showed the highest identity with its homologue from LLCV (YP_009104372.1; 69% identity; 93% coverage). ORF 3b (position 953–1687 nt) putative protein is translated through a subgenomic RNA, producing a 245 aa long protein (P3b) of 27 kDa, with conserved domains present in bromovirus CPs (PF01787, G41-E239) (Figure 2). The predicted CP of BabIV-1 is most closely related to its counterpart from LLCV (YP_009104373.1; 58% identity; 82% coverage) and contains the zinc-finger motif ³²CRLCNHHTHAGGCARCKKC₄₉. This conserved sequence is also present in the CPs of ilarviruses [8].

Complete genome analysis between BabIV-1 and ilarviruses currently listed in the ICTV revealed average nucleotide identities of 58%, 50%, and 43% for RNA segments 1, 2, and 3, respectively, with species in the ilarvirus subgroup 3. However, the identities between BabIV-1 and ilarviruses belonging to subgroups 1, 2, 4, and unclassified were in the range of 24%–40% for all genomic segments (Supplementary Table S3). Consistent with the relationships at the nucleotide level, the highest identities at the amino acid level were observed between predicted proteins of BabIV-1 and counterparts from subgroup 3 ilarviruses (Table 2).

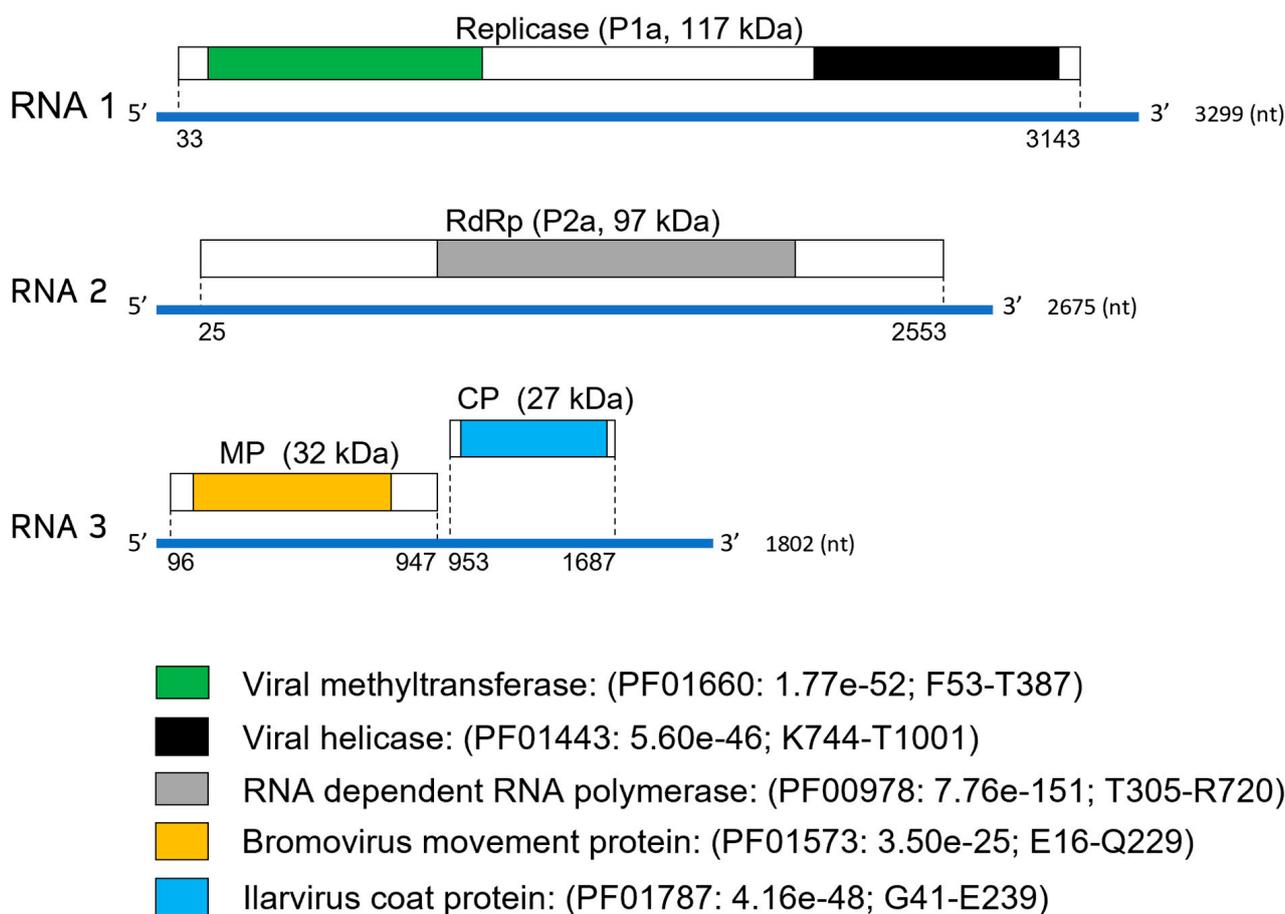


Figure 2. Genome organization of babaco ilarvirus-1. Genome positions of predicted open reading frames (ORFs) in each RNA segment are indicated with dotted lines. Conserved domains and molecular weight (in kDa) of the putative replicase, RNA-dependent-RNA-polymerase (RdRp), movement protein (MP), and coat protein (CP), respectively, are shown in different colors.

Table 2. Amino acid identities (%) between hypothetical proteins of babaco ilarvirus-1 (BabIV-1) and counterparts from closest ilarviruses. The accession numbers for the protein 1a (P1a), RNA-dependent-RNA-polymerase (RdRp), movement protein (MP), and the coat protein (CP) of subgroup 3 ilarviruses are listed in Supplementary Table S4.

Iilarviruses	P1a	RdRp	MP	CP
Subgroup 3				
Apple mosaic virus	73.96	65.63	71.33	61.61
Apple necrotic mosaic virus	74.46	71.98	72.79	70.00
Blueberry shock virus	74.53	66.16	73.58	68.56
Prunus necrotic ringspot virus	76.05	67.73	73.61	68.72
Lilac leaf chlorosis virus	78.24	64.97	81.79	72.85
Average (% identity)	75.45	67.29	74.62	68.35

Phylogenetic analyses using the amino acid sequence of each protein showed that BabIV-1 clusters with PNRSV, ApNMV, apple mosaic virus (ApMV), blueberry shock virus (BShV), and LLCV all belong to ilarvirus subgroup 3 (Figure 3).

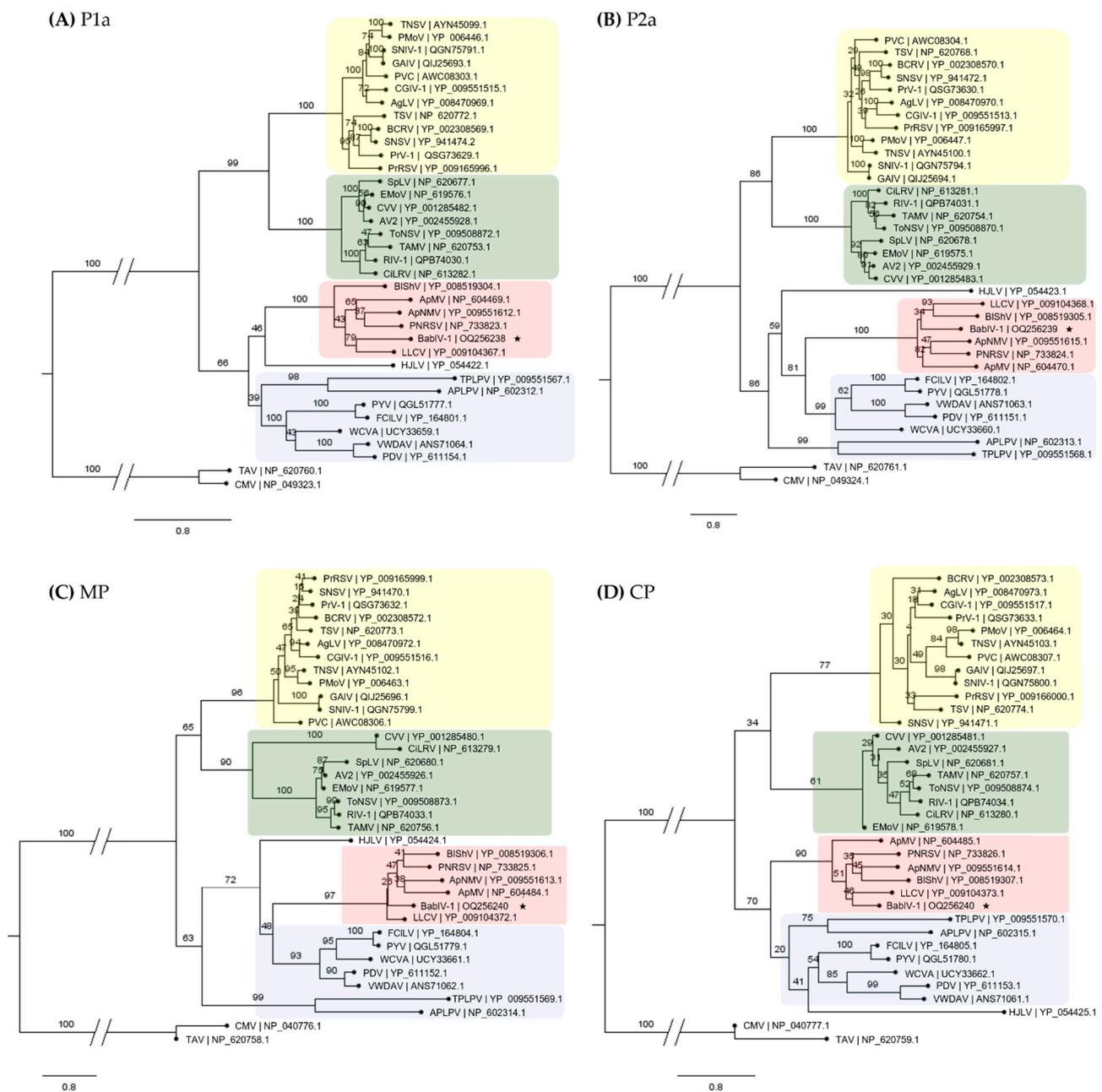


Figure 3. Phylogenetic analysis of babaco ilarvirus-1 (BabIV-1, GenBank accessions: OQ256238, OQ256239, OQ256240) and other ilarviruses based on amino acid sequences of the four proteins, P1a (A), P2a (B), movement protein (MP) (C) and coat protein (CP) (D). Colored shading denotes different clades which are consistent regardless of the protein analyzed. The analysis was performed on CIPRES with 1000 bootstraps. Bootstrap values are indicated at each node. Virus abbreviations, names and accession numbers are listed in Supplementary Table S4. *Tomato aspermy virus* (TAV) and *Cucumber mosaic virus* (CMV) were used as outgroups. Scale bar represents aminoacids substitution per site. BabIV-1 is marked with an asterisk.

3.4. Genome Organization, Sequence Comparison, and Phylogenetics of a New Alphanucleorhabdovirus

The genome of the new rhabdovirus, tentatively named babaco rhabdovirus-1 (BabRV-1), was deposited in NCBI GenBank under accession number OQ256237. The single-stranded negative sense monopartite genome of BabRV-1 is 12,802 nucleotides long. Seven ORFs were identified in the genome of BabRV-1. ORF 1 (nt positions: 156–1571) encodes a putative

51 kDa protein containing the conserved rhabdovirus nucleocapsid (N) domain (PF03216: 4.92e-23; K75-I345) with a 71.52% identity to potato yellow dwarf virus (PYDV). ORF 2 (nt positions: 1645–1953) codes for a small 12 kDa protein of unknown function with homology (32.69% aa identity) to protein X, encoded by ORF 2 of physostegia chlorotic mottle virus (PhCMoV). ORF 3 (nt positions: 2013–2852) encodes a putative 31 kDa protein with homology to the phosphoprotein (P) of several members of the *Rhabdoviridae* and shares a 58.06% identity at the amino acid level with joa yellow blotch-associated virus (JYBaV). ORF 4 (nt positions: 2915–3772) encodes a putative 32 kDa protein that contains the highly conserved motif $_{46}\text{IXD(X)}_{71}\text{G}$ belonging to the 30K superfamily of viral movement proteins (MP, Interpro IPR041344), while ORF 5 (nt positions: 3974–4747) codes for a putative matrix protein (M) of 29 kDa with a 68.87% of identity to the M protein of PYDV (Table 3).

Table 3. Amino acid identities (%) between the hypothetical proteins of babaco rhabdovirus-1 (BabRV-1) and counterparts from closest alphanucleorhabdoviruses. The accession numbers for the nucleocapsid (N), accessory ‘X’ protein, phosphoprotein (P), movement protein (MP), matrix (M), glycoprotein (G), and polymerase (L) are listed in Supplementary Table S5.

Virus	Amino Acid Identity (%)						
	N	X	P	MP	M	G	L
Joa yellow blotch-associated virus	70.76	28.95	58.06	67.13	61.87	66.19	74.36
Potato yellow dwarf virus	71.52	27.78	52.50	67.94	68.87	66.40	73.75
Physostegia chlorotic mottle virus	43.15	32.69	20.47	42.16	35.27	39.24	53.32
Eggplant mottled dwarf virus	41.70	25.47	18.52	42.16	32.56	37.69	52.91
Peach virus 1	24.17	Not applicable	15.51	10.19	14.60	21.83	33.25

The last two ORFs code for a putative glycoprotein (G) (ORF 6: nt positions: 4829–6706) with a MW of 69 kDa and the large (L) replicase (ORF 7: nt positions: 6867–12,704), with a MW of 221 kDa; these proteins share a 66.40% and 74.36% identity to PYDV and JYBaV, respectively. The latter has two conserved domains, the RdRp (PF00946: 5.58e-155; W217-L1077) and a mRNA-capping region V (PF14318: 1.34e-43; C1092- E1319), which include an essential conserved motif for mRNA capping $_{1160}\text{GXXT(X)}_{72}\text{HR}_{1235}$ [28].

The genome organization of BabRV-1, except for the accessory gene X, which is also observed in the genome of its closest relatives, adheres to the overall organization of nucleorhabdoviruses 3′-N-X-P-MP-M-G-L-5′ (Figure 4). A typical genomic feature observed in rhabdoviruses and other viruses in the order *Mononegavirales* is the presence of conserved intergenic sequences that regulate transcription [29]. In BabRV-1, a highly conserved region was identified in all the gene junctions having the consensus motif 3′-AAUUAUUUUUGGGUUG-5′ (Figure 4), which is also found in the intergenic regions of JYBaV and PYDV. Lastly, the coding region of BabRV-1 is flanked by 3′ leader (155 nt) and 5′ trailer (98 nt) stretches, which are partially complementary to each other (Figure 4).

Phylogenetic inferences based on the amino acid sequence alignment of the L protein of selected members of different genera of the *Rhabdoviridae* family clustered BabRV-1 with members of the genus *Alphanucleorhabdovirus*, with JYBaV and the potato yellow dwarf virus (PYDV) as closest relatives (Figure 5). Furthermore, BabRV-1, JYBaV, and PYDV share an identical genome organization, including the hypothetical X protein gene.

Thus far, the genus *Alphanucleorhabdovirus* includes 12 accepted virus species. The species demarcation criteria for viruses assigned to this genus state three important characteristics: (i) nucleotide sequence identity of complete genomes lower than 75%; (ii) different ecological niches as evidenced by differences in hosts and/or arthropod vectors; and (iii) ability to be distinguished in serological tests or by nucleic acid hybridization [30]. BabRV-1 satisfies two out of the three criteria, sharing 68% nucleotide identity with its closest relatives and having a natural host, babaco, different from those of PYDV and JYBaV,

which have been reported in solanaceaeous hosts. Hence, BabRV-1 should be considered a new species within the genus *Alphanucleorhabdovirus*.

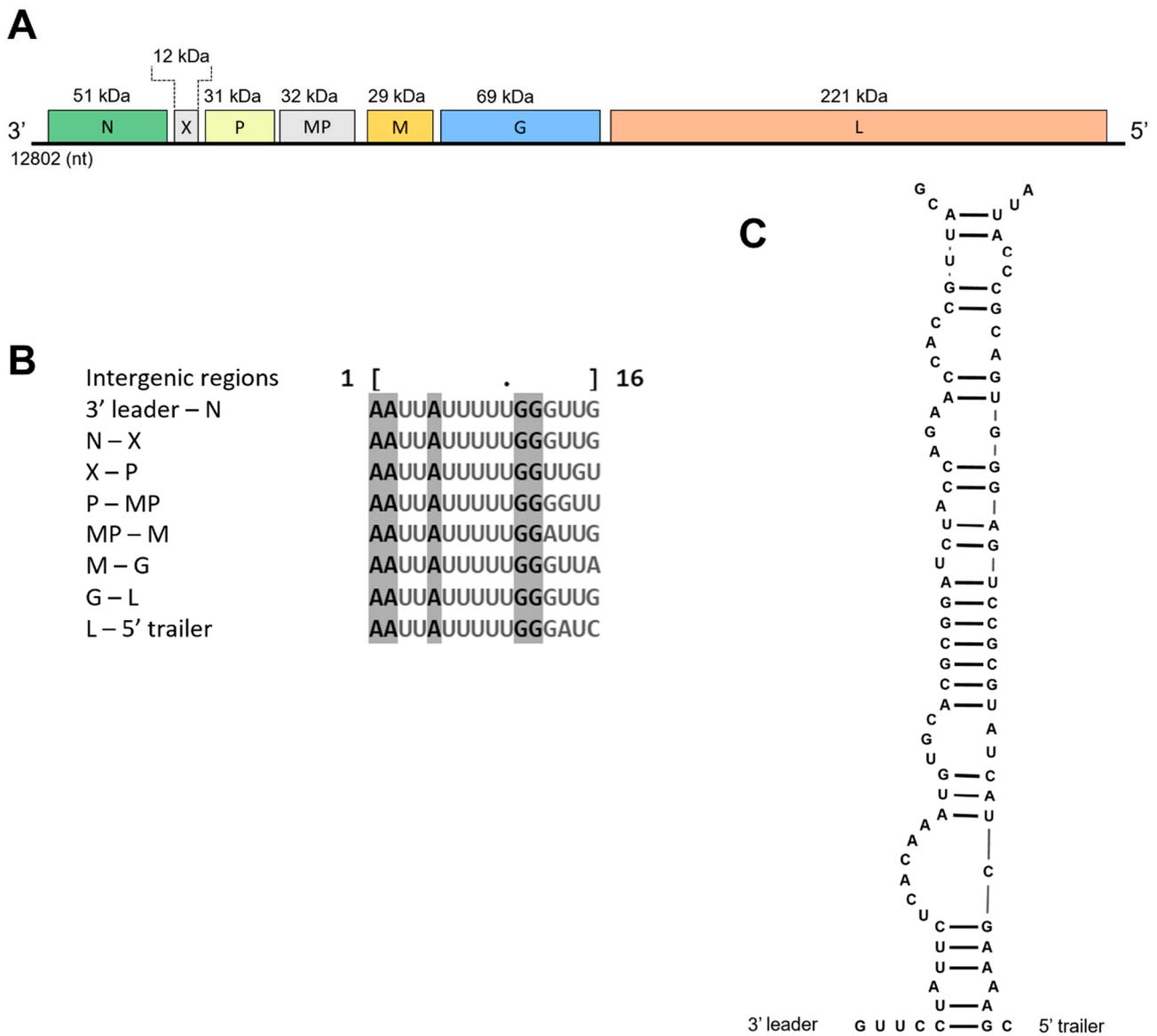


Figure 4. Genomic organization of babaco rhabdovirus-1 (BabRV-1). (A) Schematic representation of its predicted open reading frames (ORFs: nucleocapsid (N), protein X (X), phosphoprotein (P), movement protein (MP), matrix (M), glycoprotein (G), polymerase (L)). (B) Alignment of intergenic regions observed in BabRV-1 genome. (C) Partial genome complementarity of 3' leader and 5' trailer terminal sequences.

3.5. Mechanical Inoculation and Virus Survey

Mechanically inoculated plants were tested for BabIV-1 and BabRV-1 and monitored for 90 days. BabIV-1 and BabRV-1 failed to be mechanically inoculated onto papaya, babaco, or chamburo. In babaco plants ($n = 50$) obtained from the commercial nursery in the Azuay province, BabIV-1 and BabRV-1 were detected in 21% and 36%, respectively, while 7% tested positive for both viruses. BabVQ, BabCV, and BabEPV were also detected in all the plants tested.

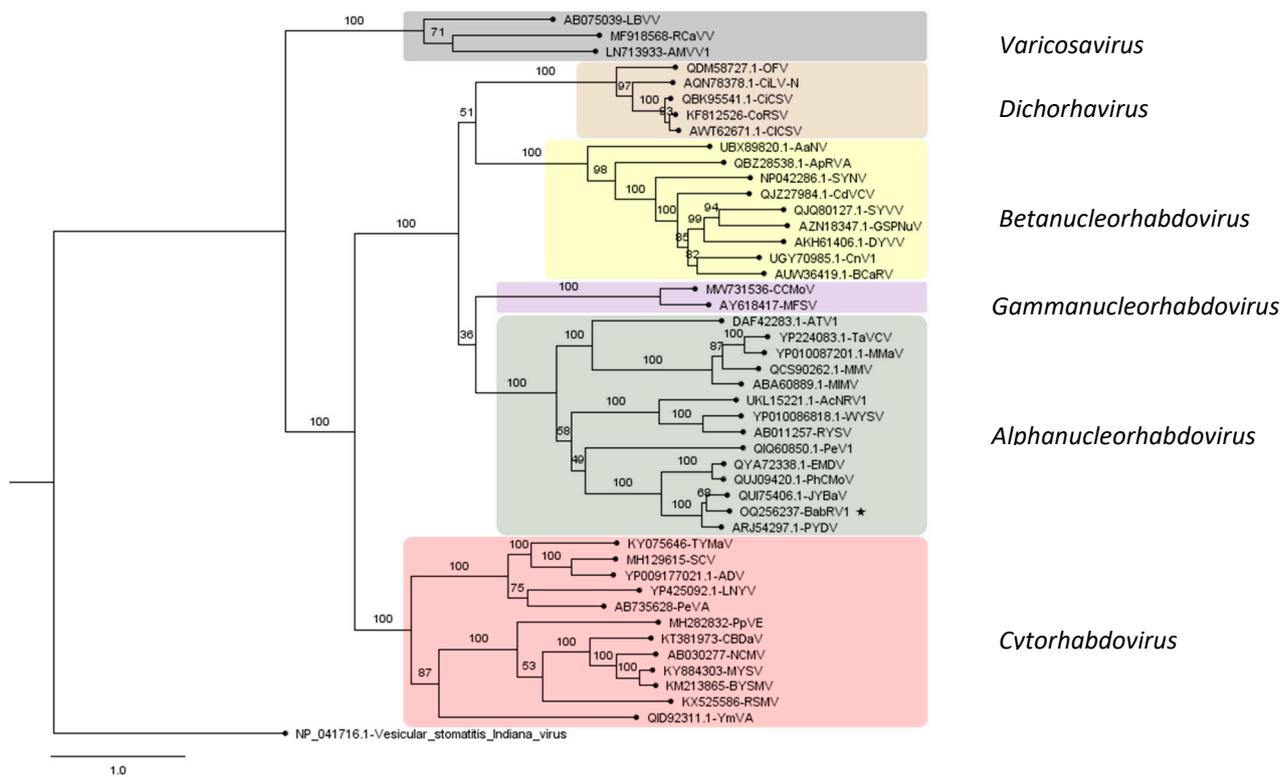


Figure 5. Phylogenetic analysis of babaco nucleorhabdovirus-1 (BabRV-1, GenBank accession: OQ256237) and other rhabdoviruses belonging to subfamily *Betarhabdovirinae*. The analysis was performed on CIPRES with 1000 bootstraps. Bootstrap values are indicated at each node. Virus abbreviations, names, and sequences are listed in Supplementary Table S5. *Vesicular stomatitis Indiana virus* was used as an outgroup. Scale bar represents amino acid substitution per site. BabRV-1 is marked with an asterisk.

4. Discussion

In the last decade, HTS has advanced the discovery of pathogenic and non-pathogenic viruses in several crop plants around the globe. In vegetatively propagated plants, such as babaco, the identification of viruses constitutes an essential step for sustainable production. Despite previous reports on the occurrence of different viruses in babaco in Ecuador [2,3], a certification scheme in babaco nurseries that prevents the spread of virus-infected material into new production areas has been neglected.

In this study, two new viruses were discovered in babaco plants obtained from a commercial nursery. The viruses, BabIV-1 and BabRV-1, belong to the *Bromoviridae* and *Rhabdoviridae* families, respectively. BabIV-1 has genomic features consistent with those of ilarviruses from subgroup 3 [31]; however, the sequence identity with closest relatives (Table 2) suggests that it should be considered a new member of the genus *Ilarvirus*. BabRV-1 possesses genomic features, including the presence of an accessory gene 'X' of unknown function located between the N and P genes, identical to JYBaV, PYDV, PhCMoV, and eggplant mottled dwarf virus (EMDV), which are members of the *Alphanucleorhabdovirus*. The amino acid sequence identity between BabRV-1 and closest relatives (Table 3) suggests its designation as a distinct new member of this genus.

BabIV-1 and BabRV-1 were found in co-infection with BabVQ, an umbra-like virus considered non-pathogenic [25], and two viruses, BabCV and BabEPV, previously regarded as 'cryptic' due to the lack of symptoms associated with them [3]. It remains to be determined whether the symptomatology observed in the co-infected plant could be attributed to either BabIV-1, BabRV-1, their interaction with each other, with BabVQ, or with additional viruses not detected in this study.

Some plant rhabdoviruses have been reported either as non-pathogenic or able to cause mild symptoms in single infections [12,32,33], whereas ilarviruses have long been associated with an array of leaf or fruit symptoms, including mosaic, vein banding, ringspot, and necrosis, among others [34–37]. A particular symptom, commonly known as ‘necrotic shock’, caused by the ilarviruses blueberry shock virus (BIShV) and strawberry necrotic shock virus (SNSV), consists of severe leaf necrosis followed by defoliation [36,38]. Although blueberry and strawberry plants infected with BIShV and SNSV, respectively, recover after the first year of symptoms, important yield losses can be produced during the infection season, and the plants remain infectious for further virus spread in the field [38]. Babaco plants infected with BabIV-1 and BabRV-1 showed premature leaf abscission (Figure 1) followed by an apparent recovery in a pattern similar to the ‘shock’ disease caused by BIShV in blueberry and SNSV in strawberry. In a survey conducted in babaco plants showing mild leaf mosaic in the same nursery where BabIV-1 and BabRV-1 were first identified, the viruses were found in 21% and 36%, respectively, with 7% of plants being positive for both. Premature leaf abscission was not observed at the time of sampling. Hence, pathogenicity studies using infectious clones are warranted to determine the role of each virus (BabIV-1 and BabRV-1) in single and mix infections in symptom expression in babaco and other model hosts.

As for transmission, being a member of the ilarvirus genus, BabIV-1 is most likely transmitted by pollen. In babaco, however, pollination is rare because fruit set occurs by parthenocarpy [39]. BabRV-1, a nucleorhabdovirus, could be transmitted by arthropod vectors, including leafhoppers, planthoppers, or aphids [12]. During our study, we were unable to establish either leafhopper or aphid colonies onto babaco plants to conduct transmission experiments; hence, the natural vector of BabRV-1 remains to be determined. Nevertheless, in commercial operations, the vegetative propagation nature of babaco is probably more efficient at virus dissemination than vector-mediated transmission.

To the best of our knowledge, this is the first report of an ilarvirus and a nucleorhabdovirus infecting babaco. Our findings contribute to the expanding virome of babaco and highlight the importance of virus discovery studies, which are a pivotal component for the establishment of virus testing and nursery certification programs for vegetatively propagated crops.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/v15061380/s1>, Table S1: Primer information for detection of babaco viruses. Table S2: List of primers used in this study for genome amplification and detection of babaco ilarvirus 1 (BabIV-1) and babaco rhabdovirus 1 (BabRV-1). Table S3: Nucleotide and amino acid identities between open reading frames (ORFs) and their predicted products of babaco ilarvirus-1 (BabIV-1) and counterparts from all ilarviruses currently available at NCBI. Table S4: List of ilarviruses used in phylogenetic analyses. Table S5: List of rhabdoviruses used in phylogenetic analyses.

Author Contributions: Conceptualization, D.F.Q.-A.; methodology, E.G.R.-P., M.G.C.-B., J.F.C.-F. and R.A.A.-Q.; validation, E.G.R.-P., M.G.C.-B. and R.A.A.-Q.; formal analysis, E.G.R.-P., M.G.C.-B. and R.A.A.-Q.; investigation, D.F.Q.-A., E.G.R.-P., M.G.C.-B. and R.A.A.-Q.; resources, D.F.Q.-A., E.S.-T. and D.M.; data curation, E.G.R.-P., M.G.C.-B. and R.A.A.-Q.; writing—original draft preparation, E.G.R.-P. and M.G.C.-B.; writing—and editing D.F.Q.-A. and R.A.A.-Q.; review, E.S.-T. and D.M.; supervision, D.F.Q.-A.; project administration, D.F.Q.-A.; funding acquisition, D.F.Q.-A. and E.S.-T. All authors have read and agreed to the published version of the manuscript.

Funding: This study was funded in part by the Ecuadorian Corporation for the Development of Research and Academia (CEDIA) under project code CEPRA XV-2021-06 and the Research Department at ESPOL.

Institutional Review Board Statement: This study was conducted in accordance with the Department of Biodiversity of the Ecuadorean Ministry of the Environment under Genetic Resource Access Permit # MAE-DNB-CM-2018-0098. This work did not involve studies of humans or animals.

Informed Consent Statement: This work did not involve studies of humans or animals.

Data Availability Statement: Virus genomic sequences generated in this work have been deposited in the GenBank of the National Center for Biotechnology Information (NCBI) under accession numbers OQ256238, OQ256239, OQ256240, and OQ256237.

Acknowledgments: The authors thank the Research Department of ESPOL for additional support and the Department of Biodiversity of the Ecuadorean Ministry of the Environment for granting Genetic Resource Access Permit # MAE-DNB-CM-2018-0098. The authors are also grateful to nursery owners in the Azuay province for sharing plant material and discussions about their work.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

1. Kyndt, T.; Romeijn-Peeters, E.; Van Droogenbroeck, B.; Romero-Motochi, J.P.; Gheysen, G.; Goetghebeur, P. Species relationships in the genus *Vasconcellea* (Caricaceae) based on molecular and morphological evidence. *Am. J. Bot.* **2005**, *92*, 1033–1044. [[CrossRef](#)] [[PubMed](#)]
2. Alvarez-Quinto, R.A.; Cornejo-Franco, J.F.; Quito-Avila, D.F. Characterization of a not so new potexvirus from babaco (*Vasconcellea × heilbornii*). *PLoS ONE* **2017**, *12*, e0189519. [[CrossRef](#)] [[PubMed](#)]
3. Cornejo-Franco, J.F.; Medina-Salguero, A.; Flores, F.; Chica, E.; Grinstead, S.; Mollov, D.; Quito-Avila, D.F. Exploring the virome of *Vasconcellea × heilbornii*: The first step towards a sustainable production program for babaco in Ecuador. *Eur. J. Plant Pathol.* **2020**, *157*, 961–968. [[CrossRef](#)]
4. Villamor, D.E.V.; Ho, T.; Al Rwahnih, M.; Martin, R.R.; Tzanetakis, I.E. High Throughput Sequencing for Plant Virus Detection and Discovery. *Phytopathology* **2019**, *109*, 716–725. [[CrossRef](#)] [[PubMed](#)]
5. Simkovich, A.; Kohalmi, S.E.; Wang, A. Iarviruses (Bromoviridae). In *Encyclopedia of Virology*, 4th ed.; Bamford, D.H., Zuckerman, M., Eds.; Academic Press: Oxford, UK, 2021; pp. 439–446.
6. Bujarski, J.; Gallitelli, D.; García-Arenal, F.; Pallás, V.; Palukaitis, P.; Reddy, M.K.; Wang, A. ICTV Report Consortium, ICTV Virus Taxonomy Profile: Bromoviridae. *J. Gen. Virol.* **2019**, *100*, 1206–1207. [[CrossRef](#)]
7. Xin, H.W.; Ji, L.H.; Scott, S.W.; Symons, R.H.; Ding, S.W. Iarviruses encode a Cucumovirus-like 2b gene that is absent in other genera within the *Bromoviridae*. *J. Virol.* **1998**, *72*, 6956–6959. [[CrossRef](#)]
8. Pallas, V.; Aparicio, F.; Herranz, M.C.; Sanchez-Navarro, J.A.; Scott, S.W. The molecular biology of ilarviruses. *Adv. Virus Res.* **2013**, *87*, 139–181.
9. Klose, M.J.; Sdoodee, R.; Teakle, D.S.; Milne, J.R.; Greber, R.S.; Walter, G.H. Transmission of Three Strains of Tobacco Streak Iarvirus by Different Thrips Species Using Virus-infected Pollen. *J. Phytopathol.* **1996**, *144*, 281–284. [[CrossRef](#)]
10. Burrell, C.J.; Howard, C.R.; Murphy, F.A. (Eds.) Chapter 27—Rhabdoviruses. In *Fenner and White's Medical Virology*, 5th ed.; Academic Press: London, UK, 2017; pp. 383–394.
11. Bejerman, N.; Dietzgen, R.G.; Debat, H. Illuminating the Plant Rhabdovirus Landscape through Metatranscriptomics Data. *Viruses* **2021**, *13*, 1304. [[CrossRef](#)]
12. Gaafar, Y.Z.A.; Richert-Pöggeler, K.R.; Maaß, C.; Vetten, H.J.; Ziebell, H. Characterisation of a novel nucleorhabdovirus infecting alfalfa (*Medicago sativa*). *Virol. J.* **2019**, *16*, 16–55. [[CrossRef](#)]
13. Dietzgen, R.G.; Kondo, H.; Goodin, M.M.; Kurath, G.; Vasilakise, N. The family *Rhabdoviridae*: Mono- and bipartite negative-sense RNA viruses with diverse genome organization and common evolutionary origins. *Virus Res.* **2017**, *227*, 158–170. [[CrossRef](#)]
14. Walker, P.J.; Dietzgen, R.G.; Joubert, D.A.; Blasdel, K.R. Rhabdovirus accessory genes. *Virus Res.* **2011**, *162*, 110–125. [[CrossRef](#)]
15. Halgren, A.; Tzanetakis, I.E.; Martin, R.R. Identification, Characterization, and Detection of Black raspberry necrosis virus. *Phytopathology* **2007**, *97*, 44–50. [[CrossRef](#)] [[PubMed](#)]
16. Thompson, J.R.; Wetzell, S.; Klerks, M.M.; Vasková, D.; Schoen, C.D.; Spak, J.; Jelkmann, W. Multiplex RT-PCR detection of four aphid-borne strawberry viruses in *Fragaria* spp. in combination with a plant mRNA specific internal control. *J. Virol. Methods* **2003**, *111*, 85–93. [[CrossRef](#)]
17. Tzanetakis, I.E.; Postman, J.D.; Martin, R.R. Identification, detection and transmission of a new vitivirus from *Mentha*. *Arch. Virol.* **2007**, *152*, 2027–2033. [[CrossRef](#)]
18. Bankevich, A.; Nurk, S.; Antipov, D.; Gurevich, A.A.; Dvorkin, M.; Kulikov, A.S.; Lesin, V.M.; Nikolenko, S.I.; Pham, S.; Prjibelski, A.D.; et al. SPAdes: A new genome assembly algorithm and its applications to single-cell sequencing. *J. Comput. Biol.* **2012**, *19*, 455–477. [[CrossRef](#)] [[PubMed](#)]
19. Dodds, J.A.; Morris, T.J.; Jordan, R.L. Plant Viral Double-Stranded RNA. *Annu. Rev. Phytopathol.* **1984**, *22*, 151–168. [[CrossRef](#)]
20. Okonechnikov, K.; Golosova, O.; Fursov, M.; the UGENE team. Unipro UGENE: A unified bioinformatics toolkit. *Bioinformatics* **2012**, *28*, 1166–1167. [[CrossRef](#)] [[PubMed](#)]
21. Edgar, R.C. MUSCLE: Multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* **2004**, *32*, 1792–1797. [[CrossRef](#)]
22. Kumar, S.; Stecher, G.; Li, M.; Niyaz, C.; Tamura, K. MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Mol. Biol. Evol.* **2018**, *35*, 1547–1549. [[CrossRef](#)]

23. Stamatakis, A. RAxML version 8: A tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* **2014**, *30*, 1312–1313. [[CrossRef](#)] [[PubMed](#)]
24. Hull, R. Mechanical Inoculation of Plant Viruses. *Curr. Protoc. Microbiol* **2009**, *13*, 16B.6.1–16B.6.4. [[CrossRef](#)] [[PubMed](#)]
25. Cornejo-Franco, J.F.; Flores, F.; Quito-Avila, D.F. An umbra-related virus found in babaco (*Vasconcellea × heilbornii*). *Arch. Virol.* **2021**, *166*, 2321–2324. [[CrossRef](#)] [[PubMed](#)]
26. Orfanidou, C.G.; Xing, F.; Zhou, J.; Li, S.; Katis, N.I.; Maliogka, V.I. Identification and Sequence Analysis of a Novel Ilarvirus Infecting Sweet Cherry. *Plants* **2021**, *10*, 514. [[CrossRef](#)]
27. Gallo-García, Y.M.; Jaramillo-Mesa, H.; Toro-Fernández, L.F.; Marín-Montoya, M.; Gutiérrez, P.A. Characterization of the genome of a novel ilarvirus naturally infecting Cape gooseberry (*Physalis peruviana*). *Arch. Virol.* **2018**, *163*, 1713–1716. [[CrossRef](#)]
28. Li, J.; Rahmeh, A.; Morelli, M.; Whelan, S.P. A conserved motif in region v of the large polymerase proteins of nonsegmented negative-sense RNA viruses that is essential for mRNA capping. *J. Virol.* **2008**, *82*, 775–784. [[CrossRef](#)]
29. Easton, A.J.; Ling, R. Mononegavirales. In *Encyclopedia of Virology*, 3rd ed.; Mahy, B.W.J., Van Regenmortel, M.H.V., Eds.; Academic Press: Oxford, UK, 2008; pp. 324–334.
30. Walker, P.J.; Freitas-Astúa, J.; Bejerman, N.; Blasdel, K.R.; Dietzgen, R.G.; Fooks, A.R.; Kondo, H.; Kurath, G.; Ramos-González, P.L.; Shi, M.; et al. ICTV Virus Taxonomy Profile: *Rhabdoviridae*. *J. Gen. Virol.* **2022**, *103*, 001689. [[CrossRef](#)]
31. James, D.; Varga, A.; Leippi, L.; Godkin, S.; Masters, C. Sequence analysis of RNA 2 and RNA 3 of lilac leaf chlorosis virus: A putative new member of the genus *Ilarvirus*. *Arch. Virol.* **2010**, *155*, 993–998. [[CrossRef](#)]
32. Medina-Salguero, A.X.; Cornejo-Franco, J.F.; Grinstead, S.; Mollov, D.; Mowery, J.D.; Flores, F.; Quito-Avila, D.F. Sequencing, genome analysis and prevalence of a cytorhabdovirus discovered in *Carica papaya*. *PLoS ONE* **2019**, *14*, e0215798. [[CrossRef](#)]
33. Reyes-Proañó, E.; Alvarez-Quinto, R.; Delgado-Jiménez, J.A.; Cornejo-Franco, J.F.; Mollov, D.; Bejerman, N.; Quito-Avila, D.F. Genome Characterization and Pathogenicity of Two New *Hyptis pectinata* Viruses Transmitted by Distinct Insect Vectors. *Phytopathology* **2022**, *112*, 2440–2448. [[CrossRef](#)]
34. Pallas, V.; Aparicio, F.; Herranz, M.C.; Amari, K.; Sanchez-Pina, M.A.; Myrta, A.; Sanchez-Navarro, J.A. Ilarviruses of *Prunus* spp.: A continued concern for fruit trees. *Phytopathology* **2012**, *102*, 1108–1120. [[PubMed](#)]
35. Parrella, G.; Troiano, E. A New Ilarvirus Found in French Hydrangea. *Plants* **2022**, *11*, 944. [[CrossRef](#)] [[PubMed](#)]
36. Tzanetakis, I.E.; Mackey, I.C.; Martin, R.R. Strawberry necrotic shock virus is a distinct virus and not a strain of *Tobacco streak virus*. *Arch. Virol.* **2004**, *149*, 2001–2011. [[CrossRef](#)]
37. Kozieł, E.; Bujarski, J.J.; Otulak, K. Molecular Biology of *Prune Dwarf Virus*—A Lesser Known Member of the *Bromoviridae* but a Vital Component in the Dynamic Virus–Host Cell Interaction Network. *Int. J. Mol. Sci.* **2017**, *18*, 2733. [[CrossRef](#)] [[PubMed](#)]
38. Thomas-Sharma, S.; Wells-Hansen, L.; Page, R.; Kartanos, V.; Saalau-Rojas, E.; Lockhart, B.E.L.; McManus, P.S. Characterization of *Blueberry shock virus*, an Emerging *Ilarvirus* in Cranberry. *Plant Dis.* **2018**, *102*, 91–97. [[CrossRef](#)] [[PubMed](#)]
39. Van Droogenbroeck, B.; Kyndt, T.; Romeijn-Peeters, E.; Van Thuyne, W.; Goetghebeur, P.; Romero-motochi, J.P.; Gheysen, G. Evidence of Natural Hybridization and Introgression between *Vasconcellea* Species (*Caricaceae*) from Southern Ecuador Revealed by Chloroplast, Mitochondrial and Nuclear DNA Markers. *Ann. Bot.* **2006**, *97*, 793–805. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.