

Article

Different Nitrate and Ammonium Ratios Affect Growth and Physiological Characteristics of *Camellia oleifera* Abel. Seedlings

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Abstract: *Camellia oleifera* Abel., is an important woody plant, and its fruit contains high-quality edible oil. Production of *C. oleifera* has significantly expanded over the last 20 years in China. Due to the lack of appropriate information on nutrient management, its production has encountered low yield and low oil quality problems. As nitrogen (N) is an essential nutrient and the most abundant in *C. oleifera* tissues, the present study investigated effects of different ratios of nitrate (NO_3^-) and ammonium (NH_4^+) on the growth of a cultivar Xianglin 27 at the seedling stage. Uniform seedlings were grown in a soil-based substrate in containers and fertigated with solutions composed of six ratios of NO_3^- and NH_4^+ , respectively for five months. Results showed that *C. oleifera* prefers both NO_3^- and NH_4^+ at a ratio of 1:1. Seedlings receiving this solution had the highest total N in leaves and total dry weight; elevated chlorophyll, soluble saccharide and protein contents as well as higher activities of peroxidase (POD), superoxide dismutase (SOD), nitrate reductase (NR), glutamine synthetase (GS), and glutamate synthase (GOGAT). Our study shows for the first time that N supply for producing *C. oleifera* should be an equal ratio of NO_3^- and NH_4^+ .

Keywords: Nitrate and ammonium ratios; *Camellia oleifera*; nitrogen forms; tea oil trees

1. Introduction

Camellia oleifera Abel., a member of the Theaceae, is an oil tree species that has been exclusively cultivated in China due to its economic value [1]. Its fruit contains high-quality oil, commonly known as tea oil or camellia oil, which has 82%–84% unsaturated fatty acids, 68%–77% monounsaturated fatty acids, 67.7%–76.7% oleic acid, and 7%–14% polyunsaturated acid. The composition is similar to that of olive oil [2], hence it is called eastern olive oil [3]. Tea oil has also been widely used for cosmetic and medicinal purposes [4]. Furthermore, *C. oleifera* is tolerant to drought [5], and it can be produced in a wide range of mountainous areas in subtropical regions. With the increasing demand for self-sufficiency in cooking oil [3], the production of *C. oleifera* has significantly increased over the last 20 years. The current cultivated area for *C. oleifera* is more than three million ha in China [6,7].

The increasing production scope of tea oil trees, however, has encountered several problems including the lack of appropriate nutrient management programs, low crop yield, and variable oil quality [1,8]. Plant growth is highly dependent on mineral nutrient supply [9]. Among the essential nutrients, nitrogen (N) is one of the most important macrolelements that significantly affects plant growth, development, and product quality [10]. Analysis of distribution of nutrients in different organs of *C. oleifera* showed the most abundant element is N ranging from 3.2 to 12 g kg⁻¹ compared to the next most abundant, potassium, ranging from 3 to 6 g kg⁻¹ [8]. Thus, N is critically important for the growth of *C. oleifera*. Commercially, N is primarily applied in the form of nitrate (NO₃⁻), ammonium (NH₄⁺) or urea (CO(NH₂)₂). Applied urea will be converted to NH₄⁺ in soils then absorbed by plants, meaning that urea, although it is an organic N source, is considered the same as NH₄⁺ during the absorption process [11]. A large body of evidence suggests that both rates and forms of N fertilizers can affect plant growth and development [11–15]. In general, NO₃⁻ is considered to be the main N source for most crops [16], whereas plants adapted to acidic soils prefer NH₄⁺-N and those adapted to high pH soils prefer NO₃⁻-N [9].

Current fertilizer programs for *C. oleifera* production have focused on the formulation of N, P₂O₅, and K₂O ratios and application rates [17,18]. Little attention has been given to the forms of N or the ratio of N forms on *C. oleifera* growth. As *C. oleifera* has adapted to red-acidic soils with pH around 5 [19], we hypothesized that *C. oleifera* would prefer NH₄⁺ over NO₃⁻ for its growth. The objective of this study was to test this hypothesis using uniform seedlings of *C. oleifera* as a model. Seedlings were fertilized with solutions comprised of six ratios of NO₃⁻ and NH₄⁺ for five months. Plant growth parameters and physiological characteristics were evaluated. Contrary to our hypothesis, *C. oleifera* prefers both NO₃⁻ and NH₄⁺ as N sources at a ratio of 1:1.

2. Materials and Methods

2.1. Experiment Site

The experiment was conducted at the experimental station of the National Engineering Research Center for Oil-tea *Camellia*, Changsha city, Hunan province, China (113°01' E and 28°06' N). The location has a subtropical monsoon climate with mild temperatures and distinct seasons. The annual lowest, highest, and average temperatures are -12 °C, 40.6 °C, and 17.3 °C, respectively. Annual average rainfall is approximately 1422 mm, the frost-free period is 275 days, and the annual average relative humidity is 80%.

2.2. Plant Materials and Experimental Treatments

Uniform seedlings of *Camellia oleifera* 'Xianglin 27' were grown from seeds in early March 2015 and transplanted to containers filled with a soil-based substrate (medium). The substrate was composed of yellow earth soil with sand, silt, and clay in a ratio of 2:3:1, perlite, and peat in a 3:1:1 ratio based on volume and had a pH of 5.8. The containers had a volume of 0.6 L with a height of 12 cm and diameter of 8 cm. Seedlings were grown in a greenhouse with a maximum photosynthetic radiation of 1000 μmol m⁻² s⁻¹, temperature ranging from 20 to 25 °C, and relative humidity of 50%–80%.

To determine effects of different ratios of NO₃⁻ and NH₄⁺ on *C. oleifera* growth, six solutions were prepared weekly and used for fertigation (i.e., both fertilization and irrigation occurred at the same time) of seedlings. All solutions consisted of 261.39 mg·L⁻¹ K₂SO₄, 136.09 mg·L⁻¹ KH₂PO₄, 221.98 mg·L⁻¹ CaCl₂, 120.37 mg·L⁻¹ MgSO₄, 1.38 mg·L⁻¹ MnSO₄, 2.86 mg·L⁻¹ H₃BO₃, 0.12 mg·L⁻¹ ZnSO₄, 0.05 mg·L⁻¹ CuSO₄, 0.017 mg·L⁻¹ Na₂MoO₄, 10.94 mg·L⁻¹ FeSO₄, and 7 μmol C₂H₄N₄ (cyanoguanidine, a nitrification inhibitor) but varied in NO₃⁻ and NH₄⁺ ratios as indicated in Table 1. N concentration in each solution was 8 mM except the control treatment (T0) devoid of N.

Table 1. Six ratios of NO_3^- and NH_4^+ used for studying effects of nitrogen forms on *Camellia oleifera* ‘Xianglin 27’ seedling growth.

Treatment	$\text{NO}_3^-:\text{NH}_4^+$ Ratio	Total N Concentration (mM)	NaNO_3 (mM)	$(\text{NH}_4)_2\text{SO}_4$ (mM)
T0	0:0	0.0	0.0	0.0
T1	1:0	8.0	8.0	0.0
T2	7:3	8.0	5.6	1.2
T3	1:1	8.0	4.0	2.0
T4	3:7	8.0	2.4	2.8
T5	0:1	8.0	0.0	4.0

After three months of growth, disease and pest free seedlings were selected and arranged in a randomized complete block design with three replications, and each replication had 10 seedlings per treatment. Seedlings were fertigated with six nutrient solutions, respectively. The fertigation started in mid-June 2015, at a rate of 150 mL per container weekly. If additional irrigation was needed, seedlings were irrigated with tap water. Besides fertigation, plants were well managed by pulling any weeds by hand.

2.3. Seedling Growth Measurements

Canopy heights of seedlings and stem diameters at the ground level were recorded using rule and Vernier caliper before fertigation and at the end of fertigation (November 2015). Canopy and stem increases were calculated based on their differences between before and after fertigation. Before the conclusion of the experiment, leaf samples were collected for analysis of physiological parameters indicated below. Seedlings were then harvested by cutting shoots from the substrate surface, and roots were recovered by inverting the containers in water, gently agitating the root system and washing the roots free from the substrate. Roots and shoots were placed in paper bags, initially dried at 105 °C for 30 min, and then dried to a constant mass at 80 °C. Root and shoot dry weights were recorded.

2.4. Analysis of Physiological Parameters

Total N contents in leaves of seedlings were determined using the Kjeldahl method [20]. For analysis of chlorophyll content, enzymatic activity, and content of soluble saccharides and proteins, leaf samples collected from each replication were immediately weighed and frozen in liquid N. Chlorophyll a and b (Chl a and Chl b) were analyzed using the acetone-ethanol-mixture method [21], soluble saccharides were assayed by anthrone colorimetry [22,23], soluble proteins were tested using the Coomassie brilliant-blue G-250 staining method [24], peroxidase (POD) activity was examined using the guaiacol method [25], superoxide dismutase (SOD) activity was analyzed by the nitroblue-tetrazolium photoreduction method [26], nitrate reductase activity was determined by spectrophotometry [27], and activities of N assimilation enzymes including nitrate reductase (NR), glutamine synthetase (GS), and glutamate synthase (GOGAT) were assayed using spectrophotometry methods [22,28]. All the analyses had three replicates.

2.5. Data Analysis

Plant growth data, dry weights, and physiological parameters mentioned above were subjected to analysis of variance using SPSS 13.0 for Windows (SPSS, Chicago, IL, USA). When significant differences occurred among treatments per parameter, means were separated using Fisher’s Protected Least Significant Differences (LSD) at $p < 0.05$ level.

3. Results

Seedlings fertigated with different ratios of NO_3^- and NH_4^+ exhibited significant differences in plant canopy heights, stem diameters as well as dry weights and physiological parameters.

3.1. Plant Growth and Dry Weights

Plant growth results including increase in canopy heights and stem diameters as well as shoot and root dry weights, are presented in Table 2. Nutrient solutions with both NO_3^- and NH_4^+ enhanced seedling growth. The greatest increase in canopy heights was observed in seedlings fertigated with an equal ratio of NO_3^- to NH_4^+ , which significantly differed from those supplied with T1, T2, and T5 solutions. The equal ratio of NO_3^- to NH_4^+ also resulted in the greatest increase in stem diameter as well as shoot, root, and total dry weight except for root dry weight of seedlings fertigated with T4 solution which was statistically similar to the seedlings fertigated with T3 solution.

Table 2. The growth and biomass accumulation of *Camellia oleifera* ‘Xianglin 27’ seedlings fertigated with six nutrient solutions varying in NO_3^- to NH_4^+ ratios.

Treatment	Canopy Height Increase (cm)	Stem Diameter Increase (mm)	Shoot Dry Weight (g)	Root Dry Weight (g)	Total Dry Weight (g)
T0 (0:0)	5.20 ± 0.52 c	1.43 ± 0.04 c	0.83 ± 0.01 e	0.68 ± 0.03 d	1.52 ± 0.03 e
T1 (1:0)	7.83 ± 1.07 b	1.67 ± 0.12 b	1.11 ± 0.06 d	0.82 ± 0.05 c	1.93 ± 0.11 d
T2 (7:3)	9.03 ± 0.23 ab	1.86 ± 0.06 ab	1.25 ± 0.06 c	0.90 ± 0.06 b	2.16 ± 0.10 c
T3 (1:1)	10.53 ± 0.55 a	2.00 ± 0.18 a	1.49 ± 0.05 a	1.05 ± 0.05 a	2.54 ± 0.10 a
T4 (3:7)	8.97 ± 1.72 ab	1.71 ± 0.08 b	1.35 ± 0.04 b	0.99 ± 0.01 a	2.34 ± 0.05 b
T5 (0:1)	7.10 ± 1.22 b	1.58 ± 0.17 bc	1.17 ± 0.07 cd	0.83 ± 0.02 c	2.00 ± 0.09 cd

Different letters within a column represent significant difference at $p < 0.05$ level tested by Fisher’s Protected Least Significant Differences (LSD).

3.2. Leaf Total Nitrogen Content

Total N contents of leaves increased from 8.6 g kg^{-1} in seedlings fertigated with a solution devoid of N (T0) to 13.5 g kg^{-1} in seedlings supplied with an equal ratio of NO_3^- to NH_4^+ (T3) and then decreased when seedlings were treated with NO_3^- and NH_4^+ at a ratio of 3:7 (Figure 1). There were no significant differences in total N content among seedlings fertigated with T0, T1 or T5 solutions.

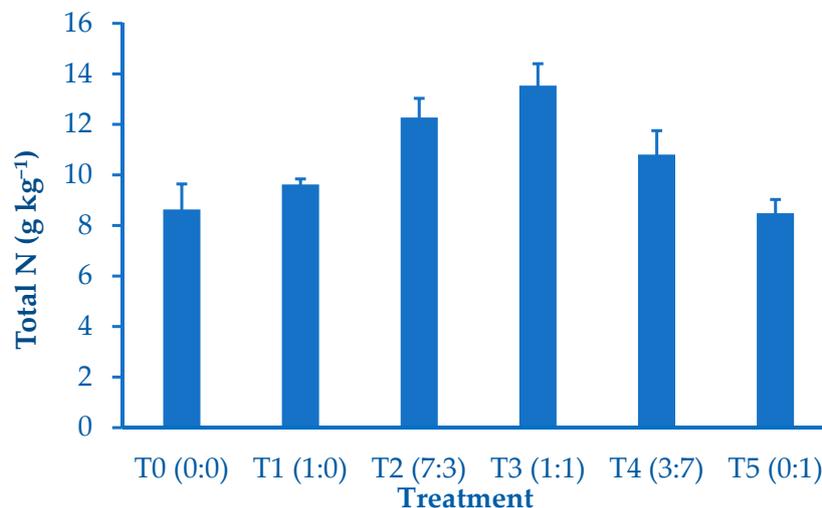


Figure 1. Total nitrogen content in leaves of *Camellia oleifera* ‘Xianglin 27’ seedlings fertigated with six nutrient solutions varying in NO_3^- to NH_4^+ ratios.

3.3. Chlorophyll Content

Chlorophyll (Chl) a contents were about three-fold higher than chl b in *C. oleifera* ‘Xianglin 27’ leaves regardless of treatments. However, chl a and b as well as total chl content were significantly affected by fertigation (Table 3). Seedlings fertigated with an equal ratio of NO_3^- and NH_4^+ had chl a, b, and total chl contents comparable to those fertigated with a solution with a 3:7 ratio of NO_3^- : NH_4^+

(T4). Leaves of seedlings supplied with an equal ratio (T3) of NO_3^- and NH_4^+ had significantly higher chl a and total chl than T0, T1, T2, and T5 and higher chl b than T0, T2, and T5.

Table 3. Leaf chlorophyll content based on fresh weight of *Camellia oleifera* ‘XL27’ seedlings fertigated with six nutrient solutions varying in NO_3^- to NH_4^+ ratios.

Treatment	Chl a ($\text{mg}\cdot\text{g}^{-1}$ FW)	Chl b ($\text{mg}\cdot\text{g}^{-1}$ FW)	Total chl ($\text{mg}\cdot\text{g}^{-1}$ FW)
T0 (0:0)	0.95 ± 0.11 b *	0.31 ± 0.03 d	1.26 ± 0.14 b
T1 (1:0)	0.99 ± 0.09 b	0.35 ± 0.02 abc	1.34 ± 0.09 b
T2 (7:3)	0.97 ± 0.03 b	0.33 ± 0.02 bcd	1.30 ± 0.05 b
T3 (1:1)	1.41 ± 0.17 a	0.36 ± 0.01 a	1.76 ± 0.17 a
T4 (3:7)	1.08 ± 0.04 ab	0.36 ± 0.01 a	1.44 ± 0.05 ab
T5 (0:1)	0.97 ± 0.03 b	0.32 ± 0.01 cd	1.29 ± 0.04 b

* The values represent mean chlorophyll (chl). Different letters within a column represent significant differences at a level of $p < 0.05$ tested using Fisher’s LSD.

3.4. Peroxidase (POD) and Superoxide Dismutase (SOD) Activities

Seedlings fertigated with an equal ratio of NO_3^- and NH_4^+ exhibited the highest activities of both SOD and POD (Table 4). But the SOD activity in T3-treated leaves was not statistically different from the other treatments except for that of T0 that was much lower than that of T3 treatment. The POD activities in leaves of seedlings receiving T2 and T3 solutions were similar, but POD activity in leaves of seedlings supplied with T3 was significantly greater than the other treatments.

Table 4. Peroxidase (POD) and Superoxide Dismutase (SOD) activities in leaves of *Camellia oleifera* ‘Xianglin 27’ seedlings fertigated with six nutrient solutions varying in NO_3^- to NH_4^+ ratios.

Treatment	SOD ($\text{U}\cdot\text{g}^{-1}$ FW)	POD ($\text{U}\cdot\text{g}^{-1}$ FW $\cdot\text{min}^{-1}$)
T0 (0:0)	314.64 ± 14.37 b	754.29 ± 248.79 b
T1 (1:0)	334.09 ± 12.02 ab	950.04 ± 252.69 b
T2 (7:3)	356.66 ± 6.54 ab	1195.94 ± 385.13 ab
T3 (1:1)	367.01 ± 15.63 a	1579.03 ± 341.98 a
T4 (3:7)	359.07 ± 0.80 ab	982.99 ± 108.93 b
T5 (0:1)	334.50 ± 28.31 ab	814.91 ± 224.00 b

Different letters within a column represent significant differences at a level of $p < 0.05$ tested using Fisher’s LSD.

3.5. Nitrate Reductase (NR), Glutamine Synthetase (GS), and Glutamate Synthase (GOGAT) Activities

The application of T3 nutrient solution resulted in the highest activity of NR although it was not significantly different from T1 and T2 treatments; it did differ significantly from T4 and T5 (Table 5). The highest activity of GS was detected in leaves of seedlings fertigated with T2 (NO_3^- to NH_4^+ at a ratio of 7:3), followed by those treated with T3 and T4 solutions. Activities of GS in seedlings fertigated with the remaining solutions were much lower than the mentioned solutions. In general, increasing the ratio of NH_4^+ caused increased activities of GOGAT in seedling leaves. Seedlings fertigated without NH_4^+ had significantly lower activities of GOGAT than those fertigated with NH_4^+ .

Table 5. Nitrate Reductase (NR), Glutamine Synthetase (GS), and Glutamate Synthase (GOGAT) activities in leaves of *Camellia oleifera* ‘Xianglin 27’ seedlings fertigated with six nutrient solutions varying in NO_3^- to NH_4^+ ratios.

Treatment	NR ($\mu\text{g}\text{NO}_3^- \cdot \text{g}^{-1}$ FW h^{-1})	GS ($\mu\text{mol}\cdot\text{g}^{-1}$ FW h^{-1})	GOGAT ($\mu\text{mol}\cdot\text{g}^{-1}$ FW h^{-1})
T0 (0:0)	29.43 ± 1.75 c	32.97 ± 0.75 d	13.80 ± 0.64 d
T1 (1:0)	36.12 ± 4.49 ab	44.77 ± 2.88 c	17.60 ± 1.24 c
T2 (7:3)	39.06 ± 1.67 ab	61.84 ± 2.98 a	21.26 ± 2.23 b
T3 (1:1)	42.51 ± 4.95 a	57.65 ± 2.28 b	25.48 ± 1.10 a
T4 (3:7)	32.98 ± 4.82 bc	54.43 ± 0.68 b	26.65 ± 1.11 a
T5 (0:1)	29.05 ± 2.53 c	48.02 ± 1.31 c	25.50 ± 0.75 a

Different letters within a column represent significant differences at a level of $p < 0.05$ tested using Fisher’s LSD.

3.6. Contents of Soluble Saccharides and Soluble Proteins

Soluble saccharide and protein contents generally increased with the increased ratios of NH_4^+ , and then decreased when NH_4^+ supply was above 50% (Table 6). The highest soluble saccharides occurred in leaves of seedlings fertigated with an equal ratio of NO_3^- and NH_4^+ . The highest soluble proteins occurred in leaves of seedlings fertigated with either T3 or T4 nutrient solutions, which were significantly different from those of the other treatments.

Table 6. Soluble saccharide and protein contents in leaves of *Camellia oleifera* ‘Xianglin 27’ seedlings fertigated with six nutrient solutions varying in NO_3^- to NH_4^+ ratios.

Treatment	Saccharides (% FW)	Soluble Proteins ($\text{mg}\cdot\text{g}^{-1}$ FW)
T0 (0:0)	3.92 ± 0.33 c	7.34 ± 0.81 d
T1 (1:0)	4.62 ± 0.30 b	9.99 ± 0.65 c
T2 (7:3)	4.96 ± 0.43 b	11.57 ± 0.90 b
T3 (1:1)	5.68 ± 0.32 a	14.27 ± 0.32 a
T4 (3:7)	5.05 ± 0.26 b	13.72 ± 0.83 a
T5 (0:1)	4.65 ± 0.24 b	11.18 ± 0.54 bc

Different letters within a column represent significant differences at a level of $p < 0.05$ tested using Fisher’s LSD.

4. Discussion

Camellia oleifera is one of four major woody oil plants in the world. Its production has significantly increased over the last 20 years in China and is expected to expand from 3.67 million ha to 4.67 million ha by 2020 [29]. The increased production requires science-based information on nutrient management. N is an essential nutrient for plant growth, and large quantities of N fertilizers are applied to ensure high crop productivity [30]. Commercial N fertilizers are comprised of NO_3^- , NH_4^+ or $\text{CO}(\text{NH}_2)_2$. N form or the ratios of NO_3^- and NH_4^+ can significantly affect plant growth and development [31]. How different ratios of NO_3^- and NH_4^+ could affect *C. oleifera* growth has not been investigated thus far. The present work studied the effects of different ratios of NO_3^- and NH_4^+ on seedling growth of a *C. oleifera* cultivar Xianglin 27. Results showed that canopy height and stem diameter increased, and shoot and root dry weights were the highest for the seedlings fertigated with the solution containing an equal ratio of NO_3^- and NH_4^+ . The increased plant biomass was accompanied with enhanced uptake of N, higher chlorophyll content, increased activity of SOD, POD, NR, GS, and GOGAT as well as the highest concentration of saccharides and soluble proteins in leaf tissue studied.

Our results differ from the conventional thought that plants adapted to acidic soils prefer NH_4^+ -N. This is based on the fact that NH_4^+ has a positive charge, and when roots take up NH_4^+ , an identically charged molecule, in this case H^+ , is released from roots to maintain a balanced pH inside the plant cells. The release of H^+ keeps the rhizosphere pH at acidic levels, thus maintaining a suitable pH environment for root growth. Another reason is that NH_4^+ is generally a major form of N in acidic soils. In a study of N forms and root-zone pH on growth and N uptake of tea (*Camellia sinensis* (L.) O. Kuntze) plants, a close relative of *C. oleifera* that is also tolerant to acidic soils, Ruan et al. [32] reported that plants fertigated with NO_3^- exhibited yellowish leaves and reduced growth compared to those receiving NH_4^+ or NO_3^- and NH_4^+ regardless of root-zone pH. Absorption of NO_3^- was 2 to 3.4 fold slower than NH_4^+ when supplied separately, and 6 to 16 fold slower when supplied simultaneously. Additionally, supply of NO_3^- reduced chlorophyll content and GS activity [32]. Coffee plants (*Coffea arabica* L.) fed with NH_4^+ also absorbed and assimilated more N than plants fed with NH_4^+ and NO_3^- or NO_3^- only [33]. Our study did not examine the specific absorption rate of NO_3^- and NH_4^+ but showed that seedling growth of *C. oleifera* was the greatest when NO_3^- and NH_4^+ were applied at the 1:1 ratio, and NO_3^- and NH_4^+ applied separately had much lower dry weight accumulation and lower contents of total N, chlorophyll, soluble saccharides and proteins as well as lower enzymatic activities of POD, SOD, NR, GS, and GOGAT. Our results agreed with those for *C. sinensis* that application of both NH_4^+ and NO_3^- increased N uptake and increased biomass

accumulation [32] but differed from those that held that application of NH_4^+ alone was equal or better than the application of NH_4^+ or NO_3^- .

Recently, increasing evidence has suggested that mixtures of NO_3^- and NH_4^+ are beneficial for plant growth compared to NO_3^- or NH_4^+ alone [31], a finding which may be attributed to several factors: (1) Nitrate is a nutrient, and plants have developed a whole set of systems for uptake, transport, and assimilation of NO_3^- . (2) There is a complementary effect between NO_3^- and NH_4^+ as reported by Lima et al. [34]. Application of NH_4^+ stimulates lateral root branching, whereas NO_3^- stimulates lateral root elongation. When NO_3^- and NH_4^+ are applied together, concomitant enhancement of branching and elongation of lateral roots occurs, suggesting that the application of NO_3^- and NH_4^+ has local, complementary effects on root growth [31]. (3) The presence of NO_3^- enhances NH_4^+ uptake and decreases NH_4^+ efflux. A study with rice (*Oryza sativa* L.) showed that an increased total NH_4^+ uptake occurred concomitantly with a decrease in total NH_4^+ efflux in the presence of NO_3^- , thus enhancing net NH_4^+ uptake [35]. (4) Nitrate is a local and systemic signal molecule. A wide range of genes can be regulated by NO_3^- , which significantly affect plant growth and development [36]. Although the present study did not investigate N effects at molecular levels, our results suggest that the increased uptake of N in seedlings fertigated with an equal ratio of NO_3^- and NH_4^+ could enhance NR and GS/GOGAT cycle, increase chlorophyll content and Rubisco activity and thus raise soluble saccharides and proteins, and subsequently plant growth, i.e., dry matter accumulation. Additionally, SOD and POD can scavenge reactive oxygen species (ROS), protect cell membrane structure, and maintain the ROS metabolism balance in plants. The increased activities of SOD and POD may safeguard plants from ROS damage. (5) Root-associated microbes could be another factor influencing N uptake, such as mycorrhizal fungi [37]. Roots of *Rhododendron fortunei* Lindl., also an acid-loving plant, can effectively use NO_3^- due to the presence of ericoid mycorrhizal fungus (*Oidiodendron maius*) that can fully utilize NO_3^- [38,39]. It is unknown if *C. sinensis* and *C. oleifera* have different rhizosphere mycorrhizal fungi or other microbes. Nevertheless, further research to test the aforementioned possibilities is needed, and information gathered from such tests will improve current nutrient management programs for *C. oleifera* production.

5. Conclusions

Camellia oleifera is a traditional woody oil plant in China and has been cultivated for more than 2000 years. Recent recognition of its healthy value as a cooking oil and its adaptation to a wide range of subtropical forest soils has renewed interest in its cultivation. As *C. oleifera* is an acid-loving plant, farmers have been using chemical fertilizers with NH_4^+ or urea as main sources of N to produce this crop. As a result, production of *C. oleifera* based on farmers' experience has not resulted in significant improvement in crop yield. Thus, there is an urgent need for science-based information on nutrient management. This study determined how different ratios of NO_3^- and NH_4^+ could affect growth of *C. oleifera* seedlings. Results show that *C. oleifera* seedlings prefer both NO_3^- and NH_4^+ with a ratio of 1:1. Seedlings receiving NO_3^- and NH_4^+ at this ratio had highest total N content and dry weight accumulation; elevated chlorophyll, soluble saccharide and protein contents as well as higher activities of POD, SOD, NR, GS, and GOGAT. Our results show that growth of *C. oleifera* seedlings is enhanced using fertilizers with an equal ratio of NO_3^- and NH_4^+ . Further study on evaluating mature plant's responses to different ratios of NO_3^- and NH_4^+ will be conducted to determine if they also require such a ratio of NO_3^- and NH_4^+ .

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