

SDPhound, a Mutual Information-Based Method to Investigate Specificity-Determining Positions

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1. XLS and HTML reports

Figure 1. Snapshot of the Excel worksheet created by SDPhound in the case of a single position run.

Results of the application of the algorithm are reported in various forms, one of them is an Excel Worksheet that contains all the relevant information related to the run. Best ranking positions and run parameters are shown as well as the estimation of the conditional probability of belonging to a specificity class given that any specific symbol, amino acid or “pigeonhole”, is found at the current position. Conditional probability is estimated from the alignment itself in the frequentist approximation. In case where the identity substitution matrix is used, a “-1.0” can appear in the conditional probability cells, indicating that the specific amino acid is not present in the alignment and therefore no information can be directly inferred for that mutation. Two typical examples are shown in Figure 1 and 2.

2. IFP Alignment

The complete alignment of the FPs is reported in the supplementary file A_FP.txt.

3. Physically based pigeonholes

The assignment of the residues to specific pigeonholes was performed according to the following groupings, derived from classical Taylor's Venn diagram [1]:

Hydrophobicity:

Pigeonhole W, hydrophilic aminoacids: R N D Q E G K P S T ;

Pigeonhole N, neutral aminoacids: A H;

Pigeonhole H, hydrophobic aminoacids: C I L M F W Y V.

Size:

Pigeonhole 1, extra extra small aminoacids: G;

Pigeonhole 2, extra small aminoacids: A S;

Pigeonhole 3, small aminoacids: C D P N T;

Pigeonhole 4, medium size aminoacids: Q E V;

Pigeonhole 5, large aminoacids: H M L I K R;

Pigeonhole 6, extra large aminoacids: F Y;

Pigeonhole 7, extra extra large aminoacids: W;

Charge:

Pigeonhole N, negative aminoacids: D E;

Pigeonhole P, positive aminoacids: R H K;

Pigeonhole L, polar aminoacids: N Q S T W Y;

Pigeonhole A, apolar aminoacids: P A G C I L M F V.

4. Extended IFP results

In the main text, we showed for space reasons only a subset of the results obtained in the runs to discriminate monomer vs multimer forms of IFPs. The tables here contain an extension to that set. In particular, Table 1 reports best ranking positions from 21 to 40, obtained in the different runs described in section 5.2 to show how many of the 33 positions experimentally identified were recognized by our approach. In Table 2 we show the full, i.e. 40 first positions, results of the same type of runs, but with inclusion of background removal correction.

Comparison of the two tables shows that, at least in this case, this procedure does not improve the

Table 1. SDPs inferred from 92 mono- and multimeric GFP homologs 40 best ranking SDPs derived with BLOSUM45 (B45), BLOSUM62 (B62), Identity (Id) and the local BLOSUM (Bloc) similarity matrices. No background correlation removal has been performed in these runs.

	B45	S	B62	S	Id	S	Bloc	S
21	78	4	83	2	21	1	223	1
22	85	3	85	3	71	2	147	*
23	6	1	78	4	6	1	125	1
24	83	2	118	4	197	2	1	4
25	118	4	5	1	1	4	92	3
26	193	4	193	4	150	2	79	3
27	30	3	6	1	206	3	127	1
28	216	3	30	3	125	1	21	1
29	5	1	216	3	127	1	85	3
30	203	4	98	3	219	4	6	1
31	11	3	21	1	189	4	218	4
32	207	4	207	4	193	4	118	4
33	218	4	195	2	75	4	11	3
34	184	4	71	2	57	4	30	3
35	195	2	11	3	92	3	189	4
36	98	3	203	4	85	3	207	4
37	71	2	184	4	161	4	203	4
38	49	3	218	4	207	4	219	4
39	21	1	147	*	78	4	161	4
40	36	4	206	3	30	3	193	4

performance of the method.

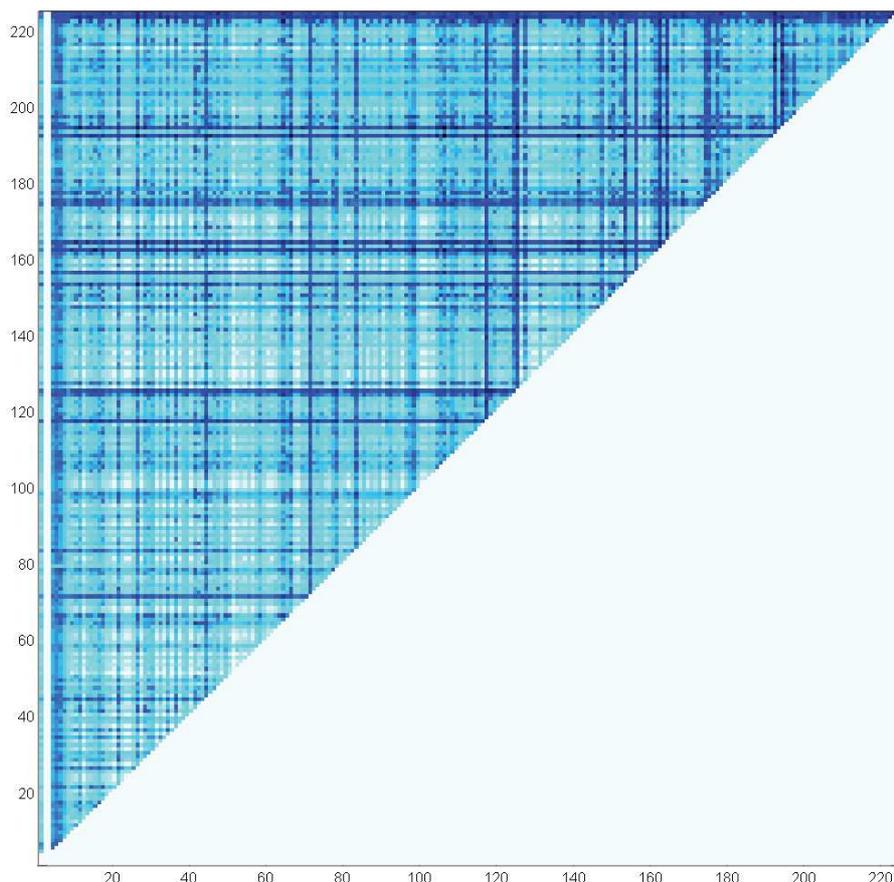
Table 2. SDPs inferred from 92 mono- and multimeric GFP homologs. Results reported are the same as Table 1 but with application of the background correlation removal and including the SDPpred results. Scores are defined as in the main text. Numbering refers to the “DsRed” sequence (1GGX PDB)

B45	S	B62	S	Id	S	Bloc	S	SDPpred	S
117	1	117	1	117	1	117	1	117	1
194	1	194	1	83	2	194	1	83	2
177	2	177	2	194	1	177	2	79	3
164	1	164	1	164	1	175	2	194	1
224	1	224	1	156	1	224	1	184	4
156	1	156	1	192	1	124	2	164	1
44	2	72	4	223	1	174	1	185	4
72	4	124	2	175	2	164	1	192	1
197	2	150	2	72	4	72	4	156	1
150	2	192	1	8	3	192	1	175	2
124	2	197	2	5	1	153	1	177	2
192	1	44	2	153	1	83	2	72	4
4	3	174	1	177	2	150	2	153	1
175	2	4	3	124	2	4	3	124	2
174	1	175	2	147	*	75	4	8	3
118	4	8	3	1	4	179	2	147	*
8	3	184	4	174	1	44	2	150	2
193	4	118	4	150	2	197	2	174	1
125	1	83	2	4	3	57	4	44	2
184	4	153	1	219	4	162	1	21	1
127	1	57	4	21	1	219	4	219	4
92	3	193	4	224	1	8	3	162	1
57	4	219	4	44	2	184	4	75	4
179	2	179	2	162	1	79	3	57	4
219	4	127	1	189	4	156	1	7	3
83	2	125	1	75	4	21	1	197	2
162	1	75	4	57	4	118	4	71	2
75	4	92	3	71	2	85	3	193	4
85	3	85	3	193	4	1	4	208	3
79	3	30	3	197	2	92	3	107	4
30	3	49	3	6	1	147	3	85	3
153	1	21	1	107	4	189	4	127	1
49	3	207	4	206	3	71	2	38	4
218	4	201	4	85	3	193	4	179	2
207	4	218	4	38	4	30	3	30	3
21	1	162	1	127	1	218	4	97	4
78	4	78	4	179	2	207	4	207	4
201	4	98	3	97	4	78	4	118	4
11	3	18	4	30	3	49	3	92	3
18	4	195	2	49	3	127	1	49	3

5. Pair correlation

Figure 4 reports a pictorial representation of the symmetric correlation matrix.

Figure 4. Pair correlation among positions is shown in a color coding ranging from white, indicating poor correlation, to intense blue. Rows and columns corresponding to positions 1,2 and 51 are white since they had a number of gaps larger than 30%.



6. MMPBSA details

The MMPBSA approach makes it possible to obtain solvation free energies of proteins through the combination of all-atoms molecular dynamics simulations of the solvated molecule and estimation of the terms accounting for solvent polarization due to solvent-solute interactions, changes in the conformational freedom of solvent upon solvation, and entropy of the solute.

In this context, the absolute free energy of the solute can be expressed as follows:

$$G = E_{MM} + G_{PB,polar} + G_{SA,nonpolar} - TS_{solute}^{tr,rot,conf},$$

where all quantities are averaged over a molecular mechanics trajectory, E_{MM} is the molecular mechanics energy, $G_{PB,polar}$ is the polarization free energy of the implicit solvent, which can be obtained through the solution of the Poisson-Boltzmann (PB) Equation, $G_{SA,nonpolar}$ is the nonpolar free energy estimated

by scaling the solvent accessible surface area (SA) by an appropriate surface tension, and $S_{solute}^{tr,rot,conf}$ is the solute translational, rotational and conformational entropy, and T is the absolute temperature.

We applied this scheme to the interesting case of IFPs in order to compute tetramerization free energies of tetrameric Wild-Type DsRed and some tetrameric, dimeric or monomeric mutants as free energy differences of the tetramer with respect to the two dimers:

$$\Delta G_{i,tetramerization} = G_{i,tetramer} - 2 \cdot G_{i,dimer}.$$

$G_{i,tetramer}$ and $G_{i,dimer}$ are, respectively, the tetramer and dimer free energies of the i^{th} mutant.

In the present case, these quantities are computed over the same molecular dynamics trajectory of the tetramer for both the tetramer and the dimers; this is acceptable since, presumably, the conformations of these proteins in their dimeric and tetrameric state do not differ significantly.

Reproducing the trend of the relative stabilities of different mutants with respect to wild-type protein (as in the “virtual screening method” described in [2]) is of particular interest to select mutations that induce the stabilization of the dimers in an otherwise tetrameric protein. This can be accomplished by calculating

$$\Delta\Delta G_{i,WT} = \Delta G_{i,tetramerization} - \Delta G_{Wild-Type,tetramerization}.$$

The entropy contribution of the solute was not taken into account in the present study. Although the conformational $T\Delta S_i^{conf}$ is an important term in driving oligomer association/dissociation, we expect that this term is scarcely affected by mutations. Indeed, previous studies have shown that the conformational $T\Delta S^{conf}$ penalty upon side-chain burial is similar among different residues, with differences generally around 0.5 kcal/mol and always smaller than 2 kcal/mol at room temperature [3], and hence negligible with respect to the $\Delta\Delta G_{i,WT}$ obtained in the present study. Moreover, translational and rotational components of the entropy for the structurally homologous IFPs lead to $T\Delta\Delta S_{i,WT}^{tr,rot} \simeq 0$.

The starting structure for the MM dynamics was obtained by adding hydrogen atoms to the X-ray crystal structure of a DsRed tetramer (PDB code: 1GGX), which was solvated in a 85 Å-box of water molecules. All amino acid mutations were produced with Insight II©(Accelrys Inc.), starting from the 1GGX PDB structure for consistency. After equilibration of the system with restraints on the motion of non-hydrogen atoms (10 ps at 50 K, 20 ps at 150 K, 240 ps at 300 K), free molecular dynamics was performed for 400 ps at 300 K. Molecular Dynamics simulations and surface areas were computed with programs of the Amber 7 package, while Poisson-Boltzmann equation was solved with Delphi 4 [4].

E_{MM} , $G_{PB,polar}$ and $G_{SA,nonpolar}$ were calculated for 40 snapshots sampled from the trajectory at 10 ps intervals. The variance reported in Figure 3 of the main article is calculated on the ΔG taken at each snapshot, as usually carried out in MMPBSA calculations [2]. Each 670 ps Molecular Dynamics run (270 ps of equilibration and 400 ps of production), providing the 40 snapshots used to calculate the free energy, required 48 hours on four Intel® Xeon™ CPU 2.40GHz processors; the subsequent MMPBSA calculation, conversely, required 20 hours on a single processor.

References and Notes

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