

Tyrosinase-Based Biosensor—A New Tool for Chlorogenic Acid Detection in Nutraceutical Formulations

Irina Georgiana Munteanu and Constantin Apetrei *

Department of Chemistry, Physics and Environment, Faculty of Sciences and Environment, Dunărea de Jos University of Galați, 47 Domneasca Street, 800008 Galați, Romania; georgiana.munteanu@ugal.ro

* Correspondence: apetreic@ugal.ro; Tel.: +40-727-580-914

Supplementary information

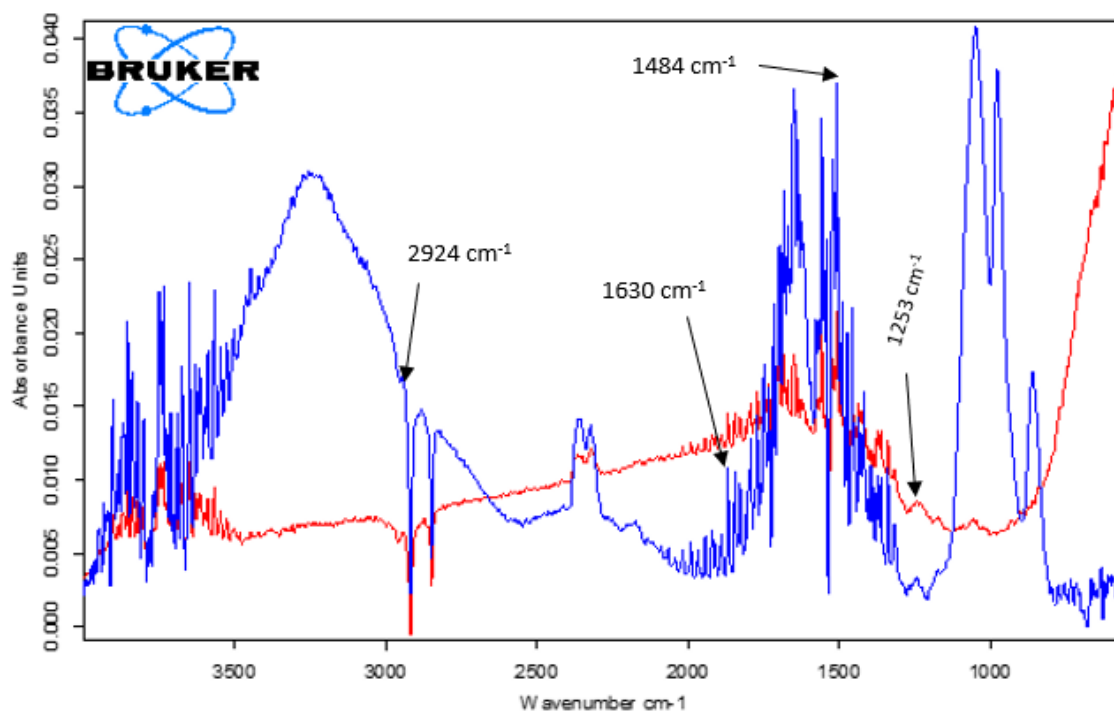


Figure S1. FTIR spectra for GPH-MnPc/SPE (red line) and for GPH-MnPc-Tyr/SPE (blue line).

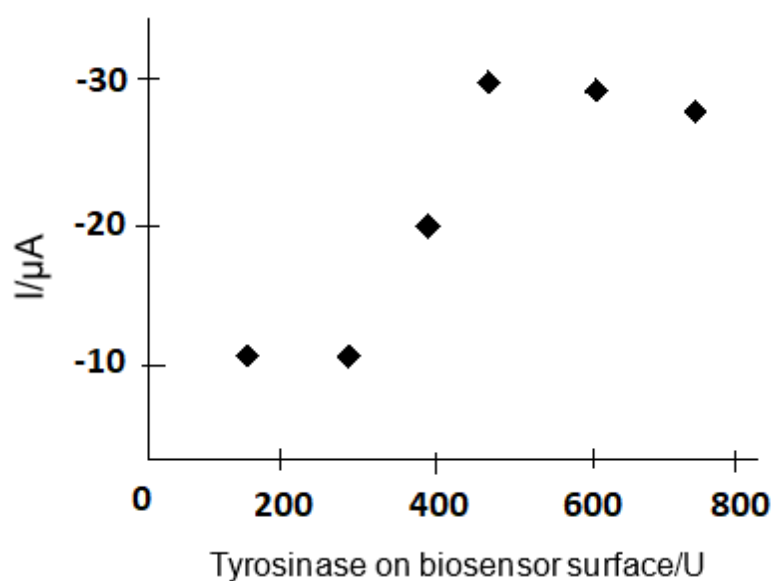


Figure S2. Effect of the amount of enzyme added on graphene and manganese phthalocyanine modified screen-printed carbon electrode on its sensitivity to catechol detection (10^{-3} M catechol, phosphate buffer solution 10^{-1} M as supporting electrolyte, at pH=7.0). Applied potential -0.08 V vs Ag/AgCl.

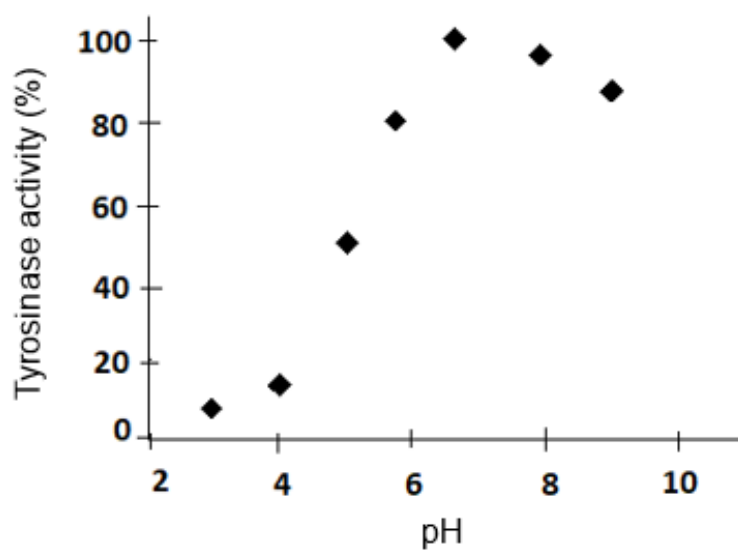


Figure S3. The effect of pH on tyrosinase activity.

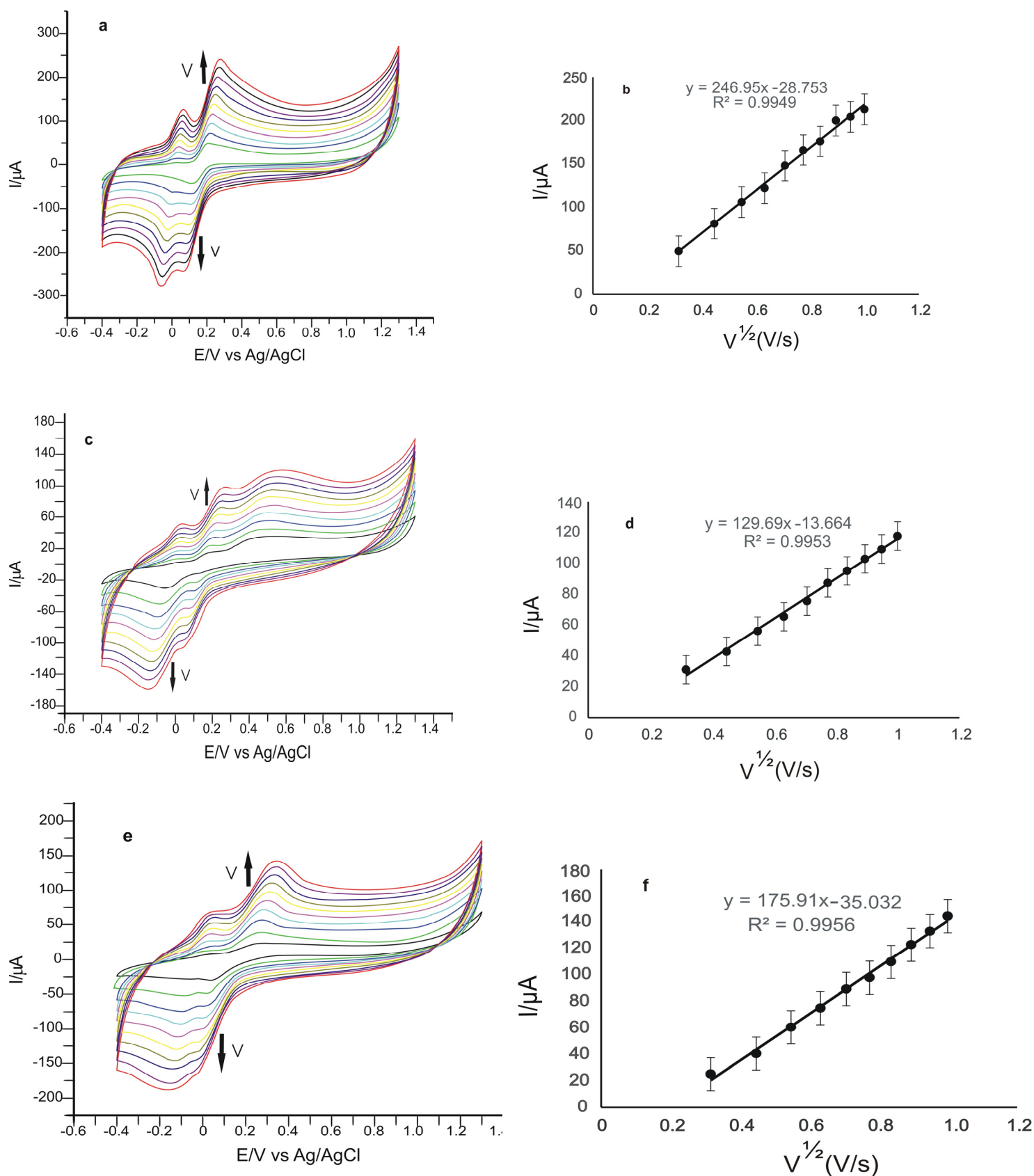


Figure S4. Cyclic voltammograms of the GPH/SPE (a), GPH-MnPc/SPE (c), GPH-MnPc-Tyr/SPE (e) immersed in 10^{-3} M catechol – 10^{-1} M PBS solution registered with scan rates between 0.1–1.0 $V \times s^{-1}$. Plot of I_{pa} vs $v^{1/2}$ in the case of GPH/SPE (b), GPH-MnPc/SPE (d), GPH-MnPc-Tyr/SPE (f).

Table S1. The values of the parameters obtained from the cyclic voltammograms of all the electrodes immersed in 10^{-3} M catechol – 10^{-1} M PBS double solution. ¹ Current of the anodic peak; ² Current of the cathodic peak; ³ Potential of the anodic peak; ⁴ Potential of the cathodic peak; ⁵ Half-wave potential; ⁶ $\Delta E = E_{pa} - E_{pc}$.

Sensor	I_{pa}^1 (μA)	I_{pc}^2 (μA)	I_{pc}/I_{pa}	E_{pa}^3 (V)	E_{pc}^4 (V)	$E_{1/2}^5$ (V)	ΔE^6 (V)
GPH-SPE	49.50	-42.55	0.85	0.204	0.117	0.160	0.087
GPH-MnPc/SPE	32.02	-30.69	0.95	0.447	-0.055	0.251	0.392
GPH-MnPc-Tyr/SPE	25.57	-32.65	1.27	0.231	-0.077	0.154	0.154