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Hydrothermal Carbonization Kinetics of Lignocellulosic Agro-Wastes: Experimental Data and Modeling

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Abstract: Olive trimmings (OT) were used as feedstock for an in-depth experimental study on the reaction kinetics controlling hydrothermal carbonization (HTC). OT were hydrothermally carbonized for a residence time τ of up to 8 h at temperatures between 180 and 250 °C to systematically investigate the chemical and energy properties changes of hydrochars during HTC. Additional experiments at 120 and 150 °C at τ = 0 h were carried out to analyze the heat-up transient phase required to reach the HTC set-point temperature. Furthermore, an original HTC reaction kinetics model was developed. The HTC reaction pathway was described through a lumped model, in which biomass is converted into solid (distinguished between primary and secondary char), liquid, and gaseous products. The kinetics model, written in MATLABTM, was used in best fitting routines with HTC experimental data obtained using OT and two other agro-wastes previously tested: grape marc and *Opuntia Ficus Indica*. The HTC kinetics model effectively predicts carbon distribution among HTC products versus time with the thermal transient phase included; it represents an effective tool for R&D in the HTC field. Importantly, both modeling and experimental data suggest that already during the heat-up phase, biomass greatly carbonizes, in particular at the highest temperature tested of 250 °C.

Keywords: hydrothermal carbonization (HTC); reaction kinetics; modeling; carbon recovery; activation energy; agro-wastes; olive trimmings

1. Introduction

In the last few decades, the need to find new environmentally friendly and sustainable technologies to produce clean energy from waste biomass has inspired scientific research to study and develop more efficient and reliable tools for waste biomass exploitation. Among these technologies, pyrolysis of agricultural and agro-industrial waste has received particular attention from the points of view of process dependent bio-char properties [1–3], pyrolysis kinetics [4,5], and innovative technology development [6]. Due to the high-energy needs of converting moist waste biomass through conventional dry pyrolysis, a wet pyrolysis technology, known also as hydrothermal carbonization (HTC), has gained increasing attention in the last few years. HTC is a wet thermochemical process carried out at relatively mild conditions: temperatures between 180 and 280 °C, autogenous saturated vapour pressure between 10 and 80 bars, and residence time ranging from a few minutes up to several hours. Under these process conditions, water acts similarly to an organic solvent due to the tremendous changes in its polarity and dielectric constant, and as a catalyst for biomass conversion via hydrolysis, dehydration, and decarboxylation reactions [7–11]. The main product of HTC is a carbon-rich solid material referred to as hydrochar, which finds application as high energy bio-fuel [12–21], as a pre-treated material for anaerobic digestion enhancement [15,22], as a soil amendment [14,23,24],

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and as an advanced carbonaceous material [25,26]. The high interest in this wet thermochemical technology, due to its high energy efficiency, relatively mild process conditions, and scalability, has also been testified by the study on its application at the industrial level [27,28]. Among different wet residual biomasses, agro-industrial wastes have received particular attention by HTC scientists due to their large production, high management and treatment costs, and the potential sanitary and pollution hazards deriving from their improper disposal. In particular, olive oil milling residues, olive pulp [16,29,30], olive stones, olive mill waste water [31], and their mixture [14] have been deeply investigated. Much less attention has been focused on the treatment of olive tree pruning residues [32]. Olive collection and seasonal olive tree pruning produce large amounts of wet lignocellulosic material not suitable for combustion in stoves and biomass boilers, and is therefore commonly abandoned and burnt in open fields with negative consequences to the soil and air quality. This paper reports a comprehensive investigation on HTC conversion of olive tree pruning residues showing the possibility to produce a valuable carbon-rich solid biofuel.

If, on the one hand, many research groups have focused their studies on HTC reaction of different biomasses primarily to determine the influence of process parameters on hydrochar's chemical and physical properties, on the other hand, the kinetics of HTC of biomasses and its modeling still lacks of a deep investigation [33,34]. The HTC reaction mechanism is very complex and involves several pathways such as hydrolysis, dehydration, decarboxylation, aromatization, and condensation [35,36]. The complexity of the HTC reaction mechanism has led some authors to develop lumped kinetics models to describe it. Liu and Balasubramanian considered HTC process as a unique first order reaction and estimated the activation energy for coconut fibres (67.5 kJ/mol) and eucalyptus leaves (59.2 kJ/mol) [37]. By using a different approach, Reza and co-workers formulated a simple reaction mechanism in which hemicellulose and cellulose degrade in parallel while lignin was assumed to be inert at the temperatures investigated (200–260 °C) [38]. The model developed by these authors allows for the determination of activation energies of hemicellulose and cellulose degradation first-order reactions, which were determined to be 30 and 73 kJ/mol, respectively [38]. HTC kinetics of biomass macro-components was also recently reported [39,40]. Jung and Kruse used an Arrhenius-type overall kinetics equation to model hydrochar mass yield and carbon and oxygen content for several types of biomasses [34]. Baratieri et al. proposed a kinetics model for HTC based on a two-step reaction mechanism [41]. They assumed that the original biomass (A) forms an intermediate product (B) that partially degrades to form the final product C (the hydrochar). In the meanwhile, two reactions in parallel to those giving compounds B and C take place leading to the formation of volatiles products. They calibrated their model using HTC experimental data of solid yields and determined the activation energies and pre-exponential factors of the reactions involved [41]. Jatzwauck and Schumpe proposed an example of a lumped model describing the HTC mechanisms [33]. According to the authors, the HTC reaction mainly occurs in three steps: in the first step, the biomass components are hydrolysed into dissolved intermediates; in the second step, the intermediates may undergo further reaction in the liquid phase or evolve into gaseous products; in the last step, alternatively to step two, the intermediates could polymerize to form secondary char. In their model, the authors also reported carbon distribution among the HTC products [33]. Such a kinetics model does not take into consideration the direct solid-solid dehydration reaction leading, as well as polymerization, to char formation. In contrast, some authors [16,42,43] experimentally demonstrated that hydrochar formation takes place through two reaction pathways: (1) solid-solid conversion, in which carbonaceous material (primary char or simply char) results predominantly from dehydration of the parent biomass; and (2) a polymerization reaction, in which a carbonaceous solid is formed via reactions of back polymerization of organics dissolved in the liquid phase.

Based on these previous studies, this work reports an original kinetics model assuming that the HTC process follows a simplified mechanism including both pathways of hydrochar (primary and secondary char) formation. The model is calibrated on HTC experimental data of olive trimmings specifically obtained for this purpose, and is further validated with additional HTC data previously

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obtained using grape marc [44] and *Opuntia ficus indica cladodes* [7]. By means of the modeling tool developed, the Arrhenius kinetics parameters (pre-exponential factor and activation energy) were estimated for the reactions constituting the model and for the various biomasses.

As a whole, this work addresses HTC reaction kinetics of lignocellulosic residual biomasses from both theoretical and experimental viewpoints, also paying attention to the heat-up transient phase necessary to increase the HTC reactor temperature to the set point value. Importantly, only in micro-autoclaves can the time necessary for the system to reach set point HTC temperature be neglected: for the bench scale and industrial scale applications, it cannot. Nevertheless, to the best knowledge of the authors, the characterization of the solid product during the heat up phase has never been addressed before: the study of such a transient phase represents a novelty in the literature.

To sum up, the present work aims to:

- (1) Analyse the differences in chemical and energy properties of hydrochar versus time at different HTC temperatures.
 - (2) Understand the evolution of hydrochar characteristics during reactor heating time.
- (3) Present an original reaction kinetics model as a tool to predict the carbon distribution among HTC products: hydrochar (primary and secondary char), gaseous, and liquid phases.

2. Materials and Methods

2.1. HTC Tests

Olive trimmings, including leaves, were collected from three different 30-year-old olive trees of the "Moresca" variety in Enna province, Italy. The biomass was dried in a ventilated oven at 105 °C for 24 h and milled to a particle size lower than 1 mm. To ensure homogeneity, raw material was sieved and a particle size range between 425 and 850 µm was selected for HTC tests. The HTC set up, the biomass preparation, and experimental procedures followed in the experimental campaign were fully described in previous studies [16,45]. For each HTC run, 5 ± 0.01 g of an oven-dried olive trimmings (OT) sample were loaded into the reactor together with 20 ± 0.01 g of deionized water with a dry biomass to water ratio (DB/W) equal to 0.25. HTC tests on OT were performed at temperatures of 180, 220, and 250 °C. Residence times were varied at 0, 0.5, 1, 3, 6, and 8 h. Residence time started to be counted when the HTC reactor reached the selected HTC set point temperature. Thus, a residence time of 0 h indicates that the HTC reactor was promptly cooled down as soon as the set temperature was reached. Cooling was performed by placing the HTC reactor on a cold stainless-steel mass at −30 °C and blowing fresh compressed air on its external walls. Once the reactor reached room temperature, its outlet valve was opened to let the gas produced flowing into a graduated cylinder filled with water [46]. By using this system, the volume of the gas evolved during HTC reaction was determined. The reactor was then opened, and the solid fraction recovered via filtration. Finally, the solid residue was dried in a ventilated oven for 8 h at 105 $^{\circ}$ C. The HTC runs were carried out in duplicate. The hydrochar yield (Y_{HC}) , or solid yield, was determined through Equation (1), where m_{HC} is the mass of hydrochar (on a dry basis) and m_B is the mass of the raw biomass (on a dry basis):

$$Y_{HC} = \frac{m_{HC}}{m_B} \tag{1}$$

The gas yield was calculated as the ratio of gas produced (mass on a dry basis) and the raw biomass (mass on a dry basis). As is well-known [28,44,47], CO_2 is the main gas produced during HTC, representing 90–95 vol.% of the total gas. For this reason, the mass of the gas produced was calculated considering it as solely consisting of CO_2 . The liquid yield was calculated as the complement to one of the sum of solid yield and gas yield.

The energy yield (EY) of hydrochars was calculated as follows:

$$E_{Y} = Y_{HC} \frac{HHV_{HC}}{HHV_{B}}$$
 (2)

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where HHV_{HC} and HHV_{B} (MJ/kg) are the higher heating values (on a dry basis) of hydrochar and raw biomass, respectively.

2.2. Analytical Determinations

Ultimate analyses were performed using a LECO 628 analyser (LECO Corporation, St. Joseph, MI, USA) equipped with sulphur module for CHN (ASTM D-5373 standard method) and S (ASTM D-1552 standard method) content determination.

Proximate analyses were carried out by a LECO Thermo-gravimetric Analyser TGA 701 (LECO Corporation, St. Joseph, MI, USA). Moisture content (M), volatile matter (VM), and ashes of solid samples were respectively determined using the following thermal methods (modified from ASTM D-3175-89 standard method): 20 °C/min ramp to 105 °C in air, held until constant weight (< $\pm 0.05\%$) (M); 16 °C/min ramp from 105 °C to 900 °C, hold time 7 min, in N₂ (VM); isothermal hold at 800 °C in air (Ash). Fixed carbon (FC) was evaluated by difference.

The higher heating values (HHV) of raw biomass and hydrochars were evaluated by means of IKA C200 calorimeter (IKA-Werke GmbH, Staufen, Germany) according to the CEN/TS 14918 standard.

2.3. HTC Reactor Heat-Up Transient Phase

In the literature, the definition of reaction time (or residence time) is not uniform. Often, HTC reaction time starts to be counted when the set HTC temperature is reached, while in some other cases [48,49] it includes the time for reactor heating. In this paper, we considered the HTC residence time as the time the reactor is maintained at the set-point constant temperature. With the reactor utilized, the heat-up time ranged between 5 and 28 min depending on the set hydrothermal temperature (Figure 1). Such heat-up time is referred here also as transient time. Constant temperature during HTC is far from reality in batch autoclave reactors, where there is a heating time, a temperature constant period, and finally a cooling time. Transient time may be neglected only if the heating is very rapid, as it could be the case when the HTC is performed in batch micro-reactors. Figure 1a shows the temperature profile during transient time for a HTC test whose set-point temperature was 250 °C, from which the heating time for different test temperatures was extrapolated.

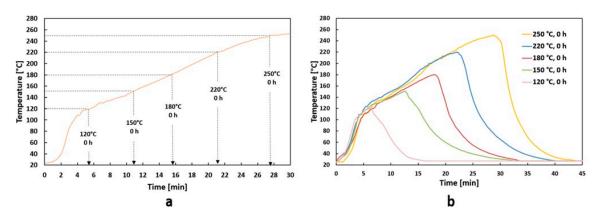


Figure 1. (a) Heating time (transient time) of olive trimmings from ambient temperature to $250\,^{\circ}$ C hydrothermal carbonization (HTC) temperature; (b) temperature trends vs. time of hydrothermal tests conducted at 120, 150, 180, 220, 250 $^{\circ}$ C at 0 h residence time.

The tests at 120 and 150 °C and 0 h of residence time were carried out to better understand biomass property changes during transient time. In such a low temperature range, only some thermal hydrolysis reactions and mass transfer phenomena due to extraction in hot water will likely occur. Thus, the results of the tests performed at 120 and 150 °C were not included in model best fitting procedures. Such results were used to validate the model as far as the predicted carbon recovery in the solid during heating, before the beginning of the HTC reaction, is concerned. Figure 1b shows

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the temperature profiles of the hydrothermal experimental trials conducted at various temperatures ($120-250\,^{\circ}$ C) at 0 h residence time. The reported temperatures profiles, acquired using a thermocouple inside the batch reactor, include both transient time up to the set hydrothermal temperature and the following immediate cooling down of the apparatus.

2.4. Kinetics Model

The HTC reaction mechanism was described schematically using a lumped component model in which the initial feedstock converts into products and intermediates in different steps (Figure 2).

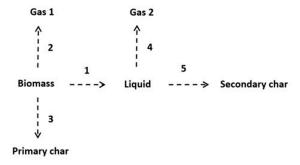


Figure 2. Simplified HTC reaction path used in the model [50].

Reaction 1 (biomass—liquid) refers to a hydrolitic processes leading to the formation of a HTC liquor rich in intermediate molecules [51] deriving from biomass structural components (extracts, lignin, cellulose, and hemicellulose). The extracts and hemicellulose constituents are known to be susceptible to hydrolysis at quite low HTC temperatures (as low as 200 °C) [52]. The hydrolysis/dissolution of these biomass components leads to oligomers and monomers formation in the aqueous liquid phase [36]. Similarly, a portion of amorphous cellulose and soluble lignin is fragmented into smaller molecules, mainly 5-Hydroxymethylfurfural (5-HMF) and phenolic derivatives [53]. The resulting water-soluble components are very reactive and could undergo a series of oligomerization/polymerization and/or condensation reactions to form a solid residue referred to as secondary char (also called polymerized char or coke—Reaction 5, liquid—secondary char). These reactions are accompanied by decarboxylation and decarbonylation reactions to form CO₂ and CO (Reaction 4, liquid—gas 2). During HTC, the non-dissolved cellulose and lignin contained in biomass undergo dehydration and decarboxylation/decarbonylation reactions to form an interconnected network called primary char (or polyaromatic char—Reaction 3, biomass—primary char) [54] and CO₂ and CO (Reaction 2, biomass—gas 1), respectively.

The reaction rate constants k_i (k_1 , k_2 , k_3 , k_4 , k_5) for the five reactions i in Figure 2 are described within the model by using Arrhenius equation (Equation (3)):

$$k_i = k_{0,i} e^{-\frac{E_a}{RT}} i = 1, \dots, 5$$
 (3)

The kinetics model consists of a system of six non-linear differential equations, Equations (4)–(9), representing the carbon molar balances for the different lumped components. C_B , C_L , C_{G1} , C_{HC1} , C_{G2} , and C_{HC2} are the molar concentration (mol/L) of carbon in biomass (B), liquid (L), gas 1 (G1), primary char (HC1), gas 2 (G2), and secondary char (HC2), respectively, while t is the reaction time. It was assumed that all the reactions involved are first order, except the one involving secondary char formation (Reaction 5, liquid \rightarrow secondary char). The order of this reaction, also called a polymerization/condensation reaction, is likely higher than 1 [55]. Thus, in Equation (9), C_L is to the power of n, with $n \ge 1$, n being the reaction order. The model was also run fixing n = 1; the relevant modeling results are reported in Appendix A.

$$\frac{\partial C_B}{\partial t} = -k_1 C_B - k_2 C_B - k_3 C_B \tag{4}$$

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$$\frac{\partial C_L}{\partial t} = k_1 C_B - k_4 C_L - k_5 C_L^n \tag{5}$$

$$\frac{\partial C_{G1}}{\partial t} = k_2 C_B \tag{6}$$

$$\frac{\partial C_{HC1}}{\partial t} = k_3 C_B \tag{7}$$

$$\frac{\partial C_{G2}}{\partial t} = k_4 C_L \tag{8}$$

$$\frac{\partial C_{HC2}}{\partial t} = k_5 C_L^n \tag{9}$$

The carbon molar concentration in component X (B, L, G1, HC1, G2, HC2), C_X, is given by:

$$C_X = \frac{n_{c,X}}{(V_B + V_W)} \tag{10}$$

where $n_{c,X}$ is the number of moles of carbon in component X, and the term $V_B + V_W$ is the sum of the volumes (expressed in L) of biomass and distilled water fed to the HTC reactor to reach the set DB/W ratio. The kinetics model was written as a MATLABTM software code (MATLAB R2016b developed by MathWorks Inc., Natick, MA, USA)—see Supplementary Materials. The differential equations system was solved numerically using Runge–Kutta method by means of a routine that estimates the kinetics parameters (k_i and n) on the basis of experimental data. In more details, the error function F (k_i , n) represented by Equation (11), was minimized in order to get k_i and n by using the Levenberg–Marquardt algorithm. This "curve-fitting" procedure is a standard tool in computing and it is extensively used for non-linear least squares problems [56,57].

$$F(k_{i}, n) = \sum_{i} \left| C_{S,j}^{exp} - C_{S,j}^{mod} \right| + \sum_{i} \left| C_{L,j}^{exp} - C_{L,j}^{mod} \right| + \sum_{i} \left| C_{G,j}^{exp} - C_{G,j}^{mod} \right|$$
(11)

Subscripts S, L, and G refer to solid, liquid, and gas phases. The superscript "exp" refers to the experimental values of the variables obtained at different HTC residence times, while the superscript "mod" refers to the corresponding values computed by the model. Subscript j is a recursive index for residence time.

The experimental data used in model best fitting were obtained using olive trimmings (OT) as described in Section 2.1, and two others ligno-cellulosic biomasses, namely *Opuntia ficus indica cladodes* (OC) and grape marc (GM), tested in previous studies [7,44]. Thus, subscript j refers to the residence times at which the experimental data were available: six for OT, three for both OC and GM.

Carbon molar concentrations (C_X) using both experimental and modeling data were calculated and computed through Equations (12)–(16):

$$C_{S,j}^{exp} = \frac{n_{c,HC}}{(V_B + V_W)} = \frac{m_{c,HC}}{M_c} \cdot \frac{1}{(V_B + V_W)} = \frac{m_{HC}\%c_{HC}}{100 M_c} \cdot \frac{1}{(V_B + V_W)} = \frac{Y_{HC}m_{B,0}\%c_{HC}}{100 M_c} \cdot \frac{1}{(V_B + V_W)}$$

$$= \frac{Y_{HC}m_{B,0}\%c_{HC}}{100 M_c} \cdot \frac{1}{(V_B + V_W)}$$
(12)

$$C_{S,j}^{\text{mod}} = C_{B,j}^{\text{mod}} + C_{\text{HC1},j}^{\text{mod}} + C_{\text{HC2},j}^{\text{mod}}$$
(13)

$$C_{L,j}^{\text{exp}} = C_{B,0}^{\text{exp}} - C_{S,j}^{\text{exp}} - C_{G,j}^{\text{exp}}$$
(14)

$$C_{G,j}^{exp} = \frac{n_{C,G}}{(V_B + V_W)} = \frac{n_{CO_2}}{(V_B + V_W)} = \frac{PV_{CO_2}}{RT} \cdot \frac{1}{(V_B + V_W)}$$
(15)

$$C_{G,j}^{\text{mod}} = C_{G1,j}^{\text{mod}} + C_{G2,j}^{\text{mod}}$$
 (16)

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Letters M, m, and Y indicate molar mass (g/mol), mass (g), and solid yield (i.e., hydrochar yield), respectively. Subscripts c, HC, and 0 refer to carbon, hydrochar, and initial value, respectively. % indicates the percentage on a dry mass basis.

In Equation (15), P and T are atmospheric pressure (1 atm) and room temperature (293 K), respectively; V_{CO2} is the measured volume of the gas, which was assumed to be 100% CO_2 , as discussed in Section 2.1; R is the gas constant (0.08206 L atm/K mol).

The solid, after the HTC reaction had started, consisted of hydrochar (Equation (12)). Conversely, in the model (Equation (13)), the solid refers to biomass whose amount decreased during HTC, while its composition was considered constant (Equation (4)), plus primary char HC1 and secondary char HC2. In the model (Equation (16)), the gas consisted of the sum of G1 and G2, while the experimental value was calculated using the ideal gas law (Equation (15)). The experimental values for the liquid were calculated by difference (Equation (14)), while the relevant model values were obtained by solving Equation (5). The available experimental data were the input values, V_B , V_W , $m_{B,0}$, and the HTC output values Y_{HC}, %c_{HC} (from hydrochar elemental analysis), and V_{CO2}. Based on the experimental data available for the HTC of OT, $F(k_i, n)$ was minimized for six reaction times (0, 0.5, 1, 3, 6, and 8 h) at each of the three HTC temperatures tested: 180, 220, and 250 °C. In the case of GM and OC, the minimization was performed at the same temperatures but by considering only three residence times (1, 3, and 8 h for GM and 0.5, 1, and 3 h for OC). It is worth mentioning that in Equation (11), the term related to the liquid phase (which could be actually omitted without compromising the correctness of the equation) allowed the software to converge faster towards the solution of the minimization problem. Once the k_i values were estimated, the activation energy $E_{a,i}$ and pre-exponential factor $k_{0,i}$ for the five simplified HTC reactions of Figure 2 were calculated through the Arrhenius plot ($\ln k_i$ vs. 1/T).

Carbon distribution among the HTC products is expressed in terms of carbon recovery (CR), defined as the ratio of carbon content in each component ($n_{c,X}$) to the carbon content in the raw biomass ($n_{c,B,0}$). The CR parameter allows for an easy understanding of the carbon distribution among the different HTC phases [58]:

$$CR_X = \frac{n_{c,X}}{n_{c,B,0}} \tag{17}$$

Similarly, oxygen recovery and hydrogen recovery are defined as the ratio of oxygen (or hydrogen) content in each component to the oxygen (or hydrogen) content in the raw biomass, on a dry basis.

3. Results and Discussion

3.1. Hydrothermal Carbonization of Olive Trimmings

The experimental results relevant to HTC runs performed on OT are summarized in Table 1. Hydrochar yields decreased with temperature and residence time except for residence times higher than 3 h. Tests performed at 180 and 250 °C and 6–8 h of residence time showed an increase in hydrochar yield compared to 3 h residence time, which could be explained by an increased contribution of back-polymerization at longer residence times [17]. The HHV values progressively increased with temperature and residence time (except a slight decrease at 8 h). Regarding the ultimate analysis, the carbon content constantly increased with both time and temperature (except for 8 h at any temperature), while hydrogen remained approximately constant, with a slight decrease at the 8 h residence time. As a result of dehydration and decarboxilation, the oxygen content followed an opposite trend with respect to carbon. As expected, the data showed an increase in fixed carbon (FC) and a decrease in volatile matter (VM) with temperature. Interestingly, the same trend was observed when increasing the residence time in the lower residence time range, but for higher residence times (6–8 h), the trend was reversed.

All the data of Table 1 in the residence time range 0–3 h presented typical trends reported in the literature. Of particular interest, in term of reaction kinetics, are the data in the time range

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3–8 h. Here, the hydrochar yield increased; the degradation of primary char was probably more than counterbalanced by the precipitation in the solid phase of polymerized/condensed organics from the liquid phase. Hydrochar was enriched in carbon from 3 to 6 h residence time, but lowered its carbon content at 8 h, when hydrogen content also reduced (and this caused the trend observed for HHV). Similary, for such a residence time range, FC decreased. This suggests that the precipitation of the so-called secondary char also led to the presence of thermally instable species, eventually with tar-like properties, whose content in carbon was significant in the initial stage of the precipitation (up until 6 h residence time), but less significant thereafter, with precipitation of more oxygenated compounds.

Table 1. Mass yields, proximate and ultimate analyses, and HHV of raw feedstock and hydrochars. Analyses were performed in duplicate. Average values are shown (Er < 2.0% for solid yields; Er < 3.2% for proximate, and Er < 0.6% for ultimate analyses; Er < 1.0% for HHVs).

Sample	Ma	ss Yields	(-)		mate An % on a c	,	Ţ		Analysi n a d.b.)		HHV (MJ/kg)	EY (-)
	Solid	Liquid	Gas	VM	FC	Ash	C	Н	N	O ²		
OT raw	-	-	-	78.4	17.6	4.0	48.3	6.1	1.5	40.0	19.8	-
120 °C, 0 h	0.93	0.07	0.00	81.3	15.0	3.7	50.6	5.9	1.6	38.1	20.1	0.94
150 °C, 0 h	0.91	0.09	0.00	82.2	14.0	3.8	49.6	6.0	1.8	38.8	20.2	0.92
180 °C, 0 h	0.88	0.11	0.01	81.2	15.7	3.1	51.3	5.9	1.6	38.1	20.5	0.91
180 °C, 0.5 h ¹	0.78	0.20	0.02	77.3	19.9	2.8	53.7	6.2	1.5	35.7	21.8	0.86
180 °C, 1 h	0.72	0.25	0.03	76.1	20.3	3.5	54.3	6.1	1.8	34.3	22.6	0.82
180 °C, 3 h	0.70	0.26	0.04	72.9	23.2	3.9	56.7	6.2	1.8	31.4	23.4	0.83
180 °C, 6 h	0.73	0.22	0.05	73.8	21.9	4.3	58.9	5.9	1.6	29.2	24.1	0.89
180 °C, 8 h	0.73	0.23	0.04	76.8	18.0	4.0	57.8	5.3	1.6	31.3	23.7	0.88
220 °C, 0 h	0.74	0.23	0.03	76.3	20.3	3.5	55.3	5.9	1.5	33.7	22.3	0.83
220 °C, 0.5 h ¹	0.71	0.23	0.06	72.9	23.0	4.1	56.3	6.2	1.6	31.7	23.9	0.86
220 °C, 1 h	0.63	0.31	0.06	70.6	25.1	4.3	59.4	6.2	1.9	28.3	24.7	0.78
220 °C, 3 h	0.58	0.33	0.09	64.4	31.4	4.2	63.0	6.3	2.2	24.4	26.4	0.77
220 °C, 6 h	0.57	0.33	0.10	66.9	28.4	4.6	65.5	6.0	2.0	21.8	26.7	0.77
220 °C, 8 h	0.61	0.28	0.11	70.1	23.6	4.4	64.1	5.5	1.9	24.1	26.7	0.82
250 °C, 0 h	0.66	0.28	0.06	72.3	23.7	4.0	58.1	6.2	1.6	30.0	23.9	0.80
250 °C, 0.5 h ¹	0.58	0.33	0.09	66.3	30.3	3.4	63.2	6.3	1.8	25.3	27.3	0.79
250 °C, 1 h	0.52	0.37	0.11	66.8	29.2	4.0	65.3	6.2	2.3	22.2	27.8	0.72
250 °C, 3 h	0.48	0.39	0.12	59.6	35.7	4.7	68.9	6.3	2.5	17.6	29.0	0.71
250 °C, 6 h	0.50	0.37	0.13	56.3	39.9	3.8	70.6	5.9	2.2	17.5	29.6	0.74
250 °C, 8 h	0.49	0.38	0.13	61.8	32.0	4.1	69.0	5.5	2.1	19.3	28.9	0.72

¹ Data from Volpe and Fiori, 2017 [16]; ² Calculated by difference; d.b.=dry basis.

3.2. Hydrothermal Carbonization during Transient Time

Figure 3 summarizes the experimental results of HTC of OT during the reactor heating up. Figure 3a shows a slight decrease in solid yield to 0.93 and 0.91 at temperatures typical of thermal hydrolysis (at 120 and 150 °C, respectively). The drop of solid yield might be explained by the solubilization of extracts that were quickly extracted from biomass by hot compressed water. With increasing temperature, at typical hydrothermal carbonization temperatures, solid mass yield dramatically dropped to 0.74 and 0.66 at 220 and 250 °C, respectively. HHV followed an inverse trend with respect to solid yield. At temperatures higher than 180 °C, there was a significant increase in HHV, where there was a substantial decrease in hydrochar yield. The decrease in hydrochar yield prevailed over the increase of HHV, and thus the energy yield decreased versus temperature.

Figure 3b shows that, even if carbon percentage in the hydrochar increased with temperature (ultimate analysis data, Table 1), its recovery lowered, due to the prevailing magnitude of the hydrochar yield decrease. The decrease of oxygen recovery versus temperature was very large; this was mainly due to the dehydration reactions occurring during HTC. At the highest temperatures, decarboxylation also occurred, which resulted in a further significant oxygen release. The hydrogen recovery mimicked the oxygen recovery in the temperature range typical of thermal hydrolysis, where dehydratation

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prevailed. Conversely, hydrogen recovery was greater that oxygen recovery at the typical HTC temperatures, where decarboxylation prevailed.

The results of proximate analysis also showed a substantial change in VM and FC at 220 and 250 °C with respect to the lower temperatures (Figure 3c).

In summary, already during the heating up transient phase, experiments testified to a clear transition between thermal hydrolysis and HTC. From the collected data, the 180–220 $^{\circ}$ C range seems the temperature interval at which this transition occurred. Already during heat up, OT carbonized to a significant extent if the temperature reached value of 220–250 $^{\circ}$ C.

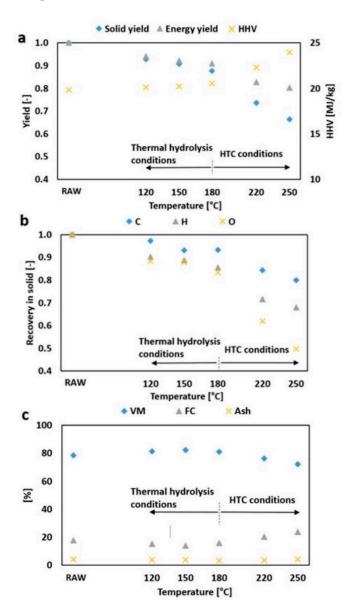


Figure 3. Transient time: (a) hydrochar yield, energy yield, and HHV vs. temperature; (b) element recovery in the hydrochar vs. temperature; and (c) proximate analysis data vs temperature.

3.3. Kinetics Model

The values of the kinetics parameters estimated using the model for OT, GM, and OC are reported in Table 2.

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Parameters	Olive Trimmings			C	Grape Marc			Opuntia Ficus Indica		
T (°C)	180	220	250	180	220	250	180	220	250	
$k_1 (s^{-1})$	0.24	0.35	0.54	0.22	0.21	0.49	0.33	0.35	0.48	
$k_2 (s^{-1})$	0.03	0.07	0.14	0.02	0.04	0.09	0.05	0.09	0.10	
$k_3 (s^{-1})$	1.05	1.13	1.40	1.00	1.14	1.41	1.04	1.14	1.41	
$k_4 (s^{-1})$	0.001	0.014	0.005	0.003	0.003	0.004	0.004	0.015	0.030	
$k_5 (s^{-1})$	0.09	0.10	0.14	0.08	0.13	0.20	0.04	0.11	0.21	
n (-)	1.10	1.51	2.01	1.10	1.51	2.00	1.11	1.51	2.00	

Table 2. Reaction rate constants (k_i) and reaction order (n) for the simplified reactions path of Figure 2.

As expected, the rate constants k_i increased as the temperature increased for all the feedstocks. Table 2 shows that k_3 was the highest at every HTC condition. Model results suggest that the conversion of biomass into primary char was the most favoured reaction. The biomass—liquid reaction was also quite fast, while the reactions involving gas phases were the slowest. Reaction rate constant k_4 was almost negligible compared to the others. The results show that the reaction order n of secondary char formation increased with temperature. This was in accordance with the results reported by some other scientists [43].

Figure 4 shows the model and experimental CR values in the solid and gas phases for OT. The vertical dotted lines represent the time for reaching the HTC temperature set point, the time at which the experimental reaction time had started to be counted. Our simulations, based on the system of differential equations represented by Equations (4)–(9), conversely considered time zero when the reactor started to be heated up to reach the set HTC temperature (about 16, 22, and 28 min to reach 180, 220, and 250 °C, respectively; see also Figure 1). While the carbon recovery increased progressively with time in the gas phase at all the HTC temperatures examined and for all the feedstocks (see Appendix A—Figures A1 and A2 for GM and OC, respectively), the carbon recovery in the solid phase decreased during reaction up to about 3 h and then tended to increase or stabilize. The model fits very well the experimental data; the errors (i.e., the differences between model predictions and experimental data), calculated by means of Equation (18), were lower than 10% in all the cases. The error values for all the feedstocks and the plots showing the carbon distribution among the HTC solid and gas phases for GM and OC are reported in Appendix A (see Table A1, Figures A1 and A2).

$$\varepsilon_{S,j} = \frac{\left| C_{S,j}^{\text{exp}} - C_{S,j}^{\text{mod}} \right|}{C_{S,j}^{\text{exp}}} \cdot 100 \tag{18}$$

Figure 5 reports the carbon recovery in all the six lumped components of the reaction scheme designed in Figure 2. The carbon recovery plots resulted from the solution of the differential equations system, Equations (4)–(9), with the optimized kinetics parameters of Table 2. Figure 5 is relevant to OT.

The carbon recovery curves are of particular interest for the assessment of the kinetics of primary and secondary char formation, as well as of the gas production from the solid and liquid phases.

The results clearly showed how the initial carbon recovery rate in the liquid (i.e., the slope of the liquid curve) increased for increasing HTC temperatures. Indeed, at 250 °C, the degradation of biomass occurred much faster when compared to lower HTC temperatures. At 250 °C, more than half of the initial carbon content in the biomass moved to HTC products just after the thermal transient (i.e., at 0 h residence time). The carbon recovery into the liquid phase reached a maximum at approximately 1.5 h of residence time, after which the gas and, to a lesser extent, the secondary char seemed predominant. The carbon recovery into the primary char reached a stable value after about 3 h for all the three temperatures. The carbon recovery in secondary char as well as in the gaseous products increased with time. Such an increase was faster in the gas at the beginning of the HTC conversion, while the increase in the secondary char was more stable during the course of HTC.

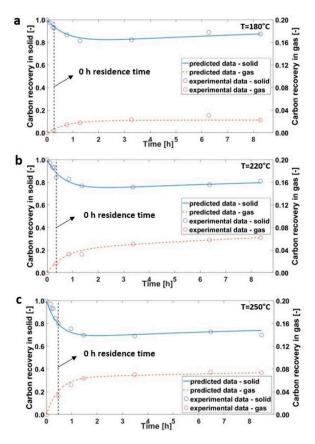


Figure 4. Carbon recovery in solid and gas for olive trimmings: (a) $180 \,^{\circ}$ C, (b) $220 \,^{\circ}$ C, and (c) $250 \,^{\circ}$ C.

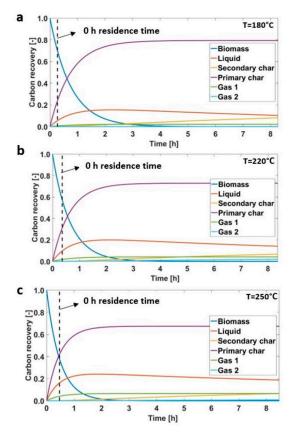


Figure 5. Carbon recovery (-) into HTC products versus residence time (h) for olive trimmings: (a) $180 \,^{\circ}$ C, (b) $220 \,^{\circ}$ C, and (c) $250 \,^{\circ}$ C.

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The Arrhenius plots for the different reaction rate constants k_i are shown in Figure 6. The activation energies $E_{a,i}$ and the pre-exponential factors $k_{0,i}$ for each HTC reactions were determined from the slope of the curves $(-E_{a,i}/R)$ and the intercept with the *y*-axis. The values of the Arrhenius parameters are reported in Table 3.

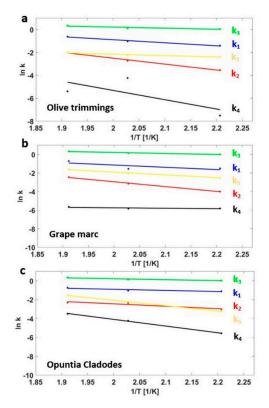


Figure 6. Arrhenius plot for determination of the activation energies $(E_{a,i})$ and pre-exponential factors $(k_{0,i})$: (a) olive trimmings, (b) grape marc, and (c) *Opuntia ficus indica cladodes*.

Table 3. Pre-exponential factors and activation energies evaluated by the Arrhenius plot for the HTC reactions for olive trimmings, grape marc, and *Opuntia ficus indica*.

Parameters	Olive Trimmings	Grape Marc	Opuntia Ficus Indica
k _{0,1} (s ⁻¹)	82.32	41.55	4.31
$k_{0.2} (s^{-1})$	2.51×10^{3}	1.69×10^{3}	14.71
$k_{0,3} (s^{-1})$	7.99	11.28	9.37
$k_{0,4} (s^{-1})$	5.38×10^{4}	0.0086	2.40×10^{4}
$k_{0,5} (s^{-1})$	1.41	52.78	$1.14 imes 10^4$
$E_{a,1}$ (kJ/mol)	22.03	20.23	9.82
$E_{a,2}(kJ/mol)$	42.93	43.14	21.33
$E_{a,3}$ (kJ/mol)	7.75	9.18	8.39
$E_{a,4}$ (kJ/mol)	67.35	4.11	58.92
E _{a,5} (kJ/mol)	10.37	24.41	7.44

The activation energies relevant to the formation of primary and secondary char ($E_{a,3}$ and $E_{a,5}$) were relatively small in comparison with the activation energies involved in liquid and gas formation, supporting what was previously reported on the relative rate of the various HTC reaction paths. However, it is important to point out that the activation energy values here obtained were relevant to our experimental settings and, consequently, reflected the experimental conditions we used in terms of water to biomass ratio and, likely even more important, reactor heating rate.

Even if common trends of the kinetic parameters are clearly identifiable in Table 3, some data appear clearly out of trend. This could be due to a couple of reasons: on the one hand, the experimental data available for grape marc and *Opuntia ficus indica* were quite limited (only three residence times

tested); on the other hand, a clear interrelation existed between pre-exponential factor and activation energy. Actually, it is worth noticing that the pre-exponential factors followed exactly the same trend as the activation energies: the higher $E_{a,i}$, the higher $k_{0,i}$. This can be explained by considering the mathematical aspects behind Arrhenius's formulation: the interrelations between $k_{0,i}$ and $E_{a,i}$ can be expressed as in Equation (19) [59]:

$$ln k_{0,i} = a E_{a,i} + b$$
(19)

Equation (19) highlights that a change in activation energy leads to a change in pre-exponential factors, and vice-versa. Such a phenomenon can be also related to the so-called compensation effect [60]. Nevertheless, in general, the results obtained with the Arrhenius plot confirmed that the reaction leading to the production of primary char was kinetically favoured, the reaction producing gas was the slowest, and the production of liquid and secondary char occurred at an intermediate rate compared to the previous ones.

4. Conclusions

Hydrothermal carbonization of olive trimmings showed that product distribution and hydrochar properties at 180–250 °C were strongly affected by temperature and residence time. The dependence appeared univocal as far as the temperature is concerned, while residence time affected in a more complex fashion. As usually found in the literature, the higher the temperature, the lower the hydrochar yield, and the higher were its HHV, carbon, and fixed carbon. The same applied when the residence time increased from zero (i.e., the feedstock was heated up to HTC temperature and then immediately cooled down) to 3 h. Increasing the residence time (HTC tests performed at 6 and 8 h) did not translate into a further decrease in hydrochar yield, which conversely tended to increase slightly. Carbon content still increased until 6 h residence time, and then stabilized, and fixed carbon content tended to decrease at the highest residence times tested (6–8 h). This particular behavior can be explained considering that in the high residence time range, polymerization/condensation occurred in the liquid phase, and the secondary char and/or tarry compounds segregated from the liquid phase and precipitated into the solid phase.

An in-depth study on the heat-up transient phase experienced by the feedstock before reaching HTC set-point temperature also allowed for very interesting results. Results clearly showed a change in properties during the transient time to 220 and 250 $^{\circ}$ C: carbonization already began during the heat-up phase in the 180–220 $^{\circ}$ C temperature range, and at a corresponding heating time of 15–20 min due to the HTC system utilized. Actually, regarding this temperature range, there was an evident increase in hydrochar HHV and a substantial decrease in hydrochar yield and oxygen content. Analyzing the oxygen recovery (a function of both hydrochar yield and oxygen content in the hydrochar), it seems that, during heat-up, dehydration prevailed until 180 $^{\circ}$ C, while decarboxylation became predominant in the temperature range 180–250 $^{\circ}$ C.

Furthermore, an innovative HTC reaction kinetics model was developed and fully explained. It consisted of a lumped component model, which accounted for reactions leading to the production of both primary and secondary char, liquid and gas phases. The model, written as a MATLABTM code and optimized with experimental data using best fitting routines, effectively simulated and predicted the carbon distribution among the hydrochar and gas phase. The model was run considering HTC experimental data relevant to three different kinds of agro-waste: olive trimmings (from this study), grape marc, and *Opuntia ficus indica*. The model, in good agreement with experimental data, showed that carbon recovery in hydrochar decreased up to 3 h for all the HTC temperatures and then tended to stabilize or even to slightly increase. In contrast, carbon recovery in gas increased with time and it was maximal (about 8%) at the highest HTC temperatures of 250 °C. The model was based on a simplified reaction scheme where biomass converted into primary char, liquid, and gas phases, and the liquid phase could react further to produce secondary char and additional gas. Through modeling, the kinetics parameters (reaction rate constant, pre-exponential factor, and activation energy) were evaluated for all the simplified HTC reactions considered, the three feedstocks analyzed and the three

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HTC temperature tested: 180, 220, and $250\,^{\circ}$ C. Biomass to primary char conversion resulted in the fastest HTC reaction, while the reactions leading to gas turned out to be the slowest. The reaction leading to liquid intermediates occurred at an in-between rate. The production of secondary char, which was not negligible, occurred with a reaction order in the range from 1 to 2, where the higher the HTC temperature, the higher the reaction order. The activation energies determined through the Arrhenius plots for primary and secondary char formation resulted relatively small compared to the other HTC reaction paths. This supports the evidence that char formation during HTC is the most favoured reaction.

For all the examined conditions (T = 180-250 °C, t = 0-8 h), the model fitting errors were lower than 10%. The kinetics model turned out to be in good agreement with the carbon recovery experimental data also during the heat-up transient time. The presented HTC reaction kinetics model is therefore a reliable tool for the prediction of carbon distribution among the HTC products of lignocellulosic biomasses.

Supplementary Materials: The following is available online at http://www.mdpi.com/1996-1073/12/3/516/s1, MATLAB code of the HTC reaction kinetics model.

Author Contributions: M.L. developed and run the HTC reaction kinetics model, discussed the results, prepared the original draft of the paper, and edited the paper in its final form. M.V. performed the HTC experimental activity, wrote concerning this, and helped in the revision of the manuscript. L.F. supervised the work, and in particular, the development of the reaction kinetics model, contributed to the discussions of the results, and revised the paper.

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Conflicts of Interest: The authors declare no conflicts of interest.

Appendix A

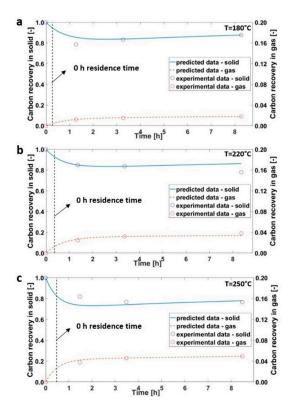


Figure A1. Carbon recovery in solid and gas phases for grape marc: (a) 180 °C, (b) 220 °C, and (c) 250 °C.

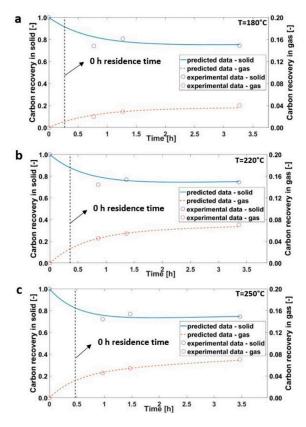


Figure A2. Carbon recovery in solid and gas phases for *Opuntia ficus indica*: (a) 180 $^{\circ}$ C, (b) 220 $^{\circ}$ C, and (c) 250 $^{\circ}$ C.

Table A1. Percentage differences between model predictions and experimental data, calculated by means of Equation (18) for carbon recovery in solid phase for olive trimmings, grape marc, and *Opuntia ficus indica*.

Process Conditions	Error (%)				
1 10cess Conditions	Olive Trimmings	Grape Marc	Opuntia Ficus Indica		
120 °C, 0 h	0.32	-	-		
150 °C, 0 h	1.15	-	-		
180 °C, 0 h	0.71	-	-		
180 °C, 0.5 h	0.62	-	11.17		
180 °C, 1 h	3.37	8.63	3.02		
180 °C, 3 h	0.55	0.95	1.32		
180 °C, 6 h	3.62	-	-		
180 °C, 8 h	0.02	0.00	-		
Avg. 180 °C	1.30	3.19	5.17		
120 °C, 0 h	1.35	-	-		
150 °C, 0 h	0.72	-	-		
180 °C, 0 h	2.12	-	-		
220 °C, 0 h	4.87	-	-		
220 °C, 0.5 h	2.94	-	10.23		
220 °C, 1 h	0.14	0.30	1.08		
220 °C, 3 h	0.24	0.03	0.66		
220 °C, 6 h	0.56	-	-		
220 °C, 8 h	1.67	10.58	-		
Avg. 220 °C	1.62	3.64	3.99		
120 °C, 0 h	4.2	-	-		
150 °C, 0 h	4.21	-	-		

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Process Conditions	Error (%)				
Trocess Conditions	Olive Trimmings	Grape Marc	Opuntia Ficus Indica		
180 °C, 0 h	4.14	-	-		
220 °C, 0 h	2.74	-	-		
250 °C, 0 h	0.25	-	-		
250 °C, 0.5 h	4.22	-	5.05		
250 °C, 1 h	0.14	10.19	4.03		
250 °C, 3 h	1.89	3.27	0.23		
250 °C, 6 h	0.00	-	-		
250 °C, 8 h	5.69	1.72	-		
Avg. 250 °C	2.75	5.06	3.10		

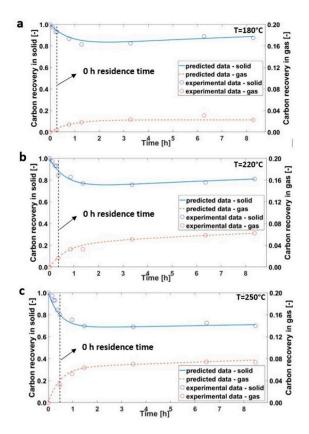


Figure A3. Carbon recovery in solid and gas phases for olive trimmings fixing n = 1: (a) 180 °C, (b) 220 °C, and (c) 250 °C.

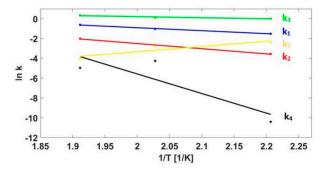


Figure A4. Arrhenius plot for determination of the activation energies $(E_{a,i})$ and pre-exponential factors $(k_{0,i})$ for olive trimmings, fixing n = 1.

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Table A2. Reaction rate constants (k_i) fixing reaction order $(n = 1)$ for the simplified reactions path of
Figure 2 for olive trimmings.

Parameters	Olive Trimmings $(n = 1)$				
T (°C)	180	220	250		
$k_1 (s^{-1})$	0.22	0.35	0.54		
$k_2 (s^{-1})$	0.03	0.07	0.13		
$k_3 (s^{-1})$	1.01	1.11	1.41		
$k_4 (s^{-1})$	0.00	0.01	0.007		
$k_5 (s^{-1})$	0.09	0.06	0.02		
n (-)	1.00	1.00	1.00		

Table A3. Pre-exponential factors and activation energies evaluated using the Arrhenius plot for the HTC reactions for olive trimmings fixing n = 1 (n.a. = not available due to mathematical constraints: k_4 resulted almost zero for T = 180 °C, such that $ln(k_4)$ became extremely low—Figure A4—and not scientifically sound; the value of k_5 was found to decrease with temperature—Figure A4, Table A2—so the value of $E_{a,5}$ became mathematically negative, which has no physical meaning).

Parameters	Olive Trimmings $(n = 1)$
$k_{0,1}$ (s ⁻¹)	163.18
$k_{0,2} (s^{-1})$	2.75×10^{3}
$k_{0,3} (s^{-1})$	10.97
$k_{0,4} (s^{-1})$	n.a.
$k_{0,5} (s^{-1})$	n.a.
$E_{a,1}$ (kJ/mol)	24.95
$E_{a,2}(kJ/mol)$	43.33
$E_{a,3}$ (kJ/mol)	9.09
$E_{a,4}$ (kJ/mol)	n.a.
$E_{a,5}$ (kJ/mol)	n.a.

References

- 1. Dinc, G.; Yel, E. Self-catalyzing pyrolysis of olive pomace. *J. Anal. Appl. Pyrolysis* **2018**, 134, 641–646. [CrossRef]
- 2. Volpe, M.; D'Anna, C.; Messineo, S.; Volpe, R.; Messineo, A. Sustainable Production of Bio-Combustibles from Pyrolysis of Agro-Industrial Wastes. *Sustainability* **2014**, *6*, 7866–7882. [CrossRef]
- 3. Volpe, M.; Panno, D.; Volpe, R.; Messineo, A. Upgrade of citrus waste as a biofuel via slow pyrolysis. *J. Anal. Appl. Pyrolysis* **2015**, *115*, 66–76. [CrossRef]
- 4. Borel, L.D.M.S.; Lira, T.S.; Ribeiro, J.A.; Ataíde, C.H.; Barrozo, M.A.S. Pyrolysis of brewer's spent grain: Kinetic study and products identification. *Ind. Crops Prod.* **2018**, *121*, 388–395. [CrossRef]
- 5. Sánchez, J.D.; Ramírez, G.E.; Barajas, M.J. Comparative kinetic study of the pyrolysis of mandarin and pineapple peel. *J. Anal. Appl. Pyrolysis* **2016**, *118*, 192–201. [CrossRef]
- 6. Luz, C.; Cordiner, S.; Manni, A.; Mulone, V. Biomass fast pyrolysis in a shaftless screw reactor: A 1-D numerical model. *Energy* **2018**, *157*, 792–805. [CrossRef]
- 7. Volpe, M.; Goldfarb, J.L.; Fiori, L. Hydrothermal carbonization of Opuntia ficus indica cladodes: Role of process parameters on hydrochar properties. *Bioresour. Technol.* **2018**, 247, 310–318. [CrossRef] [PubMed]
- 8. Kruse, A.; Funke, A.; Titirici, M.-M. Hydrothermal conversion of biomass to fuels and energetic materials. *Curr. Opin. Chem. Biol.* **2013**, *17*, 515–521. [CrossRef] [PubMed]
- 9. Kambo, H.S.; Dutta, A. A comparative review of biochar and hydrochar in terms of production, physico-chemical properties and applications. *Renew. Sustain. Energy Rev.* **2015**, *45*, 359–378. [CrossRef]
- 10. Saha, N.; Saba, A.; Reza, M.T. Effect of hydrothermal carbonization temperature on pH, dissociation constants, and acidic functional groups on hydrochar from cellulose and wood. *J. Anal. Appl. Pyrolysis* **2018**. [CrossRef]
- 11. Benavente, V.; Calabuig, E.; Fullana, A. Upgrading of moist agro-industrial wastes by hydrothermal carbonization. *J. Anal. Appl. Pyrolysis* **2015**, *113*, 89–98. [CrossRef]

Energies **2019**, 12, 516 18 of 20

12. Düdder, H.; Wütscher, A.; Vorobiev, N.; Schiemann, M.; Scherer, V.; Muhler, M. Oxidation characteristics of a cellulose-derived hydrochar in thermogravimetric and laminar flow burner experiments. *Fuel Process*. *Technol.* **2016**, *148*, 85–90. [CrossRef]

- 13. Sabio, E.; Álvarez-Murillo, A.; Román, S.; Ledesma, B. Conversion of tomato-peel waste into solid fuel by hydrothermal carbonization: Influence of the processing variables. *Waste Manag.* **2016**, *47*, 122–132. [CrossRef] [PubMed]
- 14. Volpe, M.; Wüst, D.; Merzari, F.; Lucian, M.; Andreottola, G.; Kruse, A.; Fiori, L. One stage olive mill waste streams valorisation via hydrothermal carbonisation. *Waste Manag.* **2018**, *80*, 224–234. [CrossRef] [PubMed]
- 15. Erdogan, E.; Atila, B.; Mumme, J.; Reza, M.T.; Toptas, A.; Elibol, M.; Yanik, J. Characterization of products from hydrothermal carbonization of orange pomace including anaerobic digestibility of process liquor. *Bioresour. Technol.* **2015**, *196*, 35–42. [CrossRef]
- 16. Volpe, M.; Fiori, L. From olive waste to solid biofuel through hydrothermal carbonisation: The role of temperature and solid load on secondary char formation and hydrochar energy properties. *J. Anal. Appl. Pyrolysis* **2017**, 124, 63–72. [CrossRef]
- 17. Lucian, M.; Volpe, M.; Gao, L.; Piro, G.; Goldfarb, J.L.; Fiori, L. Impact of hydrothermal carbonization conditions on the formation of hydrochars and secondary chars from the organic fraction of municipal solid waste. *Fuel* **2018**, 233, 257–268. [CrossRef]
- 18. Arauzo, P.J.; Olszewski, M.P.; Kruse, A. Hydrothermal Carbonization Brewer's Spent Grains with the Focus on Improving the Degradation of the Feedstock. *Energies* **2018**, 3226. [CrossRef]
- 19. Mäkelä, M.; Volpe, M.; Volpe, R.; Fiori, L.; Dahl, O. Spatially resolved spectral determination of polysaccharides in hydrothermally carbonized biomass. *Green Chem.* **2018**, 20, 1114–1120. [CrossRef]
- 20. Gao, L.; Volpe, M.; Lucian, M.; Fiori, L.; Goldfarb, J.L. Does Hydrothermal Carbonization as a Biomass Pretreatment Reduce Fuel Segregatuion of Coal-Biomass Blends During Oxidation? *Energy Convers. Manag.* **2019**, *181*, 93–104. [CrossRef]
- 21. Mäkelä, M.; Fullana, A.; Yoshikawa, K. Ash behavior during hydrothermal treatment for solid fuel applications. Part 1: Overview of different feedstock. *Energy Convers. Manag.* **2016**, *121*, 402–408. [CrossRef]
- 22. Codignole Luz, F.; Volpe, M.; Fiori, L.; Manni, A.; Cordiner, S.; Mulone, V.; Rocco, V. Spent coffee enhanced biomethane potential via an integrated hydrothermal carbonization-anaerobic digestion process. *Bioresour. Technol.* **2018**, 256, 102–109. [CrossRef] [PubMed]
- 23. Breulmann, M.; Van Afferden, M.; Müller, R.A.; Schulz, E.; Fühner, C. Process conditions of pyrolysis and hydrothermal carbonization affect the potential of sewage sludge for soil carbon sequestration and amelioration. *J. Anal. Appl. Pyrolysis* **2017**, 124, 256–265. [CrossRef]
- 24. Duman, G.; Toptas, A.; Ucar, S.; Yanik, J. Comparative evaluation of dry and wet carbonization of agro industrial wastes for the production of soil improver. *J. Environ. Chem. Eng.* **2018**, *6*, 3366–3375. [CrossRef]
- 25. Titirici, M.M. Hydrothermal Carbons: Synthesis, Characterization, and Applications. In *Novel Carbon Adsorbents*; Tascón, J.M.D., Ed.; Elsevier Ltd.: Oxford, UK, 2012; pp. 351–399.
- 26. Benstoem, F.; Becker, G.; Firk, J.; Kaless, M.; Wuest, D.; Pinnekamp, J.; Kruse, A. Elimination of micropollutants by activated carbon produced from fibers taken from wastewater screenings using hydrothermal carbonization. *J. Environ. Manag.* 2018, 211, 278–286. [CrossRef] [PubMed]
- 27. Lucian, M.; Fiori, L. Hydrothermal Carbonization of Waste Biomass: Process Design, Modeling, Energy Efficiency and Cost Analysis. *Energies* **2017**, *10*. [CrossRef]
- 28. Hitzl, M.; Corma, A.; Pomares, F.; Renz, M. The hydrothermal carbonization (HTC) plant as a decentral biorefinery for wet biomass. *Catal. Today* **2015**, 257, 154–159. [CrossRef]
- 29. Missaoui, A.; Bostyn, S.; Belandria, V.; Cagnon, B.; Sarh, B. Hydrothermal carbonization of dried olive pomace: Energy potential and process performances. *J. Anal. Appl. Pyrolysis* **2017**, *128*, 281–290. [CrossRef]
- 30. Christoforou, E.; Fokaides, P.A. A review of olive mill solid wastes to energy utilization techniques. *Waste Manag.* **2016**, *49*, 346–363. [CrossRef]
- 31. Poerschmann, J.; Baskyr, I.; Weiner, B.; Koehler, R.; Wedwitschka, H.; Kopinke, F.-D. Hydrothermal carbonization of olive mill wastewater. *Bioresour. Technol.* **2013**, *133*, 581–588. [CrossRef]
- 32. Volpe, M.; Fiori, L.; Volpe, R.; Messineo, A. Upgrading of Olive Tree Trimmings Residue as Biofuel by Hydrothermal Carbonization and Torrefaction: a Comparative Study. *Chem. Eng. Trans.* **2016**, *50*, 13–18. [CrossRef]

33. Jatzwauck, M.; Schumpe, A. Kinetics of hydrothermal carbonization (HTC) of soft rush. *Biomass Bioenergy* **2015**, 75, 94–100. [CrossRef]

- 34. Jung, D.; Kruse, A. Evaluation of Arrhenius-type Overall Kinetic Equations for Hydrothermal Carbonization. J. Anal. Appl. Pyrolysis 2017, 127, 286–291. [CrossRef]
- 35. Zhuang, X.; Zhan, H.; Song, Y.; He, C.; Huang, Y.; Yin, X. Insights into the evolution of chemical structures in lignocellulose and non lignocellulose biowastes during hydrothermal carbonization (HTC). *Fuel* **2019**, 236, 960–974. [CrossRef]
- 36. Funke, A.; Ziegler, F. Hydrothermal carbonization of biomass: A summary and discussion of chemical mechanisms for process engineering. *Biofuels Bioproduct Biorefinery* **2010**, 160–177. [CrossRef]
- 37. Liu, Z.; Balasubramanian, R. Hydrothermal Carbonization of Waste Biomass for Energy Generation. *Procedia Environ. Sci.* **2012**, *16*, 159–166. [CrossRef]
- 38. Reza, M.T.; Yan, W.; Uddin, M.H.; Lynam, J.G.; Hoekman, S.K.; Coronella, C.J.; Vásquez, V.R. Reaction kinetics of hydrothermal carbonization of loblolly pine. *Bioresour. Technol.* **2013**, *139*, 161–169. [CrossRef]
- 39. Borrero-lópez, A.M.; Masson, E.; Celzard, A.; Fierro, V. Modelling the reactions of cellulose, hemicellulose and lignin submitted to hydrothermal treatment. *Ind. Crops Products* **2018**, *124*, 919–930. [CrossRef]
- 40. Álvarez-Murillo, A.; Sabio, E.; Ledesma, B.; Rom, S. Generation of biofuel from hydrothermal carbonization of cellulose. Kinetics modelling. *Energy* **2016**, *94*, 600–608. [CrossRef]
- 41. Baratieri, M.; Basso, D.; Patuzzi, F.; Castello, D.; Fiori, L. Kinetic and Thermal Modeling of Hydrothermal Carbonization Applied to Grape Marc. *Chem. Eng. Trans.* **2015**, *43*, 505–510. [CrossRef]
- 42. Karayildirim, T.; Sinağ, A.; Kruse, A. Char and coke formation as unwanted side reaction of the hydrothermal biomass gasification. *Chem. Eng. Technol.* **2008**, *31*, 1561–1568. [CrossRef]
- 43. Knezevic, D.; Van Swaaij, W.; Kersten, S. Hydrothermal Conversion Of Biomass. II. Conversion Of Wood, Pyrolysis Oil, And Glucose In Hot Compressed Water. *Ind. Eng. Chem. Res.* **2010**, 49, 104–112. [CrossRef]
- 44. Basso, D.; Patuzzi, F.; Castello, D.; Baratieri, M.; Rada, C.E.; Weiss-Hortala, E.; Fiori, L. Agro-industrial waste to solid biofuel through hydrothermal carbonization. *Waste Manag.* **2016**, *47*, 114–121. [CrossRef] [PubMed]
- 45. Fiori, L.; Basso, D.; Castello, D.; Baratieri, M. Hydrothermal Carbonization of Biomass: Design of a Batch Reactor and Preliminary Experimental Results. *Chem. Eng. Trans.* **2014**, *37*, 55–60. [CrossRef]
- Basso, D.; Weiss-Hortala, E.; Patuzzi, F.; Castello, D.; Baratieri, M.; Fiori, L. Hydrothermal carbonization of off-specification compost: A byproduct of the organic municipal solid waste treatment. *Bioresour. Technol.* 2015, 182, 217–224. [CrossRef] [PubMed]
- 47. Berge, N.D.; Ro, K.S.; Mao, J.; Flora, J.R.V.; Chappell, M.A.; Bae, S. Hydrothermal Carbonization of Municipal Waste Streams. *Environ. Sci. Technol* **2011**, 45, 5696–5703. [CrossRef] [PubMed]
- 48. Lu, X.; Jordan, B.; Berge, N.D. Thermal conversion of municipal solid waste via hydrothermal carbonization: Comparison of carbonization products to products from current waste management techniques. *Waste Manag.* 2012, 32, 1353–1365. [CrossRef]
- 49. Romàn, S.; Libra, J.; Berge, N.; Sabio, E.; Ro, K.; Li, L.; Ledesma, B.; Bae, S. Hydrothermal Carbonization: Modeling, Final Properties Design and Applications: A Review. *Energies* **2018**, *11*, 216. [CrossRef]
- 50. Lucian, M.; Piro, G.; Fiori, L. A Novel Reaction Kinetics Model for Estimating the Carbon Content into Hydrothermal Carbonization Products. *Chem. Eng. Trans.* **2018**, *65*, 379–384. [CrossRef]
- 51. Libra, J.A.; Ro, K.S.; Kammann, C.; Funke, A.; Berge, N.D.; Neubauer, Y.; Titirici, M.-M.; Fühner, C.; Bens, O.; Kern, J.; et al. Hydrothermal carbonization of biomass residuals: a comparative review of the chemistry, processes and applications of wet and dry pyrolysis. *Biofuels* **2011**, *2*, 71–106. [CrossRef]
- 52. Lei, Y.-Q.; Su, Q.-H.; Tian, R. Morphology evolution, formation mechanism and adsorption properties of hydrochars prepared by hydrothermal carbonization of corn stalk. *RSC Adv.* **2016**, *109*. [CrossRef]
- Ulbrich, M.; Preßl, D.; Fendt, S.; Gaderer, M.; Splietho, H. Impact of HTC reaction conditions on the hydrochar properties and CO2 gasification properties of spent grains. *Fuel Process. Technol.* 2017, 167, 663–669. [CrossRef]
- 54. Titirici, M.M. Sustainable Carbon Materials from Hydrothermal Processes, 1st ed.; Titirici, M.-M., Ed.; John Wiley & Sons, Ltd.: London, UK, 2013; ISBN 9781118622179.
- 55. Kruse, A.; Badoux, F.; Grandl, R.; Wüst, D. Hydrothermale Karbonisierung: 2. Kinetik der Biertreber-Umwandlung. *Chemie-Ingenieur-Technik* **2012**, *84*, 509–512. [CrossRef]

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56. Almagrbi, A.M.; Hatami, T.; Glisic, S.B.; Orlovi, A.M. Determination of kinetic parameters for complex transesterification reaction by standard optimisation methods. *Hemijska Industrija* **2014**, *68*, 149–159. [CrossRef]

- 57. Urych, B. Determination of kinetic parameters of coal pyrolysis to simulate the process of underground coal gasification (UCG). *J. Sustain. Mining* **2014**, *13*, 3–9. [CrossRef]
- 58. Hwang, I.; Aoyama, H.; Matsuto, T.; Nakagishi, T.; Matsuo, T. Recovery of solid fuel from municipal solid waste by hydrothermal treatment using subcritical water. *Waste Manag.* **2012**, *32*, 410–416. [CrossRef] [PubMed]
- 59. Liu, N.; Wang, B.; Fan, W. Kinetic Compensation Effect in the Thermal Decomposition of Biomass in Air Atmosphere. *Fire Saf. Sci.* **2003**, *7*, 581–592. [CrossRef]
- 60. Fiori, L.; Valbusa, M.; Lorenzi, D.; Fambri, L. Modeling of the devolatilization kinetics during pyrolysis of grape residues. *Bioresour. Technol.* **2012**, *103*, 389–397. [CrossRef]



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