Supplementary Information

Table S1. Cyanotoxins content in aquatic invertebrates. Concentrations of microcystins are given in equivalents of MC-LR, and concentrations of saxitoxins are given in equivalents of STX. Concentrations represent the range for the different organs/tissues in [(a)] μg g^{-1} DW, [(b)] ng $individual^{-1}$, [(c)] μg gC^{-1} or [(d)] pmol $individual^{-1}$. Concentrations in μg g^{-1} WW were converted to μg g^{-1} DW multiplying by 10. MC = microcystins; NOD = nodularin; CYN = cylindrosdpermopsin; STX = saxitoxins; ANTX-a = anatoxin-a; BMAA = β-*N*-methylamino-L-alanine. I-EC = Ion-Exchange Chromatography; HPLC = High Performance Liquid Chromatography; LC-MS = Liquid Chromatography–Mass Spectrometry; ELISA = Enzyme Linked Immunosorbant Assay; PP2A = Phosphatase 2A Inhibition Assay; PP1 = Phosphatase 1 Inhibition Assay; MMPB = 3-metoxi-2methyl-4-phenylbutiric acid; EC-MS = Capillary Eletrophoresis–Mass Spectrometry; GC-MS = Gas Chromatography–Mass Spectrometry; DW = dry weight; TEH = Total Extractable Hepatotoxins; ND = not detectable.

Group of organism/Taxa	Dominant cyanobacteria	Toxin	Analytical method	Concentration	Reference
ZOOPLANKTON					
Natural community	*Microcystis* spp.	MC-LR, -RR, -YR	I-EC	75.0–1387 [(a)]	Watanabe *et al.* (1992)[1]
Natural community	*M. aeruginosa*	MC-LR	HPLC/PP2A	ND–67.0 [(a)]	Kotak *et al.* (1996)[2]
Daphnia magna (Cladocera)—Lab experiment	*M. aeruginosa*	MC	ELISA	0.2–24.5 [(a)]	Thorstrup and Christoffersen (1999)[3]
Daphnia magna (Cladocera)—Field experiment	*M. aeruginosa*	MC	ELISA	0.3–15.5 [(a)]	
D. parvula (Cladocera)	*M. aeruginosa*, *Chroococcus minor*	MC	ELISA	ND–71.2 [(b)]	Mohamed (2001)[4]
Natural community	*M. aeruginosa*	MC	ELISA	0.3–16.4 [(a)]	Ferrão-Filho *et al.* (2002)[5]
Eurytemora affinis (Copepoda)	*Nodularia spumigena*	NOD	ELISA/PP1	0.031–0.101 [(b)]	Lehtiniemi *et al.* (2002)[6]
Acartia bifilosa (Copepoda)	*N. spumigena*	NOD	ELISA/PP1	0.011–0.013 [(b)]	Kozlowsky-Suzuki *et al.* (2003)[7]
Eurytemora affinis (Copepoda)	*N. spumigena*	NOD	ELISA/PP1	0.011–0.024 [(b)]	
Acartia tonsa (Copepoda)	*N. spumigena*	NOD	Radiolabeled NOD	0.37 [(c)]	Karjalainen *et al.* (2003)[8]
Eurytemora affinis (Copepoda)	*N. spumigena*	NOD	Radiolabeled NOD	0.60 [(c)]	
Strobidium sulcatus (Ciliata)	*N. spumigena*	NOD	Radiolabeled NOD	1.55 [(c)]	
D. magna (Cladocera)	*Cylindrospermopsis raciborskii*	CYN	HPLC	0.020–0.025 [(b)]	Nogueira *et al.* (2004)[9]
D. magna (Cladocera)	*Aphanizomenon issatschenkoi*	STXs	HPLC	0.07–0.38 [(d)]	Nogueira *et al.* (2004)[10]
Natural community	*M. aeruginosa*, *Planktothrix agadhii*	MC	LC-MS/HPLC/ELISA	ND–1226 [(a)]	Ibelings *et al.* (2005)[11]
Natural community	*M. aeruginosa*	MC	PP2A/HPLC	0.7–1.02 [(a)]	Lehman *et al.* (2005)[12]
Several species	*N. spumigena*	NOD	ELISA	ND–6.2 [(a)]	Karjalainen *et al.* (2006)[13]

Table S1. *Cont.*

Eurytemora affinis (Copepoda)	*N. spumigena*	NOD	ELISA/PP1	12.1–46.4 [(a)]	
Natural community	*Microcystis* sp.	MC	ELISA	5.0–25.0 [(a)]	Smith and Haney (2006)[14]
D. pulicaria (Cladocera)	*Planktothrix agardhii*, *P. rubescens*	MC	PP2A	74.0–1099 [(a)]	Oberhaus *et al.* (2007)[15]
Natural community	*Microcystis* sp., *C. raciborskii*, *Anabaena circinalis*	MC	ELISA	5.0–63.2 [(a)]	Rocha (2007)[16]
Pleopsis polyphemoides (Cladocera)	*N. spumigena*	NOD	ELISA	ND–23.6 [(a)]	Karjalainen *et al.* (2008)[17]
Natural community (*E. affinis* and *Pseudodiaptomus forbesii*)	*Microcystis* sp.	MC	PP2A/HPLC	0.18–5.93 [(a)]	Lehman *et al.* (2008)[18]
Amphipod species	*Microcystis* sp.	MC	PP2A/HPLC	0.74–5.18 [(a)]	
Worms (unknown species)	*Microcystis* sp.	MC	PP2A/HPLC	12.0 [(a)]	
Jellyfish (unknown species)	*Microcystis* sp.	MC	PP2A/HPLC	2.2 [(a)]	
Eurytemora affinis (Copepoda)	*N. spumigena*	NOD	ELISA	0.04–0.76 [(b)]	Gorokhova and Engström-Öst (2009)[19]
Eurytemora affinis (Copepoda)	*N. spumigena*	NOD	HPLC-MS/LC-ESI-MS	0.0013–0.0143 [(b)]	Sopanen *et al.* (2009)[20]
Natural community	*Microcystis* sp.	MC	PP2A/HPLC	0.40–1.43 [(a)]	Lehman *et al.* (2010)[21]
Amphipod species	*Microcystis* sp.	MC	PP2A/HPLC	0.77 [(a)]	
CRUSTACEANS					
Cherax quadricarinatus (Decapoda)	*C. raciborskii*	CYN	HPLC	0.54–4.3 [(a)]	Saker and Eaglesham (1999)[22]
Echinogammarus ischnus (Amphipoda)—Field data	*M. aeruginosa*	MC	ELISA	0.2 [(a)]	Babcock-Jackson (2000)[23]
Echinogammarus ischnus (Amphipoda)—Lab experiment	*M. aeruginosa*	MC	ELISA	0.2–0.7 [(a)]	
Procambarus clarkii (Decapoda)	*M. aeruginosa*	MC	ELISA	0.4–2.9 [(a)]	Vasconcelos *et al.* (2001)[24]
Mysis relicta (Decapoda)	*N. spumigena*	NOD	ELISA/PP1	0.5–0.74 [(a)]	Engström-Öst *et al.* (2002)[25]
Crab—unidentified species (Decapoda)	Picoplanktonic cyanobacteria (unidentified)	MC	ELISA	ND–1.0 [(a)]	Magalhães *et al.* (2003)[26]
Shrimp—unidentified species (Decapoda)	Picoplanktonic cyanobacteria (unidentified)	MC	ELISA	ND–0.09 [(a)]	
Palaemon modestus (Decapoda)	*Microcystis* spp., *Anabaena* sp.	MC	LC-MS	ND–9.8 [(a)]	Chen and Xie (2005)[27]
Procambarus clarkii (Decapoda)	*Microcystis* spp., *Anabaena* sp.	MC	LC-MS	0.05–10.0 [(a)]	

Table S1. *Cont.*

Macrobrachium nipponensis (Decapoda)	*Microcystis* spp., *Anabaena* sp.	MC	LC-MS	ND–12.4 [(a)]	
Panaeus monodon (Decapoda)	*Oscillatoria* sp., *Pseudoanabaena* sp., *Microcystis* spp., *Aphanocapsa* sp., *Romeria* sp.	MC/NOD (TEH)	LC-MS	0.006–0.08 [(a)]	Kankaanpää *et al.* (2005)[28]
Neomysis integer (Decapoda)	*N. spumigena*	NOD	Radiolabeled NOD	0.77 [(a)]	Karjalainen *et al.* (2005)[29]
Gammarus zaddachi (Amphipoda)	*N. spumigena*	NOD	ELISA	ND–0.002 [(c)]	Korpinen *et al.* (2006)[30]
Chasmagnathus granulatus (Decapoda)	*M. aeruginosa*	MC	ELISA	0.02–0.32 [(a)]	Dewes *et al.* (2006)[31]
Neohelice granulata (Decapoda)	*Microcystis* sp.	MC	ELISA	0.3–0.34 [(a)]	Dewes *et al.* (2007)[32]
Macrobrachium nipponensis (Decapoda)	*Microcystis* spp.	MC	ELISA/LC-MS	ND–0.76 [(a)]	Song *et al.* (2007)[33]
Macrobrachium nipponensis (Decapoda)	*M. aeruginosa*	MC-LR, -LR-GSH, -LR-Cys	LC-MS	up to 4.9 [(a)]	Zhang *et al.* (2009)[34]
Panaeus duorarum (Decapoda)	Undetermined cyanobacterial blooms	BMAA	HPLC-LC/MS/MS	55–3042	Brand *et al.* (2010)[35]
Callinectes sapidus (Decapoda)	Undetermined cyanobacterial blooms	BMAA	HPLC-LC/MS/MS	ND–6976	
Callinectes sapidus (Decapoda)	*Microcystis* sp., *Anabaena* spp.	MC	ELISA	ND–8.20 [(a)]	Garcia *et al.* (2010)[36]
MOLLUSKS					
Ellipio campanatus, *Corbicula fluminea* (Bivalvia)	*Aphanizomenon flos-aquae*	STXs	HPLC	0.4–0.6 [(a)]	Sasner *et al.* (1984)[37]
Anodonta cygnea (Bivalvia)	*Oscillatoria agardhii*	MC	HPLC	70–130 [(a)]	Eriksson *et al.* (1989)[38]
Anodonta cygnea (Bivalvia)	*Oscillatoria agardhii*	MC	HPLC	30.0 [(a)]	Lindholm *et al.* (1989)[39]
Mussel—unidentified species (Bivalvia)	Unspecified	MC	CE-MS	6.0 [(a)]	Chen *et al.* (1993)[40]
Alanthyria condola (Bivalvia)	*Anabaena circinalis*	STXs	HPLC	0.8–6.2 [(a)]	Negri and Jones (1995)[41]
Mytilus galloprocincialis (Bivalvia)	*M. aeruginosa*	MC	HPLC	1.0–10.5 [(a)]	Vasconcelos (1995)[42]
Helisoma trivolvis (Gastropoda)	*M. aeruginosa*, *A. flos-aquae*,	MC	HPLC	ND–11.0 [(a)]	Kotak *et al.* (1996)[2]
Lymnaea stagnalis (Gastropoda)	*Gomphosphaeria* sp.	MC	HPLC	ND–96.0 [(a)]	
Physa gyrina (Gastropoda)	*Anabaena flos-aquae*	MC	HPLC	ND–121 [(a)]	
Anodonta grandis simpsoniana (Bivalvia)	*M. aeruginosa*, *Anabaena* spp., *Aphanizomenon flos-aquae*	MC	PP2A	0.02–1.35 [(a)]	Prepas *et al.* (1997)[43]
Unio douglasiae (Bivalvia)	*M. aeruginosa*	MC	HPLC-MS	12.0–27.0 [(a)]	Watanabe *et al.* (1997)[44]
Anodonta woodiana (Bivalvia)	*M. aeruginosa*	MC	HPLC-MS	2.1 [(a)]	

Table S1. *Cont.*

Mytilus edulis—Lab experiment (Bivalvia)	*M. aeruginosa*	MC	Limieux-GC-MS	3369 [(a)]	Williams *et al.* (1997)[45]
Mytilus edulis—Lab experiment (Bivalvia)	*M. aeruginosa*	MC	PP2A	2.0 [(a)]	
Mytilus edulis—Field sample (Bivalvia)	*M. aeruginosa*	MC	Limieux-GC-MS	ND–634 [(a)]	
Mytilus edulis—Field sample (Bivalvia)	*M. aeruginosa*	MC	PP2A	0.17–0.22 [(a)]	
Mytilus galloprocincialis (Bivalvia)	*M. aeruginosa*	MC	HPLC-MS	2.0–16.0 [(a)]	Amorim and Vasconcelos (1999)[46]
Helisoma trivolvis (Gastropoda)	*Microcystis* spp.	MC	HPLC	ND–40 [(a)]	Zurawell *et al.* (1999)[47]
Lymnaea stagnalis (Gastropoda)	*Microcystis* spp.	MC	HPLC	ND–140 [(a)]	
Physa gyrina (Gastropoda)	*Microcystis* spp.	MC	HPLC	ND–129 [(a)]	
Dreissena polymorpha (Bivalvia)—Field data	*M. aeruginosa*	MC	ELISA	0.213 [(a)]	Babcock-Jackson (2000)[23]
Dreissena polymorpha (Bivalvia)—Lab experiment	*M. aeruginosa*	MC	ELISA	up to 0.203 [(a)]	
Mytilus edulis (Bivalvia)	*Aphanizomenon* sp., *N. spumigena*, *Anabaena* sp.	NOD/MC	ELISA	ND–2.15[(a)]	Sipiä *et al.* (2001)[48]
Mytilus edulis (Bivalvia)	*Aphanizomenon flos-aquae*, *N. spumigena*	NOD/MC	ELISA/ MALDI-TOF-MS	0.040–1.49[(a)]	Sipiä *et al.* (2002)[49]
Macoma balthica (Bivalvia)	*Aphanizomenon flos-aquae*, *N. spumigena*	NOD/MC	ELISA	0.10–0.13[(a)]	
Anodonta woodiana (Bivalvia)	*Microcystis* spp.	MC	HPLC	ND–12.6 [(a)]	Yokoyama and Park (2002)[50]
Cristaria plicata (Bivalvia)	*Microcystis* spp.	MC	HPLC	ND–297 [(a)]	
Unio douglasiae (Bivalvia)	*Microcystis* spp.	MC	HPLC	ND–420 [(a)]	
Macoma balthica (Bivalvia)	*N. spumigena*	TEH/NOD-like compound	ELISA/HPLC/ MALDI-TOF-MS	0.16–30.3 [(a)]	Lehtonen *et al.* (2003)[51]
Sinotaia histrica—Field sample (Gastropoda)	*M. aeruginosa*, *Anabaena spiroides*	MC	HPLC	ND–19.5 [(a)]	Ozawa *et al.* (2003)[52]
S. histrica—Lab experiment (Gastropoda)	*M. aeruginosa*	MC	HPLC	ND–436 [(a)]	
Unio douglasiae (Bivalvia)—15 °C	*Microcystis ichytioblade*	MC	HPLC	ND–200 [(a)]	Yokoyama and Park (2003)[53]
Unio douglasiae (Bivalvia)—25 °C	*Microcystis ichytioblade*	MC	HPLC	0–630 [(a)]	
Dreissena polymorpha (Bivalvia)	*M. aeruginosa*	MC	LC-MS/MMPB	9.1–16.3 [(a)]	Dionisio-Pires *et al.* (2004)[54]
Pinctada máxima (Bivalvia)	*Trichodesmium erythraeum*	STX	HPLC	0.73 [(a)]	Negri *et al.* (2004)[55]

Table S1. *Cont.*

Anodonta cygnea (Bivalvia)	*A. issatschenkoi*	STXs	HPLC	0.26 [(a)]	Pereira *et al.* (2004)[56]
Anodonta cygnea (Bivalvia)	*C. raciborskii*	CYN	HPLC	2.9–61.5 [(a)]	Saker *et al.* (2004)[57]
Anodonta woodiana (Bivalvia)	*Microcystis* spp.	MC-LR,-RR,-YR	LC-MS/HPLC	0.007–7.88 [(a)]	Chen and Xie (2005)[58]
Hyriopsis cumingii (Bivalvia)	*Microcystis* spp.	MC-LR,-RR,-YR	LC-MS/HPLC	0.015–20.65 [(a)]	
Cristaria plicata (Bivalvia)	*Microcystis* spp.	MC-LR,-RR,-YR	LC-MS/HPLC	0.017–38.48 [(a)]	
Lamprotula leai (Bivalvia)	*Microcystis* spp.	MC-LR,-RR,-YR	LC-MS/HPLC	0.004–13.23 [(a)]	
Bellamya aeruginosa (Gastropoda)	*Microcystis* spp., *Anabaena* sp.	MC	LC-MS	ND–7.42 [(a)]	Chen *et al.* (2005)[59]
L. stagnalis—Juvenile (Gastropoda)	---	MC-LR	ELISA	0.068 [(a)]	Gérard *et al.* (2005)[60]
L. stagnalis—Adult (Gastropoda)	---	MC-LR	ELISA	0.014 [(a)]	
Dreissena polymorpha (Bivalvia)	*M. aeruginosa*, *Planktothrix agardhii*	MC	LC-MS/HPLC/ ELISA	ND–30.0 [(a)]	Ibelings *et al.* (2005)[11]
Clams (unknown species)	*M. aeruginosa*	MC	PP2A/HPLC	0.02 [(a)]	Lehman *et al.* (2005)[12]
Mytilus edulis (Bivalvia)	*N. spumigena*	NOD	LC-ESI-MS	ND–244.9 [(a)]	Svensen *et al.* (2005)[61]
Mytilus edulis (Bivalvia)	*N. spumigena*	NOD	LC-ESI-MS	0.03–13.75 [(a)]	Strogyloudi *et al.* (2006)[62]
Hydrella menziesi (Bivalvia)	*Anabaena* spp., *Microcystis* spp.	MC	LC-MS/ELISA	ND–0.97 [(a)]	Wood *et al.* (2006)[63]
Melanoides tuberculata (Gastropoda)	*C. raciborskii*	CYN	HPLC	ND–250 [(a)]	White *et al.* (2006)[64]
Anodonta sp.—Lake Kastoria (Bivalvia)	*M. aeruginosa*, *M. novacekii*	MC	PP2A	3.27 [(a)]	Gkelis *et al.* (2006)[65]
Viviparus contectus—Lake Pamvotis (Bivalvia)	*M. aeruginosa; A. flos-aquae*	MC	PP2A/ELISA	1.66–3.50 [(a)]	
Lymnaea stagnalis—Juvenile (Gastropoda)	*P. agardhii*	MC	ELISA	ND–194.8 [(a)]	Lance *et al.* (2006)[66]
Lymnaea stagnalis—Adults (Gastropoda)	*P. agardhii*	MC	ELISA	ND–135.8 [(a)]	
Cristaria plicata (Bivalvia)	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	0.004–24.69 [(a)]	Chen and Xie (2007)[67]
Hyriopsis cumingii (Bivalvia)	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	0.005–13.64 [(a)]	
Lamprotula leai (Bivalvia)	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	0.004–7.37 [(a)]	
Mytilus edulis (Bivalvia)	*N. spumigena*	NOD	LC-MS	up to 1.1 [(a)] 0.6–9.8 [(b)]	Kankaanpää *et al.* (2007)[68]
Mytilus edulis (Bivalvia)	*N. spumigena*	NOD	ELISA	0.003–0.14 [(a)]	Mazur-Marzec *et al.* (2007)[69]
Cristaria plicata (Bivalvia)	*Microcystis* spp.	MC	ELISA/LC-MS	0.045–0.73 [(a)]	Song *et al.* (2007)[33]
Bellamya aeruginosa (Gastropoda)	*Microcystis* spp.	MC	ELISA/LC-MS	0.0656–2.34 [(a)]	
Sinotaia histrica (Gastropoda)	*Microcystis* spp., *A. flos-aquae*	MC	HPLC	0.07–23.2 [(a)]	Xie *et al.* (2007)[70]

Table S1. *Cont.*

Potamopyrgus antipodarum (Gastropoda)	*P. agardhii*	MC	ELISA	ND–4.9 [(a)]	Lance *et al.* (2008)[71]
Clams (unknown species)	*Microcystis* sp.	MC	PP2A/HPLC	0.15–0.37 [(a)]	Lehman *et al.* (2008)[18]
Mytilus edulis (Bivalvia)	*N. spumigena*	NOD	LC-MS	0.012–0.08	Sipiä *et al.* (2008)[72]
Mytilus galloprocincialis (Bivalvia)	*M. aeruginosa*	MC	ELISA	ND–0.38 [(a)]	Fernandes *et al.* (2009)[73]
Sphaerium corneum (Bivalvia)	*M. viridis*, *P. agardhii*	MC	ELISA	ND–0.04 [(a)]	Gérard *et al.* (2009)[74]
Pisidum sp. (Bivalvia)	*A. flos-aquae*, *M. aeruginosa*, *P. agardhii*, *A. circinalis*	MC	ELISA	ND–0.39 [(a)]	
Potamopyrgus antipodarum (Gastropoda)	*A. flos-aquae*, *M. aeruginosa*, *P. agardhii*, *A. circinalis*	MC	ELISA	ND–0.35 [(a)]	
Planorbis planorbis (Gastropoda)	*A. flos-aquae*, *M. aeruginosa*, *P. agardhii*, *A. circinalis*	MC	ELISA	ND–0.15 [(a)]	
Radix auricularia (Gastropoda)	*A. flos-aquae*, *M. aeruginosa*, *P. agardhii*, *A. circinalis*	MC	ELISA	ND–1.04 [(a)]	
Radix ovata (Gastropoda)	*A. flos-aquae*, *M. viridis*, *M. aeruginosa*, *P. agardhii*, *A. circinalis*	MC	ELISA	ND–0.20 [(a)]	
Physella acuta (Gastropoda)	*M. viridis*, *P. agardhii*, *A. flos-aquae*, *M. aeruginosa*, *A. circinalis*	MC	ELISA	ND–242.7 [(a)]	
Aplexa hypnorum (Gastropoda)	No cyanobacteria found	MC	ELISA	ND–0.07 [(a)]	
Bellamya aeruginosa (Gastropoda)	*M. aeruginosa*	MC-LR, -LR-GSH, -LR-Cys	LC-MS	ND–9.70 [(a)]	Zhang *et al.* (2009)[34]
Bellamya aeruginosa (Gastropoda)	*M. aeruginosa*	MC-LR,-RR,-YR	LC-MS	ND–12.2 [(a)]	Zhang *et al.* (2009)[75]
Pomacea patula catemacensis (Gatropoda)	*Cylindrospermopsis catemaco*, *C. philippinensis*	CYN STXs	ELISA, HPLC, LC-MS	ND–0.00335 [(a)] ND–0.00104 [(a)]	Berry and Lind (2010)[76]
Utterbackia imbecillis (Bivalvia)	Undetermined cyanobacterial blooms	BMAA	HPLC-LC/MS/MS	251–261	Brand *et al.* (2010)[35]
Crassostra virginica (Bivalvia)	Undetermined cyanobacterial blooms	BMAA	HPLC-LC/MS/MS	281–305	
L. stagnalis (Gastropoda)	*P. agardhii*	MC	LC–ESI-MS/MS ELISA	up to 32.4 [(a)] free MC	Lance *et al.* (2010)[77]

Table S1. *Cont.*

L. stagnalis (Gastropoda)	*P. agardhii*	MC	MMPB LC–ESI-MS/MS	up to 37.5 [(a)] bound MC	
Physia acuta (Gastropoda)	Several	MC	ELISA	ND–4.32 [(a)]	Lance *et al.* (2010)[78]
Several gastropod species	Several	MC	ELISA	0.02–1.26 [(a)]	
L. stagnalis (Gastropoda)—Caged experiment	Several	MC	ELISA	0.02–0.32 [(a)]	
Potamopyrgus antipodarum (Gastropoda)—Caged experiment	Several	MC	ELISA	0.01–0.02 [(a)]	

Table S2. Cyanotoxins content in aquatic vertebrates. Concentrations of microcystins are given in equivalents of MC-LR, and concentrations of saxitoxins are given in equivalents of STX. Concentrations represent the range for the different organs/tissues in μg g^{-1} DW. Concentrations in μg g^{-1} WW were converted to μg g^{-1} DW multiplying by 10 or by 5 (fish muscle). MC = microcystins; NOD = nodularin; CYN = cylindrosdpermopsin; STX = saxitoxins; ANTX-a = anatoxin-a; BMAA = β-N-methylamino-L-alanine. HPLC = High Performance Liquid Chromatography; LC-MS = Liquid Chromatography–Mass Spectrometry; ELISA = Enzyme Linked Immunosorbant Assay; PP2A = Phosphatase 2A Inhibition Assay; PP1 = Phosphatase 1 Inhibition Assay; GC-MS = Gas Chromatography–Mass Spectrometry; WW = wet weight; DW = dry weight. ND = not detectable.

Group of organism/Taxa	Trophic guild	Dominant cyanobacteria	Toxin	Analytical method	Concentration μg g^{-1} DW	Reference
FISH						
Melanotaenia eachamensis (Melanotaeniidae)	Carnivore	*C. raciborskii*	CYN	HPLC	1.2	Saker and Eaglesham (1999)[22]
Dorosoma cepedianum (Clupeidae)	Omnivore	*M. aeruginosa*	MC	ELISA	2.97–11.91	Babcock-Jackson (2000)[23]
Perca flavescens (Percidae)	Carnivore	*M. aeruginosa*	MC	ELISA	0.46–2.30	
Sander vitreus (Percidae)	Carnivore	*M. aeruginosa*	MC	ELISA	0.50	
Morone spp. (Moronidae)	Carnivore	*M. aeruginosa*	MC	ELISA	0.62–2.33	
Semolitus atromaculatus (Cyprinidae)	Omnivore	*M. aeruginosa*	MC	ELISA	0.60–0.94	
Neogobius melanostomus (Gobiidae)—Field data	Carnivore	*M. aeruginosa*	MC	ELISA	0.48–1.87	
Neogobius melanostomus (Gobiidae)—Lab experiment	Carnivore	*M. aeruginosa*	MC	ELISA	up to 2.82	

Table S2. *Cont.*

Platichthys flesus (Pleuronectidae)	Carnivore	*Nodularia spumigena*	NOD	ELISA/PP1	0.05–0.06	Sipiä *et al.* (2000)[79]
Gadus morhua (Gadidae)	Carnivore	*Nodularia spumigena*	NOD	ELISA/PP1	ND–0.14	
Tilapia rendalli (Cichlidae)	Omnivore	*M. aeruginosa*	MC	HPLC/ELISA	ND–678	Magalhães *et al.* (2001)[80]
Platichthys flesus (Pleuronectidae)	Carnivore	*Aphanizomenon* sp., *N. spumigena*, *Anabaena* sp.	NOD/MC	ELISA	ND–0.40	Sipiä *et al.* (2001)[48]
Platichthys flesus (Pleuronectidae)	Carnivore	*Aphanizomenon flos-aquae*, *N. spumigena*	NOD/MC	ELISA/LC-MS/MS	ND–0.41	Sipiä *et al.* (2002)[49]
Gasterosteus aculeatus (Gasterosteidae)	Planktivore	*N. spumigena*	NOD	ELISA/PP1	0.15–0.80	Engström-Öst *et al.* (2002)[25]
Salmo trutta m. *trutta* L. (Salmonidae)	Carnivore	*N. spumigena*	NOD-like compound	ELISA	0.011–1.60	Kankaanpää *et al.* (2002)[81]
Fish muscle—unidentified species	Unknown	Picoplanktonic cyanobacteria (unidentified)	MC	ELISA	ND–0.02	Magalhães *et al.* (2003)[26]
Oreochromis niloticus (Cichlidae)	Omnivore	*M. aeruginosa*	MC	ELISA	0.4–8.3	Mohamed *et al.* (2003)[82]
Tilapia rendalli (Cichlidae)	Omnivore	*M. aeruginosa*	MC	ELISA	0.02–28.0	Soares *et al.* (2004)[83]
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*Microcystis viridis*	MC-RR, -LR	HPLC	49.2–115.3	Xie *et al.* (2004)[84]
Jenynsia multidentata (Anablepidae)	Omnivore	Purified toxin	MC-RR	HPLC	0.55–16.0	Cazenave *et al.* (2005)[85]
Corydoras paleatus (Callichthyidae)	Omnivore	Purified toxin	MC-RR	HPLC	0.2–196.3	
Odontesthes bonariensis (Atherinidae)	Carnivore	Undetermined cyanobacterial blooms	MC-RR	HPLC	ND–10.1	
Perca fluviatilis (Percidae)	Carnivore	*M. aeruginosa*, *Planktothrix agadhii*	MC	LC-MS/HPLC/ELISA	17.0–51.0	Ibelings *et al.* (2005)[11]
Gymnocephalus cernua (Percidae)	Omnivore	*M. aeruginosa*, *Planktothrix agadhii*	MC	LC-MS/HPLC/ELISA	9.0–194	
Osmerus eperlanus (Osmeridae)	Zooplanktivore	*M. aeruginosa*, *Planktothrix agadhii*	MC	LC-MS/HPLC/ELISA	59–874	
Platichthys flesus (Pleuronectidae)	Carnivore	*N. spumigena*	TEH/NOD	ELISA/LC-MS	ND–2.230	Kankaanpää *et al.* (2005)[86]
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	1.16–137.0	Xie *et al.* (2005)[87]
Parabramis perkinensis (Cyprinidae)	Herbivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	0.66–7.5	
Carassius auratus (Cyprinidae)	Omnivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	1.8–42.6	

Table S2. *Cont.*

Cyprinus carpio (Cyprinidae)	Omnivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	0.65–6.46	
Culter erythropterus (Cyprinidae)	Carnivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	2.0–13.7	
Culter ilishaeformis (Cyprinidae)	Carnivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	2.73–5.0	
Pseudobagrus fulvidraco (Bagridae)	Carnivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	2.0–8.0	
Coilia ectenes (Bagridae)	Carnivore	*Microcystis* sp., *Anabaena* sp.	MC	HPLC	ND–0.91	
Esox lucius (Esocidae)	Carnivore	*N. spumigena*	NOD	Radiolabeled NOD	0.41	Karjalainen *et al.* (2005)[29]
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*M. aeruginosa*	MC-LR, -RR, -YR	LC-MS	ND–97.48	Chen *et al.* (2006)[88]
Ascipenser gueldenstaedtii (Acipenseridae)—Lake Pamvotis	Carnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.30–1.44	Gkelis *et al.* (2006)[64]
Carassius auratus (Cyprinidae)—Lake Pamvotis	Omnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.12–5.40	
Carassius gibelio (Cyprinidae)—Lake Kastoria	Omnivore	*M. aeruginosa*, *M. flos-aquae*	MC	PP2A/ELISA	0.09–0.72	
Carassius gibelio (Cyprinidae)—Lake Kerkini	Omnivore	*Microcystis* spp.	MC	PP2A/ELISA	0.14–2.80	
Cyprinus carpio (Cyprinidae)—Lake Kerkini	Omnivore	*Microcystis* spp.	MC	PP2A/ELISA	0.09–0.82	
Cyprinus carpio (Cyprinidae)—Lake Pamvotis	Omnivore	*M. aeruginosa*; *A. flos-aquae*	MC	PP2A/ELISA	0.02–1.30	
Cyprinus carpio (Cyprinidae)—Lake Yliki	Omnivore	*Microcystis* spp.	MC	PP2A/ELISA	0.07–0.15	
Perca fluviatilis (Percidae)—Lake Kastoria	Carnivore	*M. aeruginosa*, *M. flos-aquae*	MC	PP2A/ELISA	0.07–1.60	
Rutilus rubilo (Cyprinidae)—Lake Pamvotis	Omnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.02–0.39	
Silurus aristoteles (Siluridae)—Lake Pamvotis	Carnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.09–0.24	
Silurus glanis (Siluridae)—Lake Pamvotis	Carnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.04–0.16	
Oreochromis niloticus (Cichlidae)	Omnivore	*M. aeruginosa*	MC	ELISA/PP2A	0.2–25.0	Mohamed and Hussein (2006)[89]
Rutilus rutilus (Cyprinidae)	Omnivore	*N. spumigena*	NOD	LC-MS/ELISA	ND–0.9	Sipiä *et al.* (2006)[90]
Platichthys flesus (Pleuronectidae)	Carnivore	*N. spumigena*	NOD	LC-MS/ELISA	0.003–1.1	
Lepomis gibbosus (Centrarchidae)—Barbadoes Pond	Zooplanktivore	*Microcystis* sp.	MC	ELISA	0.02–0.06	Smith and Haney (2006)[14]
Lepomis gibbosus (Centrarchidae)—Lab. experiment	Zooplanktivore	*Microcystis* sp.	MC	ELISA	0.01–0.11	

Table S2. *Cont.*

Oncorhyncus myskiss (Salmonidae)—Lake Rotoiti	Carnivore	*Microcystis* sp., *Anabaena* sp.	MC	LC-MS/ELISA	0.04–0.46	Wood *et al.* (2006)[63]
Oncorhyncus myskiss (Salmonidae)—Lake Rotoehu	Carnivore	*Microcystis* sp., *Anabaena* sp.	MC	LC-MS/ELISA	0.28–0.79	
Cyprinus carpio (Cyprinidae)	Omnivore	*M. aeruginosa*, *M. ichtyoblade*, *A. floes-aquae*	MC	ELISA	0.07–1.32	Adamovský *et al.* (2007)[91]
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*M. aeruginosa*, *M. ichtyoblade*, *A. floes-aquae*	MC	ELISA	0.05–1.24	
Platichthys flesus (Pleuronectidae)	Carnivore	*N. spumigena*	NOD	ELISA	ND–0.47	Mazur-Marzec *et al.* (2007)[69]
Cyprinus carpio (Cyprinidae)	Omnivore	*Anabaena* sp. (ANA 37)	ANTX-a	HPLC	0.03–0.77 [(a)]	Osswald *et al.* (2007)[92]
Gasterosteus aculeatus (Gasterosteidae)	Planktivore	*N. spumigena*	NOD	LC-MS	0.003–0.7	Sipiä *et al.* (2007)[93]
Clupea harengus(Clupeidae)	Planktivore	*N. spumigena*	NOD	LC-MS	ND–0.09	
Carassius auratus auratus (Cyprinidae)	Omnivore	*Microcystis* spp.	MC	ELISA/LC-MS	ND–3.64	Song *et al.* (2007)[33]
Carassius auratus (Cyprinidae)	Omnivore	*Microcystis* spp., *A. flos-aquae*	MC-RR	HPLC	0.49–0.82	Xie *et al.* (2007)[70]
Cyprinus carpio (Cyprinidae)	Omnivore	*Microcystis* spp., *A. flos-aquae*	MC-RR	HPLC	0.27–2.06	
Silurus glanis (Siluridae)	Omnivore	*Microcystis* spp., *A. flos-aquae*	MC-RR	HPLC	0.14	
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*M. aeruginosa*	MC-RR, -LR	HPLC-MS	13.5–91.0	Zhang *et al.* (2007)[94]
Clupea harengus membras (Clupeidae)	Planktivore	*N. spumigena*	NOD	ELISA	up to 0.22	Karjalainen *et al.* (2008)[17]
Sprattus sprattus (Clupeidae)	Planktivore	*N. spumigena*	NOD	ELISA	up to 0.10	
Gasterosteus aculeatus (Gasterosteidae)	Planktivore	*N. spumigena*	NOD	ELISA	up to 0.80	
Perca flavescens (Percidae)	Carnivore	*M. aeruginosa*	MC	ELISA	0.017–1.18	Wilson *et al.* (2008)[95]
Tilapia rendalli (Cichlidae) Furnas Reservoir	Omnivore	*Pseudanabaena mucicola*, *Microcystis panniformis*, *M. aeruginosa*	MC	PP1-HPLC	0.005–24	Deblois *et al.* (2008)[96]
Oreochromis niloticus (Cichlidae)—Furnas Reservoir	Omnivore		MC	PP1-HPLC	0.06–112	
Oreochromis niloticus (Cichlidae)—Funil Reservoir	Omnivore	*M. panniformis*, *M. viridis*, *P. mucicola*, *Anabaena* cf. *crassa*	MC	PP1-HPLC	0.02–321	

Table S2. *Cont.*

Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	ND–5.36	Chen *et al.* (2009)[97]
Carassius auratus (Cyprinidae)	Omnivore	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	0.04–0.27	
Culter ilishaeformis (Cyprinidae)	Carnivore	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	ND–0.01	
Cyprinus carpio (Cyprinidae)	Omnivore	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	0.002–0.064	
Platichthys flesus (Pleuronectidae)	Carnivore	*N. spumigena*	NOD	HPLC	ND–0.78	Persson *et al.* (2009)[98]
Platichthys flesus (Pleuronectidae)	Carnivore	*N. spumigena*	NOD	LC-MS	0.045–0.220	Vuorinen *et al.* (2009)[99]
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*M. aeruginosa*	MC-LR,-LR-GSH, -LR-Cys	LC-MS	ND–29.1	Zhang *et al.* (2009)[34]
Coilia ectenes (Bagridae)	Carnivore	*M. aeruginosa*	MC-LR, -RR, -YR	LC-MS	ND–0.0447	Zhang *et al.* (2009)[100]
Culter ilishaeformis (Cyprinidae)	Carnivore	*M. aeruginosa*	MC-LR, -RR, -YR	LC-MS	ND–0.0581	
Neosalanx taihuensis (Salangidae)	Carnivore	*M. aeruginosa*	MC-LR, -RR, -YR	LC-MS	ND–0.00756	
Hypophytalmincthtys molitrix (Cyprinidae)	Phytoplanktivore	*M. aeruginosa*	MC-LR, -RR, -YR	LC-MS	ND–4.19	
Carassius auratus (Cyprinidae)	Omnivore	*M. aeruginosa*	MC-LR, -RR, -YR	LC-MS	ND–0.2064	
Carassius gibelio (Cyprinidae)— Lakes in Greece	Omnivore	Several species	MC	ELISA	0.011–35.6	Papadimitriou *et al.* (2010)[101]
Odontesthes bonariensis (Atherinidae)	Carnivore	*Planktothrix agardhii*, *Anabaena* sp., *Microcystis* sp.	MC-LR, -RR, -YR, -LA	HPLC-MS/MS	ND–0.673	Amé *et al.* (2010)[102]
Lutjanus griseus (Lutjanidae)	Carnivore	Cyanobacterial blooms	BMAA	HPLC-C/MS/MS	ND–188	Brand *et al.* (2010)[35]
Mojarra (Gerreidae)	Carnivore	Cyanobacterial blooms	BMAA		ND–567	
Sphoeroides spengleri (Tetraodontidae)	Carnivore	Cyanobacterial blooms	BMAA		ND–455	
Sphoeroides parvus (Tetraodontidae)	Carnivore	Cyanobacterial blooms	BMAA		194–7351	
Acanthostracion quadricornis (Ostraciidae)	Omnivore	Cyanobacterial blooms	BMAA		34–47	
Archosargus rhomboidalis (Sparidae)	Omnivore	Cyanobacterial blooms	BMAA		484–2349	
Haemulon parra (Haemulidae)	Carnivore	Cyanobacterial blooms	BMAA		203–722	
Haemulon sciurus (Haemulidae)	Carnivore	Cyanobacterial blooms	BMAA		1723–3776	
Amia calva (Amiidae)	Carnivore	Cyanobacterial blooms	BMAA		554–2559	
Atractosteus spatula (Lepisosteidae)	Carnivore	Cyanobacterial blooms	BMAA		797–2290	
Micropterus salmoides (Centrarchidae)	Carnivore	Cyanobacterial blooms	BMAA		949–2388	

Table S2. *Cont.*

Geophagus brasiliensis (Cichlidae)	Omnivore	*C. raciborskii*	STXs	HPLC	0.12–0.20	Clemente *et al.* (2010)[103]
Cyprinus carpio larvae (Cyprinidae)	Omnivore	*M. aeruginosa*	MC-LR, -RR, -YR, -FR,-WR	HPLC-MS	0.17–0.27	El Ghazali *et al.* (2010)[104]
Cyprinus carpio larvae (Cyprinidae)	Omnivore	*M. aeruginosa*	MC-LR, -RR, -YR, -FR,-WR	HPLC-MS	0.22–0.55	El Ghazali *et al.* (2010)[105]
Lates niloticus (Latidae)	Carnivore	No information	MC-LR, -RR, -YR	HPLC-MS/MS	0.0097–0.28	Nyakairu *et al.* (2010)[106]
Oreochromis niloticus (Cichlidae)—Lake Mburo	Omnivore	No information	MC-LR, -RR, -YR	HPLC-MS/MS	0.731–13.12	
Oreochromis niloticus (Cichlidae)—Murchison Bay	Omnivore	No information	MC-LR, -RR, -YR	HPLC-MS/MS	0.048–14.79	
Morone saxatilis (Moronidae)	Zooplanktivore	*Microcystis* sp.	MC	PP2A	1.04–2.92	Lehman *et al.* (2010)[21]
Menidia audens (Atherinidae)	Zooplanktivore	*Microcystis* sp.	MC	PP2A	0.30–1.98	
Chirostoma spp. (Atherinidae)	Zooplanktivore	*Aphanizomenon*; *Microcystis*	MC	ELISA	0.185	Berry *et al.* (2011)[107]
Cyprinus carpio (Cyprinidae)	Omnivore	*Aphanizomenon*; *Microcystis*	MC	ELISA	0.05–0.94	
Goodea sp. (Goodeidae)	Phytoplanktivore	*Aphanizomenon*; *Microcystis*	MC	ELISA	1.57–8.67	
AMPHIBIANS						
Rana eperotica (Ranidae)—Lake Pamvotis	Carnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.26–0.47	Gkelis *et al.* (2006)[64]
Rana ridibunda (Ranidae)—Lake Gallikos	Carnivore	*M. aeruginosa*	MC	PP2A/ELISA	0.37–0.47	
Bufo marinus (Bufonidae) tadpoles	Phytoplanktivore	*Cylindrospermopsis raciborskii*	CYN	HPLC/MS-MS	0.61–8.95	White *et al.* (2007)[108]
REPITILES						
Emys orbicularis (Emydidae)	Carnivore	*Microcystis* spp.	MC-LR, -RR, -YR	PP2A/ Limieux-GC-MS	0.001–37.2	Nasri *et al.* (2008)[109]
Mauremys leprosa (Emydidae)	Omnivore	*Microcystis* spp.	MC-LR, -RR, -YR	PP2A/ Limieux-GC-MS	0.02–1,193	
Pelodiscus sinensis (Tryonichidae)	Carnivore	*Microcystis* spp.	MC-LR, -RR, -YR	LC-MS	0.011–0.021	Chen *et al.* (2009)[97]

Table S2. *Cont.*

BIRDS						
Phoeniconaias minor (Phoenocopteridae)	Omnivore	*Synechococcus bigranulatus*, *Spirulina subsalsa*, *Phormidium terebriformis*, *Oscillatoria willei*	MC ANTX-a	HPLC/MS	0.36 7.62	Krienitz *et al.* (2003)[110]
Somateria mollissima (Anatidae)	Carnivore	*N. spumigena*	NOD	LC-MS/ELISA	0.003–0.180	Sipiä *et al.* (2003)[111]
Somateria mollissima (Anatidae)	Carnivore	*N. spumigena*	NOD	LC-MS/ELISA	ND–0.198	Sipiä *et al.* (2006)[90]
Somateria mollissima (Anatidae)	Carnivore	*N. spumigena*	NOD	LC-MS/ELISA	0.003–0.052	Sipiä *et al.* (2008)[71]
Coturnix coturnix japonica (Phasianidae)	Omnivore	*Microcystis* sp.	MC	ELISA	0.0043–0.437	Pašková *et al.* (2008)[112]
Anas platyrhynchos (Anatidae)	Omnivore	*Microcystis* spp.	MC-LR, -RR,-YR	LC-MS	0.009–0.051	Chen *et al.* (2009)[97]
Nycticorax nycticorax (Ardeidae)	Omnivore	*Microcystis* spp.	MC-LR, -RR,-YR	LC-MS	0.003–0.082	

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