

Supplementary Materials:

A quorum-sensing inhibitor strain of *Vibrio alginolyticus* blocks QS-controlled phenotypes in *Chromobacterium violaceum* and *Pseudomonas aeruginosa*

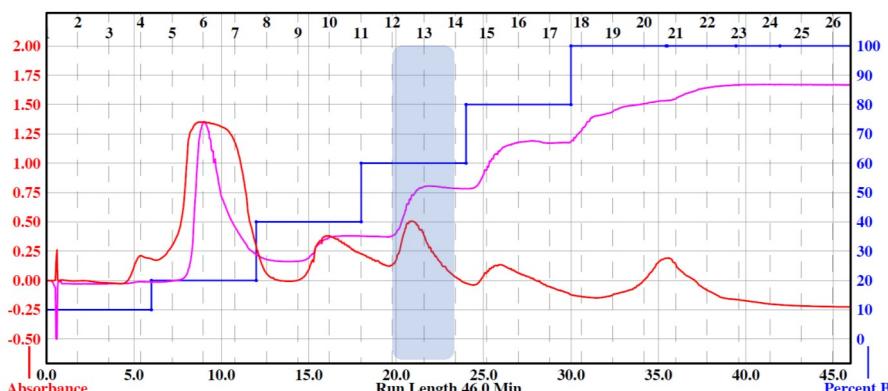
José Carlos Reina¹, Ignacio Pérez-Victoria^{2,*}, Jesús Martín² and Inmaculada Llamas^{1,3*}

¹ Department of Microbiology, Faculty of Pharmacy, University of Granada, Campus Universitario Cartuja s/n, 18071 Granada, Spain

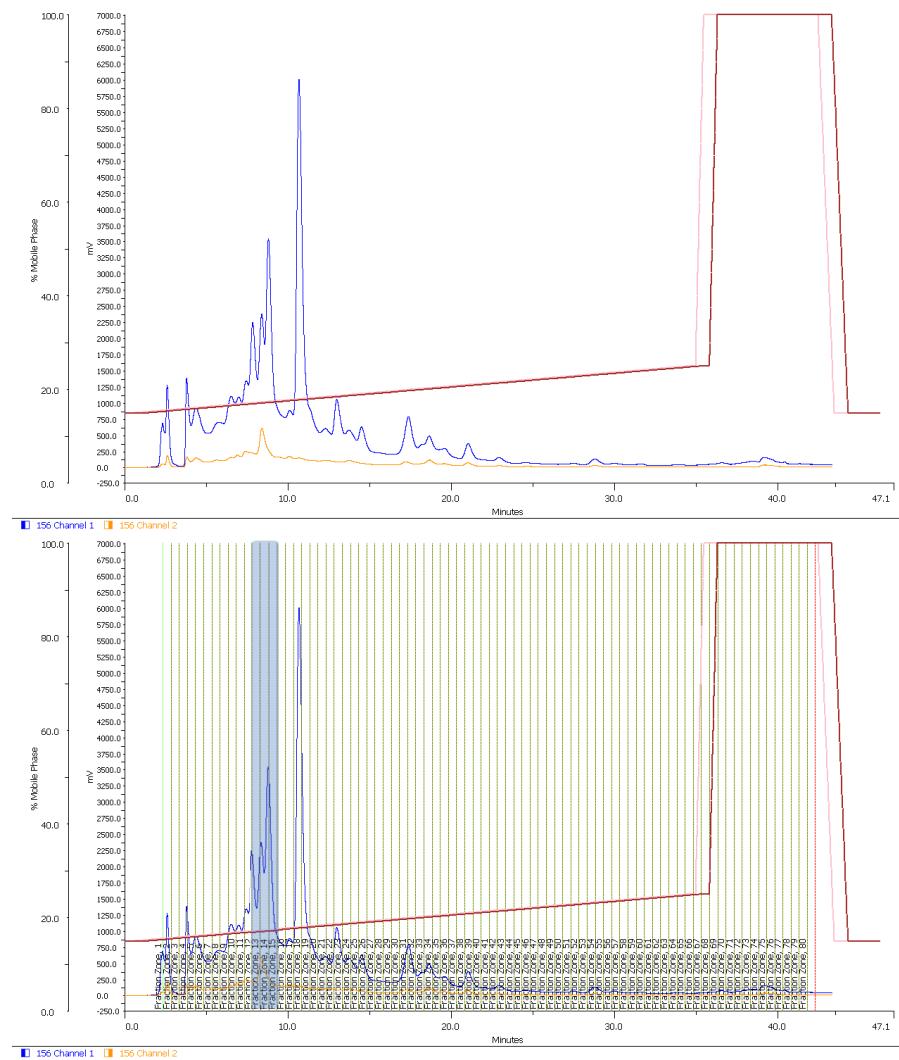
² MEDINA Foundation, Andalusian Center of Excellence for Research into Innovative Medicines, Health Science Technology Park, Avda. del Conocimiento 34, 18016 Armilla, Granada, Spain.

³ Institute of Biotechnology, Biomedical Research Center (CIBM), University of Granada, 18100 Granada, Spain

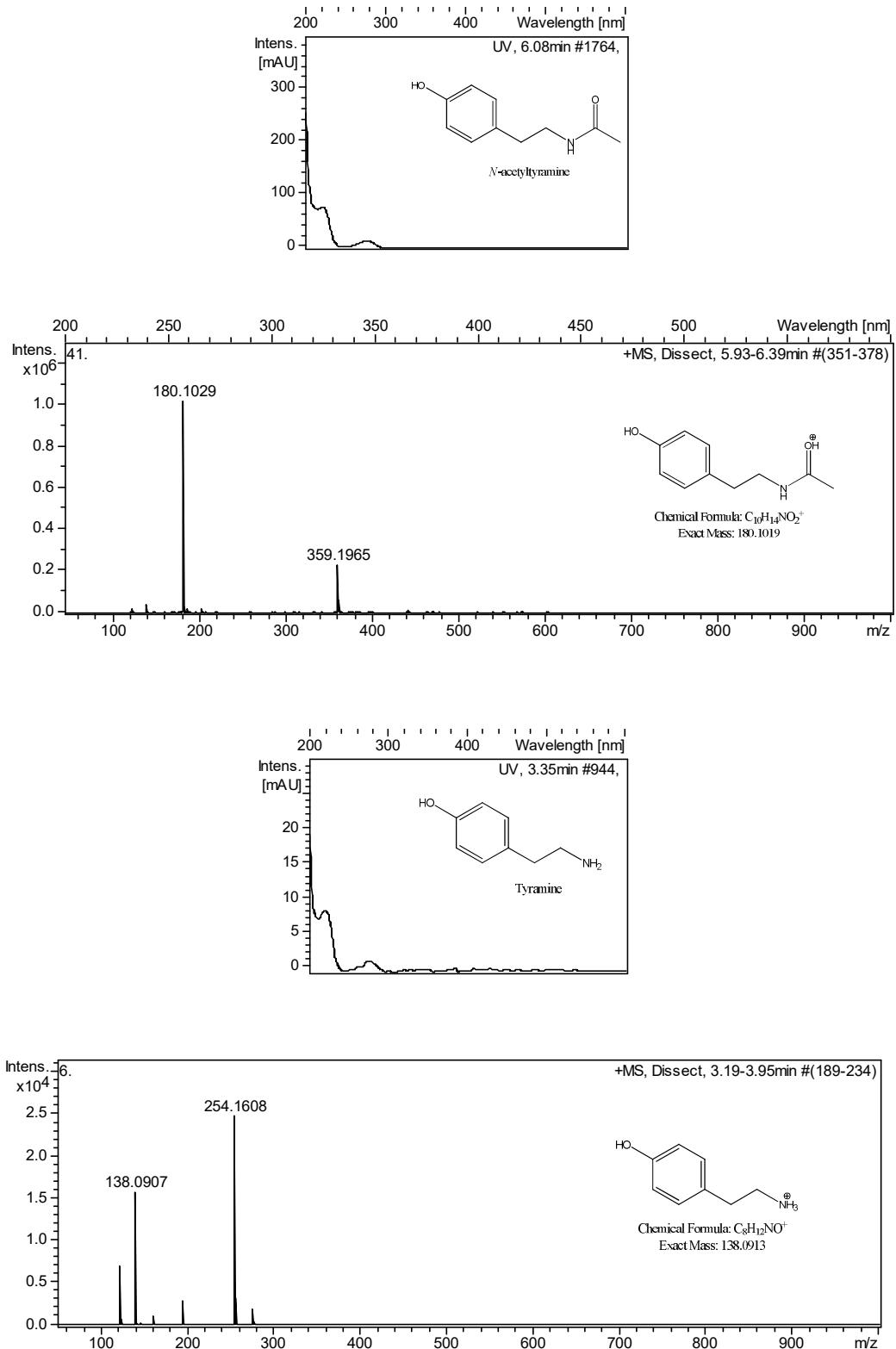
* Correspondence: illamas@ugr.es (I.L.), ignacio.perez-victoria@medinaandalucia.es (I.P.V.)



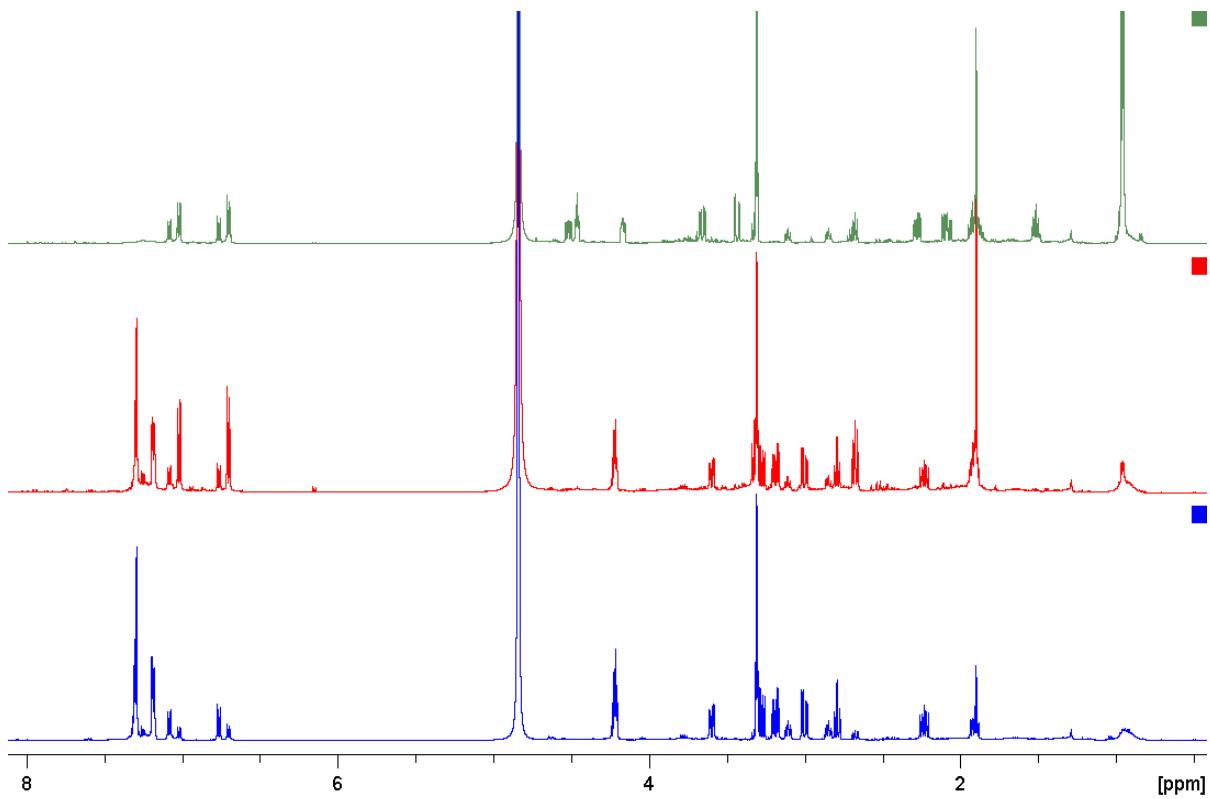
Supplementary Figure S1. Low resolution flash chromatography after solid phase extraction of the fermentation broth using SP207ss resin (Solvent A: water, Solvent B: acetone; Flow rate: 10 mL/min; Red trace: 210 nm, Purple trace: 280 nm). Fractions 12 and 13 displaying QSI activity are highlighted.



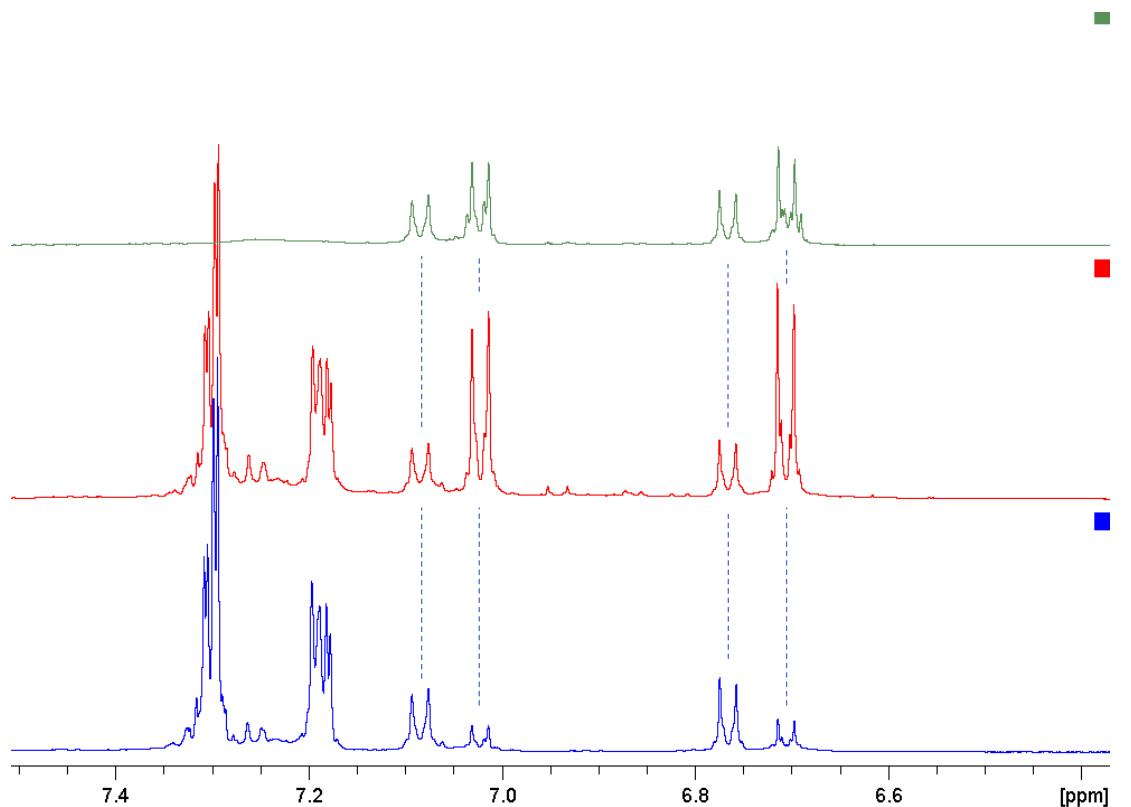
Supplementary Figure S2. Semipreparative HPLC chromatographic fractionation of the pool of the two active fractions (12 and 13) obtained after the previous low-resolution flash chromatography step (Solvent A: water, Solvent B: acetonitrile; Flow rate: 10 mL/min). Fractions 12, 13 and 14 displaying QSI activity are highlighted.



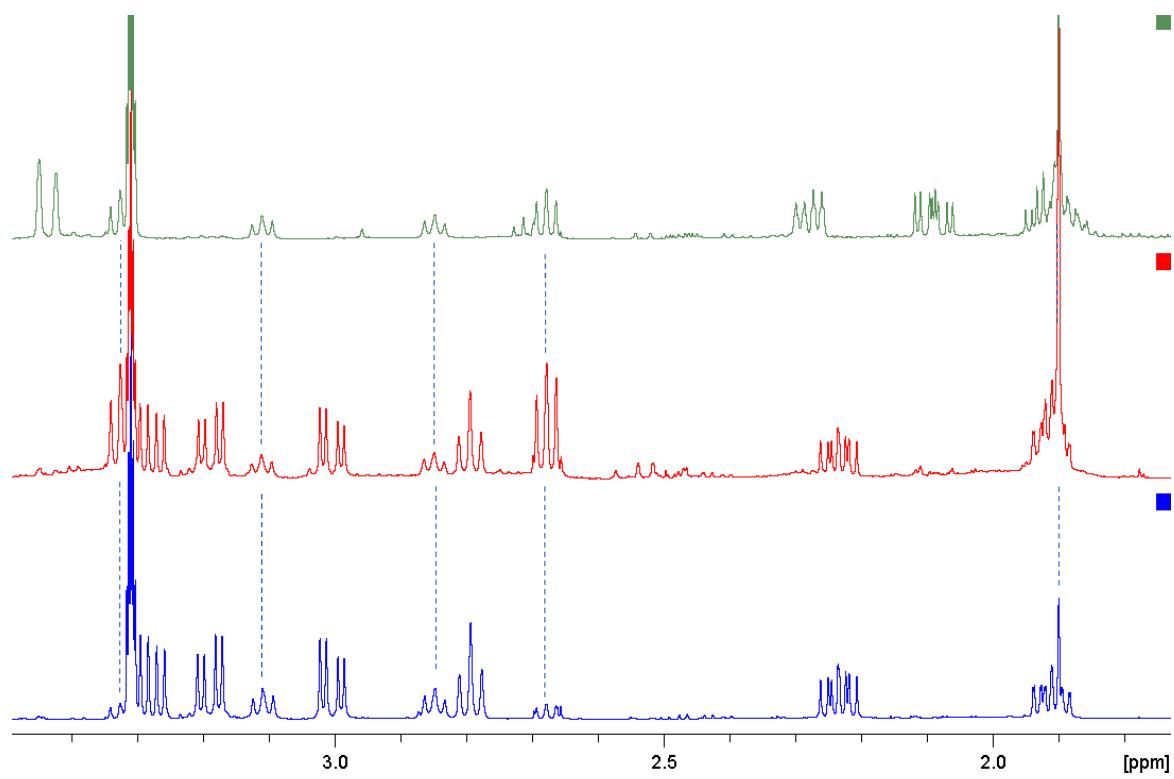
Supplementary Figure S3. UV-vis (DAD) and HRMS spectra of N-acetyltyramine (two upper spectra) and tyramine (two lower spectra) detected after LC-DAD-HRMS analysis of the three semipreparative HPLC fractions which showed QSI activity. The two compounds were observed in the three fractions, whose DAD and HRMS spectra, along with their retention time, were employed for their dereplication using our in-house databases [1,2].



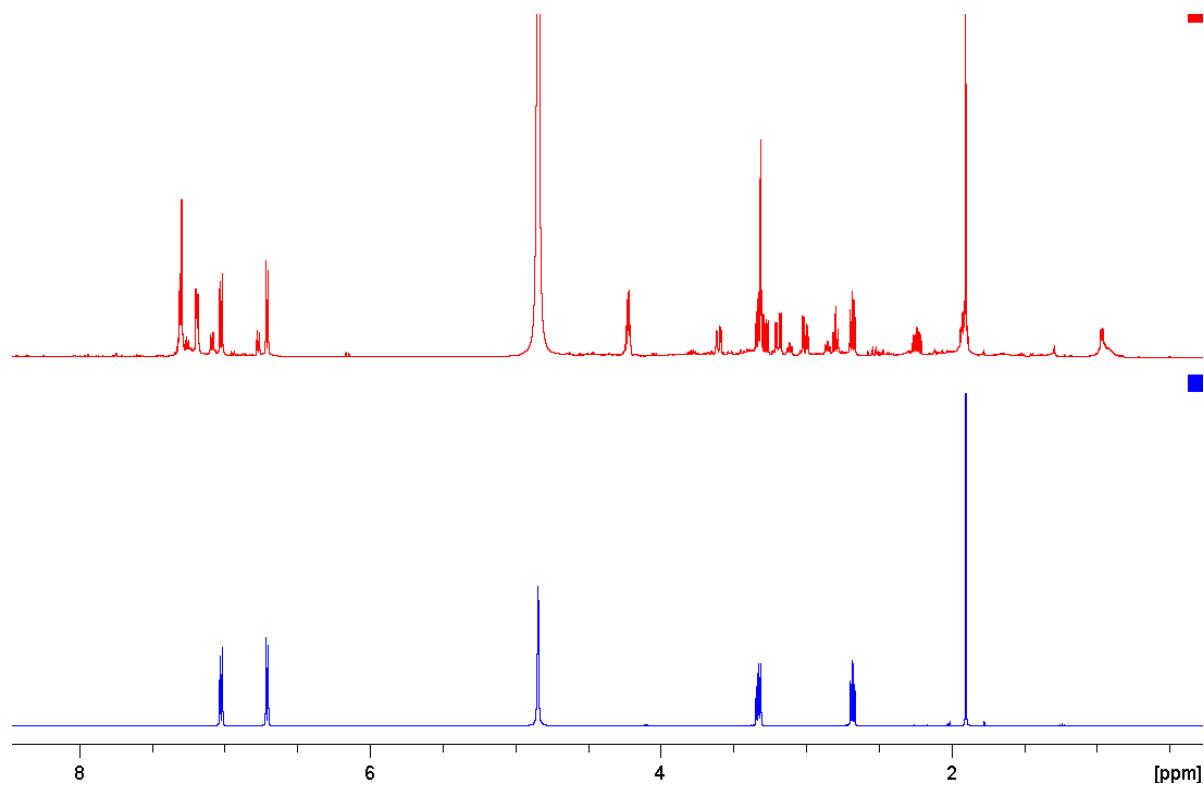
Supplementary Figure S4. ¹H NMR spectra of QSI-active HPLC fractions 12 (green), 13 (red) and 14 (blue), (500 MHz, CD₃OD).



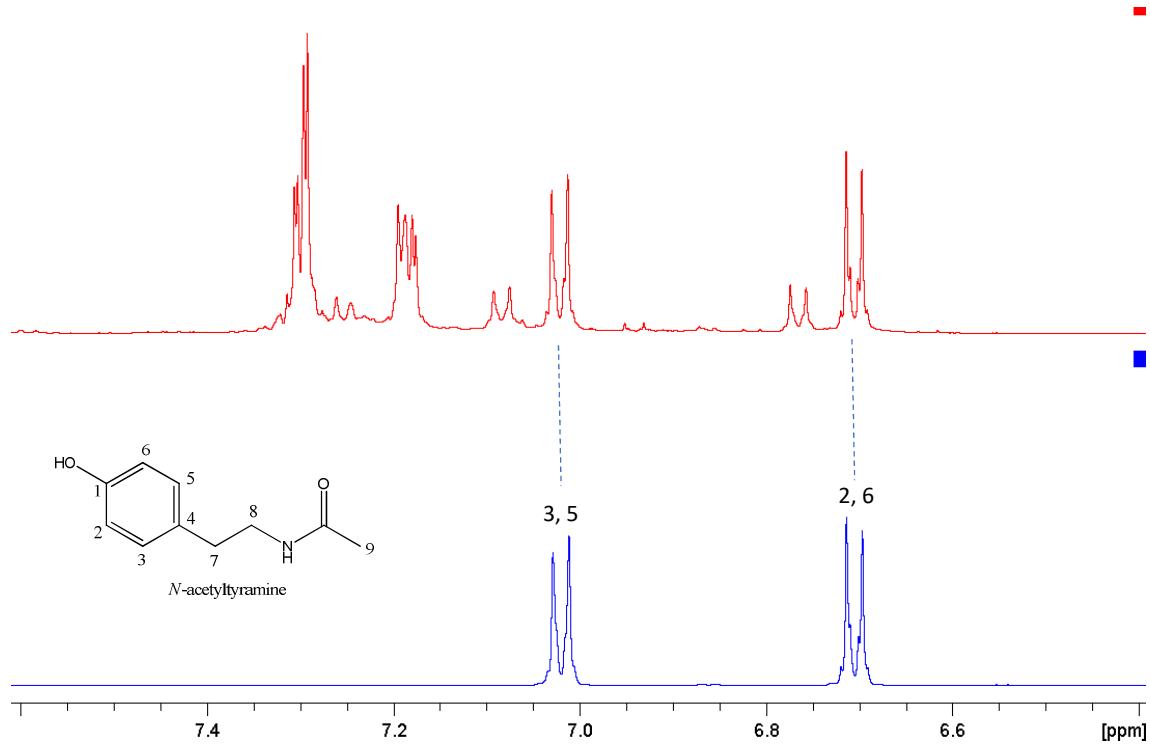
Supplementary Figure S5. Expansion of Fig. S4 spectra, indicating the common aromatic signals observed in all three spectra.



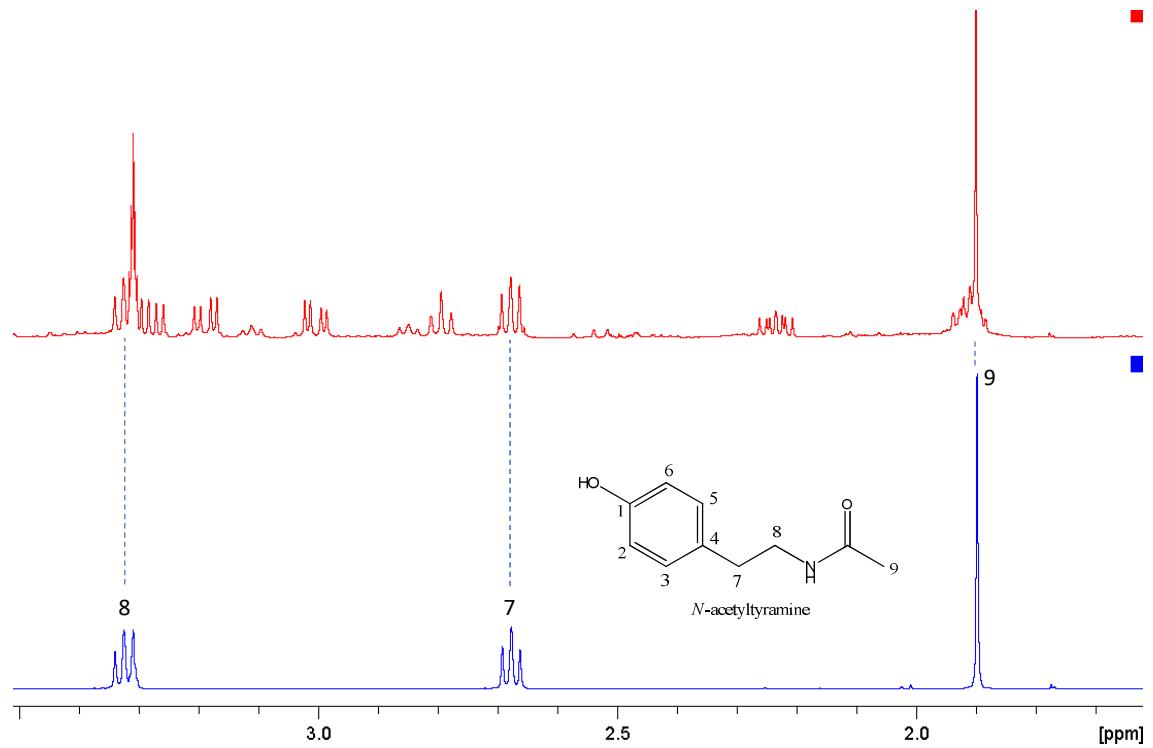
Supplementary Figure S6. A further expansion of Fig. S4 spectra, indicating the common aliphatic signals observed in all three spectra.



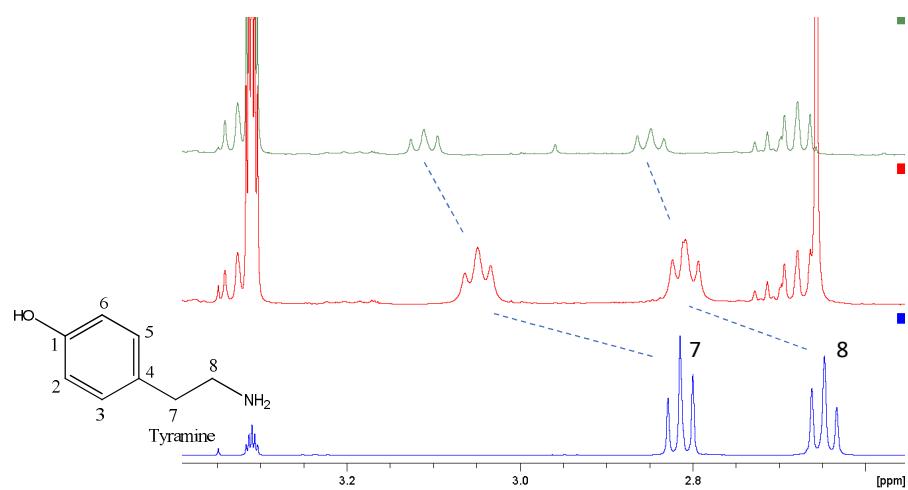
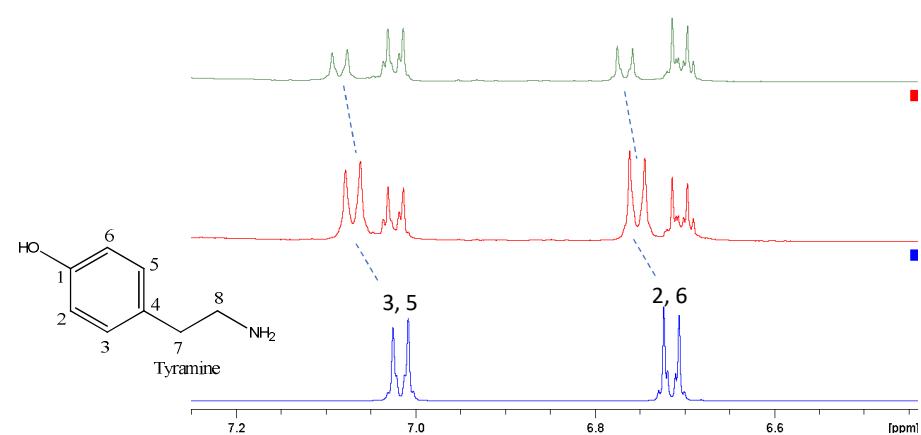
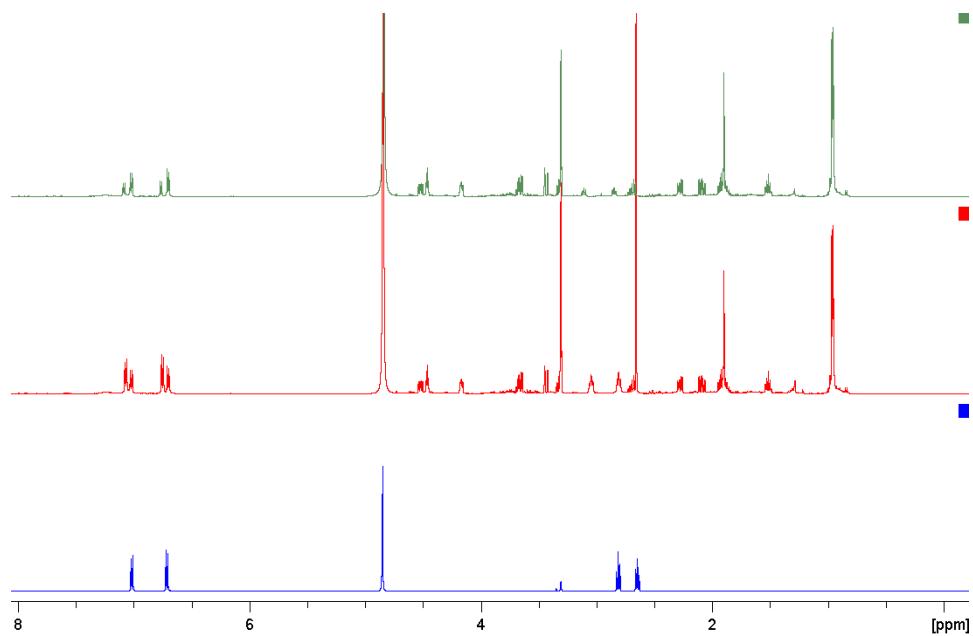
Supplementary Figure S7. ^1H NMR spectra of QSI-active HPLC fraction 13 (red) and N-acetyltyramine standard (blue) (500 MHz, CD_3OD).



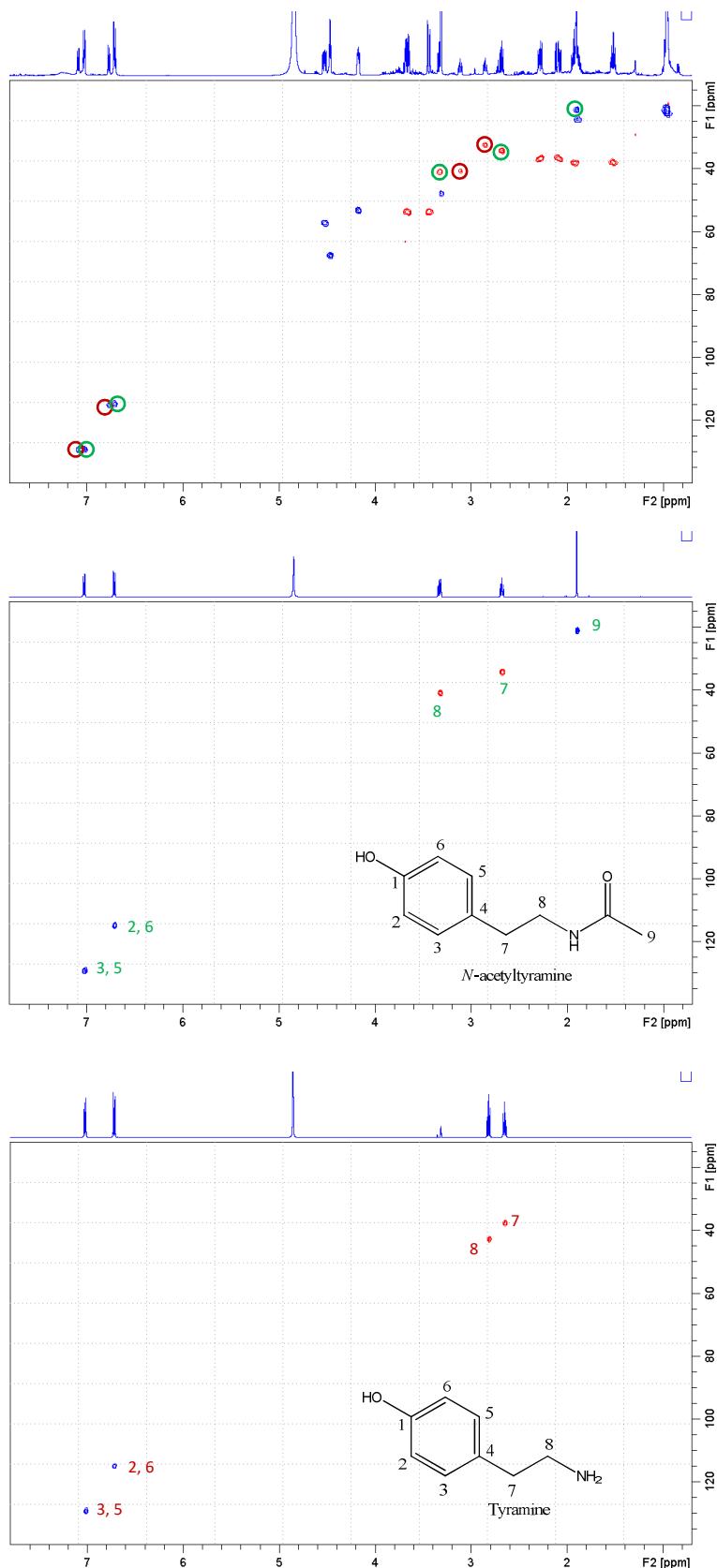
Supplementary Figure S8. Expansion of spectra in Supplementary Figure S7, indicating the identification of the aromatic signals from *N*-acetyltyramine.



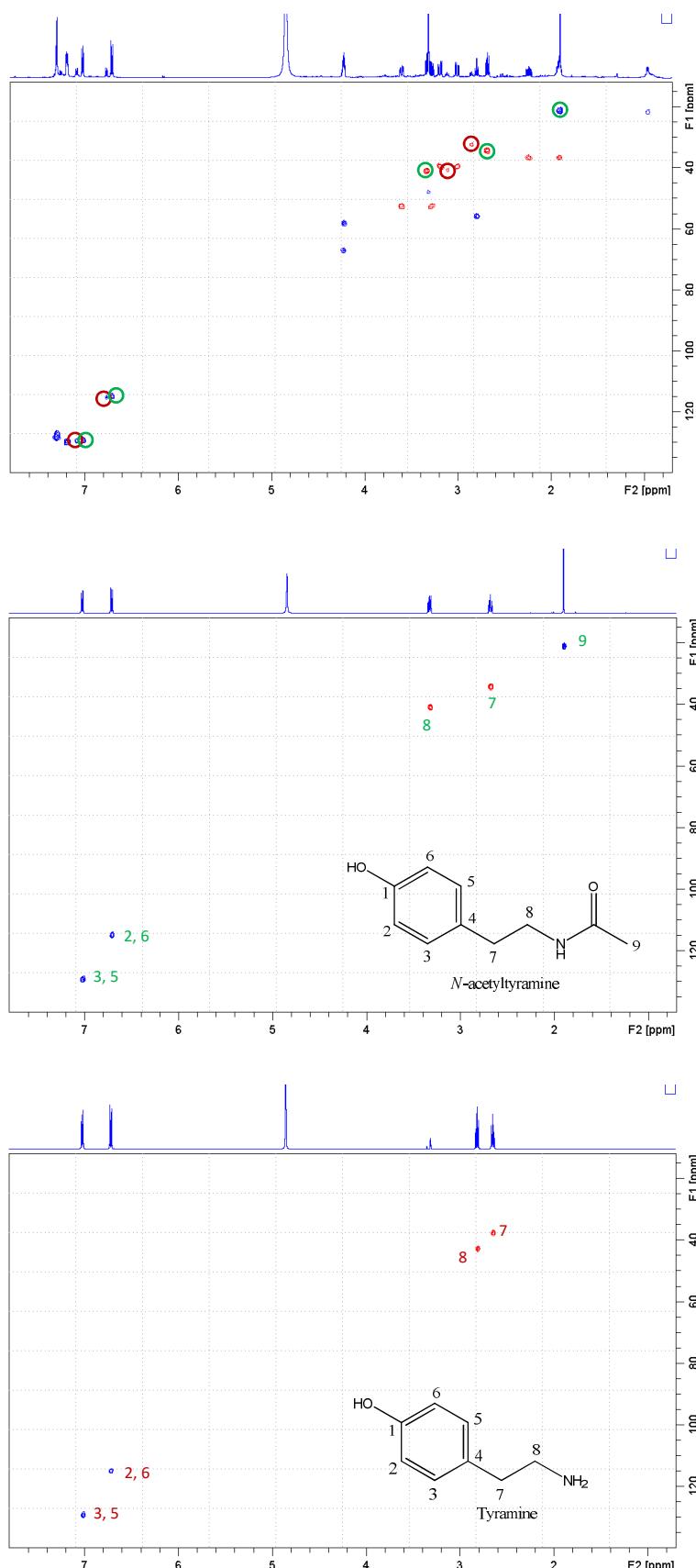
Supplementary Figure S9. A further expansion of the spectra in Supplementary Figure S7, indicating the identification of the aliphatic signals from *N*-acetyltyramine.



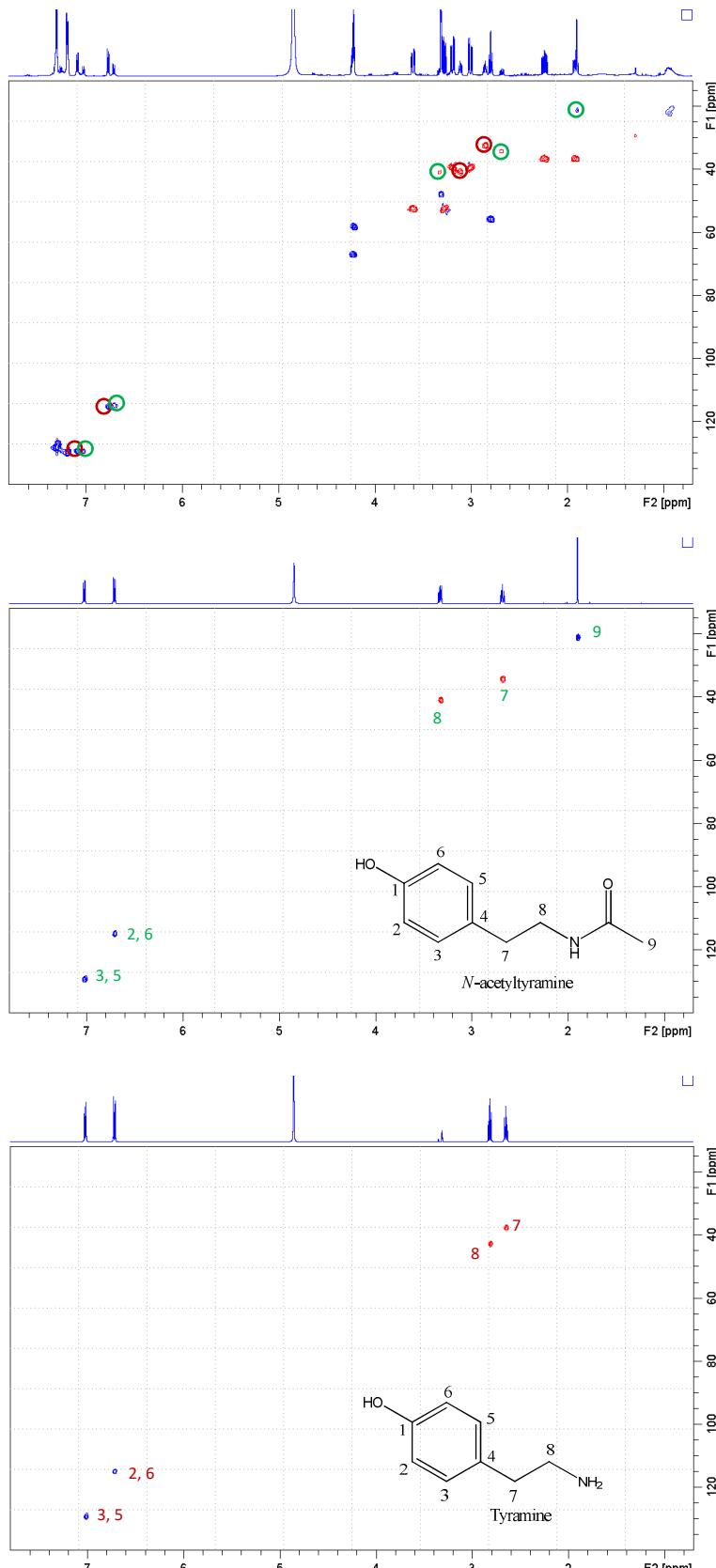
Supplementary Figure S10. ^1H NMR spectra of QSI-active HPLC fraction 12 (green), the same fraction spiked with pure tyramine (red) and tyramine standard (blue) (500 MHz, CD_3OD). The expansions indicate the identification of the aromatic (first expansion) and aliphatic (second expansion) signals from tyramine.



Supplementary Figure S11. HSQC spectra of QSI-active HPLC fraction 12 (upper), N-acetyltyramine standard (middle) and tyramine standard (lower). Signals from these two molecules are highlighted in the upper spectrum, where the non-highlighted signals correspond to cyclo-(L-Leu-L-trans-4-hydroxyproline) [3].



Supplementary Figure S12. HSQC spectra of QSI-active HPLC fraction 13 (upper), N-acetyltyramine standard (middle) and tyramine standard (lower). Signals from these two molecules are highlighted in the upper spectrum, where the non-highlighted signals correspond to cyclo-(D-Phe-L-trans-4-hydroxyproline)[4]



Supplementary Figure S13. HSQC spectra of QSI-active HPLC fraction 13 (upper), N-acetyltyramine standard (middle) and tyramine standard (lower). Signals from these two molecules are highlighted in the upper spectrum, where the non-highlighted signals correspond to cyclo-(D-Phe-L-trans-4-hydroxyproline) [4].

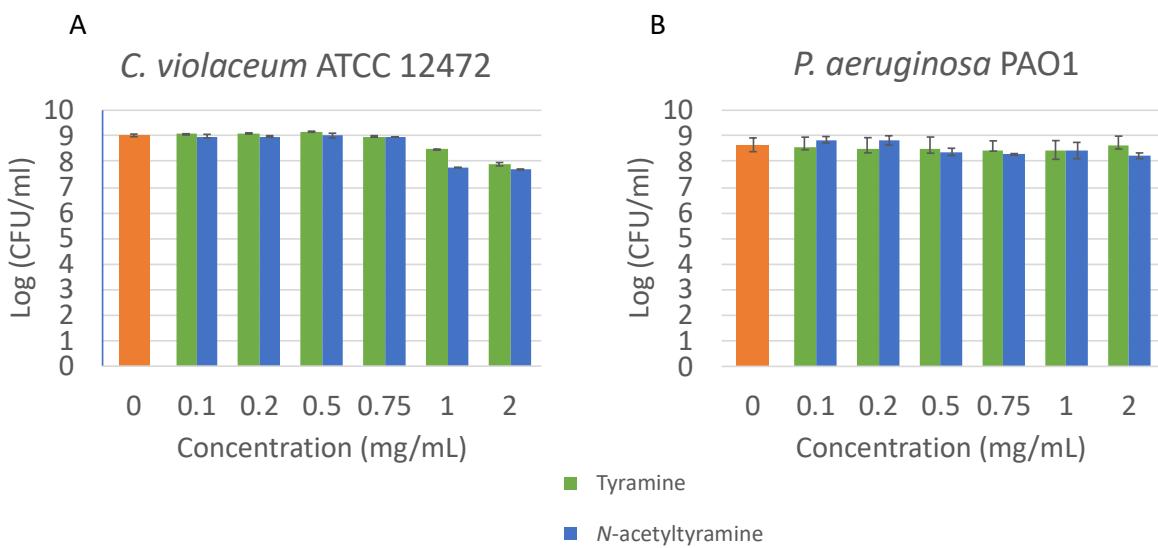


Figure S14. Growth of *C. violaceum* ATCC 12472 (A) and *P. aeruginosa* PAO1 (B) in the absence and presence of different concentrations of tyramine and *N*-acetyltyramine. Bacterial growth was measured by colony counting onto LB plates after incubating the bacteria in each condition for 24h. Data are presented as the logarithm of mean CFU \pm SD, n=3.

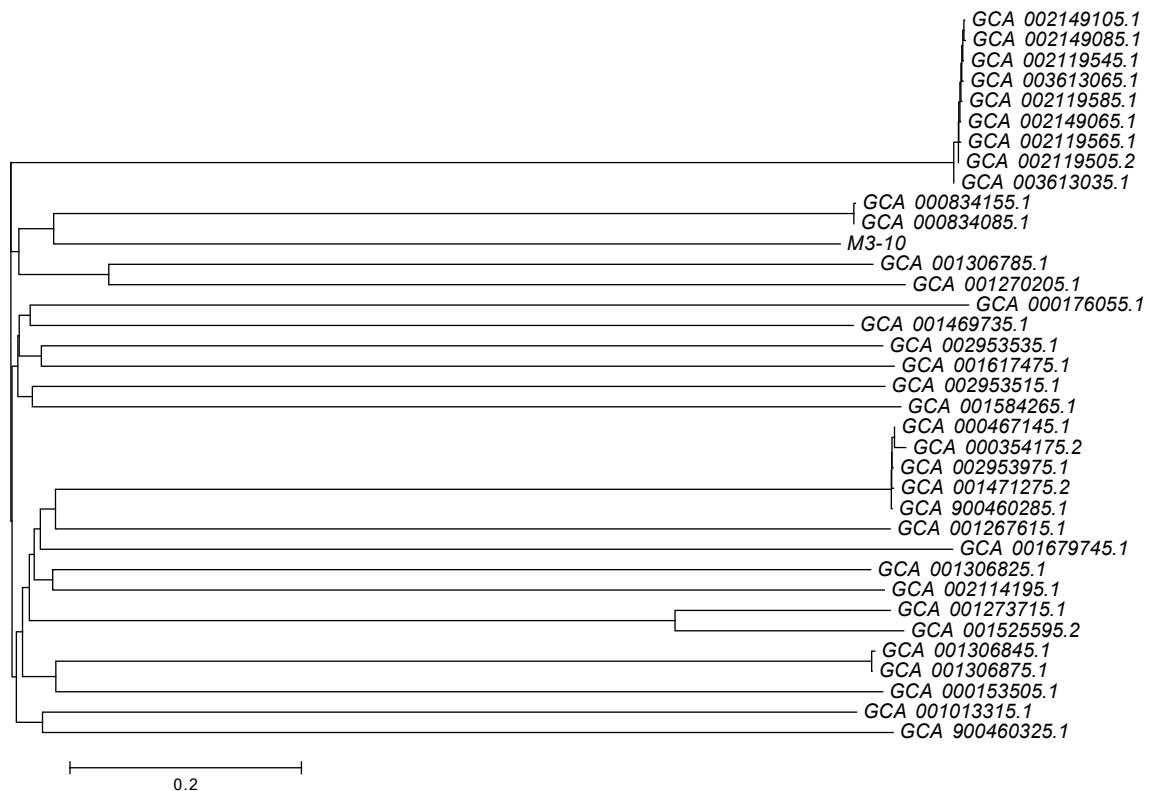
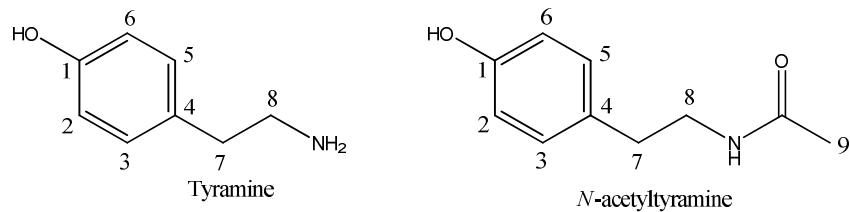


Figure S15. Neighbor-Joining distance clustering tree based on the ANI values between M3-10 and the rest of the *Vibrio alginolyticus* genomes available in NCBI.

Supplementary Table S1. NMR assignment (protonated carbons) of tyramine and *N*-acetyltyramine signals observed in the three QSI-active HPLC fractions (500 MHz, CD₃OD).

Position	Tyramine		<i>N</i> -acetyltyramine	
	δ_{C}	δ_{H} (<i>J</i> in Hz)	δ_{C}	δ_{H} (<i>J</i> in Hz)
2, 6	115.3	6.77, d (8.4)	114.9	6.71, d (8.4)
3, 5	129.3	7.08, d (8.4)	129.3	7.02, d (8.4)
7	32.4	2.85, t (7.3)	34.3	2.68, t (7.4)
8	40.7	3.11, t (7.4)	41.0	3.33, t (7.4)
9	-	-	21.1	1.90, s



Supplementary Table S2. NMR assignment (protonated carbons) of tyramine and *N*-acetyltyramine standards (500 MHz, CD₃OD).

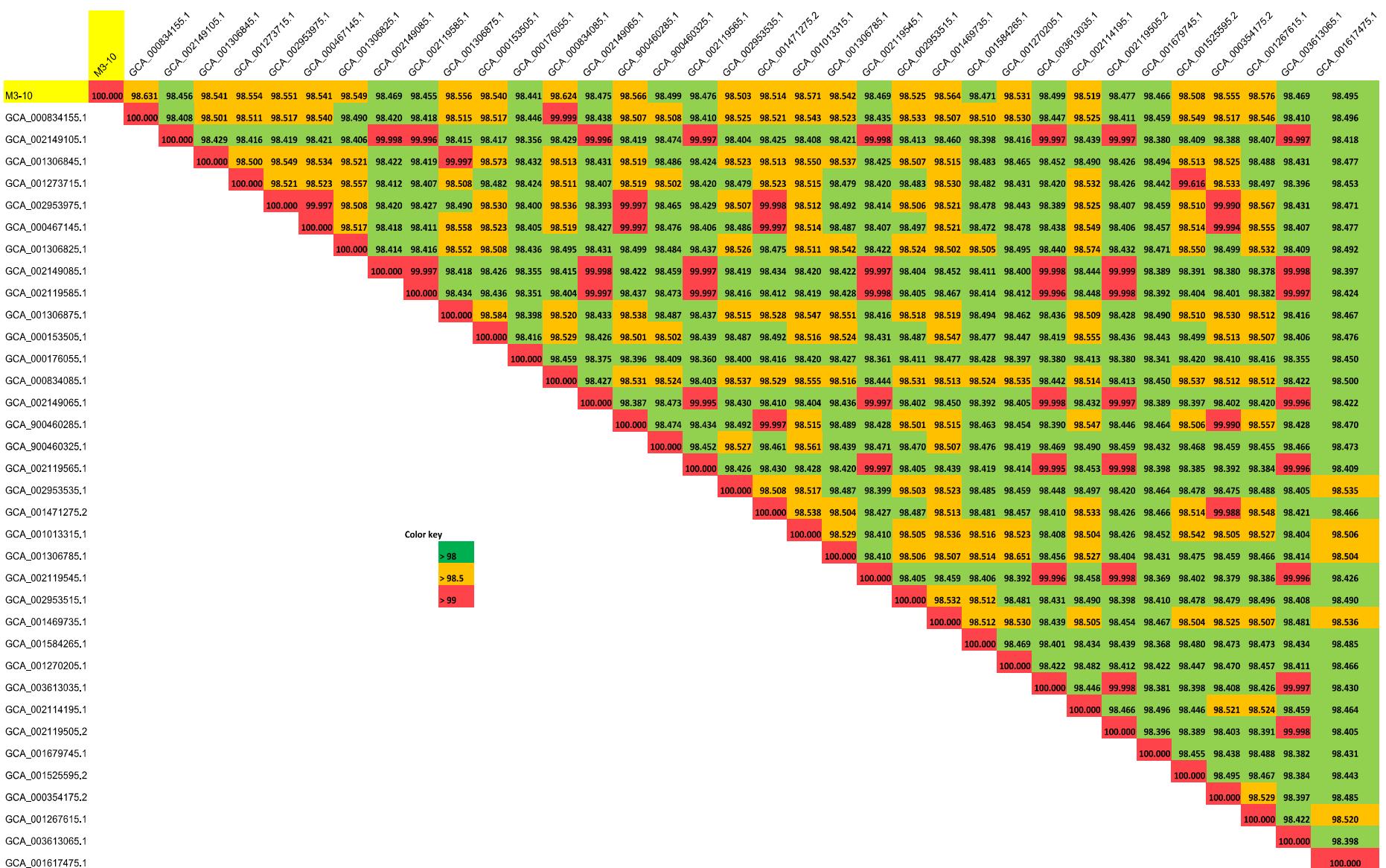
Position	Tyramine		<i>N</i> -acetyltyramine	
	δ_{C}	δ_{H} (<i>J</i> in Hz)	δ_{C}	δ_{H} (<i>J</i> in Hz)
2, 6	115.0	6.71, d (8.4)	114.9	6.71, d (8.4)
3, 5	129.3	7.01, d (8.4)	129.3	7.02, d (8.4)
7	37.7	2.65, t (7.2)	34.3	2.68, t (7.4)
8	42.9	2.81, t (7.2)	41.0	3.33, t (7.4)
9	-	-	21.1	1.90, s

Supplementary Table S3. Relative abundance (estimated by NMR) of tyramine, *N*-acetyltyramine and diketopiperazines in the three QSI-active HPLC fractions 12, 13 and 14. Qualitative QSI activity is also indicated (* weak activity, ** medium activity and *** high activity)

HPLC fraction	QSI activity	Tyramine	<i>N</i> -acetyltyramine	DKP1*	DKP2*
Fraction 12	**	1.0	1.6	5.0	0
Fraction 13	***	1.1	3.5	0	4.1
Fraction 14	*	1.1	0.4	0	3.9

*DKP1= cyclo-(L-Leu-L-*trans*-4-hydroxyproline); DKP2= cyclo-(D-Phe-L-*trans*-4-hydroxyproline)

Supplementary Table S4. ANIb values of strain M3-10 compared to the other *V. alginolyticus* strains whose genome is available



Supplementary Table S5. Protein sequence accession numbers for tyrosine decarboxylases used in the BLASTp homologue search.

1. CAI39170.2_1	2. CAI39169.2_1	3. AFP73381.1_1
4. AGW24520.1_1	5. AGW24519.1_1	6. BAE02560.1_1
7. ACS15340.1_1	8. ACS15339.1_1	9. ACS15338.1_1
10. CAY83536.1_1	11. CAY83534.1_1	12. CAY72357.1_1
13. CAY72352.1_1	14. CAY72349.1_1	15. CAY72347.1_1
16. CAY72344.1_1	17. CAY72343.1_1	18. CAY72341.1_1
19. CAY72338.1_1	20. CAY72336.1_1	21. CAY72335.1_1
22. CAY72333.1_1	23. CAY72330.1_1	24. BAE02559.1_1
25. AWT58522.1_1	26. AWT58521.1_1	27. AWT58520.1_1
28. AWT58519.1_1	29. AWT58518.1_1	30. AWT58517.1_1
31. AWT58516.1_1	32. BAI67125.1_1	33. WP_135180232.1_1
34. WP_135017352.1_1	35. E4V23_RS18420	36. WP_135043646.1_1
37. E4V46_RS09510	38. TFU21072.1_3	39. TFU11427.1_1
40. TFT82455.1_1	41. TFT56894.1_1	42. TFT37476.1_1
43. EVY62_RS19255	44. WP_129949649.1_1	45. WP_088207211.1_1
46. RZA37683.1_1	47. RZA37564.1_3	48. RYH14358.1_1
49. BHE89_RS17365	50. H049_RS0124195	51. H049_RS0123705
52. EBQ16_RS07935	53. WP_121909587.1_1	54. EA135_RS16540
55. RLZ56634.1_1	56. RLZ55416.1_1	57. C3O82_RS13145
58. RBH48265.1_1	59. RBH41953.1_1	60. WP_017628132.1_1
61. PSQ86773.1_1	62. PSQ85593.1_4	63. BRD33_05025
64. CUS67_RS12755	65. WP_104877381.1_2	66. CUM70_RS13060
67. CUM70_RS13050	68. PQF68676.1_2	69. CUS67_12755
70. PQD88208.1_1	71. PQD88205.1_1	72. C4A96_RS26915
73. C4A96_RS06470	74. C4A96_RS04445	75. C4A96_RS04265
76. PKN32413.1_1	77. CQR37_RS16425	78. PHL20111.1_1
79. OUK41848.1_2	80. OUK40079.1_2	81. OSP71327.1_1
82. OSP67774.1_2	83. WP_071425714.1_1	84. WP_080389721.1_2
85. WP_011109458.1_1	86. OIK52589.1_1	87. BHE89_18525
88. KXF72672.1_2	89. KRM71154.1_1	90. EAO52030.1_1
91. CBW46640.1_1	92. ABC68277.1_1	93. CAH04395.1_1
94. SCD69325.1_2		

Supplementary methods S1. Identification of QSI compounds in active HPLC fractions using LC-DAD-HRMS and NMR analyses.

After LC-DAD-HRMS analyses, dereplication against our in-house databases [1] identified the presence of both tyramine and *N*-acetyltyramine (Figure 1) in each of the three bioactive HPLC fractions (Figure S3). On the other hand, NMR signals from two different compounds compatible with those reported for tyramine [5] and *N*-acetyltyramine [5,6] were present in all three fractions, while signals corresponding to other compounds (diketopiperazines) were only observed in one or two of the three bioactive fractions (see Figures S4-S6). Interpretation of 2D NMR spectra (COSY, HSQC and HMBC; data not shown) confirmed the dereplication of tyramine and *N*-acetyltyramine in the three fractions and established the presence of cyclo-(L-Leu-L-trans-4-hydroxyproline) in fraction 12 [3] and cyclo-(D-Phe-L-trans-4-hydroxyproline) in fractions 13 and 14 [4]. Additionally, the identity of tyramine and *N*-acetyltyramine was definitively confirmed by directly comparing the NMR spectra (¹H and HSQC) of the three bioactive fractions with the spectra of the standards of these two related compounds acquired in the same spectrometer (see Supplementary Figures S7-S13 and Supplementary Tables S4 and S5). The relative abundance of tyramine, *N*-acetyltyramine and diketopiperazines in the three fractions was easily established from the ¹H NMR spectra (see Table S1), clearly indicating that either tyramine, *N*-acetyltyramine or both are the QSI compounds produced by the *V. alginolyticus* M3-10 strain. A QSI activity test carried out with pure standards of these two related molecules later confirmed that both are indeed QSI compounds.

Supplementary material references

1. Pérez-Victoria, I.; Martín, J.; Reyes, F. Combined LC/UV/MS and NMR Strategies for the Dereplication of Marine Natural Products. *Planta Med.* **2016**, *82*, 857–871.
2. Martín, J.; Crespo, G.; González-Menéndez, V.; Pérez-Moreno, G.; Sánchez-Carrasco, P.; Pérez-Victoria, I.; Ruiz-Pérez, L.M.; González-Pacanowska, D.; Vicente, F.; Genilloud, O.; et al. MDN-0104, an Antiplasmodial Betaine Lipid from *Heterospora chenopodii*. *J. Nat. Prod.* **2014**, *77*, 2118–2123.
3. Rustamova, N.; Bobakulov, K.; Begmatov, N.; Turak, A.; Yili, A.; Aisa, H.A. Secondary metabolites produced by endophytic *Pantoea ananatis* derived from roots of *Baccharoides anthelmintica* and their effect on melanin synthesis in murine B16 cells. *Nat. Prod. Res.* **2019**, *1*–6.
4. Ye, X.; Chai, W.; Lian, X.-Y.; Zhang, Z. Novel propanamide analogue and antiproliferative diketopiperazines from mangrove *Streptomyces* sp. Q24. *Nat. Prod. Res.* **2017**, *31*, 1390–1396.
5. Sun, J.F.; Wu, Y.; Yang, B.; Liu, Y. Chemical Constituents of Marine Sponge *Halichondria* sp. from South China Sea. *Chem. Nat. Compd.* **2015**, *51*, 975–977.
6. Gutiérrez, M.; Capson, T.L.; Guzmán, H.M.; González, J.; Ortega-Barría, E.; Quiñoá, E.; Riguera, R. Antiplasmodial metabolites isolated from the marine octocoral *Muricea austera*. *J. Nat. Prod.* **2006**, *69*, 1379–1383.