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Article

Synthesis and Antitumor Activities of Derivatives of the Marine Mangrove Fungal Metabolite Deoxybostrycin

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Abstract: Deoxybostrycin (1) is an anthraquinone compound derived from the marine mangrove fungus *Nigrospora* sp. No. 1403 and has potential to be a lead for new drugs because of its various biological properties. A series of new derivatives (2–22) of deoxybostrycin were synthesized. The *in vitro* cytotoxicity of all the new compounds was tested against MDA-MB-435, HepG2 and HCT-116 cancer cell lines. Most of the compounds exhibit strong cytotoxicity with IC₅₀ values ranging from 0.62 to 10 μ M. Compounds 19, 21 display comparable cytotoxicity against MDA-MB-435 to epirubicin, the positive control. The primary screening results indicate that the deoxybostrycin derivatives might be a valuable source of new potent anticancer drug candidates.

Keywords: deoxybostrycin derivatives; antitumor activity; marine mangrove; anthraquinone

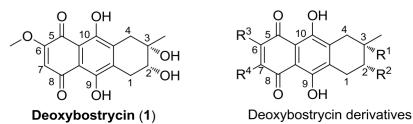
1. Introduction

Mortality and morbidity of cancer patients is the second highest among all diseases in the world, following heart disease [1-3]. Due to drawbacks of chemotherapy, such as dose limits, side effects, and low selectivity to cancer cells, discovery and development of much more effective, safe and highly selective antitumor drugs is still an urgent task.

In recent decades, much effort has been directed toward using natural products as a source of novel anticancer drugs. Recent reviews of drug discovery literature have shown that more than two thirds of the anticancer drugs approved between the 1940s and 2006 are either natural products or developed based on the knowledge gained from natural products [4,5]. In recent years, marine microorganisms have attracted great attention in the pharmaceutical community as they produce a wide variety of metabolites that are structurally unique and pharmacologically active [6,7]. Due to the structural and bioactive diversities of marine microorganism metabolites, they represent a promising resource for discovering new anticancer drugs [8,9].

Deoxybostrycin (1, Figure 1), a natural tetrahydroanthraquinone compound, isolated from the mangrove endophytic fungus Nigrospora sp. No. 1403 from the South China Sea [10], displays various biological properties including phytotoxic [11], antimalarial [12], antibacterial and cytotoxic activities [10,13]. Its structure was identified by interpretation of spectral data (IR, UV, MS, ¹H NMR, ¹³C NMR) [11,14,15]. Previous studies demonstrated that deoxybostrycin analogues can affect energy-yielding and energy-requiring processes in Ehrlich ascite cells [16] and inhibit the growth of cultured cells of Nicotiana rustica. Additionally, they act as a potent stimulator of NADH oxidation in mitochondria and as electron acceptors in an enzyme preparation of diaphorase [17]. However, there has been no study on the structural modification and anticancer activity of deoxybostrycin. In this paper, we describe the synthesis and cytotoxicity of deoxybostrycin derivatives. A series of deoxybostrycin derivatives (2–22, Schemes 1–3) were synthesized by modifying deoxybostrycin at C-2, C-3, C-6 and C-7 positions. All compounds were evaluated for their cytotoxicity against MDA-MB-435, HepG2 and HCT-116 cancer cell lines. Structure-activity relationships were discerned from the cytotoxic experimental data. As we expected, most of the deoxybostrycin derivatives exhibit strong anticancer activities against the tested cancer cell lines. Some of the derivatives showed higher cytotoxicity than the parent compound deoxybostrycin. Compounds 19 and 21 showed comparable cytotoxicity to epirubicin against MDA-MB-435 cell line with IC₅₀ values of 0.66 μ M and 0.62 μ M, respectively.

Figure 1. Structures of deoxybostrycin and deoxybostrycin derivatives.

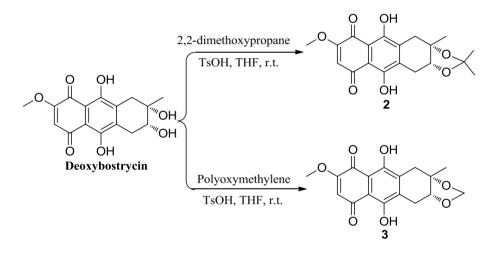


2. Results and Discussion

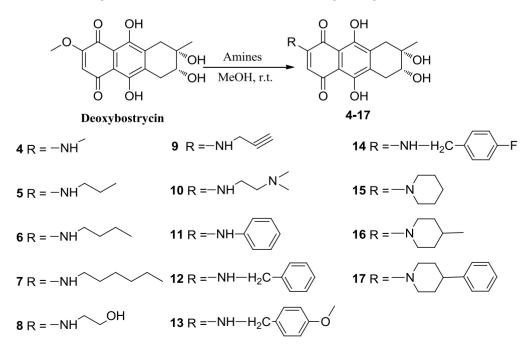
2.1. Chemistry

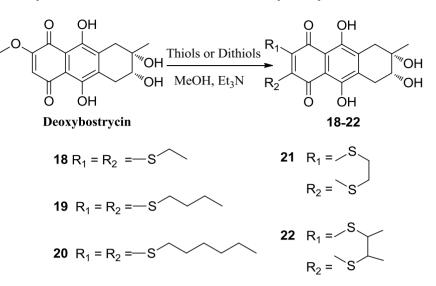
Deoxybostrycin reacted with 2,2-dimethoxypropane and polyoxymethylene in the presence of 1 equivalent of *p*-toluenesulfonic acid (TsOH) at room temperature to give 2,3-ketal derivatives 2 and 3, respectively (Scheme 1). When deoxybostrycin reacted with various amines at room temperature using methanol as solvent, a series of alkylamino and arylamino derivatives 4–17 were obtained (Scheme 2). Dithiosubstitued derivatives 18–22 were afforded by the reaction of deoxybostrycin with various thiols and dithiols at 0–5 °C in the presence of triethylamine (Scheme 3). The detailed mechanism of the nucleophilic substitution reaction of deoxybostrycin with thiols was proposed in our previous work [15]. The structures of all the synthesized compounds were characterized by IR, ¹H NMR, ¹³C NMR and HRMS (ESI).

Scheme 1. Synthesis of 2,3-ketal deoxybostrycin derivatives 2 and 3.



Scheme 2. Synthesis of 6-aminosubstituted deoxybostrycin derivatives 4–17.





Scheme 3. Synthesis of 6,7-dithiosubstituted deoxybostrycin derivatives 18–22.

2.2. Biological Activity

All synthesized compounds were evaluated for their *in vitro* cytotoxic activity against three human cancer cell lines (MDA-MB-435, HepG2 and HCT-116) by microculture tetrazolium assay (MTT) assay [18] using epirubicin as positive control.

As shown in Table 1, most of the deoxybostrycin derivatives showed good to excellent cytotoxic activity against the three tested cancer cell lines with $IC_{50} < 10 \mu M$. Some modified compounds exhibited better antitumor activities than the parent compound deoxybostrycin, and even displayed comparable activity to epirubicin. For example, the activity of compound 19 against MDA-MB-435 cell line (IC₅₀ = 0.66 μ M) showed comparable activity to epirubicin (IC₅₀ = 0.56 μ M). Similar potency was observed with compound 21 (against MDA-MB-435 and HCT-116 cells) and 22 (against MDA-MB-435 cells). Moreover, compounds 9, 11, 12, 15, 16 and 22 exhibited selectivity for MDA-MB-435 over other cell lines. The cytotoxic activities of compounds 6, 13, 14, 18 and 20 against MDA-MB-435 and HCT-116 cell lines were stronger than against HepG2 cell line. Compound 21 possessed the most potent activity against HCT-116 cell lines with an IC₅₀ value of 0.80 µM. Some results can be concluded from the SAR (structure-activity relationships) analysis based on the cytotoxic data of deoxybostrycin and its derivatives: (1) ketal 2 and 3 exhibited lower cytotoxic activities against all tested cancer cell lines than that of deoxybostrycin. The results suggested that the hydroxyl at C-2 and C-3 of deoxybostrycin was favorable for antitumor activity. Transformation of the diol to the diether decreased activity. Compound 2 with high steric hindrance at C-2 and C-3 showed nearly no cellular cytotoxic activity; (2) Compounds 4–17 derived from the replacement of methoxyl with various amines at the C-6 position generally decreased the cellular cytotoxicity with respect to the parent compound. The cytotoxic activity of Compounds 4-6 against HCT-116 cell lines showed alkylamino chain length dependence. The IC₅₀ values change from about 16 µM to 3 µM with the chain length increasing from one carbon for methylamine to six carbons for hexamine. Although compounds 9, 11, 12, 15 and 16 displayed decreased potency against HepG2 and HCT-116 cell lines, they had significantly improved selectivity for MDA-MB-435 cell. Compound 10 with a N,N-Dimethylethylenediamine substituent exhibited better cytotoxic activity against HepG2 cells than

all other amino-substituted derivatives. Compared to benzylamino derivative **12**, *p*-methoxy and *p*-fluorine substituted benzylamino derivatives **13** and **14** showed higher cytotoxic activity against HCT-116 cell line; (3) Compounds **18–22** were alkylthio-substituted derivatives of deoxybostrycin at C-6 and C-7 positions. All the dialkylthio-substituted deoxybostrycin derivatives showed excellent cytotoxic activity against all three human cancer cell lines with IC₅₀ values between 0.62 μ M and 6.49 μ M except for compounds **18** and **20** against HepG2 cell line. Among all the derivatives, compound **21** characterized with a relatively rigid 2,3-dihydro-1,4-dithiine heterocycle attached to deoxybostrycin displayed the highest potency against all the three tested cancer cell lines. Significantly, compound **21** displayed a comparable cytotoxic activity with the positive control epirubicin, for instance, compound **21** against MDA-MB-435 cell with an IC₅₀ of 0.62 μ M vs. epirubicin against MDA-MB-435 cell with an IC₅₀ of 0.56 μ M. The results suggest that the dithio-substituted deoxybostrycin derivatives benefit cytotoxic activity and serve as promising scaffolds for anti-tumor agents. These positive results serve as a valuable guideline for further research on the structural optimization, mechanism study and development of deoxybostrycin derivatives as novel anti-tumor agents.

Compounds	IC ₅₀ (μM) ^a		
	MDA-MB-435 ^b	HepG2 ^b	HCT-116 ^b
1	3.19 ± 0.92	9.99 ± 0.55	5.69 ± 0.25
2	>50	>50	26.08 ± 1.84
3	3.06 ± 0.13	12.83 ± 0.15	7.55 ± 0.45
4	11.74 ± 1.12	>50	16.57 ± 1.40
5	>50	9.98 ± 1.06	7.54 ± 0.21
6	6.79 ± 1.59	>50	3.29 ± 0.01
7	10.00 ± 1.75	>50	3.14 ± 0.16
8	>50	>50	>50
9	6.31 ± 1.40	10.90 ± 1.40	25.79 ± 0.64
10	1.52 ± 0.72	2.26 ± 0.35	3.42 ± 0.21
11	9.67 ± 1.80	>50	>50
12	5.76 ± 2.75	13.37 ± 2.72	20.70 ± 2.76
13	5.81 ± 2.89	10.09 ± 0.82	9.62 ± 0.20
14	7.25 ± 2.27	11.18 ± 0.94	7.09 ± 0.09
15	7.33 ± 1.20	>50	16.34 ± 0.90
16	7.32 ± 1.04	>50	14.92 ± 1.02
17	>50	11.81 ± 1.45	20.64 ± 1.64
18	1.96 ± 0.58	11.56 ± 1.40	6.49 ± 0.73
19	0.66 ± 0.41	5.04 ± 1.38	2.75 ± 0.23
20	2.06 ± 0.17	>50	2.94 ± 0.10
21	0.62 ± 0.23	1.98 ± 0.34	0.80 ± 0.08
22	0.97 ± 0.24	2.39 ± 0.50	2.69 ± 0.23
Epirubicin ^c	0.56 ± 0.06	0.96 ± 0.02	0.48 ± 0.03

Table 1. Cytotoxicity (IC₅₀, μ M) of compounds 1–22 against MDA-MB-435, HepG2 and HT-116 cancer cell lines.

^a IC_{50} values are taken as means \pm standard deviation from three independent experiments; ^b MDA-MB-435, human breast cancer cell line; HepG2, human liver cancer cell line; HCT-116, human colon cancer cell line; ^c Used as a positive control.

3. Experimental Section

3.1. Chemistry

Reagents were commercially available and used as received. Solvents were dried and purified using standard techniques. Melting points were measured on an X-4 micromelting point apparatus and were uncorrected. IR spectra were measured on a Bruker Vector 22 spectrophotometer using KBr pellets. NMR spectra were determined on a Varian Mercury-Plus 300 spectrometer or Bruker AV-400 NB spectrometer in CDCl₃ or DMSO- d_6 using TMS as internal standard, and coupling constants (*J*) are in Hz. ESI mass spectra were obtained on a LCQ DECA XP LC-MS mass spectrometer. Flash column chromatography was run on silica gel (Qing dao Ocean Chemical Factory, 200–300 mesh) eluted with petroleum ether-dichloromethane or dichloromethane-methanol, and C18 reversed phase silica gel (Welch Material, Inc., 45 µm) eluted with methanol-water.

3.2. Synthesis of 2,3-O-(isopropylidene) Deoxybostrycin (2)

To a solution of **1** (50 mg, 0.156 mmol) in 10 mL of tetrahydrofuran were added 2,2-dimethoxypropane (323.4 mg, 3.13 mmol) and *p*-toluenesulfonic acid (26.8 mg, 0.156 mmol). The reaction mixture was stirred for 15 h at room temperature and then diluted with water (20 mL) and extracted with dichloromethane (3 × 50 mL). The combined organic layer was washed with brine, dried over anhydrous magnesium sulfate, and concentrated *in vacuo*. The resulting residue was purified on a silica gel column using petroleum ether/dichloromethane (v/v, 1/1) as eluent to obtain 48.3 mg of compound **2** as a red solid in an 86% yield. Mp: 160–161 °C; $[\alpha]^{20}_{D} = 27.0^{\circ}$ (*c* = 1.00, CH₃OH); IR (KBr): v_{max} = 3431, 3086, 2980, 2931, 2896, 2883, 1598, 1564, 1452, 1415 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 13.12 (s, 1H), 12.71 (s, 1H), 6.17 (s, 1H), 4.39 (dd, 1H, *J* = 3.8, 2.9 Hz), 3.93 (s, 3H), 3.53 (dd, 1H, *J* = 16.7, 2.9 Hz), 3.33 (d, 1H, *J* = 16.1 Hz), 2.54 (dd, 1H, *J* = 16.7, 3.8 Hz), 2.33 (d, 1H, *J* = 16.1 Hz), 1.51 (s, 3H), 1.36 (s, 3H), 1.07 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 185.92, 179.69, 160.84, 159.33, 158.09, 139.05, 137.04, 110.04, 109.98, 108.47, 80.89, 79.38, 57.05, 33.97, 27.94, 27.58, 27.13, 26.86; ESI-MS *m/z*: 359.2 [M - H]⁻;HRMS (EI) calcd for C₁₉H₂₀O₇, 360.3579; found, 360.1207.

3.3. Synthesis of 2,3-O-(methylene) Deoxybostrycin (3)

To a solution of **1** (100 mg, 0.313 mmol) in 10 mL of tetrahydrofuran were added polyoxymethylene (45 mg) and *p*-toluenesulfonic acid (53.8 mg, 0.298 mmol). The reaction mixture was stirred for 20 h at room temperature and then with water (20 mL) and extracted with dichloromethane (3 × 50 mL). The combined organic layer was washed with brine, dried over anhydrous magnesium sulfate, and concentrated *in vacuo*. The resulting residue was purified on a silica gel column using dichloromethane-methanol (v/v, 250/1) as eluent to obtain 54 mg compound **3** as a red solid (CH₂Cl₂) in a 52% yield. Mp:150–152 °C; IR (KBr): $v_{max} = 3433$, 3072, 2967, 2953, 2908, 2886, 1594, 1479, 1444, 1420 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 13.12 (s, 1H), 12.70 (s, 1H), 6.16 (s, 1H), 4.82 (s, 1H), 4.73 (s, 1H), 4.15 (dd, 1H, *J* = 4.0, 3.0 Hz), 3.93 (s, 3H), 3.58 (dd, 1H, *J* = 16.5, 3.0 Hz), 3.48 (d, 1H, *J* = 16.1 Hz), 2.51 (dd, 1H, *J* = 16.5, 4.0 Hz), 2.35 (d, 1H, *J* = 16.1 Hz), 1.47

(s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 189.14, 186.20, 160.84, 158.97, 157.61, 138.48, 136.51, 110.32, 109.98, 108.80, 79.77, 79.62, 57.06, 32.34, 26.58, 24.97; ESI-MS *m/z*: 331.0 [M – H]⁻.

3.4. General Procedure for Preparation of Compounds (4–17)

To a solution of 1 (50 mg, 0.156 mmol) in 10 mL of methanol was added the corresponding amine (0.78 mmol). The reaction mixture was stirred at room temperature until the starting material disappeared (for aniline, the reaction mixture was stirred at 50 $^{\circ}$ C). The solvent was removed under reduced pressure. The resulting residue was subsequently purified using first silica gel chromatography with dichloromethane-methanol as eluent, then C18 reversed phase silica gel chromatography with methanol-water as eluent.

3.4.1. 6-(Methylamino) 1-Deoxy-6-demethoxybostrycin (4)

A red solid (MeOH) in a 40% yield; mp: 221–223 °C; IR (KBr): $v_{max} = 3375$, 3338, 2929, 2901, 2853, 2814, 1582, 1514, 1452, 1418 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 14.23 (s, 1H), 12.36 (s, 1H), 7.95 (q, 1H, *J* = 5.0 Hz), 5.55 (s, 1H), 4.75 (d, 1H, *J* = 5.1 Hz), 4.41 (s, 1H), 3.63 (dt, 1H, *J* = 7.3, 5.1 Hz), 2.83 (d, 3H, *J* = 5.0 Hz), 2.85 (dd, 1H, *J* = 18.8, 5.1 Hz), 2.77 (d, 1H, *J* = 17.9 Hz), 2.68 (dd, 1H, *J* = 18.8, 7.3 Hz), 2.57 (d, 1H, *J* = 17.9 Hz), 1.20 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.31, 183.47, 156.34, 154.56, 151.05, 139.24, 132.89, 109.17, 107.89, 98.98, 70.74, 69.38, 35.99, 30.70, 29.62, 25.93; ESI-MS *m*/*z*: 318.1 [M - H]⁻; HRMS (EI) calcd for C₁₆H₁₇NO₆, 319.1056; found, 319.1059.

3.4.2. 6-(*n*-Propylamino) 1-Deoxy-6-demethoxybostrycin (5)

A red solid (MeOH) in a 44% yield; mp: 215–216 °C; IR (KBr): $v_{max} = 3385$, 3285, 3085, 2966, 2934, 2876, 1578, 1508, 1445, 1412 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 14.23 (s, 1H), 12.37 (s, 1H), 7.86 (t, 1H, J = 5.7 Hz), 5.63 (s, 1H), 4.74 (d, 1H, J = 5.1 Hz), 4.40 (s, 1H), 3.63 (dt, 1H, J = 7.3, 5.1 Hz), 3.18 (dt, 2H, J = 5.7, 7.2 Hz), 2.85 (1H, dd, J = 18.7, 5.1 Hz), 2.79 (d, 1H, J = 17.9 Hz), 2.68 (dd, 1H, J = 17.9 Hz), 2.57 (d, 1H, J = 17.9 Hz), 1.61 (sextet, 2H, J = 7.2 Hz), 1.20 (s, 3H), 0.91 (t, 3H, J = 7.4 Hz); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.34, 183.54, 156.35, 154.50, 150.14, 139.28, 132.84, 109.18, 107.79, 98.96, 70.74, 69.37, 44.26, 36.00, 30.69, 25.93, 21.22, 11.82; ESI-MS *m/z*: 346.2 [M – H]⁻; HRMS (EI) calcd for C₁₈H₂₁NO₆, 347.1369; found, 347.1370.

3.4.3. 6-(*n*-Butylamino) 1-Deoxy-6-demethoxybostrycin (6)

A red solid (MeOH) in a 40% yield; mp: 215–217 °C; IR (KBr): $v_{max} = 3386, 3287, 3085, 2960, 2934, 2872, 1580, 1509, 1445, 1412 cm⁻¹; ¹H NMR (400 MHz, DMSO-$ *d* $₆): <math>\delta$ 14.22 (s, 1H), 12.36 (s, 1H), 7.85 (t, 1H, *J* = 6.0 Hz), 5.61 (s, 1H), 4.74 (d, 1H, *J* = 5.1 Hz), 4.40 (s, 1H), 3.63 (dt, 1H, *J* = 7.4, 5.2 Hz), 3.21 (dt, 2H, *J* = 6.0, 7.1 Hz), 2.85 (dd, 1H, *J* = 18.7, 5.2 Hz), 2.79 (d, 1H, *J* = 17.9 Hz), 2.68 (dd, 1H, *J* = 18.7, 7.4 Hz), 2.57 (d, 1H, *J* = 17.9 Hz), 1.57 (pentet, 2H, *J* = 7.1 Hz), 1.35 (sextet, 2H, *J* = 7.3 Hz), 1.20 (s, 3H), 0.91 (t, 3H, *J* = 7.3 Hz); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.30, 183.53, 156.35, 154.50, 150.09, 139.29, 132.83, 109.18, 107.79, 98.91, 70.74, 69.37, 42.33, 36.00,

30.68, 29.90, 25.94, 20.16, 14.13; ESI-MS m/z: 360.2 $[M - H]^-$; HRMS (EI) calcd for C₁₉H₂₃NO₆, 361.1525; found, 361.1522.

3.4.4. 6-(*n*-Hexylamino) 1-Deoxy-6-demethoxybostrycin (7)

A red solid (MeOH) in a 52% yield; mp: 214–215 °C; IR (KBr): $v_{max} = 3384$, 3287, 3086, 2955, 2932, 2870, 2858, 1576, 1509, 1448, 1410 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 14.25 (s, 1H), 12.38 (s, 1H), 7.88 (t, 1H, *J* = 6.1 Hz), 5.63 (s, 1H), 4.75 (d, 1H, *J* = 5.1 Hz), 4.41 (s, 1H), 3.64 (dt, 1H, *J* = 7.2, 5.2 Hz), 3.21 (dt, 2H, *J* = 6.1, 7.0 Hz), 2.86 (dd, 1H, *J* = 18.7, 5.2 Hz), 2.80 (d, 1H, *J* = 18.0 Hz), 2.69 (dd, 1H, *J* = 18.7, 7.2 Hz), 2.58 (d, 1H, *J* = 18.0 Hz), 1.57 (m, 2H), 1.37–1.25 (m, 6H), 1.19 (s, 3H), 0.87 (t, 3H, *J* = 6.8 Hz); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.35, 183.58, 156.36, 154.51, 150.13, 139.28, 132.88, 109.21, 107.82, 98.90, 70.73, 69.38, 42.61, 35.97, 31.39, 30.71, 27.75, 26.59, 25.91, 22.49, 14.35; ESI-MS *m*/*z*: 388.1 [M - H]⁻; HRMS (EI) calcd for C₂₁H₂₇NO₆, 389.1838; found, 389.1834.

3.4.5. 6-(2'-Hydroxyethylamino) 1-Deoxy-6-demethoxybostrycin (8)

A red solid (MeOH) in a 30% yield; mp: 232–234 °C; IR (KBr): $v_{max} = 3441$, 3389, 3340, 3298, 3062, 2966, 2945, 2926, 2875, 1571, 1528, 1445, 1419 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 14.21 (s, 1H), 12.36 (s, 1H), 7.63 (t, 1H, *J* = 5.9 Hz), 5.70 (s, 1H), 4.89 (br t, 1H), 4.75 (d, 1H, *J* = 4.2 Hz), 4.41 (s, 1H), 3.66–3.59 (m, 3H), 3.28 (q, 2H, *J* = 5.9 Hz), 2.86 (dd, 1H, *J* = 18.7, 5.0 Hz), 2.80 (d, 1H, *J* = 18.0 Hz), 2.69 (dd, 1H, *J* = 18.7, 7.4 Hz), 2.58 (d, 1H, *J* = 18.0 Hz), 1.20 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.46, 183.39, 156.43, 154.57, 150.35, 139.33, 132.99, 109.12, 107.78, 99.37, 70.73, 69.38, 58.95, 45.34, 36.01, 30.70, 25.92; ESI-MS *m/z*: 348.1 [M – H][–]; HRMS (EI) calcd for C₁₇H₁₉NO₇, 349.1162; found, 349.1158.

3.4.6. 6-(Prop-2'-yn-1'-ylamino) 1-Deoxy-6-demethoxybostrycin (9)

A red solid (MeOH) in a 56% yield; mp: 214–216 °C; IR (KBr): $v_{max} = 3390, 3242, 3242, 3070, 2968, 2932, 2912, 2851, 1586, 1503, 1447, 1387 cm⁻¹; ¹H NMR (400 MHz, DMSO-$ *d* $₆): <math>\delta$ 14.03 (s, 1H), 12.36 (s, 1H), 8.08 (t, 1H, *J* = 5.8 Hz), 5.74 (s, 1H), 4.76 (d, 1H, *J* = 5.1 Hz), 4.42 (s, 1H), 4.08 (dd, 2H, *J* = 5.8, 2.3 Hz), 3.64 (dt, 1H, *J* = 7.3, 5.2 Hz), 3.27 (t, 1H, *J* = 2.3 Hz), 2.86 (dd, 1H, *J* = 18.8, 5.1 Hz), 2.80 (d, 1H, *J* = 18.0 Hz), 2.69 (dd, 1H, *J* = 18.8, 7.3 Hz), 2.59 (d, 1H, *J* = 18.0 Hz), 1.20 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.54, 183.10, 156.66, 154.94, 149.46, 139.31, 133.53, 109.10, 107.64, 101.23, 79.13, 75.25, 70.71, 69.36, 36.05, 31.83, 30.67, 25.92; ESI-MS *m/z*: 342.1 [M – H]⁻; HRMS (EI) calcd for C₁₈H₁₇NO₆, 343.1056; found, 343.1052.

3.4.7. 6-[(2'-(Dimethylamino)ethyl)amino]-1-deoxy-6-demethoxybostrycin (10)

A red solid (MeOH) in a 60% yield; mp: 215–217 °C; IR (KBr): $v_{max} = 3287$, 3087, 2974, 2944, 2863, 2818, 1575, 1512, 1444, 1411 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 14.19 (s, 1H), 12.29 (s, 1H), 7.47 (t, 1H, J = 5.4 Hz), 5.67 (s, 1H), 4.75 (d, 1H, J = 4.6 Hz), 4.41 (s, 1H), 3.63 (dt, 1H, J = 7.4, 5.3 Hz), 3.37–3.26 (m, 4H), 2.86 (dd, 1H, J = 18.7, 5.2 Hz), 2.79 (d, 1H, J = 18.1 Hz), 2.68 (dd, 1H, J = 18.7, 7.4 Hz), 2.59 (d, 1H, J = 18.1 Hz), 2.25 (s, 6H), 1.20 (s, 3H); ¹³C NMR (100 MHz, 2.25 (s, 6H), 1.20 (s, 3H); ¹³C NMR (100 MHz).

DMSO-*d*₆): δ 186.44, 183.21, 156.45, 154.65, 149.81, 139.40, 133.07, 109.10, 107.78, 99.47, 70.73, 69.37, 56.39, 45.28, 36.01, 30.70, 25.92; ESI-MS *m*/*z*: 375.1 [M - H]⁻; HRMS (EI) calcd for C₁₉H₂₄N₂O₆, 376.1634; found, 376.1628.

3.4.8. 6-(Phenylamino) 1-Deoxy-6-demethoxybostrycin (11)

A red solid (MeOH) in a 38% yield; mp: 220–222 °C; IR (KBr): $v_{max} = 3359$, 3283, 3060, 2980, 2940, 2857, 2821, 1584, 1543, 1444, 1382 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.92 (s, 1H), 12.49 (s, 1H), 9.46 (s, 1H), 7.48–7.24 (m, 5H), 6.00 (s, 1H), 4.76 (br s, 1H), 4.43 (s, 1H), 3.64 (br t, 1H), 2.86 (dd, 1H, J = 18.6, 5.0 Hz), 2.82 (d, 1H, J = 18.2 Hz), 2.69 (dd, 1H, J = 18.8, 7.4 Hz), 2.60 (d, 1H, J = 18.2 Hz), 1.20 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.93, 183.08, 156.96, 154.94, 147.89, 139.26, 138.24, 133.89, 129.83, 126.18, 124.48, 109.21, 107.80, 101.70, 70.72, 69.38, 36.14, 30.66, 25.93; ESI-MS m/z: 380.1 [M – H]⁻; HRMS (EI) calcd for C₂₁H₁₉NO₆, 381.1212; found, 381.1204.

3.4.9. 6-(Benzylamino) 1-Deoxy-6-demethoxybostrycin (12)

A red solid (MeOH) in 29% yield; mp: 220–222 °C; IR (KBr): $v_{max} = 3382$, 3264, 3085, 2973, 2934, 2900, 2870, 1574, 1511, 1444, 1411 cm⁻¹; ¹H NMR (300 MHz, DMSO-*d*₆): δ 12.37 (s, 1H), 8.49 (t, 1H, J = 6.4 Hz), 7.38–7.18 (m, 5H), 5.53 (s, 1H), 4.75 (d, 1H, J = 5.3 Hz), 4.46 (d, 2H, J = 6.4 Hz), 4.42 (s, 1H), 3.61 (dt, 1H, J = 7.5, 5.3 Hz), 2.82 (dd, 1H, J = 18.9, 5.3 Hz), 2.77 (d, 1H, J = 17.4 Hz), 2.66 (dd, 1H, J = 18.9, 7.5 Hz), 2.57 (d, 1H, J = 17.4 Hz,), 1.17 (s, 3H); ESI-MS *m/z*: 394.1 [M – H]⁻; HRMS (EI) calcd for C₂₂H₂₁NO₆, 395.1369; found, 395.1361.

3.4.10. 6-(p-Methoxybenzylamino) 1-Deoxy-6-demethoxybostrycin (13)

A red solid (MeOH) in a 44% yield; mp: 223–225 °C; $[\alpha]^{20}_{D} = -250.0^{\circ}$ (c = 1.00, CH₃OH); IR (KBr): $v_{max} = 3373$, 3069, 2979, 2938, 2861, 2834, 1583, 1507, 1445, 1387 cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6): δ 14.11 (s, 1H), 12.39 (s, 1H), 8.40 (t, 1H, J = 6.5 Hz), 7.33–6.87 (m, 4H), 5.57 (s, 1H), 4.74 (d, 1H, J = 5.1 Hz), 4.40 (br s, 2H), 4.39 (s, 1H), 3.73 (s, 3H), 3.63 (dt, 1H, J = 7.2, 5.2 Hz), 2.84 (dd, 1H, J = 18.8, 5.2 Hz), 2.80 (d, 1H, J = 18.1 Hz), 2.68 (dd, 1H, J = 18.8, 7.2 Hz), 2.58 (d, 1H, J = 18.1 Hz), 1.19 (s, 3H); ¹³C NMR (100 MHz, DMSO- d_6): δ 186.33, 183.51, 158.95, 156.45, 154.65, 149.93, 139.26, 133.12, 129.42, 129.04, 129.04, 114.42, 114.42, 109.18, 107.72, 100.10, 70.71, 69.36, 55.52, 45.25, 35.99, 30.68, 25.91; ESI-MS m/z: 424.1 [M – H] ⁻; HRMS (EI) calcd for C₂₃H₂₃NO₇, 425.1475; found, 425.1464.

3.4.11. 6-(p-Fluorobenzylamino) 1-Deoxy-6-demethoxybostrycin (14)

A red solid (MeOH) in a 47% yield; mp: 234–236 °C; IR (KBr): $v_{max} = 3272$, 3085, 2976, 2936, 2873, 2856, 1577, 1510, 1443, 1411 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 14.07 (s, 1H), 12.40 (s, 1H), 8.46 (t, 1H, J = 6.5 Hz), 7.44–7.14 (m, 4H), 5.57 (s, 1H), 4.74 (d, 1H, J = 5.1 Hz), 4.46 (d, 2H, J = 6.5 Hz), 4.41 (s, 1H), 3.63 (dt, 1H, J = 7.3, 5.2 Hz), 2.85 (dd, 1H, J = 18.8, 5.2 Hz), 2.80 (d, 1H, J = 18.1 Hz), 2.68 (dd, 1H, J = 18.8, 7.2 Hz), 2.58 (d, 1H, J = 18.1 Hz), 1.19 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 186.43, 183.44, 160.63, 156.47, 154.68, 149.98, 139.24, 133.80, 133.20,

129.77, 129.69, 115.74, 109.19, 107.72, 100.21, 70.71, 69.36, 45.00, 35.99, 30.68, 25.90; ESI-MS *m/z*: 412.1 [M – H]⁻; HRMS (EI) calcd for $C_{22}H_{20}FNO_6$, 413.1275; found, 413.1267.

3.4.12. 6-(Piperidin-1-yl) 1-Deoxy-6-demethoxybostrycin (15)

A red solid (MeOH) in a 30% yield; mp: 188–190 °C; IR (KBr): $v_{max} = 3369$, 3065, 2935, 2856, 1593, 1551, 1448, 1409 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.86 (s, 1H), 12.66 (s, 1H), 5.98 (s, 1H), 4.74 (d, 1H, J = 5.1 Hz), 4.39 (s, 1H), 3.63 (dt, 1H, J = 7.3, 5.2 Hz), 3.55 (br s, 4H), 2.83 (dd, 1H, J = 18.7, 5.2 Hz), 2.79 (d, 1H, J = 18.2 Hz), 2.67 (dd, 1H, J = 18.7, 7.3 Hz), 2.57 (d, 1H, J = 18.2 Hz), 1.65 (br s, 6H), 1.19 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 185.37, 184.10, 157.65, 155.43, 154.35, 138.14, 134.29, 110.61, 109.80, 107.87, 70.72, 69.40, 50.89,36.22, 30.49, 25.93, 24.15; ESI-MS *m/z*: 372.1 [M – H]⁻; HRMS (EI) calcd for C₂₀H₂₃NO₆, 373.1525; found, 373.1522.

3.4.13. 6-(4'-Methylpiperidin-1-yl) 1-Deoxy-6-demethoxybostrycin (16)

A red solid (MeOH) in a 32% yield; mp: 198–200 °C; $[\alpha]^{20}{}_{D} = -44.8^{\circ}$ (c = 1.00, CH₃OH); IR (KBr): $v_{max} = 3407$, 3087, 2948, 2923, 2872, 1593, 1573, 1543, 1455 cm⁻¹; ¹H NMR (400 MHz, DMSO- d_6): δ 13.85 (s, 1H), 12.66 (s, 1H), 5.98 (s, 1H), 4.74 (d, 1H, J = 5.1 Hz), 4.39 (s, 1H), 4.09 (br d, 2H), 3.63 (dt, 1H, J = 7.3, 5.2 Hz), 3.01 (br t, 2H), 2.81 (dd, 1H, J = 18.7, 5.2 Hz), 2.79 (d, 1H, J = 18.2 Hz), 2.67 (dd, 1H, J = 18.7, 7.3 Hz), 2.56 (d, 1H, J = 18.2 Hz), 1.75 (m, 2H), 1.67 (m, 1H), 1.31 (m, 2H), 1.19 (s, 3H), 0.94 (d, 3H, J = 6.3 Hz); ¹³C NMR (100 MHz, DMSO- d_6): δ 185.35, 184.07, 157.69, 155.47, 154.27, 138.16, 134.30, 110.59, 110.00, 107.86, 70.72, 69.40, 50.14, 36.22, 34.08, 30.50, 25.94; ESI-MS m/z: 386.1 [M – H]⁻; HRMS (EI) calcd for C₂₁H₂₅NO₆, 387.1682; found, 387.1678.

3.4.14. 6-(4'-Phenylpiperidin-1-yl) 1-Deoxy-6-demethoxybostrycin (17)

A red solid (MeOH) in a 28% yield; mp: 227–228 °C; IR (KBr): $v_{max} = 3332$, 3084, 2939, 2927, 2876, 2847, 1593, 1567, 1537, 1454, 1407 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.85 (s, 1H), 12.70 (s, 1H), 7.36–7.17 (m, 5H), 6.07 (s, 1H), 4.75 (d, 1H, *J* = 5.1 Hz), 4.41 (s, 1H), 4.27 (br d, 2H), 3.64 (dt, 1H, *J* = 7.3, 5.2 Hz), 3.14 (br t, 2H), 2.87 (m, 2H), 2.79 (d, 1H, *J* = 18.2 Hz), 2.69 (dd, 1H, *J* = 18.7, 7.2 Hz), 2.58 (d, 1H, *J* = 18.2 Hz), 1.81 (m, 4H), 1.20 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 185.38, 183.96, 157.85, 155.65, 154.29, 145.87, 138.20, 134.46, 128.90,127.19, 127.19,126.71, 110.63, 110.37, 107.91, 70.72, 69.41, 50.50,41.85, 36.24, 33.23,30.50, 25.94; ESI-MS *m/z*: 448.2 [M – H]⁻; HRMS (EI) calcd for C₂₆H₂₇NO₆, 449.1838; found, 449.1837.

3.5. General Procedure for Preparation of Compounds (18–22)

To a solution of 1 (50 mg, 0.156 mmol) and triethylamine (8 equivalents) in 10 mL of methanol was added the corresponding thiol (0.624 mmol, the butane-2,3-dithiol was racemate). The reaction mixture was stirred at 0-5 °C until the starting material disappeared. The solvent was removed under reduced pressure. The resulting residue was subsequently purified using first silica gel chromatography with dichloromethane-methanol as eluent, and then C18 reversed phase silica gel column with methanol-water as eluent to obtain the corresponding products.

A red solid (MeOH) in a 35% yield; mp: 186–188 °C; IR (KBr): $v_{max} = 3378$, 2976, 2959, 2925, 2855, 1611, 1574, 1489, 1427, 1408 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.11 (s, 2H), 4.79 (br s, 1H), 4.46 (s, 1H), 3.63 (br t, 1H), 3.26 (q, 4H, *J* = 7.4 Hz), 2.83 (dd, 1H, *J* = 18.1, 5.4 Hz), 2.79 (d, 1H, *J* = 18.7 Hz), 2.66 (dd, 1H, *J* = 19.0, 7.4 Hz), 2.58 (d, 1H, *J* = 18.7 Hz), 1.21 (m, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 175.21, 165.23, 165.19, 145.47, 138.77, 138.58, 109.40, 70.60, 69.34, 36.27, 30.32, 29.26,25.81, 15.63; ESI-MS *m/z*: 409.0 [M – H]⁻; HRMS (EI) calcd for C₁₉H₂₂O₆S₂, 410.0858; found, 410.0855.

3.5.2. 6,7-Bis(n-butylthio) 1-Deoxy-6-demethoxybostrycin (19)

A red solid (MeOH) in a 39% yield; mp: 178–180 °C; $[\alpha]^{20}_{D} = -17.4^{\circ}$ (*c* = 1.00, CH₃OH); IR (KBr): $v_{max} = 3317, 2958, 2929, 2870, 1599, 1438, 1422, 1408 cm⁻¹; ¹H NMR (400 MHz, DMSO-$ *d* $₆): <math>\delta$ 13.12 (s, 2H), 4.80 (br s, 1H), 4.46 (s, 1H), 3.63 (br t, 1H), 3.25 (t, 4H, *J* = 7.2 Hz), 2.83 (dd, 1H, *J* = 19.0, 5.6 Hz), 2.78 (d, 1H, *J* = 18.7 Hz), 2.66 (dd, 1H, *J* = 19.0, 7.4 Hz), 2.57 (d, 1H, *J* = 18.7 Hz), 1.56–1.47 (m, 4H), 1.38 (sextet, 4H, *J* = 7.3 Hz), 1.20 (s, 3H), 0.86 (t, 6H, *J* = 7.3 Hz); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 175.10, 165.35, 165.31, 145.78, 138.79, 138.62, 109.41, 70.60, 69.34, 36.28, 34.64, 32.33, 30.31, 25.81, 21.62, 13.89;; ESI-MS *m/z*: 465.2 [M - H]⁻; HRMS (EI) calcd for C₂₃H₃₀O₆S₂, 466.1484; found, 466.1483.

3.5.3. 6,7-Bis(*n*-hexylthio) 1-Deoxy-6-demethoxybostrycin (20)

A red solid (MeOH) in a 55% yield; mp: 170–172 °C; IR (KBr): $v_{max} = 3322, 2956, 2925, 2853, 1599, 1439, 1422, 1408 cm⁻¹; ¹H NMR (400 MHz, DMSO-$ *d* $₆): <math>\delta$ 13.11 (s, 2H), 4.79 (br s, 1H), 4.45 (s, 1H), 3.62 (br t, 1H), 3.23 (t, 4H, *J* = 7.2 Hz), 2.82 (dd, 1H, *J* = 19.2, 5.4 Hz), 2.77 (d, 1H, *J* = 18.7 Hz), 2.65 (dd, 1H, *J* = 19.2, 7.3 Hz), 2.56 (d, 1H, *J* = 18.7 Hz), 1.57–1.47 (m, 4H), 1.40–1.33 (m, 4H), 1.27–1.21 (m, 8H), 1.20 (s, 3H), 0.87–0.80 (m, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 175.22, 165.19, 165.14, 145.83, 138.74, 138.59, 109.36,70.60, 69.32, 36.29, 34.97,31.19, 30.29, 30.20, 28.12, 25.81, 22.43, 14.28; ESI-MS *m/z*: 521.1 [M – H][–]; HRMS (EI) calcd for C₂₇H₃₈O₆S₂, 522.2110; found, 522.2107.

3.5.4. 6,7-(Ethan-1',2'-yl-dithio) 1-Deoxy-6-demethoxybostrycin (21)

A red solid (MeOH) in a 47% yield; mp: 226–227 °C; IR (KBr): $v_{max} = 3529$, 3479, 2974, 2922, 2849, 2824, 1588, 1515, 1449, 1415 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 12.64 (s, 2H), 4.80 (d, 1H, J = 5.2 Hz), 4.45 (s, 1H), 3.64 (dt, 1H, J = 7.5, 5.2 Hz), 3.34 (s, 4H), 2.85 (dd, 1H, J = 18.9, 5.4 Hz), 2.80 (d, 1H, J = 18.4 Hz), 2.66 (dd, 1H, J = 18.9, 7.7 Hz), 2.59 (d, 1H, J = 18.4Hz), 1.21 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 180.71, 156.93, 156.86, 140.79, 137.10, 136.93, 107.63, 107.60, 70.63, 69.29, 36.48, 30.33, 26.68, 25.89; ESI-MS *m/z*: 379.1 [M – H]⁻; HRMS (EI) calcd for C₁₇H₁₆O₆S₂, 380.0388; found, 380.0380.

3.5.5. 6,7-(Butan-2',3'-yl-dithio) 1-Deoxy-6-demethoxybostrycin (22)

A red solid (MeOH) in a 78% yield; mp: 225–227 °C; $[\alpha]^{20}_{D} = -188.7^{\circ}$ (*c* = 1.00, CH₃OH); IR (KBr): $v_{max} = 3484$, 3406, 2969, 2930, 2873, 2818, 1582, 1515, 1443, 1413 cm⁻¹; ¹H NMR (400 MHz, DMSO-*d*₆): δ 12.68 (s, 1H), 12.66 (s, 1H), 4.79 (d, 1H, *J* = 5.2 Hz), 4.44 (s, 1H), 3.68–3.44 (m, 3H), 2.84 (dd, 1H, *J* = 18.6, 5.2 Hz), 2.80 (d, 1H, *J* = 18.5 Hz), 2.66 (dd, 1H, *J* = 18.6, 7.7 Hz), 2.58 (d, 1H, *J* = 18.5 Hz), 1.32 and 1.30 (each d, 3H, *J* = 6.0 Hz), 1.21 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 180.96, 180.86, 156.73, 156.65, 139.83, 139.29, 136.96, 136.84, 107.75, 70.59, 69.26, 36.48, 30.30, 25.90, 23.33, 18.08; ESI-MS *m/z*: 407.1 [M – H]⁻; HRMS (EI) calcd for C₁₉H₂₀O₆S₂, 408.0698; found, 408.0701.

3.6. Antitumor Activity in Vitro

3.6.1. Cell Culture

MDA-MB-435, HepG2 and HCT-116 cells were cultured in Dulbecco's modification Eagle's medium (DMEM, Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (FBS, Hyclone, Logan, UT, USA), 2 mM L-glutamine, 100 μ g/mL streptomycin and 100 U/mL penicillin (Invitrogen). The cells were incubated at 37 °C in a humidified atmosphere of 5% CO₂.

3.6.2. Assessment of Antitumor Activity by MTT Assay

Cells were seeded in 96-well flat-bottom plates at a density of 1×10^4 cells/mL, and cultured in a humidified incubator (5% CO₂) at 37 °C for 24 h, followed by exposure to various concentrations of compounds tested for 48 h. Subsequently, 20 µL of MTT reagent (Genview, Houston, TX, USA, 5 mg/mL) dissolved in PBS (pH 7.4) was added to each well and mixed, the cells were then incubated for an additional 4 h. Culture supernatant was moved, 150 µL of DMSO (Sangon Biotech, Shanghai, China) was added to each well to fully dissolve the MTT-formazan crystals. Cell growth inhibition was determined by measuring the absorbance (Abs) at $\lambda = 570$ nm using a microplate reader and calculated according to the following equation:

$$[Growth inhibition = (1 - OD of treated cells/OD of control cells) \times 100\%]$$
(1)

The half maximal inhibitory concentrations (IC_{50}) were obtained from liner regression analysis of the concentration-response curves plotted for each tested compound.

4. Conclusions

In this paper, 21 derivatives of deoxybostrycin were designed, synthesized and evaluated for their anti-tumor activity against MDA-MB-435, HepG2 and HCT-116 cell lines. The bioassay results indicated that most of these derivatives possess good anti-tumor activities. The substitution pattern on the anthraquinone ring affected anticancer activity remarkably. It was confirmed that a methoxyl at C-6 is not necessary for cytotoxic activity. However, acetonide formed at C-2, C-3 in compound **2** strongly reduces cytotoxic activity. Replacement of the methoxyl at C-6 with amines does not improve the anti-tumor activity. Introduction of alkylthio groups at C-6 and C-7 positions of the deoxybostrycin

improved the cytotoxicity greatly. In particular, compounds **19**, **21** and **22** displayed the highest cellular cytotoxicity against MDA-MB-435 and distinguished themselves as potential anti-tumor agents.

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References

- 1. Heron, M. Deaths: Leading causes for 2005. Natl. Vital Stat. Rep. 2009, 58, 1–97.
- 2. Heron, M. Deaths: Leading causes for 2004. Natl. Vital Stat. Rep. 2007, 56, 1–96.
- 3. David, V.; Jaime, R.A.; Cristina, T.; Pedro, C.B.; Jaime, V.A. Studies on quinones. Part 46. Synthesis and *in vitro* antitumor evaluation of aminopyrimidoisoquinolinequinones. *Eur. J. Med. Chem.* **2010**, *45*, 5234–5242.
- 4. Thomas, E. Cancer therapy with natural products and medicinal plants. *Planta Med.* **2010**, *76*, 1035–1036.
- Wang, L.; Li, D.; Xu, S.; Cai, H.; Yao, H.; Zhang, Y.; Jiang, J.; Xu, J. The conversion of oridonin to spirolactone-type or enmein-type diterpenoid, synthesis and biological evaluation of ent-6,7-seco-oridonin derivatives as novel potential anticancer agents. *Eur. J. Med. Chem.* 2012, 52, 242–250.
- 6. Schwartsmann, G.; Da Rocha, A.B.; Mattei, J.; Lopes, R. Marine-derived anticancer agents in clinical trials. *Expert. Opin. Inv. Drug* **2003**, *12*, 1367–1383.
- 7. D'Incalci, M.; Simone, M.; Tavecchio, M.; Damia, G.; Garbi, A.; Erba, E. New drugs from the sea. *J. Chemother.* **2004**, *16*, 86–89.
- 8. O'Hanlon, L.H. Scientists are searching the seas for cancer drugs. *J. Natl. Cancer Inst.* **2006**, *98*, 662–663.
- 9. Banerjee, S.; Wang, Z.; Mohammad, M.; Sarkar, F.H.; Mohammad, R.M. Efficacy of selected natural products as therapeutic agents against cancer. *J. Nat. Prod.* **2008**, *71*, 492–496.
- Xia, X.K.; Li, Q.; Li, J.; Shao, C.L.; Zhang, J.Y.; Zhang, Y.G.; Liu, X.; Lin, Y.C.; Liu, C.H.; She, Z.G. Two new derivatives of griseofulvin from the mangrove endophytic fungus *Nigrospora* sp. (Strain No. 1403) from *Kandelia candel* (L.) Druce. *Planta Med.* 2011, 77, 1735–1738.
- 11. Charudattan, R.; Rao, K.V. Bostrycin and 4-deoxybostrycin: Two nonspecific phytotoxins produced by *Alternaria eichhorniae*. *Appl. Environ. Microbiol.* **1982**, *43*, 846–849.
- Sommart, U.; Rukachaisirikul, V.; Sukpondma, Y.; Phongpaichit, S.; Sakayaroj, J.; Kirtikara, K. Hydronaphthalenones and a dihydroramulosin from the endophytic fungus PSU-N24. *Chem. Pharm. Bull.* 2008, *56*, 1687–1690.

- Ge, H.M.; Song, Y.C.; Shan, C.Y.; Ye, Y.H.; Tan, R.X. New and cytotoxic anthraquinones from *Pleospora* sp. IFB-E006, an endophytic fungus in *Imperata cylindrical. Planta Med.* 2005, *71*, 1063–1065.
- 14. Nota, T.; Take, T.; Watanabe, T.; Abe, J. The structure of bostrycin. *Tetrahedron* **1970**, *26*, 1339–1346.
- Chen, H.; Zhong, L.L.; Long, Y.H.; Li, J.; Wu, J.H.; Liu, L.; Chen, S.P.; Lin, Y.C.; Li, M.F.; Zhu, X.; She, Z.G. Studies on the synthesis of derivatives of marine-derived bostrycin and their structure-activity relationship against tumor cells. *Mar. Drugs* 2012, *10*, 932–952.
- 16. Miko, M.; Drobnica, L.; Chance, B. Inhibition of energy metabolism in Ehrlich ascites cells treated with dactylarin *in vitro*. *Cancer Res.* **1979**, *39*, 4242–4251.
- 17. Haraguchi, H.; Abo, T.; Fukuda, A.; Okamura, N.; Yagi, A. Mode of phytotoxic action of Altersolanols. *Phytochemistry* **1996**, *431*, 989–992.
- Xie, G.E.; Zhu, X.; Li, Q.; Gu, M.H.; He, Z.J.; Wu, J.H.; Li, J.; Lin, Y.C.; Li, M.F.; She, Z.G.; *et al.* SZ-685C, a marine anthraquinone, is a potent inducer of apoptosis with anticancer activity by suppression of the Akt/FOXO pathway. *Br. J. Pharmacol.* 2010, *159*, 689–697.

Samples Availability: Available from the authors.

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