



Review

Long-Acting Anti-HIV Drugs Targeting HIV-1 Reverse Transcriptase and Integrase

Kamal Singh 1,2,3,*, Stefan G. Sarafianos 4 and Anders Sönnerborg 1,3,5

- Department of Molecular Microbiology and Immunology, University of Missouri, Columbia, MO 65211, USA
- Bond Life Sciences Center, University of Missouri, Columbia, MO 65211, USA
- Division of Clinical Microbiology, Department of Laboratory Medicine, Karolinska Institute, Huddinge 14186, Stockholm, Sweden; Anders.Sonnerborg@ki.se
- Laboratory of Biochemical Pharmacology, Department of Pediatrics, Emory University School of Medicine, Atlanta, GA 30322, USA; stefanos.sarafianos@emory.edu
- Division of Infectious Diseases, Department of Medicine Huddinge, Karolinska Institute, Huddinge 14186, Stockholm, Sweden
- * Correspondence: singhka@missouri.edu; Tel.: +1-573-882-9024

Received: 17 March 2019; Accepted: 18 April 2019; Published: 20 April 2019



Abstract: One of the major factors contributing to HIV-1 drug resistance is suboptimal adherence to combination antiretroviral therapy (cART). Currently, recommended cART for HIV-1 treatment is a three-drug combination, whereas the pre-exposure prophylaxis (PrEP) regimens consist of one or two antivirals. Treatment regimens require adherence to a once or twice (in a subset of patients) daily dose. Long-acting formulations such as injections administered monthly could improve adherence and convenience, and thereby have potential to enhance the chances of expected outcomes, although long-lasting drug concentrations can also contribute to clinical issues like adverse events and development of drug resistance. Globally, two long-acting antivirals have been approved, and fifteen are in clinical trials. More than half of investigational long-acting antivirals target HIV-1 reverse transcriptase (HIV-1 RT) and/or integrase (HIV-1 IN). Here, we discuss the status and potential of long-acting inhibitors, including rilpivirine (RPV), dapivirine (DPV), and 4-ethynyl-2-fluoro-2-deoxyadenosine (EFdA; also known as MK-8591), which target RT, and cabotegravir (CAB), which targets IN. The outcomes of various clinical trials appear quite satisfactory, and the future of long-acting HIV-1 regimens appears bright.

Keywords: HIV-1; long-acting formulation; reverse transcriptase; integrase; antivirals

1. Introduction

Revolutionary advances in combination antiretroviral therapy (cART) have rendered HIV from a fatal to chronic disease. If managed efficiently cART significantly improves the life expectancy of a patient [1]. The cART restores (or maintains) CD4 cell counts and suppresses viral load. Currently recommended first-line treatments for HIV infection include a three-drug coformulation in a once-daily fixed-dose single-pill regimen. This formulation virologically suppresses HIV in more than 80% of patients [2–4]. Some components have also been recommended for pre-exposure prophylaxis (PrEP) [5]. The approved PrEP regimens are tenofovir disoproxil fumarate (TDF) monotherapy or the combination of TDF and emtricitabine (2',3'-dideoxy-5-fluoro-3'-thiacytidine or FTC) [5]. The results of clinical trials have shown a reduction in the risk of HIV acquisition by more than 85% in uninfected individuals on PrEP medication [6,7]. In spite of such success, desired therapeutic outcomes of cART are hampered by compromised adherence in both resource-rich and low- and middle-income countries. Long-acting (LA) treatment strategies, especially parenteral, have been successful in facilitating the adherence and

minimizing the lapse in medication in other fields of medicine [8–10]. Hence, parallel strategies to improve adherence to anti-HIV regimens have been sought.

To date, one LA antiviral (ibalizumab, a humanized IgG4 antibody) has been approved by the United States Food and Drug Administration. Ibalizumab inhibits HIV infection at post attachment steps by binding to domain 2 of the CD4 receptor and blocking binding of HIV to the cell. Ibalizumab, together with optimized background therapy, is recommended for treatment-experienced patients with multidrug-resistant viruses who are failing current therapy [2]. Another LA drug, which is an entry inhibitor interfering with the binding to gp41 and thereby the fusion step (albuvirtide, a 32-amino acid long analog of gp41) is approved exclusively in China. An additional fifteen antivirals are currently being evaluated in preclinical or advanced clinical trials. Of these, six target HIV-1 reverse transcriptase (HIV-1 RT), two inhibit HIV-1 integrase (HIV-1 IN), three are entry inhibitors, two block capsid (CA) assembly/disassembly processes, and the remaining two inhibit HIV protease (PR). Here, we discuss only RT and IN inhibitors including: EFdA (4-ethynyl-2-fluoro-2-deoxyadenosine) also known as MK-8591, a nucleoside RT inhibitor that inhibits HIV by multiple mechanisms; rilpivirine (RPV), a second-generation non-nucleoside RT inhibitor (NNRTI); and cabotegravir (CAB), a second-generation integrase strand transfer inhibitor (INSTI).

2. LA Antivirals Targeting HIV-1 RT

Currently, three RTIs are in clinical trials and another three are in preclinical development (Table 1). HIV-1 RT is a multifunctional enzyme, composed of two subunits (p66 and p51) that has RNA- and DNA-dependent DNA polymerase activities, as well as an RNase H activity [11]. HIV-1 RT has been the most sought target for anti-HIV drug development. Thus, approximately half of the approved antivirals target RT. There are two classes of HIV-1 RT inhibitors: nucleos(t)ide RT inhibitors (NRTIs) and non-nucleoside RT inhibitors (NNRTIs). The NRTIs are competitive inhibitors that bind at the dNTP (deoxynucleotide triphosphate) binding site [12]. All NRTIs currently approved for the treatment of HIV infection lack a 3'OH and, thus, act as chain terminators [11–13]. The NNRTIs are allosteric RT inhibitors, which bind ~10 Å away from the polymerase active site in a pocket known as the NNRTI binding pocket (NNIBP) [14]. The NNRTIs reposition nucleic acid binding [15] and restrict conformational changes required for the catalysis of DNA synthesis by HIV-1 RT [12].

Table 1. Long-acting antivirals targeting HIV-1 RT and HIV-1 IN. NRTI, nucleos(t)ide RT inhibitors; NNRTI, non-nucleoside RT inhibitors; INSTI, integrase strand transfer inhibitors; EFdA, 4-ethynyl-2-fluoro-2-deoxyadenosine; TAF, tenofovir alafenamide fumarate; DPV, dapivirine; RPV, rilpivirine; RAL, raltegravir.

Drug Class	Drug	Formulation	Clinical Trial Stage
NRTI	EFdA	Implant (vaginal film, subcutaneous polyethylene vinyl acetate membrane)	Phase II
	TAF	Implant (multipurpose intravaginal ring, subdermal polyvinyl acid membrane, subcutaneous thin-film polycaprolactone) Injectable (subcutaneous nanosuspension)	Preclinical
	GS-9131	Injectable (intravenous propylene or polyethylene glycol in citric acid)	Preclinical
NNRTI	DPV	Implant (vaginal ring)	Phase III
	RPV	Injectable (subcutaneous/intramuscular nanosuspension) Implant (microarray patch) Topical (nanoformulation)	Phase III
	Elsulfavirine	Injectable (subcutaneous/intramuscular nanosuspension)	Preclinical
INSTI	CAB	Injectable (intramuscular nanosuspension)	Phase III
	RAL	Injectable (subcutaneous nanosuspension)	Preclinical

Pharmaceuticals **2019**, 12, 62 3 of 14

2.1. Long-Acting NRTIs

2.1.1. EFdA (Islatravir)

EFdA is an extremely potent NRTI [16]. The EFdA-triphosphate binding affinity of HIV-1 RT can be greater than the natural dNTP substrate [17,18]. In contrast to all approved NRTIs, EFdA retains a 3′-OH group (Figure 1). EFdA also contains a 4′-ethynyl and a 2-fluoro group (Figure 1). Our early studies suggested that incorporation of EFdA-monophosphate in the elongating DNA chain blocks the translocation of RT along the template strand [19,20]. Therefore, we termed EFdA as a translocation-deficient RT inhibitor (TDRTI) [17]. Early molecular modeling [17] and more recent crystallographic studies [20] suggested that the 4′-ethynyl moiety of EFdA binds in a hydrophobic pocket contributing to the strong RT affinity for EFdA. Biochemical studies showed that EFdA inhibits RT through multiple mechanisms: it can block DNA synthesis either as a delayed or as an obligate chain terminator, and EFdA-MP-terminated primers can be protected from excision; also, EFdA-MP is often misincorporated by RT, leading to mismatched primers that are also hard to extend and protected from excision [18,20]. EFdA exhibits picomolar range antiviral activity in activated peripheral blood mononuclear cells (PBMCs) and MT4 cells and can be used in combinations with clinically-used antiretroviral drugs [21]. Importantly, tenofovir-resistant K65R HIV is hypersusceptible to EFdA ([19] and reviewed in [22]).

Figure 1. Nucleos(t)ide reverse transcriptase inhibitors in clinical trials or in preclinical development as long-acting (LA) antivirals.

GS-9131

EFdA has a high genetic barrier to resistance in culture, which has been attributed to the strong and generally conserved interactions between EFdA and the HIV-1 RT polymerase active site [23,24]. In vitro drug susceptibility assays showed that EFdA had significantly greater efficacy compared to other NRTIs [23]. Moreover, EFdA inhibits both WT and RTI-resistant viruses in a subtype-independent manner [25].

With respect to the potential of EFdA/MK-8591 as an LA antiviral, a single injection in rats of an extended release parenteral formulation of MK-8591 (in bioerodible poly(lactic acid) (PLA) and poly(caprolactone) (PCL) and nonerodible ethylene co-vinyl acetate (EVA)) was shown to result in levels of the inhibitor adequate for efficient antiviral effect for more than 180 days [26,27]. EFdA is suitable for formulation in polymer implants [28] and vaginal delivery films [29,30]. Studies have shown a strong potential of EFdA also as a microbicide [30,31]. These studies, combined with studies on macaque animal models [22,32,33], together with the results of phase Ia and Ib human clinical

Pharmaceuticals **2019**, 12, 62 4 of 14

trials [34,35], suggest that EFdA has strong potential as an LA and PrEP regimen or it can be combined with other antiretrovirals for treatment or maintenance therapy.

2.1.2. Tenofovir Alafenamide Fumarate

Tenofovir alafenamide fumarate (TAF) (Figure 1) is chemically related to tenofovir disoproxil fumarate (TDF), which has been part of first-line therapy after its approval in 2001. TDF as monotherapy or co-formulated with emtricitabine (Truvada®) has been approved for PrEP. Reports have shown that a one-tenth dose of TAF has better potency than its predecessor (TDF) [36]. Furthermore, although TAF is more potent than TFV (tenofovir) in vitro, the antiviral susceptibilities to TAF and TFV are highly correlated, indicating that the two compounds have virtually the same resistance profile when assessed as fold change from the wild-type [37]. However, the increased cell loading of TFV with TAF versus TDF observed in vivo suggests that TAF may retain activity against TDF-resistant mutant viruses [37]. Double-blind phase III clinical trial data suggested that TAF-containing regimens have a favorable long-term renal and bone safety profile [38]. A subdermal polyvinyl alcohol implant linearly delivered TFV in plasma at measurable concentrations for more than six weeks [39]. A thin-film polymer devise (TFPD) containing TFV LA demonstrated linear release of 60 days depending on the rate of release [40], and TAF + FTC-loaded nanoparticles showed protection from HIV-1 (Day 4: 80%, Days 7 and 14: 60%, respectively) in humanized BLT (bone marrow-liver-thymus) mice [41]. A release rate of TAF hemifumarate (TAF₂) ranging between 0.35 and 0.40 mg/day from multipurpose intravaginal rings (IVRs) has also been reported. These rings also contained acyclovir (ACV) and etonogestrel (ENG) in combination with ethinyl estradiol (EE) [42]. In PBMCs, a high concentration of TFV-diphosphate (median, 512 fmol/10⁶ cells) was present up to 35 days. In animal studies, the concentration of TFV-diphosphate was at least 10-fold greater than that associated with effective PrEP in humans taking standard daily doses of oral TDF [39].

2.1.3. GS-9131 (Rovafovir Etalafenamide)

GS-9131 (Figure 1) is a phosphonoamidate prodrug with 2'-fluoro modification [43,44] that has demonstrated antiviral activity against viruses containing K65R, L74V, and M184V resistance mutations [42] with a very robust barrier to resistance and a unique resistance profile [43,45]. In early preclinical studies, nucleotide prodrug GS-9131 has shown a favorable toxicity, resistance, and pharmacokinetic profile [46]. An injectable formulation of GS-9131 is currently in preclinical development. The potential of GS-9131 as an LA drug awaits the availability of more clinical results.

2.2. Long-Acting NNRTIs

2.2.1. Dapivirine

Dapivirine (DPV), a diaryl pyrimidine derivative, is a second-generation NNRTI (Figure 2). It is closely related to approved NNRTIs etravirine (ETR) and RPV [47]. DPV is currently under development as a topical microbicide to prevent sexual transmission of HIV in the form of a vaginal ring. The safety and pharmacokinetic profile results of the first clinical trial in a cohort of 16 women who used a DPV-containing vaginal ring for 28 days supported its use as a sustained-release topical microbicide for HIV-1 prevention in women [48]. The vaginal ring contained 25 mg DPV in a platinum catalyzed silicone elastomer matrix [48]. The concentration of DPV in the vaginal fluid on Day 28 exceeded more than 3900-fold IC₉₉ in a tissue explant infection model [48]. Another clinical trial included 25 mg DPV or 100 mg maraviroc (MVC) alone or in combination. The inhibition of HIV-1 infection was assessed via ex vivo challenge of cervical tissue samples. The results showed that only DPV had concentration-dependent inhibition of HIV in cervical tissues [49]. The results of the phase III clinical trial ASPIRE (Antiretroviral Strategy to Promote Improvement and Reduce Drug Exposure) showed that the incidence of HIV-1 infection among women who used a silicone elastomer vaginal matrix ring containing 25 mg DPV was decreased by 27% [50]. A follow-up phase III trial

Pharmaceuticals **2019**, 12, 62 5 of 14

(MTN-020/ASPIRE) [51] showed that 19% (6/32) women who acquired HIV-1 during the previous trial experienced virological failure. Results of a number of clinical trials addressing the impact of DPV ring on adherence [52], social concerns [53,54], pharmacokinetics in lactating women [55], pregnancy [56], and ring size [57] have been reported. Several other trials are in advanced stages. The outcomes of reported clinical trials are not necessarily encouraging. Nonetheless, the results of continuing trials could possibly pave the way for the use of the DPV ring as a microbicide.

2.2.2. Rilpivirine

Rilpivirine (RPV) (Figure 2) alone or in combination with cabotegravir (CAB) is one of the most studied antivirals as potential LA agents. RPV is an NNRTI that has been approved in first-line HIV therapy for patients with a viral load of less than 100,000 copies/mL. Phase III clinical trials showed non-inferiority of RPV-containing combination with respect to efavirenz (EFV)-containing therapy [58–60]. There are three co-formulations of RPV (TDF/FTC/RPV, TAF/FTC/RPV, and dolutegravir/RPV) available for oral use.

Encouraging pharmacokinetic properties of RPV LA formulation (in surfactant poloxamer 338) were reported in preclinical and phase I clinical trials [60–64]. These studies prompted phase II clinical trials HPTN (HIV prevention and trial network) 076 [65] and LATTE2 (Long-Acting antireTroviral Treatment Enabling 2) [66]. HPTN 076 was a double-blind, 2:1 randomized trial that was intended to evaluate the safety of 1200 mg of RPV LA compared to placebo [65] in 136 sexually-active and low-risk HIV-uninfected women from Cape Town, South Africa, Harare, Zimbabwe, Newark, New Jersey, and the Bronx, New York. RPV LA was administered in two gluteal, intramuscular injections every eight weeks for a 48-week period, followed by 28-day self-administered 25 mg of daily oral RPV. After withdrawals and discontinuations, 64 women received RPV LA, and 34 received placebo [65]. The results of HPTN 076 showed that RPV LA, 1200 mg intramuscular (IM) every eight weeks was tolerable, safe, and acceptable. RPV plasma concentration at Week 76 was above the protein-adjusted 90% inhibitory concentration (PA-IC₉₀) of RPV LA (12.5 ng/mL) in more than 92% of participants [65]. The primary goal of the HPTN 076 clinical trial was the assessment of RPV LA as PrEP.

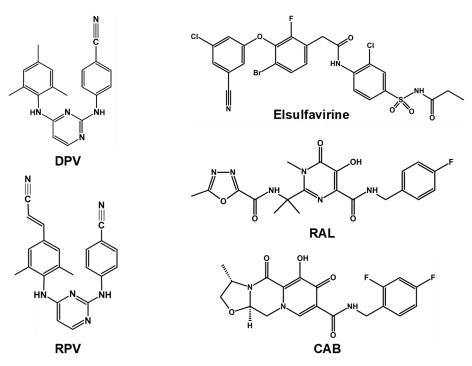


Figure 2. Non-nucleoside reverse transcriptase inhibitors (NNRTIs) and integrase strand transfer inhibitors (INSTIs) in clinical trials and preclinical development as LA antivirals.

Pharmaceuticals **2019**, 12, 62 6 of 14

In the phase IIb LATTE trial, a combination of 25 mg of RPV and selected doses of CAB were used once-daily oral as maintenance therapy for 72 weeks, after an induction period of 24 weeks by CAB with two NRTIs or efavirenz (EFV) plus two NRTIs [67]. The results showed that (i) CAB with 2 NRTIs had potent antiviral activity in the induction phase, and (ii) CAB plus RPV maintenance therapy provided antiviral activity similar to EFV plus two NRTIs until the end of Week 96 [67]. The results of this study demonstrated the potential of RPV LA, which prompted the LATTE2 clinical trial [66]. The LATTE2 trial contained CAB LA and RPV. Since CAB inhibits HIV-1 integrase, a discussion of CAB LA and its potential as an LA agent is discussed in the following section.

2.2.3. Elsulfavirine

Elsulfavirine (VM-1500) is an NNRTI and prodrug of active compound VM-1500A [68] (Figure 2). It received global approval in Russia for HIV-1 treatment in June 2017. Elsulfavirine has in vitro antiviral efficacy in the nM range ($EC_{50} = 1.2 \text{ nM}$), displays antiviral activity against a broad range of NNRTI resistance viruses, and does not have cross-resistance to other NNRTIs [69,70]. Elsulfavirine has shown safety and efficacy comparable to efavirenz (EFV) when combined with TDF and FTC [68]. In a preclinical study, 10 mg/kg injectable elsulfavirine or VM-1500A in beagle dogs showed that VM1500A plasma levels were above 50 ng/mL for at least four weeks, providing a proof-of-concept for its development as an LA antiviral [68].

3. LA Antivirals Targeting HIV-1 IN

Four INSTIs (raltegravir (RAL), elvitegravir (EVG), dolutegravir (DTG), and bictegravir (BIC)) have been approved for HIV treatment. One of these, RAL and another INSTI cabotegravir (CAB) (yet to be approved) (Figure 2), are at different stages of clinical trials for assessment of their feasibility as LA drugs. As the name implies, integrase strand transfer inhibitors (INSTIs) inhibit HIV-1 IN, a 32-kDa protein that integrates genomic DNA into the host genome. HIV-1 IN functions as an oligomer, which most likely forms upon DNA binding [71,72]. HIV-1 IN has two activities: a 3'-end processing (3'EP) and a strand-transfer (ST) activity. Both activities are conducted by the same active site. Hence, INSTIs, in spite of their nomenclature, also inhibit 3'EP, albeit with low efficacy [73].

3.1. Cabotegravir

CAB is an investigational second-generation INSTI, which is currently in advanced clinical trials for both treatment and prevention. CAB has a higher resistance barrier than first-generation INSTIs (RAL and EVG), but lower than the other two second-generation INSTIs (DTG and BIC) [74]. A phase IIa clinical trial (HPTN 077) [75] enrolled 200 low-risk healthy individuals in two cohorts. Cohort 1 (n = 110) received 800 mg CAB LA intramuscularly (n = 82) or placebo (n = 28) every 12 weeks after an initial four weeks receiving 30 mg of CAB in mannitol, polysorbate 20, polyethylene glycol 3350, and water once daily. Cohort 2 (n = 90) received 600 mg CAB LA intramuscularly (n = 69) or placebo (n = 20) every eight weeks after receiving 30 mg of CAB for four weeks, once daily. The results of this study revealed that (i) CAB LA was well-tolerated, and (ii) CAB LA 600 mg every eight weeks met pharmacokinetic targets for study participants [75].

In the phase IIb LATTE2 clinical trial, 256 HIV-infected treatment-naive patients initially received oral cabotegravir 30 mg plus abacavir (ABC) (600 mg) and lamivudine (3TC) 300 mg once daily for 20 weeks. After the 20-week period, the patients with viral suppression (plasma HIV-1 RNA <50 copies/mL) were randomized in the ratio of 2:2:1 to receive intramuscular CAB LA (400 mg) plus RPV (600 mg) at four-week or eight-week intervals (CAB LA 600 mg plus RPV 900 mg) or the continuation of CAB plus ABC/3TC. The study concluded that injectable combination of CAB LA and RPV every four or eight weeks was as effective as daily CAB/ABC/3TC oral therapy. At Week 96, 84–94% of patients had HIV RNA suppressed to <50 copies/mL. The injectable two-drug combination (CAB LA and RPV) was tolerable and safe [66]. Currently, three Phase III clinical trials are ongoing. These are (i) FLAIR (First Long-Acting Injectable Regimen) (NCT02938520), ATLAS (Antiretroviral Therapy as

Pharmaceuticals **2019**, 12, 62 7 of 14

Long Acting Suppression) (NCT02951052), and ATLAS-2M (NCT03299049). Very recently, the results from ATLAS and FLAIR have been presented. In the ATLAS study, once monthly CAB LA + RPV LA was found noninferior to continued three-drug oral cART at Week 48 and generally well tolerated with infrequent virological failures [76]. In the FLAIR study, monthly injections of CAB+RPV were noninferior to DTG/ABC/3TC at Week 48 and generally well tolerated with few virological failures [77]. Of note is that in a few HIV-1 subtype A1 (HIV-1A1) strains derived from Russian patients failing therapy, primary INSTI mutations had developed.

3.2. Raltegravir

RAL is a first-generation INSTI. It has been recommended by the European AIDS Clinical Society, the United States Department of Health and Human Services, and the International Antiviral Society, USA panel [2,3], as part of PrEP following HIV exposure. A long-acting preparation of RAL (RAL LA) in 5% polyethylene glycol 3350, 0.2% polysorbate 80, and 5% mannitol in water was administered subcutaneously to humanized BLT (bone marrow-liver-thymus) mice and rhesus macaques in a preclinical study [78]. The results showed favorable pharmacokinetic properties in rhesus macaques and potent antiretroviral activity in infected humanized BLT mice together with long-term protection from repeated vaginal HIV challenges in uninfected BLT mice [78].

4. Challenges of Subtype-Specific Polymorphisms and Pre-Existing Resistance Mutations

Subtype-specific polymorphisms and pre-existing resistance mutations can influence the efficacy of antiretrovirals [14,71,79–81]. For example, polymorphism E138A in HIV-1 RT is more common in subtype C (HIV-1C) (6–8%) than HIV-1B (0–2.3%) [82]. Mutation E138A reduces the susceptibility of RPV to varying degrees [82–84], leaving the possibility that RPV LA formulation in HIV-1C patients may not yield the desired outcome.

A detailed phylogenetic analyses showed two distinct genetic clusters in *pol*, which were also maintained in *gag*, *int/vif*, and *env* [85]. These two clusters were linked to either C181 or Y181 in RT, suggesting that C181 group O strains are naturally resistant to NNRTIs.

The resistance profile of CAB is still emerging. However, some reports have documented CAB resistance mutations. In a study conducted with SIVmac251-infected rhesus macaques, mutations I31L, Q91R, E92Q/G/M, T97A/I, G106S, G118R, H156G/R, and V172L and a duplication of five residues at position 232 have been implicated in CAB resistance [86,87]. Except E92Q/G/M and G118R, all mutations are unique to CAB resistance [88]. A recent in vitro resistance selection study showed the emergence of H51N, L74M/I, Q146L, Q148R/K S153Y, S147G, and R263K mutation under CAB pressure [74]. Polymorphisms I31L and L74I/M are present to a varying extent in different HIV-1 subtypes [89]. For example, polymorphism L74I has been reported in more than 93% in HIV-1A1 strains from Russia and countries of the former Soviet Union [90–92]. Hence, subtype-specific polymorphisms in HIV-1 IN can affect the outcome of the CAB LA formulation.

The emergence of NRTI resistance mutations in different subtypes can potentially reduce the long-acting formulation of certain NRTIs. For example, the K65R mutation is associated with both TAF and TDF resistance. Reports have shown that the K65R mutation is selected faster in HIV-1C than in HIV-1B and HIV-1A [79,82]. Globally, HIV-1C is the most prevalent subtype. A high prevalence of K65R has been reported in HIV-1C [93–95]. Despite a low prevalence of K65R transmitted resistance mutation [96], a recent comprehensive study suggested a substantial potential for onward transmission to uninfected individuals [97]. Hence, these reports emphasize drug resistance surveillance in untreated individuals for an effective TAF LA formulation. In this context, the EFdA LA formulations may provide better results, as K65R viruses are hypersusceptible to EFdA [19].

5. Clinical Challenges with LA Anti-HIV Compounds

Although the preclinical and clinical data presented so far are promising, some clinical issues remain to be addressed. These issues include side effects, drug-drug interactions, and pregnancy,

Pharmaceuticals **2019**, 12, 62 8 of 14

and the pharmacokinetics with long-lasting drug concentrations is not only a prerequisite for the promising clinical benefits, but can also lead to the development of drug resistance. The dosing interval must be selected based on the trough of the compound at the end of the injection interval to ensure that it remains above the concentration required to inhibit HIV-1 efficiently in order to avoid selective pressure for the development of drug resistance. Furthermore, even if the injection interval can be defined so that suboptimal drug concentrations are avoided, there is a potential for emergence of viral resistance as drug concentrations decline during protracted periods of sub-therapeutic exposure after ART discontinuation. Thus, even if LA agents could increase adherence in many patients, an advanced risk for resistance development still persists if patients are non-adherent to the schedule of optimized dosing intervals.

6. Conclusions

In conclusion, here, we presented the status of potential LA antivirals that target HIV-1 RT and HIV-1 IN. So far, the results from clinical trials appear encouraging, but more data are required before LA antivirals can be commonplace. Nonetheless, the future of the LA formulations appears bright. Once the LA combinations are approved, improved adherence to and convenience of cART are expected to be achieved.

Author Contributions: K.S. wrote the manuscript. K.S., S.G.S., and A.S. did the literature survey and edited the manuscript.

Funding: This research was funded by the National Institute of Health R01 Grants GM118012 and AI076119. The study was also supported by the Swedish Research Council (2016-01675, A.S.) and the Stockholm County Council (ALF 20160074, A.S.). In addition, K.S. acknowledges support from the Bond Life Sciences Center grant (DU108).

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Teeraananchai, S.; Kerr, S.J.; Amin, J.; Ruxrungtham, K.; Law, M.G. Life expectancy of HIV-positive people after starting combination antiretroviral therapy: A meta-analysis. *HIV Med.* **2017**, *18*, 256–266. [CrossRef] [PubMed]
- 2. Gulick, R.M.; Flexner, C. Long-acting HIV drugs for treatment and prevention. *Annu. Rev. Med.* **2019**, 70, 137–150. [CrossRef]
- 3. Gunthard, H.F.; Saag, M.S.; Benson, C.A.; del Rio, C.; Eron, J.J.; Gallant, J.E.; Hoy, J.F.; Mugavero, M.J.; Sax, P.E.; Thompson, M.A.; et al. Antiretroviral drugs for treatment and prevention of HIV infection in adults: 2016 recommendations of the international antiviral society-USA panel. *JAMA* **2016**, *316*, 191–210. [CrossRef] [PubMed]
- 4. Saag, M.S.; Benson, C.A.; Gandhi, R.T.; Hoy, J.F.; Landovitz, R.J.; Mugavero, M.J.; Sax, P.E.; Smith, D.M.; Thompson, M.A.; Buchbinder, S.P.; et al. Antiretroviral drugs for treatment and prevention of HIV infection in adults: 2018 recommendations of the international antiviral society-USA panel. *JAMA* 2018, 320, 379–396. [CrossRef] [PubMed]
- 5. Preexposure Prophylaxis for the Prevention of HIV Infection in the United States—2017 Update: A Clinical Practice Guideline. Available online: https://www.cdc.gov/hiv/pdf/risk/prep/cdc-hiv-prep-guidelines-2017. pdf (accessed on 12 March 2019).
- 6. Molina, J.M.; Capitant, C.; Spire, B.; Pialoux, G.; Cotte, L.; Charreau, I.; Tremblay, C.; Le Gall, J.M.; Cua, E.; Pasquet, A.; et al. On-demand preexposure prophylaxis in men at high risk for HIV-1 infection. *N. Engl. J. Med.* 2015, 373, 2237–2246. [CrossRef]
- 7. McCormack, S.; Dunn, D.T.; Desai, M.; Dolling, D.I.; Gafos, M.; Gilson, R.; Sullivan, A.K.; Clarke, A.; Reeves, I.; Schembri, G.; et al. Pre-exposure prophylaxis to prevent the acquisition of HIV-1 infection (proud): Effectiveness results from the pilot phase of a pragmatic open-label randomised trial. *Lancet* **2016**, *387*, 53–60. [CrossRef]
- 8. Winner, B.; Peipert, J.F.; Zhao, Q.; Buckel, C.; Madden, T.; Allsworth, J.E.; Secura, G.M. Effectiveness of long-acting reversible contraception. *N. Engl. J. Med.* **2012**, *366*, 1998–2007. [CrossRef]

9. Rattan, J.; Noznesky, E.; Curry, D.W.; Galavotti, C.; Hwang, S.; Rodriguez, M. Rapid contraceptive uptake and changing method mix with high use of long-acting reversible contraceptives in crisis-affected populations in chad and the democratic republic of the congo. *Glob. Health Sci. Pract.* **2016**, *4* (Suppl. 2), S5–S20. [CrossRef]

- 10. Fok, W.K.; Blumenthal, P.D. HIV and contraception. *Curr. Opin. Obstet. Gynecol.* **2017**, 29, 419–426. [CrossRef] [PubMed]
- 11. Sarafianos, S.G.; Marchand, B.; Das, K.; Himmel, D.M.; Parniak, M.A.; Hughes, S.H.; Arnold, E. Structure and function of HIV-1 reverse transcriptase: Molecular mechanisms of polymerization and inhibition. *J. Mol. Biol.* **2009**, *385*, 693–713. [CrossRef] [PubMed]
- 12. Singh, K.; Marchand, B.; Kirby, K.A.; Michailidis, E.; Sarafianos, S.G. Structural aspects of drug resistance and inhibition of HIV-1 reverse transcriptase. *Viruses* **2010**, 2, 606–638. [CrossRef] [PubMed]
- 13. Menendez-Arias, L. Molecular basis of human immunodeficiency virus drug resistance: An update. *Antivir. Res.* **2010**, *85*, 210–231. [CrossRef] [PubMed]
- 14. Singh, K.; Flores, J.A.; Kirby, K.A.; Neogi, U.; Sonnerborg, A.; Hachiya, A.; Das, K.; Arnold, E.; McArthur, C.; Parniak, M.; et al. Drug resistance in non-b subtype HIV-1: Impact of HIV-1 reverse transcriptase inhibitors. *Viruses* **2014**, *6*, 3535–3562. [CrossRef] [PubMed]
- 15. Das, K.; Martinez, S.E.; Bauman, J.D.; Arnold, E. HIV-1 reverse transcriptase complex with DNA and nevirapine reveals non-nucleoside inhibition mechanism. *Nat. Struct. Mol. Biol.* **2012**, *19*, 253–259. [CrossRef]
- 16. Kawamoto, A.; Kodama, E.; Sarafianos, S.G.; Sakagami, Y.; Kohgo, S.; Kitano, K.; Ashida, N.; Iwai, Y.; Hayakawa, H.; Nakata, H.; et al. 2'-deoxy-4'-c-ethynyl-2-halo-adenosines active against drug-resistant human immunodeficiency virus type 1 variants. *Int. J. Biochem. Cell Biol.* 2008, 40, 2410–2420. [CrossRef] [PubMed]
- 17. Michailidis, E.; Marchand, B.; Kodama, E.N.; Singh, K.; Matsuoka, M.; Kirby, K.A.; Ryan, E.M.; Sawani, A.M.; Nagy, E.; Ashida, N.; et al. Mechanism of inhibition of HIV-1 reverse transcriptase by 4'-ethynyl-2-fluoro-2'-deoxyadenosine triphosphate, a translocation-defective reverse transcriptase inhibitor. *J. Biol. Chem.* 2009, 284, 35681–35691. [CrossRef]
- 18. Michailidis, E.; Huber, A.D.; Ryan, E.M.; Ong, Y.T.; Leslie, M.D.; Matzek, K.B.; Singh, K.; Marchand, B.; Hagedorn, A.N.; Kirby, K.A.; et al. 4'-ethynyl-2-fluoro-2'-deoxyadenosine (efda) inhibits HIV-1 reverse transcriptase with multiple mechanisms. *J. Biol. Chem.* 2014, 289, 24533–24548. [CrossRef] [PubMed]
- 19. Michailidis, E.; Ryan, E.M.; Hachiya, A.; Kirby, K.A.; Marchand, B.; Leslie, M.D.; Huber, A.D.; Ong, Y.T.; Jackson, J.C.; Singh, K.; et al. Hypersusceptibility mechanism of tenofovir-resistant HIV to efda. *Retrovirology* **2013**, *10*, 65. [CrossRef] [PubMed]
- 20. Salie, Z.L.; Kirby, K.A.; Michailidis, E.; Marchand, B.; Singh, K.; Rohan, L.C.; Kodama, E.N.; Mitsuya, H.; Parniak, M.A.; Sarafianos, S.G. Structural basis of HIV inhibition by translocation-defective rt inhibitor 4'-ethynyl-2-fluoro-2'-deoxyadenosine (efda). *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 9274–9279. [CrossRef]
- 21. Hachiya, A.; Reeve, A.B.; Marchand, B.; Michailidis, E.; Ong, Y.T.; Kirby, K.A.; Leslie, M.D.; Oka, S.; Kodama, E.N.; Rohan, L.C.; et al. Evaluation of combinations of 4'-ethynyl-2-fluoro-2'-deoxyadenosine with clinically used antiretroviral drugs. *Antimicrob. Agents Chemother.* 2013, 57, 4554–4558. [CrossRef]
- 22. Markowitz, M.; Sarafianos, S.G. 4'-ethynyl-2-fluoro-2'-deoxyadenosine, mk-8591: A novel HIV-1 reverse transcriptase translocation inhibitor. *Curr. Opin. HIV AIDS* **2018**, *13*, 294–299. [CrossRef]
- 23. Takamatsu, Y.; Das, D.; Kohgo, S.; Hayashi, H.; Delino, N.S.; Sarafianos, S.G.; Mitsuya, H.; Maeda, K. The high genetic barrier of efda/mk-8591 stems from strong interactions with the active site of drug-resistant HIV-1 reverse transcriptase. *Cell Chem. Biol.* 2018, 25, 1268–1278. [CrossRef] [PubMed]
- 24. Maeda, K.; Desai, D.V.; Aoki, M.; Nakata, H.; Kodama, E.N.; Mitsuya, H. Delayed emergence of HIV-1 variants resistant to 4'-ethynyl-2-fluoro-2'-deoxyadenosine: Comparative sequential passage study with lamivudine, tenofovir, emtricitabine and bms-986001. *Antivir. Ther.* **2014**, *19*, 179–189. [CrossRef] [PubMed]
- 25. Njenda, D.T.; Aralaguppe, S.G.; Singh, K.; Rao, R.; Sonnerborg, A.; Sarafianos, S.G.; Neogi, U. Antiretroviral potency of 4'-ethnyl-2'-fluoro-2'-deoxyadenosine, tenofovir alafenamide and second-generation nnrtis across diverse HIV-1 subtypes. *J. Antimicrob. Chemother.* **2018**, *73*, 2721–2728. [CrossRef] [PubMed]
- Grobler, J.; McHale, C.; Freddo, C.; Dreyer, D.; Sun, L.; Vavrek, M.; Breidinger, S.; Fillgrove, K.; Hazuda, D.;
 Lai, M.-T. Mk-8591 concentrations at sites of HIV transmission and replication. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Seattle, WA, USA, 13–16 February 2017.

27. Barrett, S.E.; Teller, R.S.; Forster, S.P.; Li, L.; Mackey, M.A.; Skomski, D.; Yang, Z.; Fillgrove, K.L.; Doto, G.J.; Wood, S.L.; et al. Extended-duration mk-8591-eluting implant as a candidate for HIV treatment and prevention. *Antimicrob. Agents Chemother.* **2018**, *62*, e1058-18. [CrossRef] [PubMed]

- 28. Flexner, C. Antiretroviral implants for treatment and prevention of HIV infection. *Curr. Opin. HIV AIDS* **2018**, *13*, 374–380. [CrossRef] [PubMed]
- 29. Zhang, W.; Hu, M.; Shi, Y.; Gong, T.; Dezzutti, C.S.; Moncla, B.; Sarafianos, S.G.; Parniak, M.A.; Rohan, L.C. Vaginal microbicide film combinations of two reverse transcriptase inhibitors, efda and csic, for the prevention of HIV-1 sexual transmission. *Pharm. Res.* **2015**, *32*, 2960–2972. [CrossRef] [PubMed]
- 30. Zhang, W.; Parniak, M.A.; Mitsuya, H.; Sarafianos, S.G.; Graebing, P.W.; Rohan, L.C. Preformulation studies of efda, a novel nucleoside reverse transcriptase inhibitor for HIV prevention. *Drug Dev. Ind. Pharm.* **2014**, 40, 1101–1111. [CrossRef] [PubMed]
- 31. Zhang, W.; Parniak, M.A.; Sarafianos, S.G.; Cost, M.R.; Rohan, L.C. Development of a vaginal delivery film containing efda, a novel anti-HIV nucleoside reverse transcriptase inhibitor. *Int. J. Pharm.* **2014**, 461, 203–213. [CrossRef]
- 32. Murphey-Corb, M.; Rajakumar, P.; Michael, H.; Nyaundi, J.; Didier, P.J.; Reeve, A.B.; Mitsuya, H.; Sarafianos, S.G.; Parniak, M.A. Response of simian immunodeficiency virus to the novel nucleoside reverse transcriptase inhibitor 4'-ethynyl-2-fluoro-2'-deoxyadenosine in vitro and in vivo. *Antimicrob. Agents Chemother.* 2012, 56, 4707–4712. [CrossRef]
- 33. Stoddart, C.A.; Galkina, S.A.; Joshi, P.; Kosikova, G.; Moreno, M.E.; Rivera, J.M.; Sloan, B.; Reeve, A.B.; Sarafianos, S.G.; Murphey-Corb, M.; et al. Oral administration of the nucleoside efda (4'-ethynyl-2-fluoro-2'-deoxyadenosine) provides rapid suppression of HIV viremia in humanized mice and favorable pharmacokinetic properties in mice and the rhesus macaque. *Antimicrob. Agents Chemother.* 2015, 59, 4190–4198. [CrossRef] [PubMed]
- 34. Friedman, E.; Schuermann, D.; Rudd, D.J.; Fox-Bosetti, S.; Zhang, S.; Robberechts, M.; Hueser, H.; Hazuda, D.J.; Iwamoto, M.; Grobler, J. A single monotherapy dose of mk-8591, a novel nrti, suppresses HIV for 10 days. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Boston, MA, USA, 22–25 February 2016.
- 35. Grobler, J.; Friedman, E.; Barrett, S.E.; Wood, S.L.; Ankrom, W.; Fillgrove, K.L.; Lai, M.-T.; Gindy, M.; Iwamoto, M.; Hazuda, D.J. Long-acting oral and parenteral dosing of mk-8591 for HIV treatment or prophylaxis. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Boston, MA, USA, 22–25 February 2016.
- 36. Ruane, P.J.; DeJesus, E.; Berger, D.; Markowitz, M.; Bredeek, U.F.; Callebaut, C.; Zhong, L.; Ramanathan, S.; Rhee, M.S.; Fordyce, M.W.; et al. Antiviral activity, safety, and pharmacokinetics/pharmacodynamics of tenofovir alafenamide as 10-day monotherapy in HIV-1-positive adults. *J. Acquir. Immune Defic. Syndr.* **2013**, 63, 449–455. [CrossRef]
- 37. Margot, N.A.; Johnson, A.; Miller, M.D.; Callebaut, C. Characterization of HIV-1 resistance to tenofovir alafenamide in vitro. *Antimicrob. Agents Chemother.* **2015**, *59*, 5917–5924. [CrossRef]
- 38. Sax, P.E.; Wohl, D.; Yin, M.T.; Post, F.; DeJesus, E.; Saag, M.; Pozniak, A.; Thompson, M.; Podzamczer, D.; Molina, J.M.; et al. Tenofovir alafenamide versus tenofovir disoproxil fumarate, coformulated with elvitegravir, cobicistat, and emtricitabine, for initial treatment of HIV-1 infection: Two randomised, double-blind, phase 3, non-inferiority trials. *Lancet* 2015, 385, 2606–2615. [CrossRef]
- 39. Gunawardana, M.; Remedios-Chan, M.; Miller, C.S.; Fanter, R.; Yang, F.; Marzinke, M.A.; Hendrix, C.W.; Beliveau, M.; Moss, J.A.; Smith, T.J.; et al. Pharmacokinetics of long-acting tenofovir alafenamide (gs-7340) subdermal implant for HIV prophylaxis. *Antimicrob. Agents Chemother.* **2015**, *59*, 3913–3919. [CrossRef] [PubMed]
- 40. Schlesinger, E.; Johengen, D.; Luecke, E.; Rothrock, G.; McGowan, I.; van der Straten, A.; Desai, T. A tunable, biodegradable, thin-film polymer device as a long-acting implant delivering tenofovir alafenamide fumarate for HIV pre-exposure prophylaxis. *Pharm. Res.* **2016**, *33*, 1649–1656. [CrossRef] [PubMed]
- 41. O'Neill, A.M. Perceptual rotations on children's rorschachs. J. Clin. Psychol. 1989, 45, 809–813. [CrossRef]
- 42. Smith, J.M.; Moss, J.A.; Srinivasan, P.; Butkyavichene, I.; Gunawardana, M.; Fanter, R.; Miller, C.S.; Sanchez, D.; Yang, F.; Ellis, S.; et al. Novel multipurpose pod-intravaginal ring for the prevention of HIV, hsv, and unintended pregnancy: Pharmacokinetic evaluation in a macaque model. *PLoS ONE* **2017**, *12*, e0185946. [CrossRef] [PubMed]

43. Cihlar, T.; Ray, A.S.; Boojamra, C.G.; Zhang, L.; Hui, H.; Laflamme, G.; Vela, J.E.; Grant, D.; Chen, J.; Myrick, F.; et al. Design and profiling of gs-9148, a novel nucleotide analog active against nucleoside-resistant variants of human immunodeficiency virus type 1, and its orally bioavailable phosphonoamidate prodrug, gs-9131. *Antimicrob. Agents Chemother.* **2008**, *52*, 655–665. [CrossRef]

- 44. Ray, A.S.; Vela, J.E.; Boojamra, C.G.; Zhang, L.; Hui, H.; Callebaut, C.; Stray, K.; Lin, K.Y.; Gao, Y.; Mackman, R.L.; et al. Intracellular metabolism of the nucleotide prodrug gs-9131, a potent anti-human immunodeficiency virus agent. *Antimicrob. Agents Chemother.* 2008, 52, 648–654. [CrossRef] [PubMed]
- 45. White, K.L.; Margot, N.; Stray, K.; Yu, H.; Stepan, G.; Boojamra, C.; Mackman, R.; Ray, A.; Miller, M.D.; Cilhar, T. Gs-9131 is a novel nrti with activity against nrti-resistant HIV-1. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Seattle, WA, USA, 13–16 February 2017.
- 46. Mackman, R.L.; Ray, A.S.; Hui, H.C.; Zhang, L.; Birkus, G.; Boojamra, C.G.; Desai, M.C.; Douglas, J.L.; Gao, Y.; Grant, D.; et al. Discovery of gs-9131: Design, synthesis and optimization of amidate prodrugs of the novel nucleoside phosphonate HIV reverse transcriptase (rt) inhibitor gs-9148. *Bioorg. Med. Chem.* 2010, 18, 3606–3617. [CrossRef]
- 47. Das, K.; Clark, A.D., Jr.; Lewi, P.J.; Heeres, J.; De Jonge, M.R.; Koymans, L.M.; Vinkers, H.M.; Daeyaert, F.; Ludovici, D.W.; Kukla, M.J.; et al. Roles of conformational and positional adaptability in structure-based design of tmc125-r165335 (etravirine) and related non-nucleoside reverse transcriptase inhibitors that are highly potent and effective against wild-type and drug-resistant HIV-1 variants. *J. Med. Chem.* **2004**, *47*, 2550–2560.
- 48. Nel, A.; Haazen, W.; Nuttall, J.; Romano, J.; Rosenberg, Z.; van Niekerk, N. A safety and pharmacokinetic trial assessing delivery of dapivirine from a vaginal ring in healthy women. *AIDS* **2014**, *28*, 1479–1487. [CrossRef]
- 49. Chen, B.A.; Panther, L.; Marzinke, M.A.; Hendrix, C.W.; Hoesley, C.J.; van der Straten, A.; Husnik, M.J.; Soto-Torres, L.; Nel, A.; Johnson, S.; et al. Phase 1 safety, pharmacokinetics, and pharmacodynamics of dapivirine and maraviroc vaginal rings: A double-blind randomized trial. *J. Acquir. Immune Defic. Syndr.* 2015, 70, 242–249. [CrossRef] [PubMed]
- 50. Baeten, J.M.; Palanee-Phillips, T.; Brown, E.R.; Schwartz, K.; Soto-Torres, L.E.; Govender, V.; Mgodi, N.M.; Matovu Kiweewa, F.; Nair, G.; Mhlanga, F.; et al. Use of a vaginal ring containing dapivirine for HIV-1 prevention in women. *N. Engl. J. Med.* **2016**, *375*, 2121–2132. [CrossRef]
- 51. Riddler, S.A.; Balkus, J.E.; Parikh, U.M.; Mellors, J.W.; Akello, C.; Dadabhai, S.; Mhlanga, F.; Ramjee, G.; Mayo, A.J.; Livant, E.; et al. Clinical and virologic outcomes following initiation of antiretroviral therapy among seroconverters in the mtn-020/aspire phase iii trial of the dapivirine vaginal ring. *Clin. Infect. Dis.* 2018. [CrossRef] [PubMed]
- 52. Mensch, B.S.; Richardson, B.A.; Husnik, M.; Brown, E.R.; Kiweewa, F.M.; Mayo, A.J.; Baeten, J.M.; Palanee-Phillips, T.; van der Straten, A.; MTN-020/ASPIRE Study Team. Vaginal ring use in a phase 3 microbicide trial: A comparison of objective measures and self-reports of non-adherence in aspire. *AIDS Behav.* 2019, 23, 504–512. [CrossRef]
- 53. Chitukuta, M.; Duby, Z.; Katz, A.; Nakyanzi, T.; Reddy, K.; Palanee-Phillips, T.; Tembo, T.; Etima, J.; Musara, P.; Mgodi, N.M.; et al. Negative rumours about a vaginal ring for HIV-1 prevention in sub-saharan africa. *Cult. Health Sex.* **2019**, 1–16. [CrossRef] [PubMed]
- 54. Palanee-Phillips, T.; Roberts, S.T.; Reddy, K.; Govender, V.; Naidoo, L.; Siva, S.; Gafoor, Z.; Pather, A.; Matovu, F.; Hlahla, K.; et al. Impact of partner-related social harms on women's adherence to the dapivirine vaginal ring during a phase iii trial. *J. Acquir. Immune Defic. Syndr.* 2018, 79, 580–589. [CrossRef] [PubMed]
- 55. Noguchi, L.M.; Hoesley, C.; Kelly, C.; Scheckter, R.; Bunge, K.; Nel, A.; Marzinke, M.A.; Hendrix, C.W.; Dezzutti, C.S.; Hillier, S.L.; et al. Pharmacokinetics of dapivirine transfer into blood plasma, breast milk, and cervicovaginal fluid of lactating women using the dapivirine vaginal ring. *Antimicrob. Agents Chemother.* **2019**, *63*, e01930-18. [CrossRef] [PubMed]
- 56. Makanani, B.; Balkus, J.E.; Jiao, Y.; Noguchi, L.M.; Palanee-Phillips, T.; Mbilizi, Y.; Moodley, J.; Kintu, K.; Reddy, K.; Kabwigu, S.; et al. Pregnancy and infant outcomes among women using the dapivirine vaginal ring in early pregnancy. *J. Acquir. Immune Defic. Syndr.* **2018**, *79*, 566–572. [CrossRef]
- 57. Murphy, D.J.; Desjardins, D.; Boyd, P.; Dereuddre-Bosquet, N.; Stimmer, L.; Caldwell, A.; Le Grand, R.; Kelly, C.; van Roey, J.; Malcolm, R.K. Impact of ring size and drug loading on the pharmacokinetics of a combination dapivirine-darunavir vaginal ring in cynomolgus macaques. *Int. J. Pharm.* **2018**, *550*, 300–308. [CrossRef]

58. Cohen, C.; Wohl, D.; Arribas, J.; Henry, K.; Van Lunzen, J.; Bloch, M.; Towner, W.; Wilkins, E.; Wang, H.; White, K.; et al. Star study: Single tablet regimen emtricitabine/rilpivirine/tenofovir df is non-inferior to efavirenz/emtricitabine/tenofovir df in art-naïve adults. *J. Int. AIDS Soc.* **2012**, *15* (Suppl. 4), 18221. [CrossRef]

- 59. Cohen, C.J.; Andrade-Villanueva, J.; Clotet, B.; Fourie, J.; Johnson, M.A.; Ruxrungtham, K.; Wu, H.; Zorrilla, C.; Crauwels, H.; Rimsky, L.T.; et al. Rilpivirine versus efavirenz with two background nucleoside or nucleotide reverse transcriptase inhibitors in treatment-naive adults infected with HIV-1 (thrive): A phase 3, randomised, non-inferiority trial. *Lancet* 2011, 378, 229–237. [CrossRef]
- 60. Jackson, A.G.; Else, L.J.; Mesquita, P.M.; Egan, D.; Back, D.J.; Karolia, Z.; Ringner-Nackter, L.; Higgs, C.J.; Herold, B.C.; Gazzard, B.G.; et al. A compartmental pharmacokinetic evaluation of long-acting rilpivirine in HIV-negative volunteers for pre-exposure prophylaxis. *Clin. Pharmacol. Ther.* **2014**, *96*, 314–323. [CrossRef]
- 61. McGowan, I.; Dezzutti, C.S.; Siegel, A.; Engstrom, J.; Nikiforov, A.; Duffill, K.; Shetler, C.; Richardson-Harman, N.; Abebe, K.; Back, D.; et al. Long-acting rilpivirine as potential pre-exposure prophylaxis for HIV-1 prevention (the mwri-01 study): An open-label, phase 1, compartmental, pharmacokinetic and pharmacodynamic assessment. *Lancet HIV* **2016**, *3*, e569–e578. [CrossRef]
- 62. Spreen, W.; Williams, P.; Margolis, D.; Ford, S.L.; Crauwels, H.; Lou, Y.; Gould, E.; Stevens, M.; Piscitelli, S. Pharmacokinetics, safety, and tolerability with repeat doses of gsk1265744 and rilpivirine (tmc278) long-acting nanosuspensions in healthy adults. *J. Acquir. Immune Defic. Syndr.* 2014, 67, 487–492. [CrossRef] [PubMed]
- 63. Verloes, R.; Deleu, S.; Niemeijer, N.; Crauwels, H.; Meyvisch, P.; Williams, P. Safety, tolerability and pharmacokinetics of rilpivirine following administration of a long-acting formulation in healthy volunteers. HIV Med. 2015, 16, 477–484. [CrossRef] [PubMed]
- 64. Williams, P.E.; Crauwels, H.M.; Basstanie, E.D. Formulation and pharmacology of long-acting rilpivirine. *Curr. Opin. HIV AIDS* **2015**, *10*, 233–238. [CrossRef] [PubMed]
- 65. Bekker, L.-G.; Li, S.S.; Tolly, B.; Marzinke, M.A.; Mgodi, N.; Justman, J.E.; Swaminathan, S.; Adeyeye, A.; Farrior, J.H.; Sista, N. Hptn076:Tmc278 ls safe, tolarable, and acceptable for HIV preexposure prophylaxix. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Seattle, WA, USA, 13–16 February 2017.
- 66. Margolis, D.A.; Gonzalez-Garcia, J.; Stellbrink, H.J.; Eron, J.J.; Yazdanpanah, Y.; Podzamczer, D.; Lutz, T.; Angel, J.B.; Richmond, G.J.; Clotet, B.; et al. Long-acting intramuscular cabotegravir and rilpivirine in adults with HIV-1 infection (latte-2): 96-week results of a randomised, open-label, phase 2b, non-inferiority trial. *Lancet* 2017, 390, 1499–1510. [CrossRef]
- 67. Margolis, D.A.; Brinson, C.C.; Smith, G.H.R.; de Vente, J.; Hagins, D.P.; Eron, J.J.; Griffith, S.K.; Clair, M.H.S.; Stevens, M.C.; Williams, P.E.; et al. Cabotegravir plus rilpivirine, once a day, after induction with cabotegravir plus nucleoside reverse transcriptase inhibitors in antiretroviral-naive adults with HIV-1 infection (latte): A randomised, phase 2b, dose-ranging trial. *Lancet Infect. Dis.* **2015**, *15*, 1145–1155. [CrossRef]
- 68. Al-Salama, Z.T. Elsulfavirine: First global approval. Drugs 2017, 77, 1811–1816. [CrossRef] [PubMed]
- 69. Namasivayam, V.; Vanangamudi, M.; Kramer, V.G.; Kurup, S.; Zhan, P.; Liu, X.; Kongsted, J.; Byrareddy, S.N. The journey of HIV-1 non-nucleoside reverse transcriptase inhibitors (nnrtis) from lab to clinic. *J. Med. Chem.* **2018**. [CrossRef]
- 70. Rai, M.A.; Pannek, S.; Fichtenbaum, C.J. Emerging reverse transcriptase inhibitors for HIV-1 infection. *Expert Opin Emerg Drugs* **2018**, 23, 149–157. [CrossRef] [PubMed]
- 71. Hill, K.J.; Rogers, L.C.; Njenda, D.T.; Burke, D.H.; Sarafianos, S.G.; Sonnerborg, A.; Neogi, U.; Singh, K. Strain-specific effect on biphasic DNA binding by HIV-1 integrase. *AIDS* **2019**, *33*, 588–592. [CrossRef] [PubMed]
- 72. Passos, D.O.; Li, M.; Yang, R.; Rebensburg, S.V.; Ghirlando, R.; Jeon, Y.; Shkriabai, N.; Kvaratskhelia, M.; Craigie, R.; Lyumkis, D. Cryo-em structures and atomic model of the HIV-1 strand transfer complex intasome. *Science* 2017, 355, 89–92. [CrossRef]
- 73. Neogi, U.; Singh, K.; Aralaguppe, S.G.; Rogers, L.C.; Njenda, D.T.; Sarafianos, S.G.; Hejdeman, B.; Sonnerborg, A. Ex-vivo antiretroviral potency of newer integrase strand transfer inhibitors cabotegravir and bictegravir in HIV type 1 non-b subtypes. *AIDS* **2018**, *32*, 469–476.

74. Oliveira, M.; Ibanescu, R.I.; Anstett, K.; Mesplede, T.; Routy, J.P.; Robbins, M.A.; Brenner, B.G.; Montreal Primary, H.I.V.C.S.G. Selective resistance profiles emerging in patient-derived clinical isolates with cabotegravir, bictegravir, dolutegravir, and elvitegravir. *Retrovirology* **2018**, *15*, 56. [CrossRef]

- 75. Landovitz, R.J.; Li, S.; Grinsztejn, B.; Dawood, H.; Liu, A.Y.; Magnus, M.; Hosseinipour, M.C.; Panchia, R.; Cottle, L.; Chau, G.; et al. Safety, tolerability, and pharmacokinetics of long-acting injectable cabotegravir in low-risk HIV-uninfected individuals: Hptn 077, a phase 2a randomized controlled trial. *PLoS Med.* **2018**, 15, e1002690. [CrossRef]
- 76. Swindells, S.; Andrade-Villanueva, J.-F.; Gary, J.; Richmond, G.J.; Rizzardini, G.; Baumgarten, A.; Maria Del Mar Masia Del Mar, M.; Latiff, G.; Pokrovsky, V.; Mrus, J.M.; et al. Long-acting cabotegravir+rilpivirine maintenance therapy: Atlas week 48 results. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Seattle, WA, USA, 4–7 March 2019.
- 77. Orkin, C.; Arastéh, K.; Hernández-Mora, M.C.; Pokrovsky, V.; Overton, E.T.; Overton, M.-P.; Oka, S.; D'Amico, R.; Dorey, D.; Griffith, S.K.; et al. Long-acting cabotegravir + rilpivirine for HIV maintenance: Flair week 48 results. In Proceedings of the Conference on Retroviruses and Opportunistic Infections, Seattle, WA, USA, 4–7 March 2019.
- 78. Kovarova, M.; Swanson, M.D.; Sanchez, R.I.; Baker, C.E.; Steve, J.; Spagnuolo, R.A.; Howell, B.J.; Hazuda, D.J.; Garcia, J.V. A long-acting formulation of the integrase inhibitor raltegravir protects humanized blt mice from repeated high-dose vaginal HIV challenges. *J. Antimicrob. Chemother.* **2016**, *71*, 1586–1596. [CrossRef]
- 79. Brenner, B.G. Resistance and viral subtypes: How important are the differences and why do they occur? *Curr. Opin. HIV AIDS* **2007**, *2*, 94–102. [CrossRef]
- 80. Lessells, R.J.; Katzenstein, D.K.; de Oliveira, T. Are subtype differences important in HIV drug resistance? *Curr. Opin. Virol.* **2012**, 2, 636–643. [CrossRef]
- 81. Wainberg, M.A.; Brenner, B.G. The impact of HIV genetic polymorphisms and subtype differences on the occurrence of resistance to antiretroviral drugs. *Mol. Biol. Int.* **2012**, 2012, 256982. [CrossRef] [PubMed]
- 82. Sluis-Cremer, N.; Jordan, M.R.; Huber, K.; Wallis, C.L.; Bertagnolio, S.; Mellors, J.W.; Parkin, N.T.; Harrigan, P.R. E138a in HIV-1 reverse transcriptase is more common in subtype c than b: Implications for rilpivirine use in resource-limited settings. *Antivir. Res.* **2014**, *107*, 31–34. [CrossRef]
- 83. Giannini, A.; Vicenti, I.; Materazzi, A.; Boccuto, A.; Dragoni, F.; Zazzi, M.; Saladini, F. The HIV-1 reverse transcriptase e138a natural polymorphism decreases the genetic barrier to resistance to etravirine in vitro. *J. Antimicrob. Chemother.* **2019**, *74*, 607–613. [CrossRef]
- 84. Tambuyzer, L.; Nijs, S.; Daems, B.; Picchio, G.; Vingerhoets, J. Effect of mutations at position e138 in HIV-1 reverse transcriptase on phenotypic susceptibility and virologic response to etravirine. *J. Acquir. Immune Defic. Syndr.* **2011**, *58*, 18–22. [CrossRef] [PubMed]
- 85. Tebit, D.M.; Lobritz, M.; Lalonde, M.; Immonen, T.; Singh, K.; Sarafianos, S.; Herchenroder, O.; Krausslich, H.G.; Arts, E.J. Divergent evolution in reverse transcriptase (rt) of HIV-1 group o and m lineages: Impact on structure, fitness, and sensitivity to nonnucleoside rt inhibitors. *J. Virol.* **2010**, *84*, 9817–9830. [CrossRef]
- 86. Charpentier, C.; Descamps, D. Resistance to HIV integrase inhibitors: About r263k and e157q mutations. *Viruses* **2018**, *10*, 41. [CrossRef] [PubMed]
- 87. Radzio, J.; Council, O.; Cong, M.-E.; Mitchell, J.; Ellis, S.; Huang, W.; Spreen, W.; Heneine, W.; Garcia-Lerma, G. Resistance emergence in macaques administered cabotegravir la during acute infection. In Proceedings of the Conference on Retroviruses and Opportunistic Infections 2017, Seattle, WA, USA, 13–16 February 2017.
- 88. Wensing, A.M.; Calvez, V.; Gunthard, H.F.; Johnson, V.A.; Paredes, R.; Pillay, D.; Shafer, R.W.; Richman, D.D. 2017 update of the drug resistance mutations in HIV-1. *Top. Antivir. Med.* **2017**, 24, 132–133.
- 89. Rogers, L.; Obasa, A.E.; Jacobs, G.B.; Sarafianos, S.G.; Sönnerborg, A.; Neogi, U.; Singh, K. Structural implications of genotypic variations in HIV-1 integrase from diverse subtypes. *Front. Microbiol.* **2018**, *9*, 1754. [CrossRef]
- 90. Gashnikova, N.M.; Astakhova, E.M.; Gashnikova, M.P.; Bocharov, E.F.; Petrova, S.V.; Pun'ko, O.A.; Popkov, A.V.; Totmenin, A.V. HIV-1 epidemiology, genetic diversity, and primary drug resistance in the tyumen oblast, russia. *Biomed. Res. Int.* **2016**, 2016, 2496280. [CrossRef] [PubMed]
- 91. Lapovok, I.; Laga, V.; Kazennova, E.; Bobkova, M. HIV type 1 integrase natural polymorphisms in viral variants circulating in fsu countries. *Curr. HIV Res.* **2017**, *15*, 318–326. [CrossRef] [PubMed]

92. Gupta, R.K.; Chrystie, I.L.; O'Shea, S.; Mullen, J.E.; Kulasegaram, R.; Tong, C.Y. K65r and y181c are less prevalent in haart-experienced HIV-1 subtype a patients. *AIDS* **2005**, *19*, 1916–1919. [CrossRef]

- 93. Doualla-Bell, F.; Avalos, A.; Brenner, B.; Gaolathe, T.; Mine, M.; Gaseitsiwe, S.; Oliveira, M.; Moisi, D.; Ndwapi, N.; Moffat, H.; et al. High prevalence of the k65r mutation in human immunodeficiency virus type 1 subtype c isolates from infected patients in botswana treated with didanosine-based regimens. *Antimicrob. Agents Chemother.* **2006**, *50*, 4182–4185. [CrossRef] [PubMed]
- 94. Skhosana, L.; Steegen, K.; Bronze, M.; Lukhwareni, A.; Letsoalo, E.; Papathanasopoulos, M.A.; Carmona, S.C.; Stevens, W.S. High prevalence of the k65r mutation in HIV-1 subtype c infected patients failing tenofovir-based first-line regimens in south africa. *PLoS ONE* **2015**, *10*, e0118145. [CrossRef] [PubMed]
- 95. Smit, E.; White, E.; Clark, D.; Churchill, D.; Zhang, H.; Collins, S.; Pillay, D.; Sabin, C.; Nelson, M.; Winston, A.; et al. An association between k65r and HIV-1 subtype c viruses in patients treated with multiple nrtis. *J. Antimicrob. Chemother.* **2017**, 72, 2075–2082. [CrossRef] [PubMed]
- 96. Chan, P.A.; Huang, A.; Kantor, R. Low prevalence of transmitted k65r and other tenofovir resistance mutations across different HIV-1 subtypes: Implications for pre-exposure prophylaxis. *J. Int. AIDS Soc.* **2012**, *15*, 17701. [CrossRef]
- 97. TenoRes Study, G. Global epidemiology of drug resistance after failure of who recommended first-line regimens for adult HIV-1 infection: A multicentre retrospective cohort study. *Lancet Infect. Dis.* **2016**, *16*, 565–575.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).