

Electrophilic Substitution at C(7) of a Protected 7-Deaza-2'-deoxyguanosine – The 2'-Deoxyribonucleoside Parent Analogue of Queuosine [#]

Natalya Ramzaeva and Helmut Rosemeyer *

Organische Chemie I – Bioorganische Chemie, Institut für Chemie, Fachbereich Biologie/Chemie, Universität Osnabrück, Barbarastr. 7, D-49069 Osnabrück, Germany

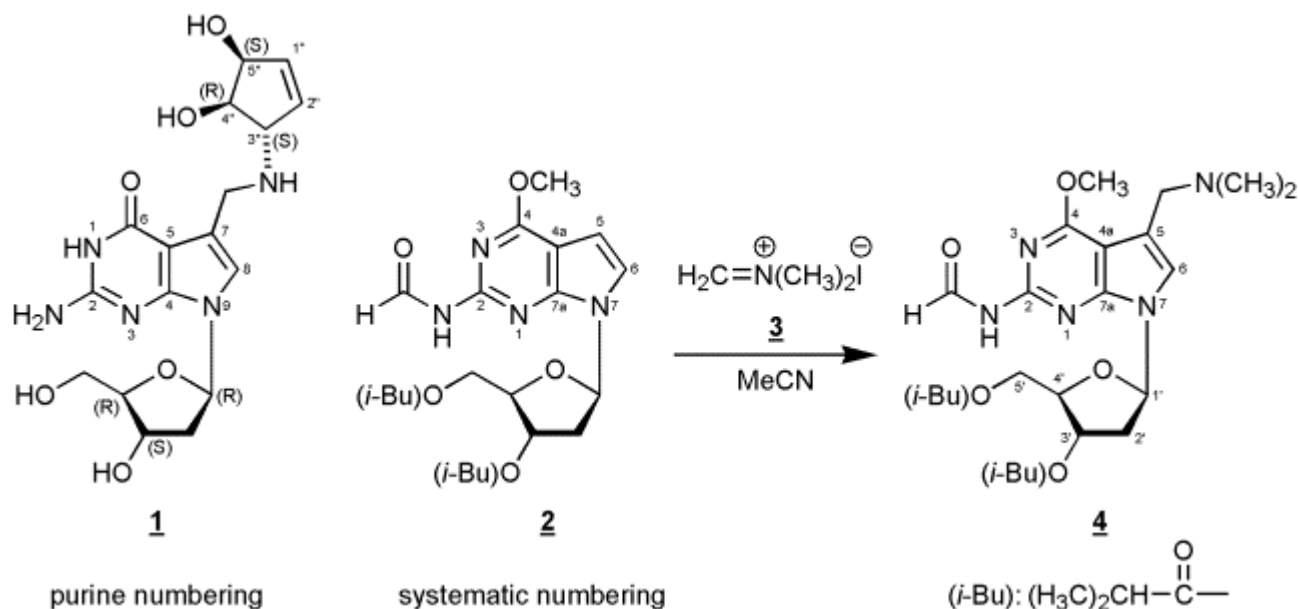
* Author to whom correspondence should be addressed. E-Mail: Helmut.Rosemeyer@uos.de

Received: 28 March 2007 / Accepted: 4 April 2007 / Published: 23 April 2007

Abstract: In this manuscript we report on the regioselective electrophilic substitution at C(7) of 7-[2-deoxy-3,5-bis-O-(2-methylpropanoyl)- β -D-erythro-pentofuranosyl]-2-(formylamino)-4-methoxy-7H-pyrrolo[2,3-d]pyrimidine (**2**) – a protected precursor of 7-deaza-2'-deoxyguanosine - using N,N-dimethyl-methyleniminium iodide (Eschenmoser's salt) yielding the Mannich compound **4**.

Keywords: 7-Deazapurine, Mannich bases, Eschenmoser's Salt

Most naturally occurring 7-deazapurine ribonucleosides [1,2] – both of the adenosine as well as of the guanosine type – carry substituents at the 7-position, and many of them which are found in tRNA represent Mannich bases such as the nucleoside “Q” {2-amino-5-(4,5-cis-dihydroxy-1-cyclopenten-3-yl-trans-aminomethyl-(7- β -D-ribofuranosyl)-3,7-dihydro-4H-pyrrolo[2,3-d]pyrimidin-4-one, Queuosine, 1}. Many attempts have been made to direct a Mannich side chain regioselectively into position 7. In contrast to 7-deazaadenosine derivatives [3,4] this is not an easy task for 7-deazaguanosine precursors [5] and requires chemical detours [6]. For example, regioselective C(7) Mannich alkylation (morpholine, HCOH/HOAc) can be performed at 3',5'-bis-O-toluoylated 6-methoxy-2-(methylthio)-7-deazapurine 2'-deoxy- β -D-ribonucleoside, followed by a three-step aglycone conversion to 7-deazaguanine and deprotection of the glycone [7].



It has been reported for 7-deazaguanines that the position of electrophilic substitution strongly depends on the particular substituent pattern of the base as well as of the reaction conditions [8-10]. Of decisive importance is the observation that a free 2-amino group directs the electrophilic attack into the undesired 8-position (position 6 using systematic numbering) of the 7-deazaguanine moiety. This is the result of mesomeric stabilization of the σ -complex formed during electrophilic attack at the 8-position. On the other hand, 2-acylamino-7-deazaguanine derivatives form the desired 7-substituted compounds [8]

In this communication we disclose that reaction of the fully protected 7-deaza-2'-deoxy-7-deazaguanosine derivative **2** [8] with *N,N*-dimethyl-methyleniminium iodide (Eschenmoser's salt) afforded the C(7) alkylated Mannich compound **4** in moderate yield. This reaction presents a new and alternative route to 7-deazapurine nucleosides with a Mannich side chain in position 7. The structure of **4** was unequivocally assigned by ¹H- and ¹³C-NMR spectroscopy as well as by ¹H-NMR NOE difference spectroscopy (see Experimental Procedure). ¹³C-NMR resonances were assigned applying DEPT-135 and [C,H]HETCOR spectra.

A further transformation of the tertiary amino group of **4** into other Mannich compounds can be accomplished after quaternization. Furthermore, the synthetic or enzymatic incorporation of Mannich bases derived from compounds such as compound **4** allows the introduction of reporter groups in a favourable position of a DNA molecule because the Mannich side chain protrudes into the major groove of a B-DNA double helix.

Experimental Procedure

7-[2-Deoxy-3,5-bis-O-(2-methylpropanoyl)-β-D-erythro-pentofuranosyl]-5-(dimethylaminomethyl)-2-(formylamino)-4-methoxy-7H-pyrrolo[2,3-d]pyrimidine (4).

Compound **2** (305 mg, 0.7 mmol) was dissolved in acetonitrile (3 mL) and *N,N*-dimethyl-methyleniminium iodide (**3**, Eschenmoser's Salz, 260 mg, 1.4 mmol) was added. After heating to 80°C for 24h, the reaction mixture was evaporated to a small volume, and compound **4** (67 mg, 20 %) was isolated as an amorphous solid by thick-layer chromatography (chloroform/methanol, 9:1, two developments, *R_f*, 0.4), elution from the silica gel with methanol and centrifugation. ¹H-NMR ((D₆)DMSO, 500.1 MHz): δ 10.73 (1H, d, *H*C=O, *J* = 9.9 Hz); 9.44 (1H, d, NH, *J* = 9.3 Hz); 7.31 (1H, s, H-C(6)); 6.49 (1H, dd, H-C(1'), *J* = 6.4, 8.0 Hz); 5.36 (1H, dd, H-C(3'), *J* = 2.8, 3.6 Hz); 4.28 - 4.17 (3H, m, H-C(4'), H₂-C(5'));

4.03 (3H, s, OCH₃); 3.67 (2H, s, CH₂); 2.86 (1H, m, iBu-CH); 2.63 – 2.57 (2H, m, H₂-C(2')); 2.47 (1H, m, iBu-CH); 2.28 (6H, s, N(CH₃)₂); 1.15 and 1.09 (12H, 4 iBu-CH₃). ¹H-NMR NOE Data ((D₆)DMSO): proton irradiated: H-C(1'). NOE observed: H-C(6), 1.5 %; H_α-C(2'), 2.5 %; H-C(4'), 1.5 %. ¹³C-NMR ((D₆)DMSO, 125.8 MHz): δ 175.6, 175.8 (2 C=O, iBu); 163.4 (C-4); 163.3 (C=O, formyl); 152.4 (C-7a); 152.0 (C-2); 121.6 (C-6); 111.4 (C-5); 101.5 (C-4a); 82.9 (C-1'); 81.1 (C-4'); 74.2 (C-3'); 63.6 (C-5'); 53.8 (CH₂); 53.7 (OCH₃); 44.1 and 33.1 (4 Me, iBu); 35.9 (C-2'); 18.6 (2 Me, N(CH₃)₂).

References and Notes

1. Suhadolnik, R. J., *Nucleosides As Biological Probes*. John Wiley & Sons, New York **1979**, pp. 158-169.
2. Limbach, P. A.; Crain, P. F.; McCloskey, J. A. *Nucleic Acids Res.* **1994**, *22*, 2183.
3. Watanabe, S. I.; Ueda, T. *Nucleosides Nucleotides* **1983**, *2*, 113.
4. West, R. A. *J. Org. Chem.* **1961**, *26*, 4959.
5. Seela, F. Lüpke, U. *Chem. Ber.* **1977**, *110*, 1462.
6. Seela, F.; Richter, R. *Chem. Ber.* **1978**, *111*, 2925.
7. Seela, F.; Chen, Y.; Zulauf, M. *Synthesis* **1997**, 1067.
8. Ramzaeva, N.; Seela, F. *Helv. Chim. Acta* **1995**, *78*, 1083.
9. Akimoto, H.; Imamiya, E.; Hitaka, T.; Nomura, H.; Nishimura, S. *J. Chem. Soc. Perkin Trans 1* **1988**, 1637.
10. Benghiat, E.; Crooks, P. A. *J. Heterocycl. Chem.* **1983**, *20*, 1023.

Purine numbering has been used within the General Part of the manuscript, systematic numbering within the Experimental Part.

© 2007 by MDPI (<http://www.mdpi.org/>). Reproduction is permitted for noncommercial purposes.