

Supplementary Materials

Exploring Binding Pockets in the Conformational States of the SARS-CoV-2 Spike Trimers for Screening of Allosteric Inhibitors Using Molecular Simulations and Ensemble-Based Ligand Docking

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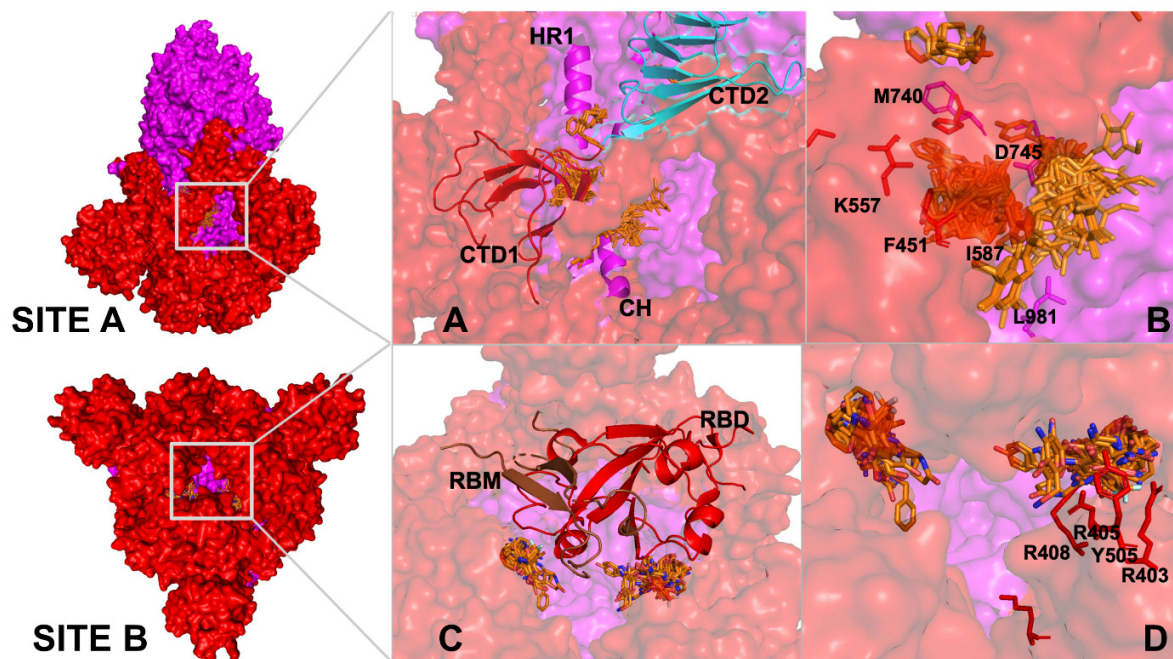


Figure S1. Structural analysis and closeups of the binding pockets and clusters of ligand poses (in default stick colors) of top compounds at site A and B. (A) The binding pocket at Site A is situated between the S1 (red) and S2 (magenta) subunits. (B) A close-up of the two most common binding clusters in site A. Bound ligands are depicted in orange sticks. (C) The binding pocket at Site B and two major clusters of binding poses. The bound ligands are shown in default colored sticks. (D) A close-up of the two most common binding clusters in site A. The S1 subunit is shown in red surface and S2 in magenta surface. The bound ligands are shown in default colored sticks. The binding site residues that are shown are in red sticks and annotated with residue name/number in bold black color.

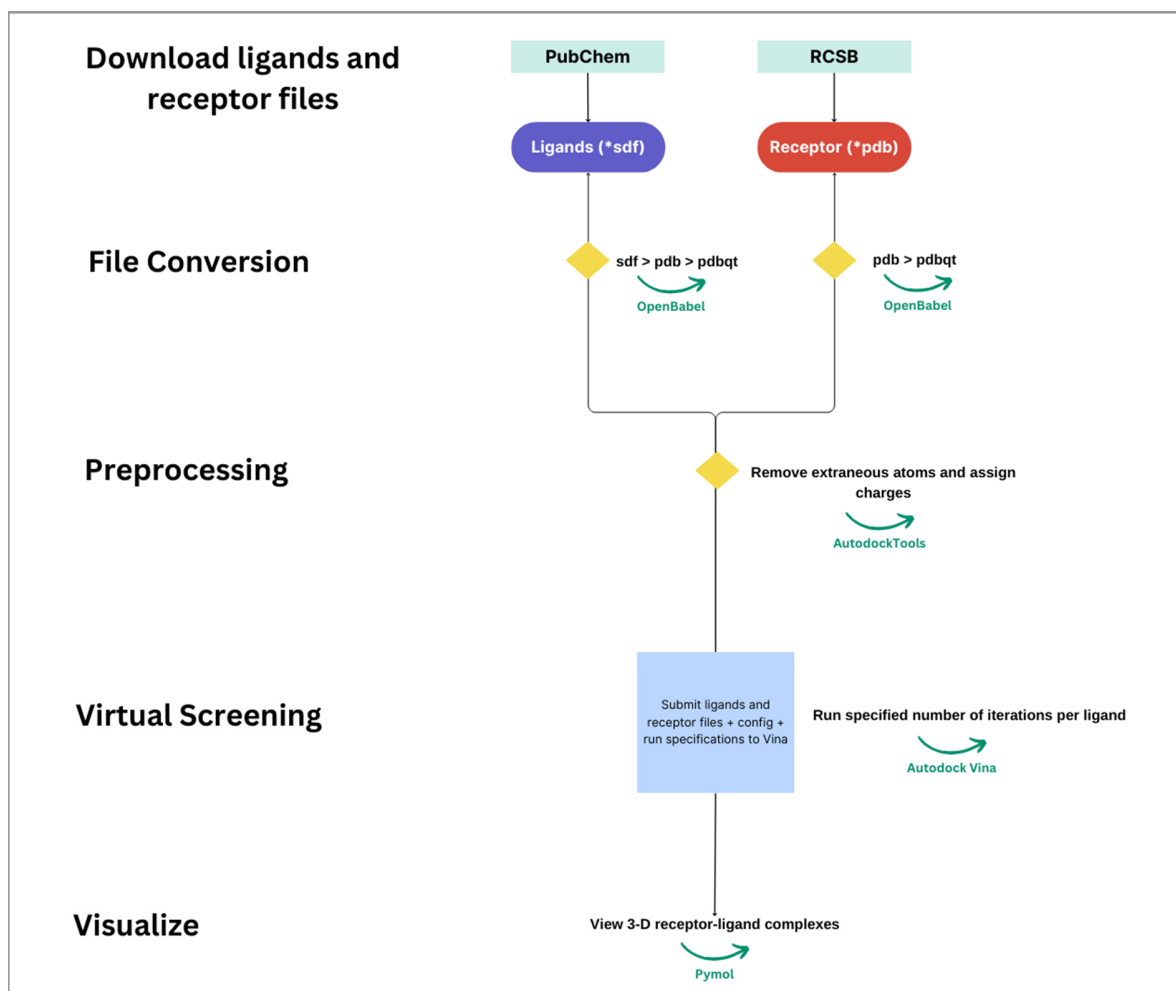


Figure S2. A schematic flow-chart of the virtual screening protocol used in this study.

Auto dock Scripts

A.1 Iteration Script

```
1 #!/usr/bin/perl
2 print "Ligand_file:\t";
3 $ligfile=<STDIN>;
4 chomp $ligfile;
5 open (FH,$ligfile)||die "Cannot open file\n";
6 @arr_file=<FH>;
7
8 for($i=0;$i<@arr_file;$i++)
9 {
10 print "@arr_file[$i]\n";
11 @name=(split(/\./,@arr_file[$i]))[0];
12 }
13 for($i=0;$i<@arr_file;$i++)
14 {
15     chomp @arr_file[$i];
16     print "@arr_file[$i]\n";
17     mkdir @name;
18     for($j=1; $j<=3; $j++)
19     {
20         system("vina --config conf.txt --ligand @arr_file[$i] --log @name/@arr_file[$i]_[$j]_log.log");
21     }
22 }
```

A.2 Output Script

```
1 #!/bin/bash
2
3 echo 'num, zincid, filename,mode,affinity,dist_rmsd,best_mode_rmsd' | tee -a output.csv
4
5 ligfile="$1"
6 while read -r line; do
7     name=$(echo "$line" | cut -f 1 -d '.')
8     echo "$name"
9     for i in $name/*.log; do
10         echo -n $name, | tee -a output.csv
11         id=$(grep -A1 "<zinc_id>" $name.sdf | tail -n 1)
12         echo -n $id, | tee -a output.csv
13         echo -n $i | tee -a output.csv
14         result=$(grep -A3 'mode' $i | tail -n 1 | tr -s " " | sed 's/ /, /g')
15         if [ -z "$result" ]; then echo 'NA,NA,NA,NA' | tee -a output.csv; else echo $result | tee -a output.csv; fi
16     done
17 done < "$ligfile"
```

A.3 Cluster Scripts

```
1 #!/bin/bash
2
3 ligfile="$1"
4 while read -r line; do
5     name=$(echo "$line" | cut -f 1 -d '.')
6     echo "$name"
7     cd $name
8     for ((i=79; i<101; i++)); do
9         echo "$i"
10         vina --config ../conf_6m0j_fullgrid.txt --ligand $line --out [$name]_[$i]_out.pdbqt --log [$name]_[$i]_log.log
11     done
12     cd ..
13 done < "$ligfile"
```

```
#!/bin/bash

jobid="$1"

cat > job_${1}.sbatch << "END"

#SBATCH --time=16-0
#SBATCH --ntasks-per-node=1
#SBATCH --cpus-per-task=1
#SBATCH --mem-per-cpu=40G
#SBATCH --partition=cpu-long.q
#SBATCH --output=slurm_${1}.out
#SBATCH --job-name=serialvina_${1}
#SBATCH --mail-type=END,FAIL
#SBATCH --mail-user=grgupta@chapman.edu

module load anaconda3/current
source /cm/shared/apps/anaconda3/etc/profile.d/conda.sh
conda activate myprojenv

(time ./vina_iterate.sh $1.txt) &> $1_time.txt

END
```

```
#!/usr/bin/env bash

#SBATCH --time=16-0
#SBATCH --ntasks-per-node=1
#SBATCH --cpus-per-task=1
#SBATCH --mem-per-cpu=40G
#SBATCH --partition=cpu-long.q
#SBATCH --output=slurm_zinc_1.out
#SBATCH --job-name=zincvina_x1
#SBATCH --mail-type=END,FAIL
#SBATCH --mail-user=grgupta@chapman.edu

module load anaconda3/current
source /cm/shared/apps/anaconda3/etc/profile.d/conda.sh
conda activate myprojenv

(time ./vina_iterate.sh zinc_1.txt) &> zinc_1_time.txt
```