

Supplementary materials

Add and Go: FRET Acceptor for Live-Cell Measurements Modulated by Externally Provided Ligand

Alexey S. Gavrikov¹, Nina G. Bozhanova¹, Mikhail S. Baranov^{1,2}, Alexander S. Mishin^{1*}

¹ Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences, Miklukho-Maklay st. 16/10, Moscow, 117997, Russia

² Pirogov Russian National Research Medical University, Ostrovitianov 1, Moscow, 117997, Russia

*correspondence: mishin@ibch.ru

1. Supplementary materials and methods

1.1 Constructs sequences

>H2B-mNeonGreen-DiB1

MPEPAKSAPAPKKGSKKAVTKAQKKGGKKRKRSRKESYSIYVYKVLKQVHPDTGISS
KAMGIMNSFVNDIFERIAGEASRLAHYNKRSTITSREIQTAVRLLLPGELAKHAVSEGT
KAITKYTSAKDPPVATMVSKGEEDNMASLPATHELHIFGSINGVDFDMVGQGTGNPN
DGYEELNLKSTKGDQLQFSPWILVPHIGYGFHQYLPYPDGMSPFQAAMVDGSGYQVH
RTMQFEDGASLTVNYRYTYEGSHIKGEAQVKGTGFPADGPVMTNSLTAADWCRSKK
TYPNDKTIISTFKWSYTTGNGKRYRSTARTTYTFAKPMAANYLKNQPMYVFRKTELK
HSKTELNFKEWQKAFTDVMGMDELYKGDPPVATMASSPTPPRGVTVVNNFDCKRYL
GTWYEIARFDHRFERGLEKVTATYSLRDDGGLNVINKGYNPDRGMWQQSEGKAYFT
GAPTRAALKVSFFGPFYGGYNVIALDREYRHALVCGPDRDYLWINSRTPPTISDEVKE
MLAVATREGFDVSKFIWVQQPGSGS

>Vimentin-DiB1

MSTRSVSSSSYRRMFGGPGTASRPSSSRSYVTTSTRTYSLGSALRPSTSRSLYASSPGG
VYATRSSAVRLRSSVPGVRLQLQDSVDFSLADAINTEFKNTRTNEKVELQELNDRFANY
IDKVRFLEQQNKILLAELEQLKGQGSRLGDLYEEEMRELRRQVDQLTNDKARVEVE
RDNLAEDIMRLREKLQEEMLQREEAENTLQSFRQVDNASLARLDLERKVESLQEEI
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EWYKSKFADLSEAANRNDALRQAKQESTEYRRQVQSLTCEVDALKGTNESLERQ
MREMEENFAVEAANYQDTIGRLQDEIQNMKEEMARHLREYQDLLNVKMALDIEIAT
YRKLLGEEESRISLPLPNFSSLNLRETNLDSLPLVDTHSKRTLLIKTVETRDGQVINETS
QHDDLEGDPPVATGMASSPTPPRGVTVVNNFDCKRYLGTWYEIARFDHRFERGLE
KVTATYSLRDDGGLNVINKGYNPDRGMWQQSEGKAYFTGAPTRAALKVSFFGPFYGG
YNVIALDREYRHALVCGPDRDYLWINSRTPPTISDEVKEMLA VATREGFDVSKFIWV
QQPGSGS

>Tension sensor

MPVFHTRTIESILEPVAQQISHLVIMHEEGEVDGKAIPDLTAPVSAVQAAVSNLVRVVK
ETVQTTEDQILKRDMPPAFIKVENACTKLVRAAQMLQADPYSVPARDYLIDGSRGILS
GTSDLLLTFDEAEVRKIIRVCKGILEYLTVAEVVETMEDLVTYTKNLGPGMTKMAKM
IDERQQELTHQEHRVMLVNSMNTVKELLPVLISAMKIFVTTKNTKSQGIEEALKNRNF
TVEKMSAEINEIIRVLQLTSWDEDAWASKDTEAMKRALALIDSKMNQAKGWLDPN
APPGDAGEQAIRQILDEAGKAGELCAGKERREILGTCKTLGQMTDQLADLRARGQG
ATPMAMQKAQQVSQGLDLLTAKVENAARKLEAMTNSKQAIKIDAAQNWLADPN
GGSEGEHIRGIMSEARKVAELCEEPKERDDILRSLGEISALTAKLSDLRRHGKGDSP
ARALAKQIATSLQNLQSKTNRAVANTRPVKAAVHLEGKIEQAQRWIDNPTVDDRGV
GQAAIRGLVAEGRRLANVMMGPYRQDLLAKCDRVDQLAAQLADLAARGESESPQA
RAIAAQLQDSLKDLKARMQEAMTQEVSDVFSDTTTPIKLLAVAATAPSDTPNREEVFE
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NPGNQAAAYEHFETMKNQWIDNVEKMTGLVDEAIDTKSLLDASEEAIKKDLKCKVA
MANMQPQMLVAGATSIARRANRILLVAKREVENSEDPKFREAVKAAASDELKTIKSPM
VMDAKAVAGNISDPGLQKSFLDSGYRILGAVAKVREAFQPQEPDFPPPPDLEHLHLT
DELAPPKPLPEGEVPPPRPPPEEKDEVEMVSKGEEDNMAASLPATHELHIFGSINGVD
FDMVGQGTGNPNDGYEELNLKSTKGDLQFSPWILVPHIGYGFHQYLPYPDGMSPFQ
AAMVDGSGYQVHRTMQFEDGASLTVNYRYTYEGSHIKGEAQVKGTGFPADGPVMT
NSLTAADWCRSKKTYPNDKTIISTFKWSYTTGNGKRYRSTARTTYTFAKPMAANYLK
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GPPGAGPPGAGPPGAGPPGAGPPGAMASSPTPPRGVTVVNNFDCKRYLGTWYEIA
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LKVSFFGPFYGGYNVIALDREYRHALVCGPDRDYLWINSRTPPTISDEVKQEMLAVATR
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IIAAAKRMALLMAEMSLVRGGSGNKRALIQCAKDIKASDEVTRLAKEVAKQCTD
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VKETVREAEAAASIKIRTDAGFTLRWVRKTPWYQGS

>YAP-mNeonGreen

MDPGQQPPPQPAPQGGQPPSQPPQGGPPSGPGQPAPAATQAAPQAPPAGHQIVHV
RGDSETDLEALFNAVMPNPKTANVPQTVPMRLRKLKLPDSFFKPPPEPKSHSRQASTDAGT
AGALTPQHVAHSSPASLQLGAVSPGTLTPTGVVSGPAATPTAQHLRQSSFEIPDDVPL
PAGWEMAKTSSGQRYFLNHIDQTTTWQDPRKAMLSQMNVTAPTSPPVQQNMMNSA
SGPLPDGWEQAMTQDGEIYYINHKNKTTSWLDPRLDPRFAMNQRISSAPVKQPPPL
APQSPQGGVMGGSNSNQQQQMRLQQLQMEKERLRLKQQELLRQAMRNINPSTANS
PKCQELALRSQLEQDGGTQNPVSSPGMSQELRTMTTNSSDPFLNSGTYSRDEST
DSGLSMSSYSVPRTPDDFLNSVDEMDTGDITINQSTLPSQQNRFPDYLEAIPGTNVDLG
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WILVPHIGYGFHQYLPYPDGMSPFQAAMVDGSGYQVHRTMQFEDGASLTVNYRYTY
EGSHIKGEAQVKGTGFPADGPVMTNSLTAADWCRSKKTYPNDKTIISTFKWSYTTGN
GKRYRSTARTTYTFAKPMAANYLKNQPMYVFRKTELKHSKTELNFKEWQKAFTDV
MGMDELYKG

>14-3-3-DiB1

MERASLIQKAKLAEQAERYEDMAAFMKGAVEKGEELSCEERNLLSVAYKNVVGQ
RAAWRVLSSIEQKSNEEGSEEKGPEVREYREKVELTELQGVCDTVLGLLDSHLIKEAG
DAESRVFYLMKMGDYRYLAEVATGDDKKRIIDSARSAYQEAMDISKKEMPPTNPIR
LGLALNFSVFHYEIANSPPEAISLAKTTFDEAMADLHTLSEDSYKDSTLIMQLLRDNL
TLWTADNAGEEGGEAPQEPQSDPPVATMASSPTPPRGVTVVNNFDCKRYLGTWYEIA
RFDHRFERGLEKVTATYSLRDDGGLNVINKGYNPDRGMWQQSEGKAYFTGAPTRAA
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2. Supplementary figures and movies

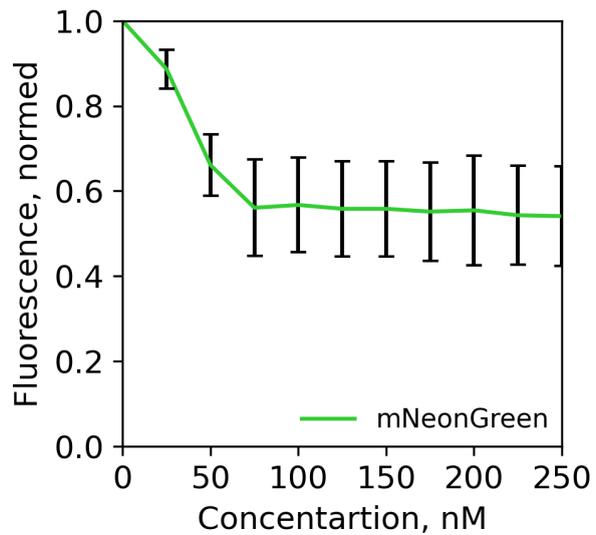


Figure S1. Live-cell titration of modulated FRET acceptor. Fluorescence intensity of mNeonGreen in live HeLa cells transiently transfected with H2B-mNeonGreen-DiB1 in the presence of different concentrations of compound **3**; error bars represent standard deviation (technical replicates = 17).

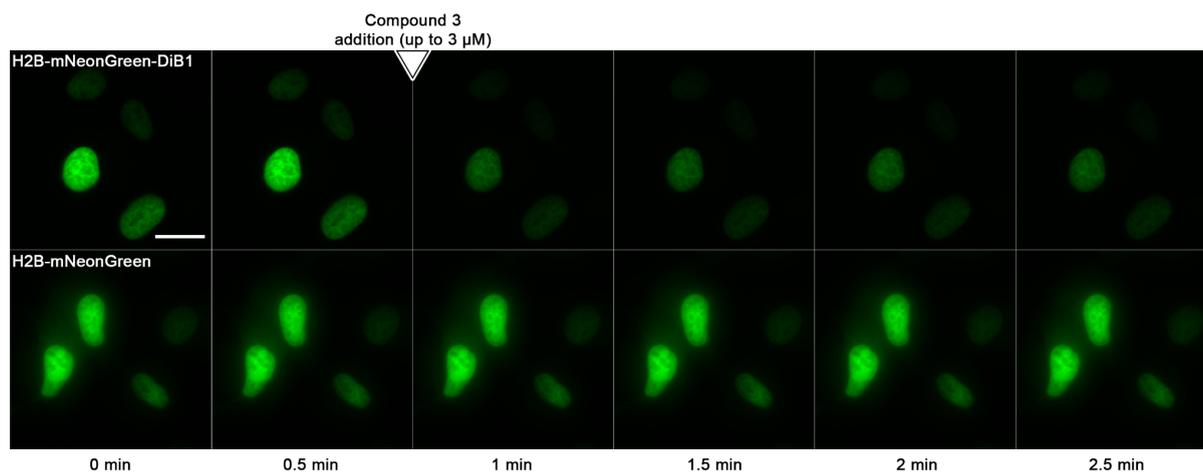


Figure S2. Live-cell image sequence for FRET acceptor activation. Widefield fluorescence imaging of live HeLa Kyoto cells transiently transfected with H2B-mNeonGreen (lower row) and H2B-mNeonGreen-DiB1 (upper row) constructs before and after compound **3** addition up to 3 μM concentration; Scale bar is 20 μm .

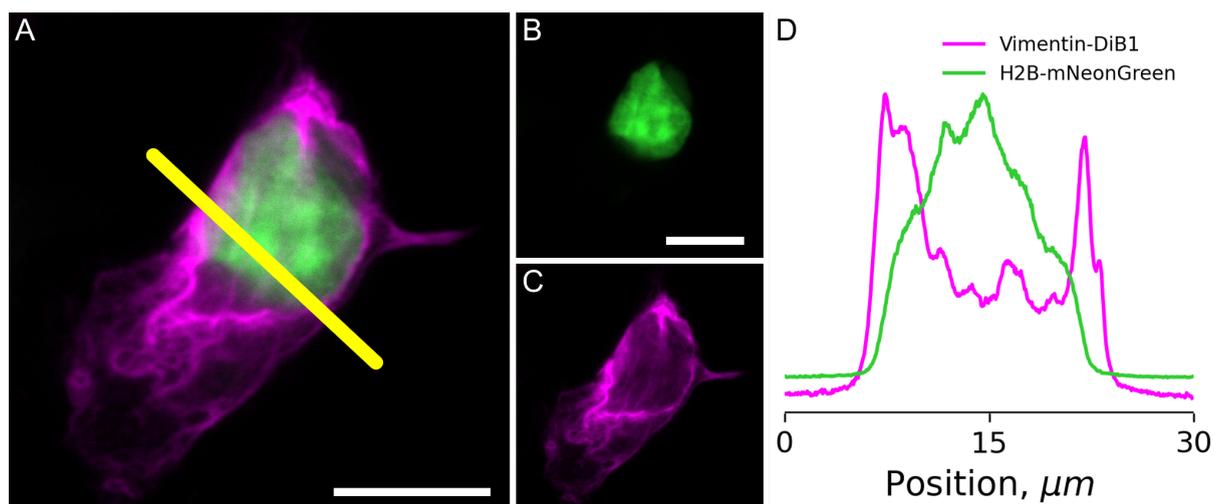


Figure S3. Imaging of non-interacting proteins labeled with modulated FRET pair.

Live-cell imaging of DiB1 and mNeonGreen on non-interacting proteins. Widefield fluorescence imaging of live HeLa Kyoto cells transiently cotransfected with H2B-mNeonGreen and vimentin-DiB1 in the presence of 100 nM of compound **3**. **(A)** Composed image of labeled nucleus and vimentin filaments. **(B, C)** Separate channels of panel **(A)**. **(D)** Fluorescence intensity profile (indicated by yellow line on panel **(A)**); Scale bars are 20 μm .

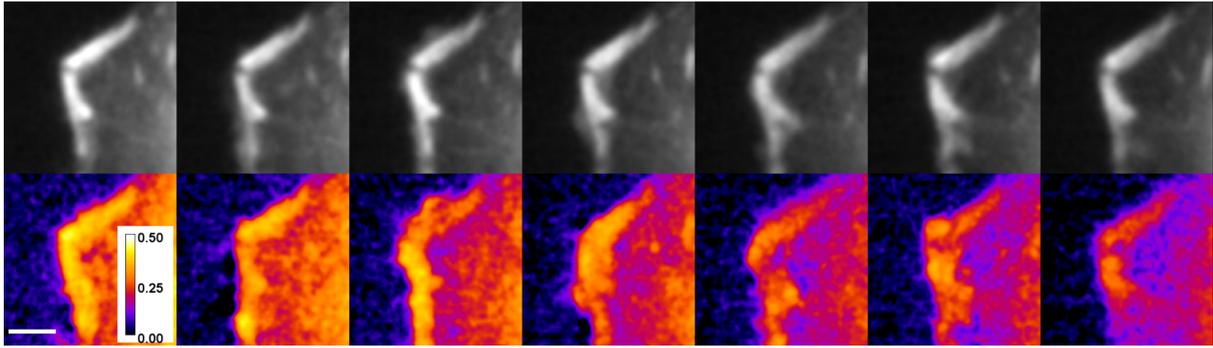


Figure S4. Tension sensor with single-channel FRET detection. Widefield fluorescence imaging of dynamic tension changes in focal contacts of live HeLa Kyoto cells transiently transfected with tension sensor construct. Frame interval – 6 minute. A cyclic FRET acceptor activation (staining with compound **3**, followed by washing with HHBS buffer) was performed on every time point. Upper row – widefield images before staining, lower row – FRET efficiency images; pseudocolor indicates FRET efficiency after the addition of 3 μM compound **3**; Scale bar is 3 μm .

Supplementary Movie S1. Cyclic FRET activation. Repeated cycles of FRET activation in live HeLa cells transiently transfected with H2B-mNeonGreen-DiB1 construct. Green lines represent average intensities of mNeonGreen (donor) fluorescence in labeled nuclei. Acceptor activation was achieved by the addition of HHBS buffer supplied with 3 μ M of compound **3**. Washing was performed with HHBS buffer; Scale bar is 10 μ m. Video plays at 30 fps.

Supplementary Movie S2. Tension sensor with single-channel FRET detection. Widefield fluorescence imaging of dynamic tension changes in focal contacts of live HeLa Kyoto cells transiently transfected with tension sensor construct. Frame interval - 6 minutes. A cyclic FRET acceptor activation (staining with compound **3**, followed by washing with HHBS buffer) was performed on every time point. Upper image - widefield microscopy, lower image - FRET efficiency image; pseudocolor indicates FRET efficiency after the addition of 3 μ M compound **3**; Scale bar is 3 μ m. Video plays at 2 fps.