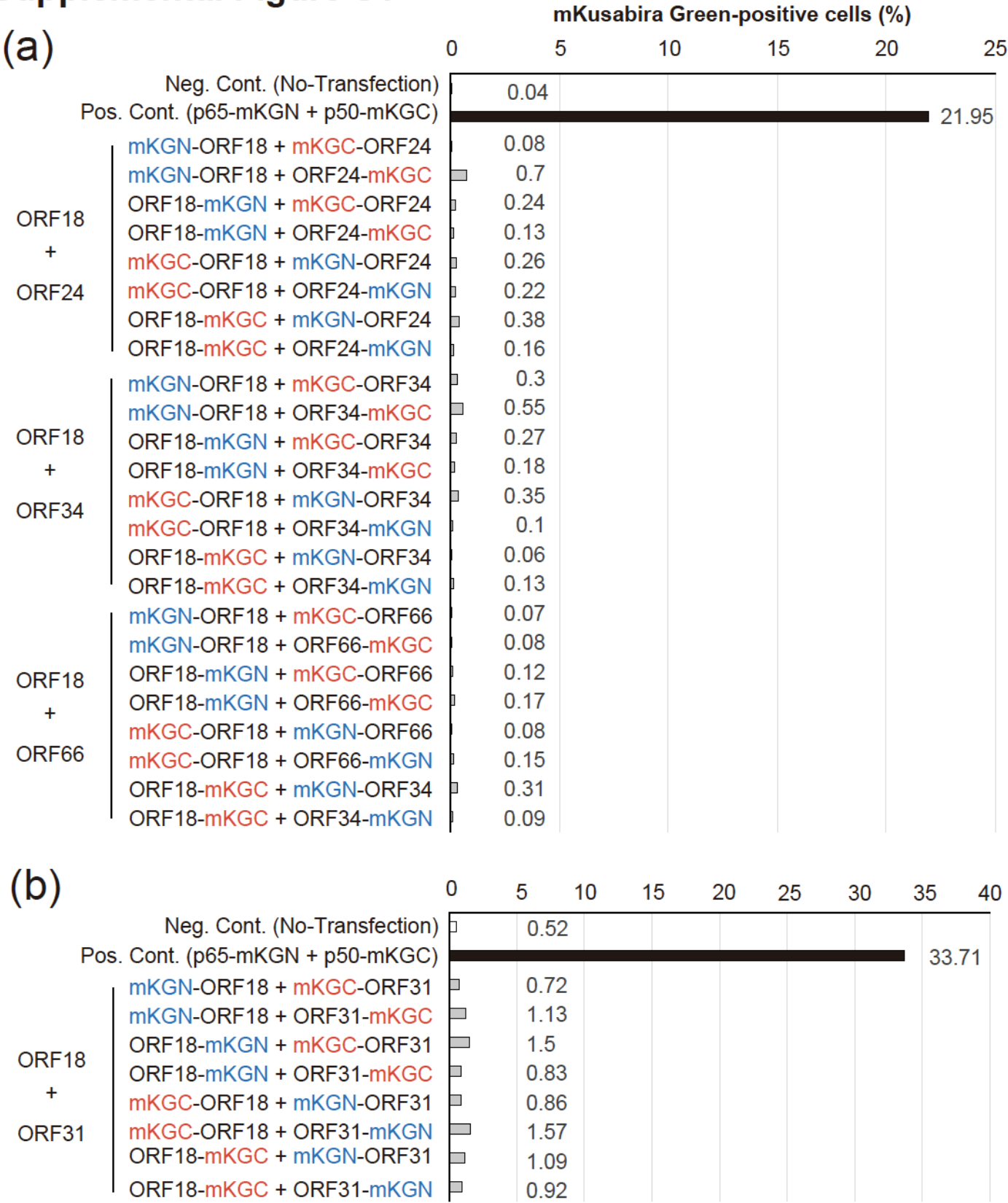


# Supplemental Figure S1



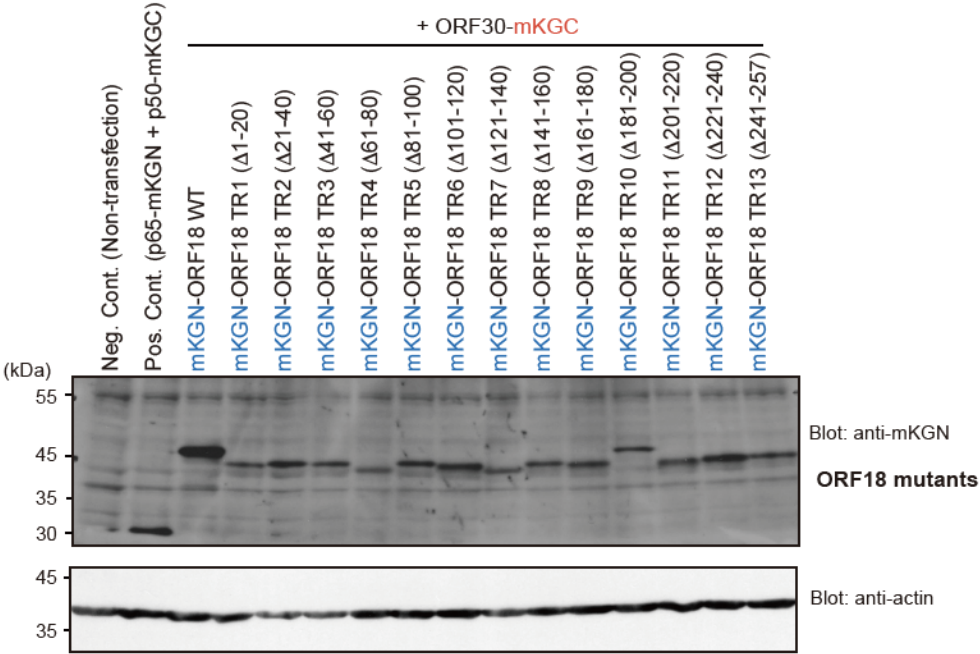
**Supplemental Figure S1.**

(a and b) Screening and optimization of the BiFC assay to determine vPIC component interactions.

Each indicated combination of the expression plasmids were co-transfected into 293T cells by the calcium phosphate method and single samples were assessed by flow cytometry.

The negative control (Neg. Cont.) consisted of non-transfected cells, and the positive control (Pos. Cont.) comprised cells co-transfected with p65-mKGN (pCONT-1) and p50-mKGC (pCONT-2) expression plasmids.

# Supplemental Figure S2

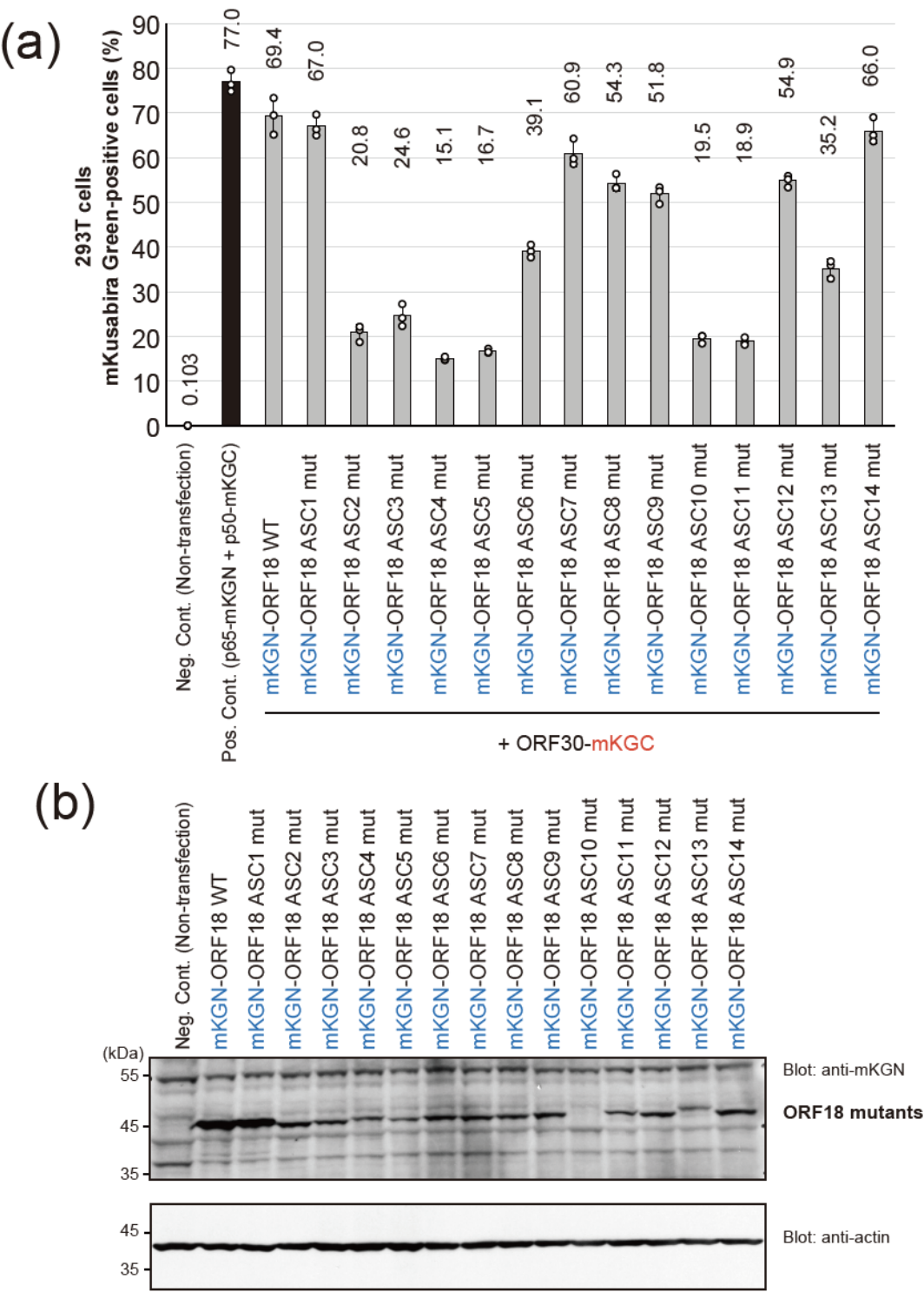


## Supplemental Figure S2.

Confirmation of ORF18 truncated mutant protein expression.

Each indicated combination of the expression plasmids were simultaneously and independently co-transfected in order to conduct a BiFC assay (as described in Fig. 2) and a Western blot (described here). The co-transfected cells were lysed and subjected to Western blotting using anti-mKGN primary antibodies. An antibody that recognizes actin (anti-actin) was used as a loading control. The original blotting data are shown in Supplemental Figure S5.

# Supplemental Figure S3

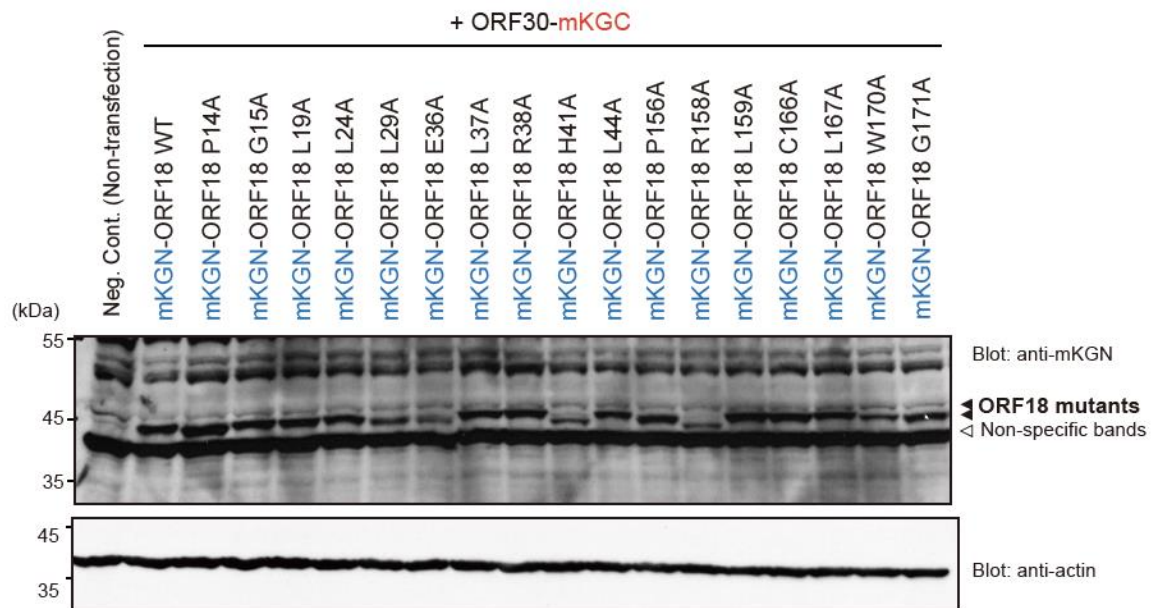


Supplemental Figure S3.

(a) BiFC assay of ORF18 block alanine-scanning mutants (ASC1mut-ASC14mut). Each mKGN-ORF18 block alanine-scanning mutant expression plasmid was co-transfected into 293T cells with ORF30-mKGC. The 293T cells were transfected with a lipofection method and three independent samples were assessed by flow cytometry. The negative control (Neg. Cont.) was non-transfected cells and the positive control (Pos. Cont.) comprised cells co-transfected with p65-mKGN (pCONT-1) and p50-mKGC (pCONT-2). Each bar and error bar indicate the average and standard deviation, respectively.

(b) Protein expression of each ORF18 block alanine-scanning mutant (ASC1mut-ASC14mut). Each mKGN-ORF18 block alanine-scanning mutant expression plasmid was co-transfected into 293T cells with ORF30-mKGC. The samples were subjected to Western blotting using anti-mKGN primary antibody. An antibody that recognizes actin (anti-actin) was used as a loading control. The original blotting data are shown in Supplemental Figure S5.

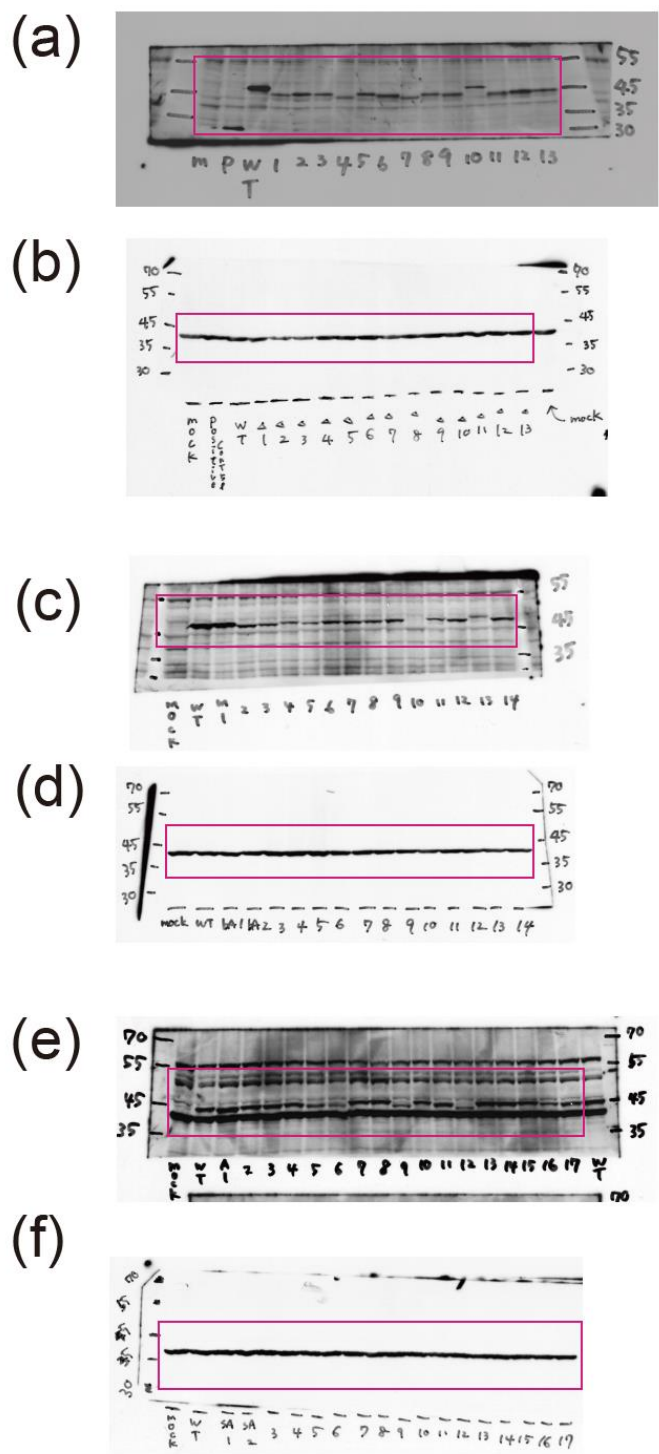
# Supplemental Figure S4



## Supplemental Figure S4.

S4. Protein expression of each ORF18 single alanine mutant. Each indicated combination of the expression plasmids were simultaneously and independently co-transfected in order to conduct a BiFC assay (as described in Fig. 4b) and a Western blot (described here). The cells were lysed and subjected to Western blotting using anti-mKGN primary antibodies. An antibody that recognizes actin (anti-actin) was used as a loading control. The original blotting data are shown in Supplemental Figure S5.

# Supplemental Figure S5



## Supplemental Figure S5

Original Western blotting data (X-ray film) of Supplemental Figure S2, S3, and S4.

(a) Original anti-mKGN blotting data of Supplemental Figure S2 upper panel.

(b) Original anti-actin blotting data of Supplemental Figure S2 lower panel.

(c) Original anti-mKGN blotting data of Supplemental Figure S3b upper panel.

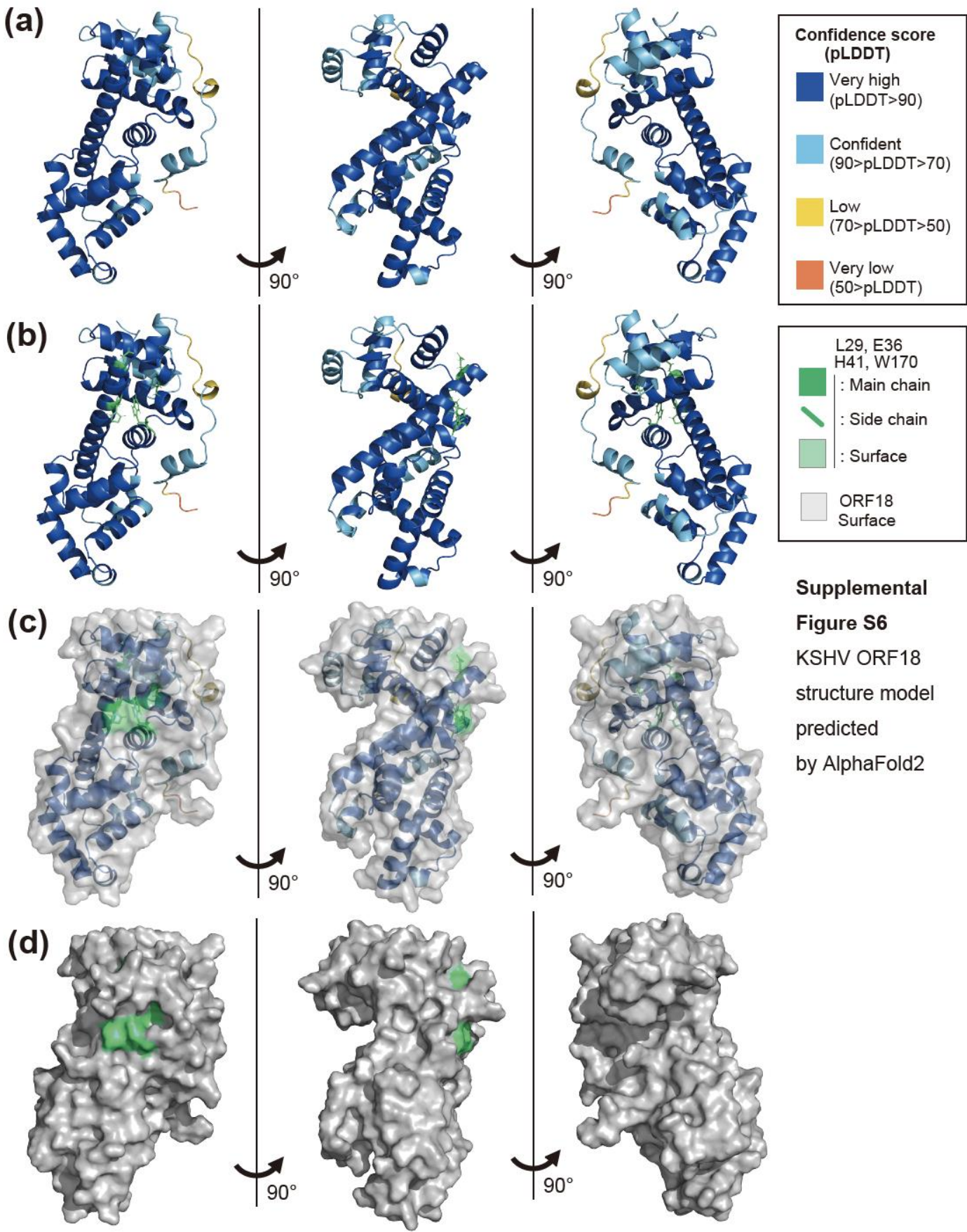
(d) Original anti-actin blotting data of Supplemental Figure S3b lower panel.

(e) Original anti-mKGN blotting data of Supplemental Figure S4 upper panel.

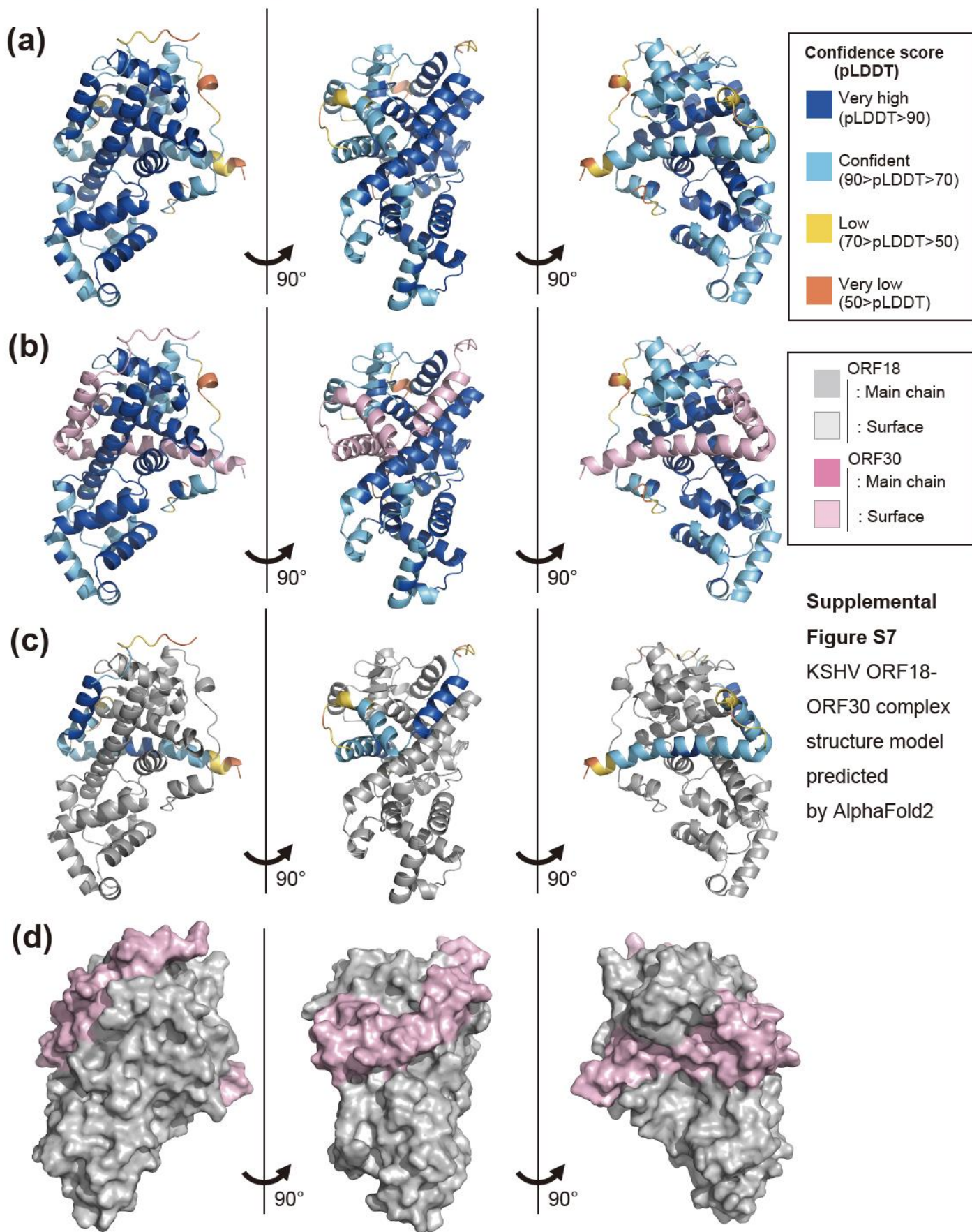
(f) Original anti-actin blotting data of Supplemental Figure S4 lower panel.



Supplemental Figure S6

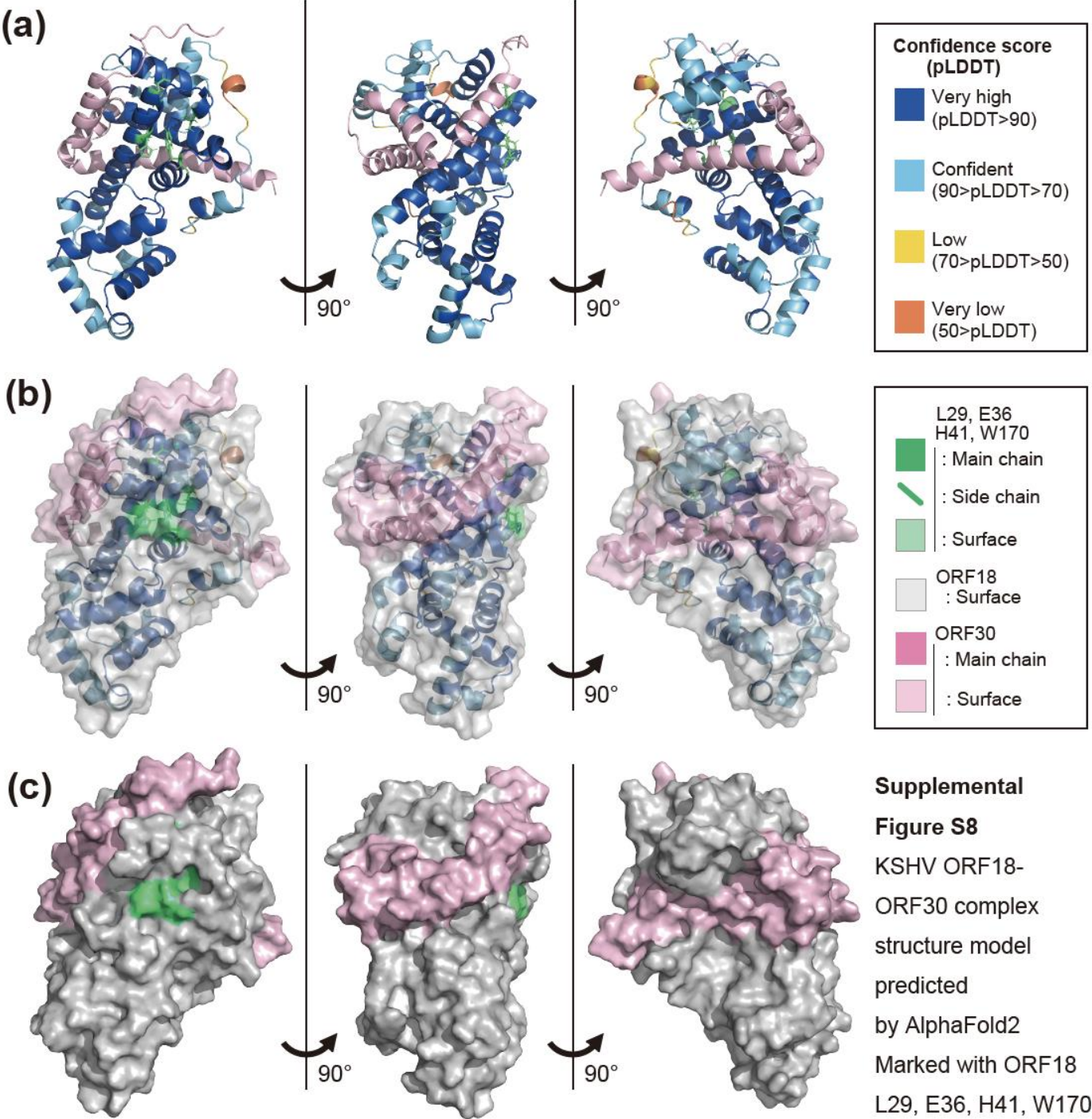


# Supplemental Figure S7



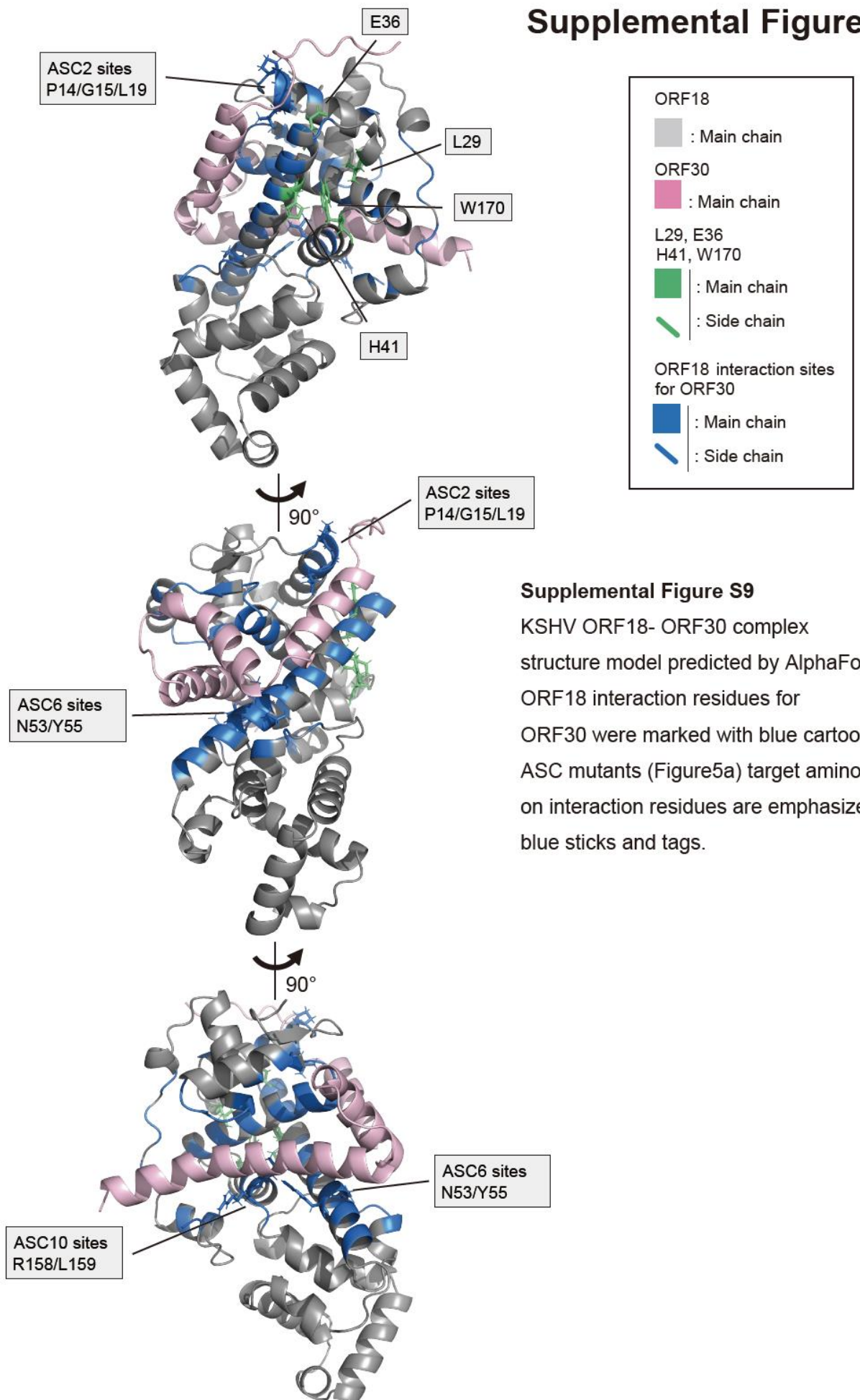


# Supplemental Figure S8





# Supplemental Figure S9

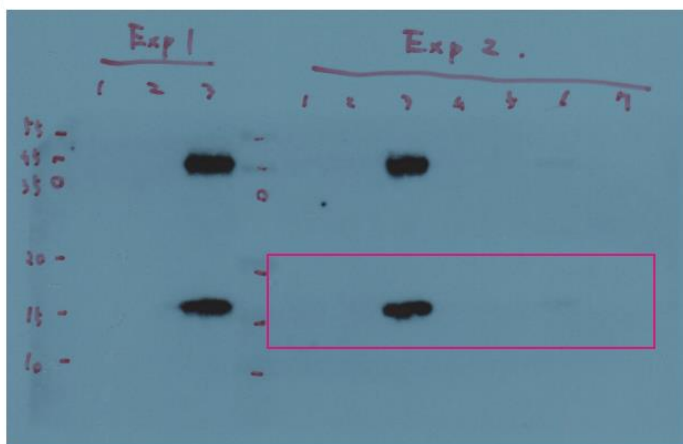


## Supplemental Figure S9

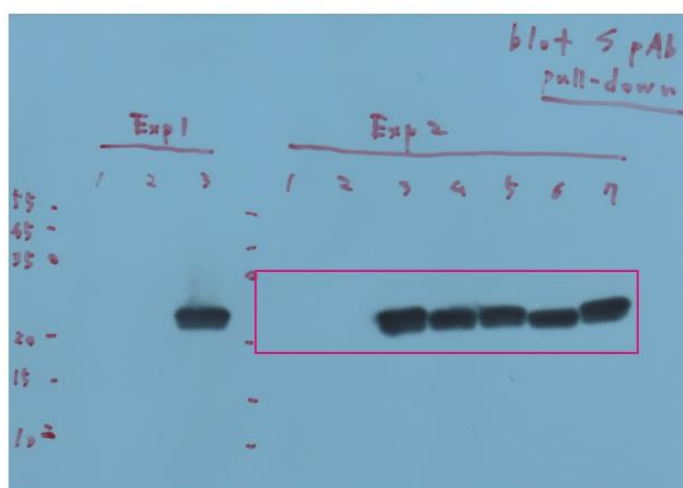
KSHV ORF18- ORF30 complex  
structure model predicted by AlphaFold2;  
ORF18 interaction residues for  
ORF30 were marked with blue cartoon.  
ASC mutants (Figure5a) target amino-acids  
on interaction residues are emphasized as  
blue sticks and tags.

# Supplemental Figure S10

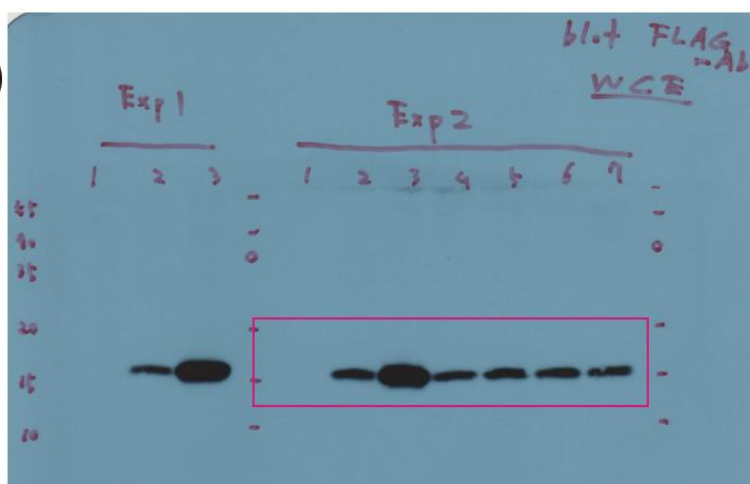
(a)



(b)



(c)



## Supplemental Figure S10

Original Western blotting data (X-ray film) of Figure 5b

(a) Original anti-FLAG blotting data of Figure 5b upper panel.

(b) Original anti-Stag blotting data of Figure 5b Middle panel.

(c) Original anti-FLAG blotting data of Figure 5b lower panel.

**Supplemental TABLE S1: Primers for construction of expression plasmids**

Plasmid name	Primer name	Primer sequences (5' -> 3')	
<i>[VPK components]</i>			
ph-ORF18-mKGN	S2_EcoRI-kshvORF18	catcGAATTCgatgctcgaaatactggtgtagac	<i>*a</i>
ph-ORF18-mKGC	As(stp)_kshvORF18-NotI	taaGCGGCGCGTaaacgcggtgtgtttaaac	<i>*a</i>
ph-mKGN-ORF18	As2(Nstp)_kshvORF18-NotI	gttGCGGCGCGTaaacgcggtgtgtttaaac	<i>*a</i>
ph-mKGC-ORF18	As2(Nstp)_kshvORF18-NotI	atcGAATTCgatgggtgaacgactggatctcgac	<i>*a</i>
ph-ORF30-mKGN	S2_EcoRI-kshvORF30	tgagCGGCGCGTcaatttcgacacggtgtgtac	<i>*a</i>
ph-ORF30-mKGC	As(stp)_kshvORF30-NotI	aaaGCGGCGCGTcttcgcacggtgtgtac	<i>*a</i>
ph-mKGN-ORF30	As2(Nstp)_kshvORF30-NotI	catGGATCCatgacgcgcgcgcgcgcgcgc	<i>*a</i>
ph-mKGC-ORF30	S_BamHI-kshvORF24	ctaaAAGCTTTtagaccagcgcgcgcgcgcgc	<i>*a</i>
ph-ORF24-mKGN	As(stp)_kshvORF24-HindIII	gtcAAGCTTgaaccgcgcgcgcgcgcgcgc	<i>*a</i>
ph-mKGN-ORF24	As2(Nstp)_kshvORF24-HindIII	catcGAATTCgatgcacaaacagaaagactctgc	<i>*a</i>
ph-ORF31-mKGN	S2_EcoRI-kshvORF31	tagGCGGCGCGTctacgtatctcttctgtagacgt	<i>*a</i>
ph-ORF31-mKGC	As(stp)_kshvORF31-NotI	acgGCGGCGCGTctacgtatctcttctgtagacgt	<i>*a</i>
ph-mKGN-ORF31	As2(Nstp)_kshvORF31-NotI	catcCTCGAGaatgttctgttgtagctgcgc	<i>*a</i>
ph-ORF34-mKGN	S_XhoI-kshvORF34	ctaaAAGCTTTtagagttggttgatctctcc	<i>*a</i>
ph-ORF34-mKGC	As(stp)_kshvORF34-HindIII	actcAAGCTTtaggttggttgatctctcc	<i>*a</i>
ph-mKGN-ORF34	As2(Nstp)_kshvORF34-HindIII	catcCTCGAGaatgttctgttgtagctgcgc	<i>*a</i>
ph-ORF66-mKGN	S_XhoI-kshvORF66	ctgaAAGCTTTcaggaggaacactccgcacag	<i>*a</i>
ph-ORF66-mKGC	As(stp)_kshvORF66-HindIII	ctccAAGCTTtgaaggaacactccgcacag	<i>*a</i>
ph-mKGN-ORF66	As2(Nstp)_kshvORF66-HindIII		
<i>[Expression control plasmids]</i>			
pCIneo-mCherry	S_EcoRI_mCherry As(stp)_mCherry_Sall	catcGAATTCgatgtagcaaggcgag tagGTCGACtactgtacagctgctcatg	<i>*a</i> <i>*a</i>
<i>[ORF18 single mutants]</i>			
ph-mKGN-ORF18-P14A	S_1_KSHV_ORF18_P14A As_1_KSHV_ORF18_P14A	ccgctgagcGcAGggcttagagagctcatg ctaaagccTGCgtacgcgcgtgcgtctc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-G15A	S_2_KSHV_ORF18_G15A As_2_KSHV_ORF18_G15A	ctgagccgcGCGcttagagagctcatggtg ctctcaagCGCcggtgcgcgcgcgcgcgc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L19A	S_3_KSHV_ORF18_L19A As_3_KSHV_ORF18_L19A	gcttagagagcGCcatgtggtgcttttgcacaaatag gccacatGGCctctctaaagccccgcgtc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L24A	S_4_KSHV_ORF18_L24A As_4_KSHV_ORF18_L24A	gtggcgcttTGCAaaataagtaatttaataac cttattingTGCaaagcgccacatgagctc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L29A	S_5_KSHV_ORF18_L29A As_5_KSHV_ORF18_L29A	gaatGCAaatacaattccacgccaag cgtggaatgattTGCattctattttgcacaaaag	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-E36A	S_6_KSHV_ORF18_E36A As_6_KSHV_ORF18_E36A	catccacgcacaaGCActgcgtttttattcaattg cagTGcttgggcgtggaatgattttaattc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L37A	S_7_KSHV_ORF18_L37A As_7_KSHV_ORF18_L37A	ccaagagGCCcggttttattcatttggttc gaataaaacGGCctcttgcgcgfggaatg	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-R38A	S_8_KSHV_ORF18_R38A As_8_KSHV_ORF18_R38A	caagaagctGCTtttattcatttggtctc gaataaaAGCcaagctctggtggaatg	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-H41A	S_9_KSHV_ORF18_H41A As_9_KSHV_ORF18_H41A	gtttttatGCAttggtctctgcagagatg gagaaccaaTGcaataaaacagcagctctg	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L44A	S_10_KSHV_ORF18_L44A As_10_KSHV_ORF18_L44A	catttggttGCCgcagagatgacaaatttc catctcgcGCAcacaacaaatgaataaaacg	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-P156A	S_11_KSHV_ORF18_P156A As_11_KSHV_ORF18_P156A	gtggtgcGCAgcagagctgctctgtgag cagctgcGTCgagcacactcccaatgattaaac	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-R158A	S_12_KSHV_ORF18_R158A As_12_KSHV_ORF18_R158A	ccctgcGCTctgctctctggtgggcgcgc cgcaaggagAGCgcaggggngcncacttcc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L159A	S_13_KSHV_ORF18_L159A As_13_KSHV_ORF18_L159A	cctgcaggGCGctctctgagcgaggctac cgcaaggagCGCctgcaggggngcncacttcc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-C166A	S_14_KSHV_ORF18_C166A As_14_KSHV_ORF18_C166A	ggctacGCActgccttttggggcagcgc caaaaaggcagTGCgtagccgcgcgcgaaggag	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-L167A	S_15_KSHV_ORF18_L167A As_15_KSHV_ORF18_L167A	ggctactgcGCTgcttttggggcagcgatg caaaaaggcAGCgcagttagccgcgcgcgaag	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-W170A	S_16_KSHV_ORF18_W170A As_16_KSHV_ORF18_W170A	cgctcttGCAggcagcagatgaacacgaac catcgtgcGTCaaaggcggagcagtagcc	<i>*b</i> <i>*b</i>
ph-mKGN-ORF18-G171A	S_17_KSHV_ORF18_G171A As_17_KSHV_ORF18_G171A	cgctcttttggGCAgcagatgaacacgaacg catcgtGGCcaaaaggcggagcagtag	<i>*b</i> <i>*b</i>

Plasmid name	Primer name	Primer sequences (5' -> 3')	
<i>[ORF18 ASC mutants]</i>			
ph-mKGN-ORF18_ASC1	S mm1 EcoRI KSHV_ORF18	catcgaattcgatgctcGCAaaatacgtgtgtagac	<i>*c</i>
ph-mKGN-ORF18_ASC2	S mm2 KSHV_ORF18	CAGCGcttagagagGCGCatggtgcgttttgca	<i>*b</i>
	As mm2 KSHV_ORF18	ctctcaagCGCTGcgtacggcgtcgtgctc	<i>*b</i>
ph-mKGN-ORF18_ASC3	S mm3 KSHV_ORF18	GCAaaataagaatGCAaatacaattcagcccaag	<i>*b</i>
	As mm3 KSHV_ORF18	TGCActctatttttTGCAaagcacaatgaagctc	<i>*b</i>
ph-mKGN-ORF18_ASC4	S mm4 KSHV_ORF18	caaGCAGCGCTttattcattttgcttc	<i>*b</i>
	As mm4 KSHV_ORF18	ACGGCGTGCgttcgtggaatgtatttaattc	<i>*b</i>
ph-mKGN-ORF18_ASC5	S mm5 KSHV_ORF18	GCAttggtGCTcgagagatgaacttc	<i>*b</i>
	As mm5 KSHV_ORF18	GGCAaccaaTGCaaataacgcagctcttgg	<i>*b</i>
ph-mKGN-ORF18_ASC6	S mm6 KSHV_ORF18	GCTgtgGCCGCTtaaggaggactactgc	<i>*b</i>
	As mm6 KSHV_ORF18	CGCGGCGcaAGCtagaccgaattgtacatc	<i>*b</i>
ph-mKGN-ORF18_ASC7	S mm7 KSHV_ORF18	GCAgtgGCTGCAGCGCTtttttaattctggg	<i>*b</i>
	As mm7 KSHV_ORF18	CAGCcaactGCTgttctggggcgatcgcg	<i>*b</i>
ph-mKGN-ORF18_ASC8	S mm8 KSHV_ORF18	GCGGCAAGCTctgggaatgtgtctccc	<i>*b</i>
	As mm8 KSHV_ORF18	gAGCTGCCGCAaatttccgtcacacagg	<i>*b</i>
ph-mKGN-ORF18_ASC9	S mm9 KSHV_ORF18	ggagacAGCTTGCAAGCattaaacaaaagttccg	<i>*b</i>
	As mm9 KSHV_ORF18	CAtgccGCTGctctctggtgcgggctac	<i>*b</i>
ph-mKGN-ORF18_ASC10	S mm10 KSHV_ORF18	CGCAGGcaTGcGgcacacttccagatttaac	<i>*b</i>
	As mm10 KSHV_ORF18	AGGTGgagcacaacgggacgcgc	<i>*b</i>
ph-mKGN-ORF18_ASC11	S mm11 KSHV_ORF18	CAGCTgecttGACAGCAgcagatgacacgaacg	<i>*b</i>
	As mm11 KSHV_ORF18	CaaagcgAGCTCGctagccgcgcgcagagag	<i>*b</i>
ph-mKGN-ORF18_ASC12	S mm12 KSHV_ORF18	CAcgtgcagGCTGCTcttctgcgcagagcttttc	<i>*b</i>
	As mm12 KSHV_ORF18	GCcaagctGCTggttctcgtctcccaaaaag	<i>*b</i>
ph-mKGN-ORF18_ASC13	S mm13 KSHV_ORF18	GCAattgtacGCGatagctcgggctgtttatg	<i>*b</i>
	As mm13 KSHV_ORF18	CGctagcaaaTGCAagctctgcgcgaagag	<i>*b</i>
ph-mKGN-ORF18_ASC14	S mm14 KSHV_ORF18	ccGCAcgtGCTatgccaagaggtctctg	<i>*b</i>
	As mm14 KSHV_ORF18	AGCAcgtTGCggagactcagtgacaaatg	<i>*b</i>
<i>[ORF18 truncated mutants]</i>			
ph-mKGN-ORF18A1	S EcoRI_d1_KSHV-ORF18	gccacGAATTCgtggcgctttttcaaaatagaatttaataac	<i>*a</i>
ph-mKGN-ORF18A2	S_d2_KSHV-ORF18-2	GAGGCTCTATGcaattggttctcagaggttac	<i>*d</i>
	As_d2_KSHV-ORF18-2	gaacaaatgCATGAGGCTCTCTTAAGCCCC	<i>*d</i>
ph-mKGN-ORF18A3	S_d3_KSHV-ORF18	GCTTTTATTCaagccaatgcgggagactac	<i>*d</i>
	As_d3_KSHV-ORF18	caattgcagGATAAAACGACAGCTCTTTGGGC	<i>*d</i>
ph-mKGN-ORF18A4	S_d4_KSHV-ORF18	GTTAAAGGGAGGCTTgaaagctgtgtatgatgg	<i>*d</i>
	As_d4_KSHV-ORF18	cgagcttcaAGCCTCCCTTAACAGGTACAC	<i>*d</i>
ph-mKGN-ORF18A5	S_d5_KSHV-ORF18	AGGTGgagcacaacgggacgcgc	<i>*d</i>
	As_d5_KSHV-ORF18	ggtagctcCACCTCCGACGGAACCTTG	<i>*d</i>
ph-mKGN-ORF18A6	S_d6_KSHV-ORF18	GCTGCTGTGTggtgcgaattatctgttcaac	<i>*d</i>
	As_d6_KSHV-ORF18	gataaattgcagACACAGCAGCATCTCGCTG	<i>*d</i>
ph-mKGN-ORF18A7	S_d7_KSHV-ORF18	CTTTGAGGGGCAacgtgtggaacgaacttttg	<i>*d</i>
	As_d7_KSHV-ORF18	caaggtGCCCCCTCAAGAGCCCCAC	<i>*d</i>
ph-mKGN-ORF18A8	S_d8_KSHV-ORF18-2	CCCCGGAAActgcggcgctactg	<i>*d</i>
	As_d8_KSHV-ORF18-2	ccgcaagTTCCGGGGCGATGCGAAC	<i>*d</i>
ph-mKGN-ORF18A9	S_d9_KSHV-ORF18	CAGAGTGCTCTcttctgcacaaagctttc	<i>*d</i>
	As_d9_KSHV-ORF18	gcgaagaagAGGACGCTGCAGGGGAG	<i>*d</i>
ph-mKGN-ORF18A10	S_d10_KSHV-ORF18	CTGGGTGGCTAggtgtctgtatgttgggc	<i>*d</i>
	As_d10_KSHV-ORF18	cagaagactACGCCACCCAGCGCTTCGTG	<i>*d</i>
ph-mKGN-ORF18A11	S_d11_KSHV-ORF18	CTTATGCCACAGcgcgaatccgcctcatgtac	<i>*d</i>
	As_d11_KSHV-ORF18	gatgtgcgGCTGTGGCATAAGACGCC	<i>*d</i>
ph-mKGN-ORF18A12	S_d12_KSHV-ORF18	GCACTCTGTcgtgcgaagcgcgcgc	<i>*d</i>
	As_d12_KSHV-ORF18	cttcggagagACAGACTGGCTCCACCG	<i>*d</i>
ph-mKGN-ORF18A13	As(stp)_NotI_d13_KSHV-ORF18	taaGCGGCGCGTaaagcggaataacccg	<i>*a</i>
<i>[Sub-cloning into 2xS tag expression plasmids]</i>			
pCIneo-2xS-ORF18 WT/mut	S_EcoRI_ORF18 As_ORF18(stp)_MluI	catGAATTCatgcctggaataactggtgtggtg taaACCGCTTaaacgcgtgtgtgttaaac	<i>*a</i> <i>*a</i>

*\*a* : Uppercase indicates restriction enzyme site

*\*b* : Uppercase indicates mutagenesis sites

*\*c* : Uppercase indicates mutagenesis sites, underlined lowercase indicates restriction enzyme site

*\*d* : Uppercase indicates N-terminal protein coding sequences, lowercase indicates C-terminal protein coding sequences