

# Staphylococcal *saoABC* operon codes for a DNA-binding protein SaoC implicated in the response to nutrient deficit

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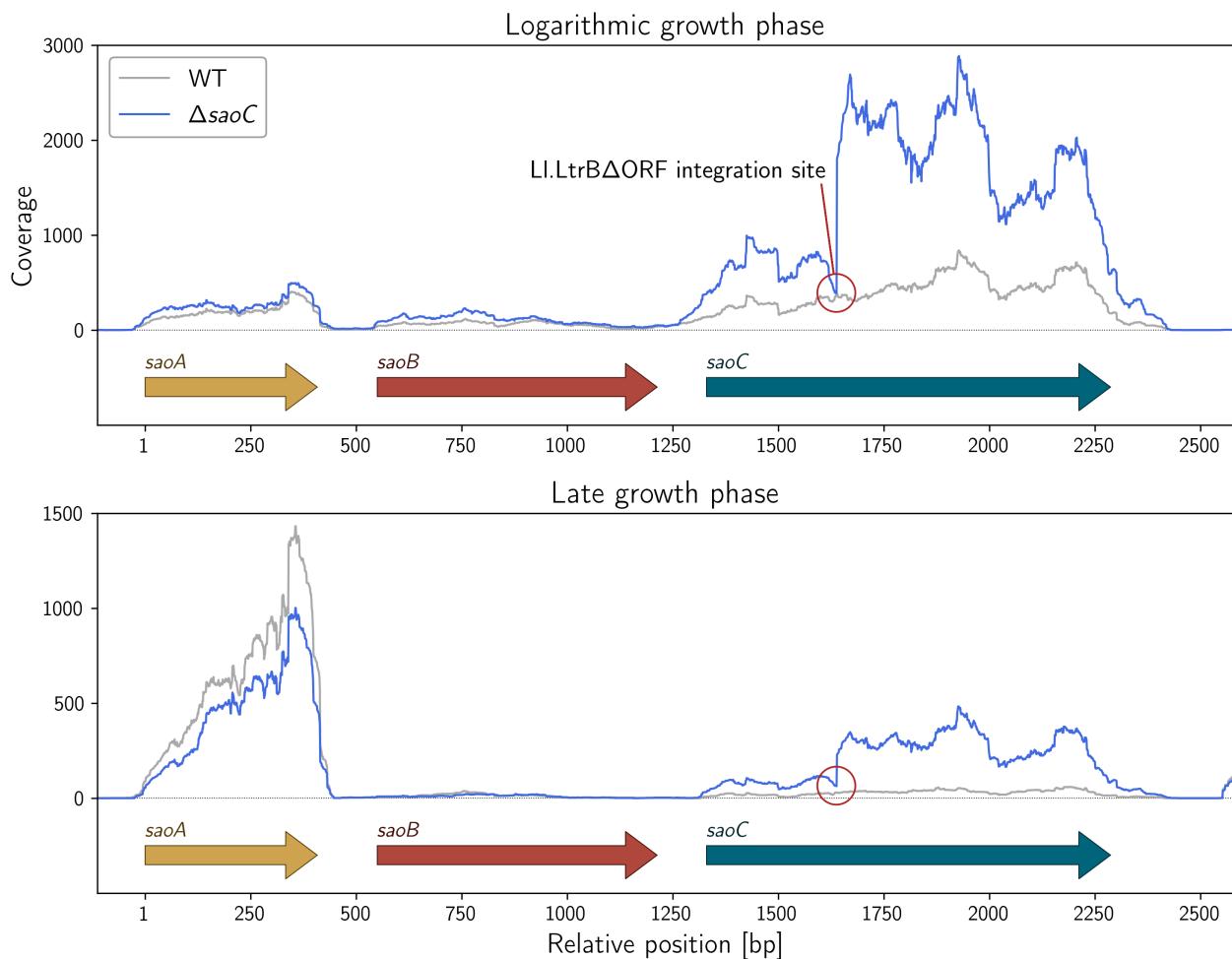
**List S1.** Occurrence of *saoABC* operon among staphylococcal species. The *saoABC* operon has been found in 47 out of 51 staphylococcal species. Negative species are underlined. It should be noted that in case of 4 species, in genomes of which we were unable to find the operon, the number of available genomic sequences was low (<10). Results for these species might be thus unreliable and in the future *saoABC* operon will likely be found in those species as well.

- |                            |                            |                                 |                               |
|----------------------------|----------------------------|---------------------------------|-------------------------------|
| 1. <i>S. agnetis</i>       | 14. <i>S. delphini</i>     | 27. <i>S. lentus</i>            | 40. <i>S. saccharolyticus</i> |
| 2. <i>S. argensis</i>      | 15. <i>S. devriesei</i>    | 28. <i>S. lugdunensis</i>       | 41. <i>S. saprophyticus</i>   |
| 3. <i>S. argenteus</i>     | 16. <i>S. edaphicus</i>    | 29. <i>S. lutrae</i>            | 42. <i>S. schleiferi</i>      |
| 4. <i>S. arlettae</i>      | 17. <i>S. epidermidis</i>  | 30. <i>S. massiliensis</i>      | 43. <i>S. schweitzeri</i>     |
| 5. <i>S. aureus</i>        | 18. <i>S. equorum</i>      | 31. <i>S. microti</i>           | 44. <i>S. sciuri</i>          |
| 6. <i>S. auricularis</i>   | 19. <i>S. felis</i>        | 32. <i>S. muscae</i>            | 45. <i>S. simiae</i>          |
| 7. <i>S. capitis</i>       | 20. <i>S. fleurettii</i>   | 33. <i>S. nepalensis</i>        | 46. <i>S. simulans</i>        |
| 8. <i>S. caprae</i>        | 21. <i>S. gallinarum</i>   | 34. <i>S. pasteuri</i>          | 47. <i>S. stepanovicii</i>    |
| 9. <i>S. carnosus</i>      | 22. <i>S. haemolyticus</i> | 35. <i>S. petrasii</i>          | 48. <i>S. succinus</i>        |
| 10. <i>S. chromogenes</i>  | 23. <i>S. hominis</i>      | 36. <i>S. pettenkoferi</i>      | 49. <i>S. vitulinus</i>       |
| 11. <i>S. cohnii</i>       | 24. <i>S. hyicus</i>       | 37. <i>S. piscifermentans</i>   | 50. <i>S. warneri</i>         |
| 12. <i>S. condimenti</i>   | 25. <i>S. intermedius</i>  | 38. <i>S. pseudointermedius</i> | 51. <i>S. xylosus</i>         |
| 13. <i>S. cornubiensis</i> | 26. <i>S. kloosii</i>      | 39. <i>S. rostri</i>            |                               |

**Table S1.** Primers used in the study. Restriction sites are double-underlined, start and stop codons as well as recombinase attachment sites (*attB1/attB2*) are in bold, ribosome-binding sites (RBSs) are underlined, group II Ll.LtrB-ΔORF intron target sequences are in bold and underlined.

Name	Sequence	Use
pD-saoC-F	ATAC <u><u>ATATGTTAACTAAAGAATTGCACAAC</u></u> G	pETDuet-saoC (NdeI/XhoI)
pD-saoC-R	AGACT <u><u>CGAGTTACAGGTTGAATAAACG</u></u>	
pC-saoB-F	<b>CCGG<u><u>TGACTAACTAACTAATGGGAGGTGT</u></u></b> TCTTATGG	pCN51-saoB (SalI/BamHI)
pC-saoB-R	<b>CGCG<u><u>GATCCTAAAATGGTTACGTAAATCC</u></u></b> ATTTC	
pC-saoC-F	<b>CCGG<u><u>TGACTAACTAACTAATGCAGAAAGG</u></u></b> <u>ACTTAAAGTATG</u>	pCN51-saoC (SalI/BamHI)
pC-saoC-F	<b>CGCG<u><u>GATCCTACAGGTTGAATAAACGTGCG</u></u></b>	
pK-saoA-attB1-UF	<b>GGGG<u><u>AACAAGTTGTACAAAAAAGCAGGCT</u></u></b> TGATGCCCATTTGAAACTCTTTTC	pKOR-del-saoA ( <i>attB1/SacII/attB2</i> )
pK-saoA-UR	AGGT <u><u>CCCCCGCGGAGTTGTGCCCTTAAAAA</u></u> ATTTG	
pK-saoA-DF	AGGT <u><u>CCCCCGCGGTATCACACTGAATTCAA</u></u> ATGAGATTAC	
pK-saoA-attB2-DR	<b>GGGG<u><u>ACCACTTGTACAAGAAAGCTGGGT</u></u></b> TTCTCTAAATGTTGAACAAATTACGGAC	
pK-saoC-attB1-F	<b>GGGG<u><u>AACAAGTTGTACAAAAAAGCAGGCT</u></u></b> GTCAAGAACATAACTCAGCATATG	pKOR-del-saoC ( <i>attB1/SacII/attB2</i> )
pK-saoC-UR	AGGT <u><u>CCCCCGCGGACTTAAAGTCCTTCTGCA</u></u> TATAC	
pK-saoC-DF	AGGT <u><u>CCCCCGCGGACTTGTGATTGAACACAA</u></u> ATTTAATAC	
pK-saoC-attB2-R	<b>GGGG<u><u>ACCACTTGTACAAGAAAGCTGGGT</u></u></b> ATCAGTGTAAAGAAAAGCATTGAC	
pN-IBS1-saoB	AAAAAA <u><u>AGCTTATAATTATCCTTA<u><u>TATTACCAA</u></u></u> <u>ACCGTGC</u>CCCCAGATAGGGTG</u>	pNL9164-saoB
pN-EBS1-saoB	CAGATTGTACAAATGTGGTGATAACAGATAAG <b>T<u><u>CCAACCTT</u></u></b> TAACCTACCTTCTTG	
pN-EBS2-saoB	TGAACGCAAGTTCTAATT <u><u>TCGGTT<u><u>TAATATC</u></u></u> GATAGAGGAAAGTGTCT</u>	
pN-IBS1-saoC	AAAAAA <u><u>AGCTTATAATTATCCTTA<u><u>GTAGACGA</u></u></u> <b>TT<u><u>TA</u></u></b>GTGC</u> CCCCAGATAGGGTG	pNL9164-saoC
pN-EBS1-saoC	CAGATTGTACAAATGTGGTGATAACAGATAAG <b>TC<u><u>GATTACG</u></u></b> TAACTACCTTCTTG	
pN-EBS2-saoC	TGAACGCAAGTTCTAATT <u><u>TCGATT<u><u>TCTACTC</u></u></u> GATAGAGGAAAGTGTCT</u>	

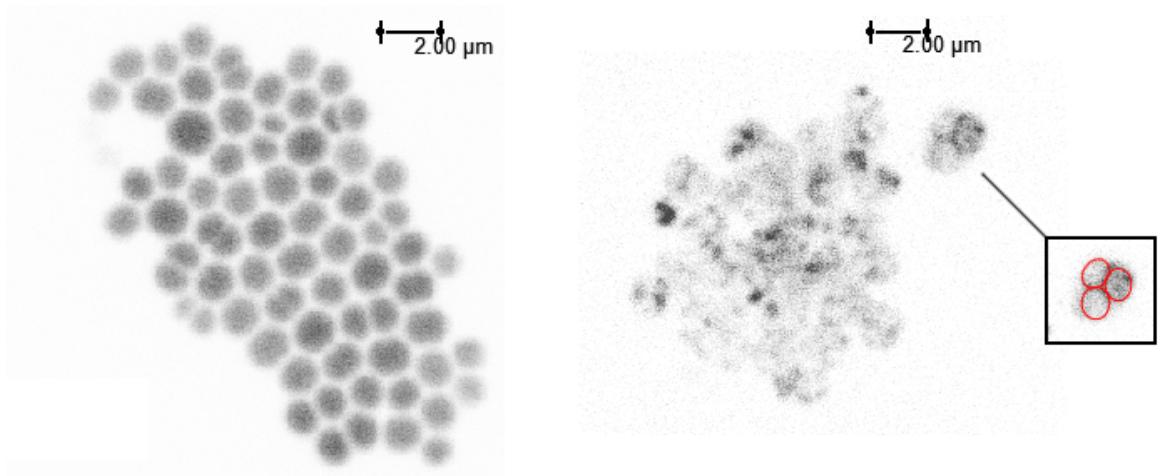
pC-P-saoABC-F	GCT <u>GCATGCATAACTGCTTAGGTAATCTT</u>	pCN35-P-saoABC-gfpmut2-term
pC-P-saoABC-R	AGA <u>GTGACCCAGGTGAATAAACGTG</u>	(PaeI/SalI/BamHI/EcoRI)
pC-gfpmut2-F	CAG <u>GGTCACAGTAAAGGAGAAGAACTTTTC</u>	
pC-gfpmut2-R	CGGG <u>GATCCTATTGTATAGTTCATCCATG</u>	
pC-term-F	AGAG <u>GATCCACTTGATTTAACACAAATT</u>	
pC-term-R	CCT <u>GAATTAGGAATACACAGGAGCCA</u>	
RT-saoA-F	TGTTAGTTGAGCGGTTCGTG	RT-qPCR
RT-saoA-R	GCTTGGA <u>CCCTTAGATGCT</u>	
RT-saoB-F	AGATTTAACACA <u>AGATAACGGCCTC</u>	
RT-saoB-R	GTGTGACC <u>CATCATTAGTTGGATGAA</u>	
RT-saoC-F	ATCAAC <u>ACAGACGCTAACAGCGGA</u>	
RT-saoC-R	GCT <u>CAACGCTGGCTTTCA</u>	
RT-gyrB-F	CAACAA <u>TGAACCCCTGAGCACC</u>	
RT-gyrB-R	CGGTTT <u>CTACAACGTCACCC</u>	
RT-23SRNA-F	CCGAAG <u>CTGTGGATTGTCCT</u>	
RT-23SRNA-R	GCT <u>ACTCACACCGGCATTCT</u>	
saoA-prom-F	AAGTT <u>CATCAAATAAACTGTGACCATC</u>	DNA pull-down
saoA-prom-R	TGTT <u>CACCATCTCTCCTGTTAAC</u>	
saoC-prom-F	TTC <u>CATCCAACTAATGATGGTCACAC</u>	
saoC-prom-R	TCTAATT <u>CATCAAACGCTAGATCATAAC</u>	
saoC-prom-fr1.2-F	ATT <u>TATGCTTATTAGAAGAAATGGATTACG</u>	
saoC-prom-fr1-R	TGAA <u>ACGTCAGTCTGCTTCATAC</u>	
saoC-prom-fr2-R	TATA <u>CTGACGTTGTCTACGGG</u>	
saoC-prom-fr3-F	TT <u>AAAATATGTATGAAGCAAGACTGAC</u>	
saoC-prom-fr3-R	CGT <u>GTGCAAATTCTTAGTTAACATAC</u>	
saoC-rand-prom-F	CAA <u>ATAATAGGTACAAATCTAAAGACGG</u>	Control for DNA pull-down
saoC-rand-prom-R1	CT <u>CTCACTATTTTATACGCCTTTATG</u>	
saoC-rand-prom-R2	T <u>GACTATAAGTAGCTAAACATAATGTATTG</u>	



**Figure S1.** Coverage plots of reads mapped specifically to *saoABC* operon. The reads were derived from all RNA-Seq replicas either for the logarithmic (upper plot) or the late phase of growth (lower plot). In both cases clearly elevated coverage of *saoC* (disrupted by LI.LtrB $\Delta$ ORF intron) might be observed for  $\Delta saoC$  mutant, which corroborate the claim that functional SaoC acts as a transcriptional repressor by binding to the conserved regulatory sequence upstream its own gene.

MbtS	1	-----MNLS-KQIKK---YRERDGYSQEYLAEKLYVSRQS-----	31
		: .        :   . :   .   .   .   :	
SaoC	1	MLTKEFAQRVELSEKQVRKIVQHLEERGYQ---LSKTEYRGREATDFKEE	47
MbtS	32	-----ISNWENDKSLPDIHN	46
		: .   :     :   . .	
SaoC	48	DIELFKDIADKVQQTNSYDLAFDELEKEKDFLQVIVKN--DDKNLPTNQN	95
MbtS	47	LLMMCELFNVTLDDLVKGTIPFVPDIKAQRSNLWTVVMLIFMTLAAILM	96
		: . . :	
SaoC	96	VAQL-----	99
MbtS	97	GPLVVYWNWTGVTVAIILGIGFYASMKIEDLK-----	129
		:       :	
SaoC	100	-----VEDLRLEIQKMREERHLLGQMM	121
MbtS	130	-Kvh-----	132
		:	
SaoC	122	NQVHQQQQELKELQNQLTSKIDSNSESLKAIQTSQEAQASQAKVLA	171
MbtS	133	-----KMDNYDRIVAFMNGKDPSEVQT-----	154
		.   : .   . .   :   . . . : .	
SaoC	172	ESTNKVEKNAVTEDKADSKDSKVAGVNTSTDAKTDTKADNAGDGTTCVD	221
MbtS	154	-----	154
SaoC	222	KEDQISATEAIEKASVEQSNGNAAETSNEATVDAEAQHDAEQQVAEAHA	271
MbtS	155	-----TKARNTMTNALSIIS-----VIGILSLI	177
		.     .   . .   :   : .   : .   : .   : .   : .   : .   :	
SaoC	272	EASKQATSNDSLEAKAENDSTASQSEMSEPKPQEEKKGFFARLFNL---	317
MbtS	178	IFLSVYLANKFL	189
SaoC	317	-----	317

**Figure S2.** Sequence alignment of MbtS, a transcription factor recently described for *S. aureus* [1], and SaoC. Sequence similarity between these two proteins is limited.



**Figure S3.** Confocal microscopy picture of *S. aureus* RN4220 transformed with pCN68 for free GFP expression (left) and pCN35-P-saoABC-gfpmut2-term carrying a copy of complete *saoABC* operon with *gfpmut-2* sequence insert for expression of SaoC-GFP fusion protein (right). In the latter case fluorescence signal forms ring-shaped halos, which indicates peripheral localisation of SaoC-GFP, likely at the inner surface of cell membrane.

CAAATAATAGGTACAAATCTAAAGACGGTAAGTGCTAGCCAACCTTGTATGCATTGT  
AATAACTCGTAAATAATAATTAACAAATACATTATGTTAGCTACTTATAGTCAC  
CATAGCAAATAGTCGCTTCTAAGATAATCCACCAAATTATTACTTATAGGTACATTGT  
AAGCTAGTCCGAGGAGTTAAAAAGAGTGTGCCATGATTAGGGATTGTAACCAGTCA  
AAGGTCAACTTCTTATATGGATTGAAACAAGAAATAGTGTTCAGTTCTCAATA  
CGTCATAGTTGATGTGCTATCTCAATTCACAGTATCTATTCTGAATTGGACATGGCTG  
AAGAAGGGAAAAGTTGAGGGTAGTCACAAAGTTAAAGAATATTACATGGAATGCCAGA  
ATGAACATAAAATCATAGGCTCCTTATGAAGTATCCCTCAGCGAACTGTCACACGGTAAC  
TAAAGAGAAGAATGATCATAAAAAGGCGTATAAAAATAGTGAGAGT

**Sequence S1.** Shuffled *saoC* promoter sequence (*saoC*-rand-prom) used as a control for DNA pull-down experiments.  
Hybridisation sites for primers are underlined.

## References

1. Yeo, W.S.; Anokwute, C.; Marcadis, P.; Levitan, M.; Ahmed, M.; Bae, Y.; Kim, K.; Kostrominova, T.; Liu, Q.; Bae, T. A Membrane-Bound Transcription Factor Is Proteolytically Regulated by the Aaa+ Protease Ftsh in *Staphylococcus Aureus*. *J. Bacteriol.* **2020**, *202*, e00019-20. <https://doi.org/10.1128/JB.00019-20>.