

**Supplementary Figure 1.** Sequences of proteins described in this work. The color code is the same used in Figure 1. Catalytic domains are denoted in bold. The HA epitope tag is indicated in dark blue background.

#### Ppz1-HA

MGNSSSKSSKKDSHSNSSSRNPRQVSRTETSHSVKSAKSNKSSRSRSLPSSSTTNTNSNVPD  
PSTPSKPNLEVNHQRHSSHTNRYHFPSSSSHSHSNSQNELLTTPSSSSTKRPSTSRRSSYNTKAA  
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TAYSTPLNSPGLSKLTDHSGEYFTSNSTSSLNHHSSRDIYPSKHISNDDDIENSSQLSNIHASM  
ENVNDKNNNITDSKKDPNEEFNDIMQSSGNKNAPKKFKKPIDIDETIQKLLDAGYAAKRTKNVC  
LKNNEILQICIKAREIFLSQPSLLELSPPVKIVGDVHGQYGDLLRLFTKCGFPPSSNYLFLGDY  
VDRGKQSLETILLLLFCYKIKYPENFFLLRGNHECANVTRVYGFYDECKRRCNIKIWKTFIDTFN  
TLPLAAIVAGKIFCVHGGLSPLVNSMDEIRHVVRPTDVPDFGLINDLLWSDPTDSPNEWEDNER  
GVSFCYNKVAINKFLNKFGLVLCRAHMVVEDGYEFFNDRSLVTVFSAPNYCGEFDNWGAVMSV  
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#### Ppz2-HA

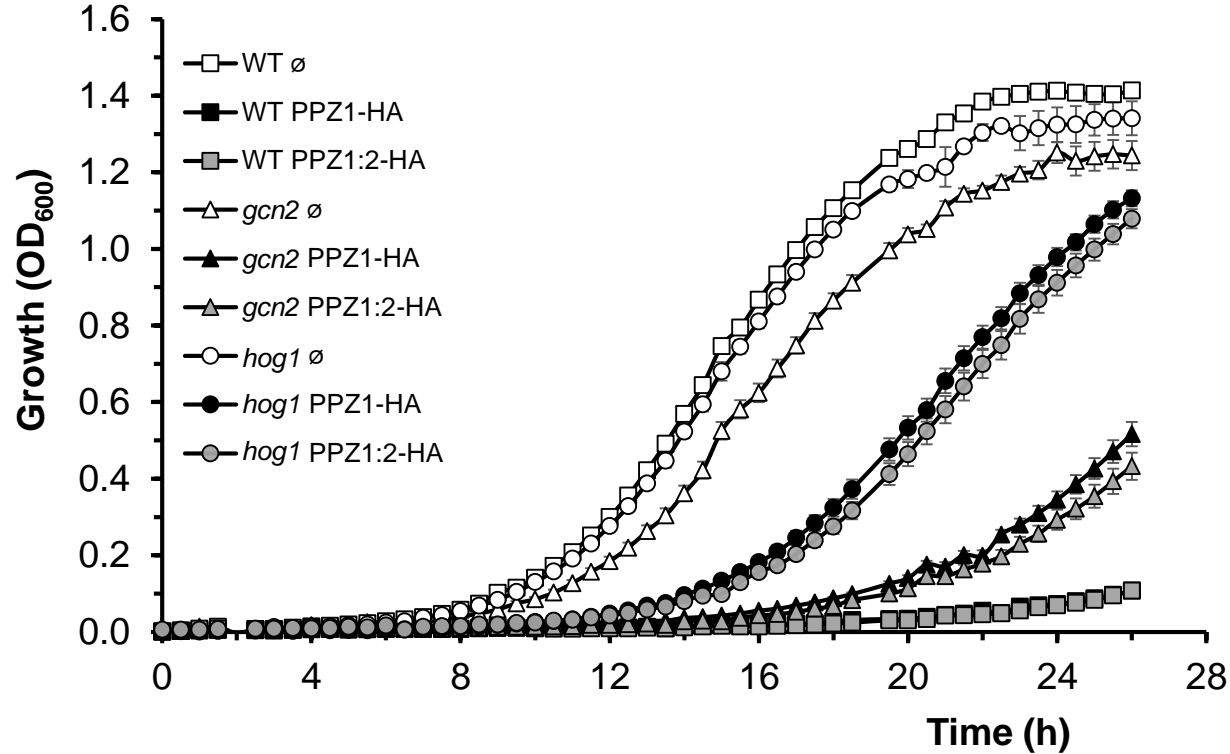
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SKRTPLRRHNSLQPEKGVTFGSSTSSKLRRRSNTLPASYPLNAEAGNGSDYFSNRNSHASS  
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YLSAYPTLAHRDSSSSLSPRGKGQRSSSSSSSSQRIYVSPSPPTGDFVHGSCADGDNGSRTNTM  
VEMKRKKPVRPVDIDEIIQRLLDAGYAAKRTKNVCLKNSEIIQICHKARELFLAQPALLELSPS  
VKIVGDVHGQYADLLRLFTKCGFPPMANYLFLGDYVDRGKQSLETILLLLCYKIKYPENFFLLR  
GNHECANVTRVYGFYDECKRRCNIKIWKTFVDTFNTLPLAAIVTGKIFCVHGGLSPLVNSMDEI  
RHVSRPTDVPDFGLINDLLWSDPTDSSNEWEDNERGVSFYCNKVAINKFLNKFGLVLCRAHMV  
VEDGYEFFNDRSLVTVFSAPNYCGEFDNWGAVMTVSEGLLCSFELLDPLDSTALKQVMKKGRQE  
RKLANRYPYDVPDYA\*

#### Ppz1:2-HA

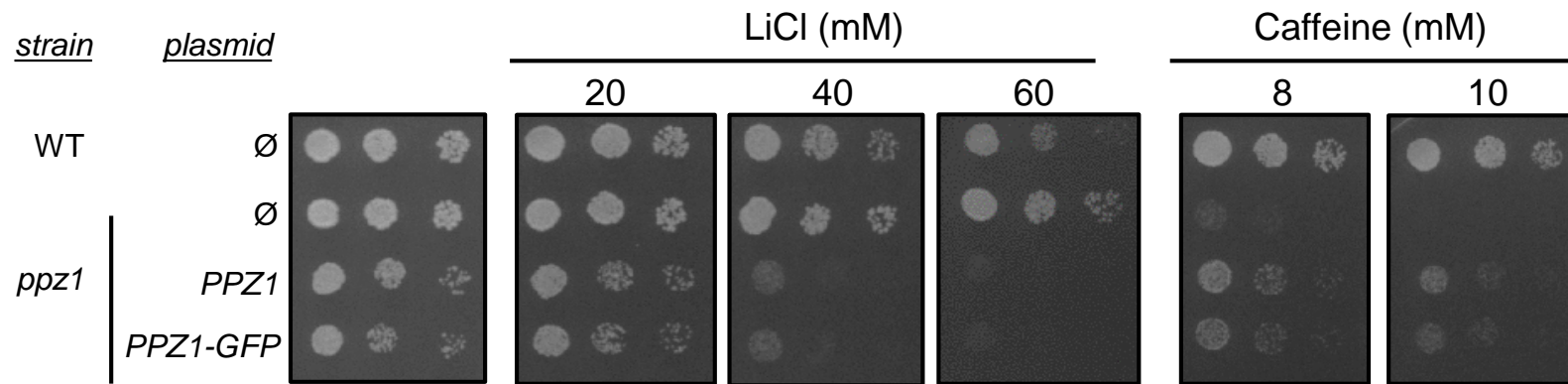
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TLPLAAIVTGKIFCVHGGLSPLVNSMDEIRHVSRTDVPDFGLINDLLWSDPTDSSNEWEDNER  
GVSFYCNKVAINKFLNKFGLVLCRAHMVVEDGYEFFNDRSLVTVFSAPNYCGEFDNWGAVMTV  
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## Ppz2:1

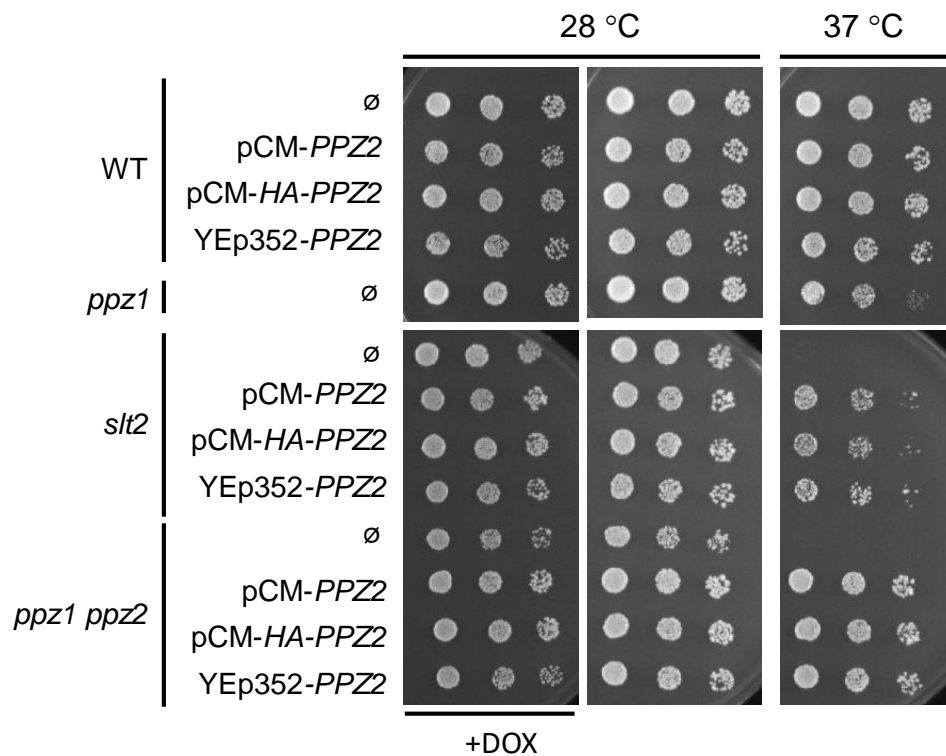
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SKRTPLRRHNSLQPEKGVTFGSSTSSKLRRRSDNTLPASYPLNAEAGGNGSDYFSNRSNSHASS  
RKSSFSGSTGNTAYSTPLHSPALRKMSRDNDSDGDNVNGRGTSPIPNLNIDKPPSPASSASKRE  
YLSAYPTLAHRDSSSSLSPRGKGQRSSSSSSSSSQRIYVSPSPPTGDFVHGSCADGDNGSRTNTM  
SGNKNAPKKFKKPIDIDETIQKLLDAGYAAKRTKNVCLKNNEILQICIKAREIFLSQPSLLELS  
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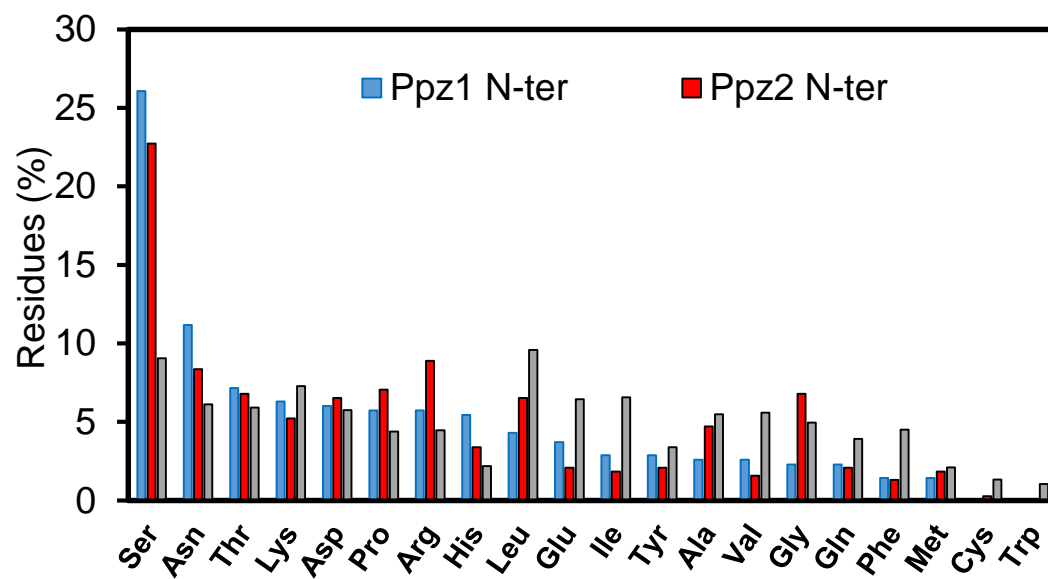
**Figure S2. Growth profiles of cells overexpressing Ppz1-HA and Ppz1:2-HA and carrying the *gcn2* or the *hog1* mutations.** BY4741 cells (WT) and its *gcn2* and *hog1* derivatives were transformed with the indicated pCM190-derived plasmids and grown in liquid medium as described in Material and Methods (except initial OD was 0.002). The OD<sub>600</sub> was determined every 30 min. Data is presented as the mean  $\pm$  SEM of three independent experiments made by triplicate.



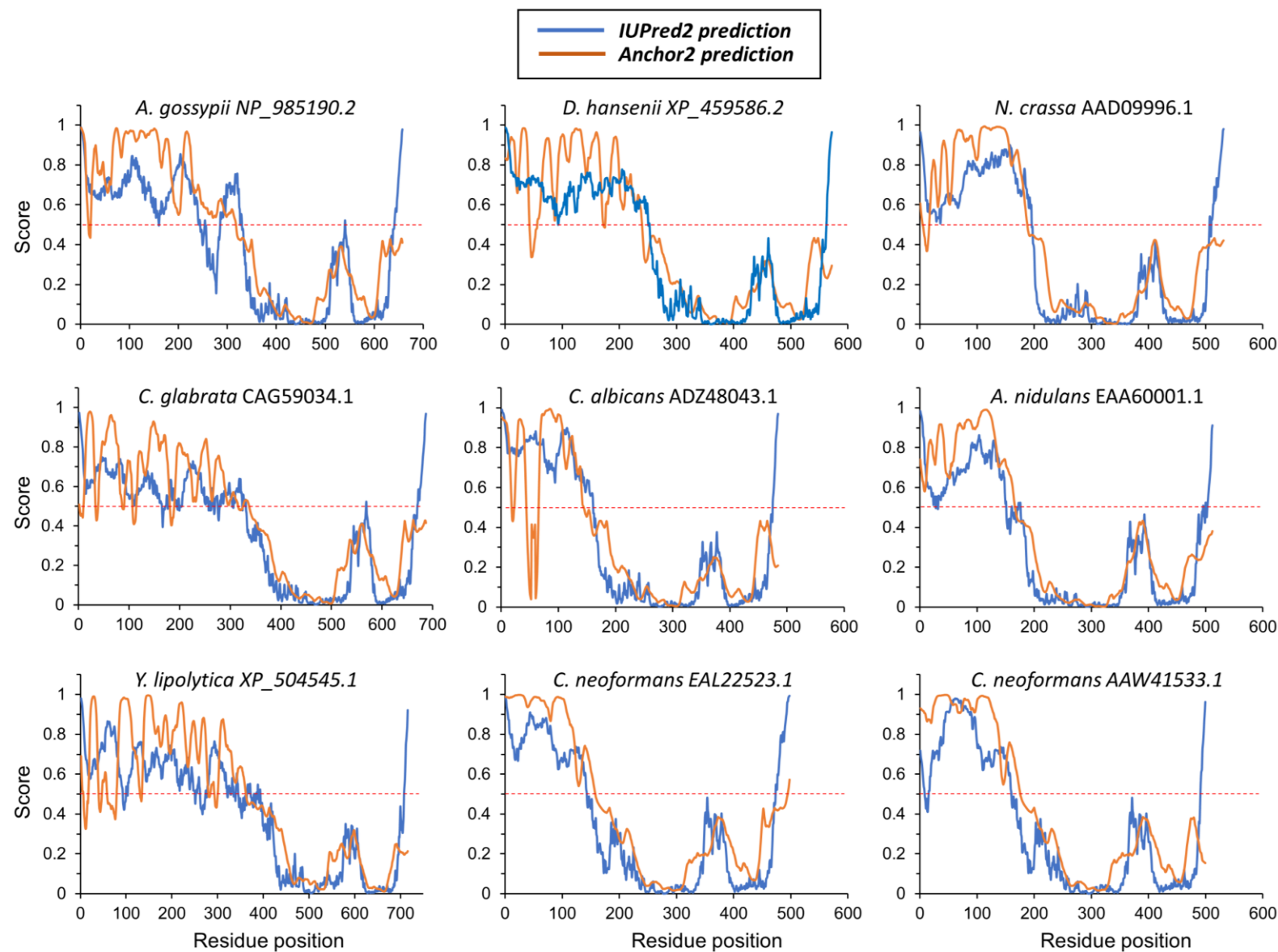
**Figure S3. Functional analysis of the C-terminally GFP-tagged Ppz1 phosphatase.** The indicated plasmids (YEplac195-based) were introduced in the wild type (WT) DBY746 strain and its isogenic *ppz1* derivative and tested on SC (lacking uracil) plates for tolerance to LiCl and caffeine. Plates were incubated for 3 days. Ø, empty YEplac195 plasmid.



**Figure S4. Functional analysis of the N-terminally HA-tagged Ppz2 phosphatase.** The indicated plasmids (pCM is pCM190) were introduced in the wild type (WT) BY4741 strain and its isogenic derivatives and tested for tolerance to high temperature. Plates were incubated for 3 days. Ø, empty pCM190 plasmid .



**Figure S5.** Percent aminoacidic composition of the N-terminal region of Ppz1 (349 residues) and Ppz2 (383 residues) in comparison with the whole yeast proteome (grey bars).



**Figure S6.** Prediction of Intrinsically Disordered (IDRs) and folding upon binding regions for Ppz phosphatases from diverse fungi. A) Prediction of IDRs according the IUPred2 software. B) Prediction of disordered binding regions based in the Anchor2 software. The 0.5 cut-off (discontinuous line) corresponds to 5% false positive prediction on IDRs or ordered protein segments.