



Review

The Non-Coding RNA *GAS5* and Its Role in Tumor Therapy-Induced Resistance

George I. Lambrou ^{1,*} , Kyriaki Hatziagiapiou ¹ and Apostolos Zaravinos ^{2,*}

¹ Choremeio Research Laboratory, First Department of Pediatrics, National and Kapodistrian University of Athens, Thivon & Levadeias 8, 11527 Goudi, Athens, Greece; khatziag@med.uoa.gr

² Department of Basic Medical Sciences, College of Medicine, Member of QU Health, Qatar University, 2713 Doha, Qatar

* Correspondence: glamprou@med.uoa.gr (G.I.L.); azaravinos@qu.edu.qa (A.Z.);
Tel.: +30-210-7467427 (G.I.L.); +974-4403-7819 (A.Z.)

Received: 28 September 2020; Accepted: 13 October 2020; Published: 15 October 2020



Abstract: The growth arrest-specific transcript 5 (*GAS5*) is a >200-nt lncRNA molecule that regulates several cellular functions, including proliferation, apoptosis, invasion and metastasis, across different types of human cancers. Here, we reviewed the current literature on the expression of *GAS5* in leukemia, cervical, breast, ovarian, prostate, urinary bladder, lung, gastric, colorectal, liver, osteosarcoma and brain cancers, as well as its interaction with various miRNAs and its effect on therapy-related resistance in these malignancies. The general consensus is that *GAS5* acts as a tumor suppressor across different tumor types and that its up-regulation results in tumor sensitization to chemotherapy or radiotherapy. *GAS5* seems to play a previously unappreciated, but significant role in tumor therapy-induced resistance.

Keywords: *GAS5*; malignancy; proliferation; invasion; metastasis; tumor growth

1. Introduction

The massive and rapid increase in the amount of human genome-scale DNA sequencing and the parallel development of methods to exploit these data drive the biomedical research today in a significant transition. The three billion base pairs of human DNA do not provide information about the function of the genes, or how cells grow, divide, form organisms, how mistakes in them are reflected in diseases, and how to develop a drug. Thus, it is important to construct a catalogue of expressed or suppressed genes for each cellular function, in order to understand how each component works within living cells. The transcriptome, i.e., the genes that are transcribed into mRNA, determines the phenotype and function of each cell [1]. Thus, alterations in gene expression are highly dynamic; they drive cellular phenotypic characteristics, DNA replication and cell division, as well as how a cell responds to an extracellular stimulus or perturbation. The understanding of when, where and to what extent a gene is expressed, can elucidate the regulatory mechanisms and biological pathways that lead to, maintain or reverse multiple drug-resistance in cancer treatment. Almost twenty years ago, the non-coding RNAs (ncRNAs) were discovered [2,3] and provided a breakthrough in our understanding of the functionality of the human genome. To take advantage of the large and rapidly increasing body of genome-scale sequence information, new technologies are required to exploit this information by characterizing biological processes and by studying the synchronous expression of a high number of genes. A variety of techniques has evolved to monitor, rapidly and efficiently, the transcript abundance of all genes in an organism [4].

Thousands of genes have been identified through high throughput methodologies, and a plethora of them have been studied for their role in tumor progression, as well as therapy-induced resistance.

2. The Long Non-Coding RNA Repressor *GAS5*

The ncRNAs are essential players in many cellular processes, from normal development to oncogenic transformation, offering an additional level of regulatory complexity in the transcription of mammalian genes [5–7]. They can be divided into microRNAs (miRNAs), piwi-interacting (piRNAs), small nucleolar (snoRNAs), long non-coding (lncRNAs) and other types of ncRNAs [7,8] and are implicated in various aspects of growth, such as neuronal, muscle and germline development [9–11]. One such lncRNA is the growth arrest-specific 5 (*GAS5*), which was originally found to accumulate in growth-arrested cells, acting as a decoy hormone response element for the glucocorticoid receptor (GR) and hence, blocking the upregulation of gene expression by activated GR [12–14] (Figure 1). *GAS5* has a 5' upstream oligopyrimidine tract sequence (5'TOP class genes) [15,16]. Serum starvation or treatment with inhibitors of protein translation can attenuate the translation of these 5'TOP RNAs and affect their degradation [17], leading to high numbers of spliced, mature *GAS5* RNA molecules [16].

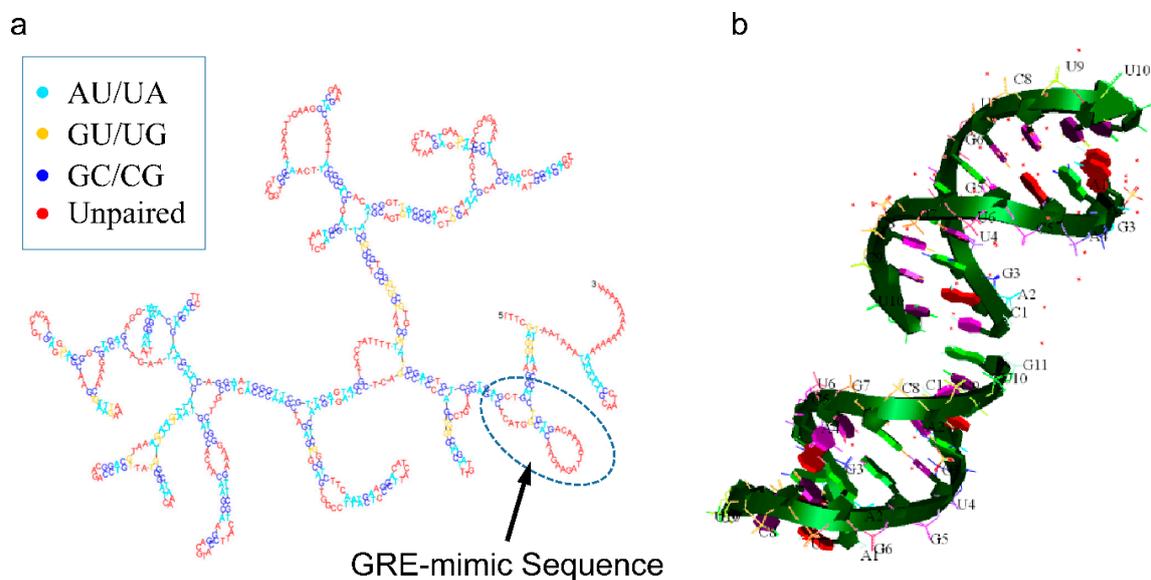


Figure 1. Secondary structure of the growth arrest-specific 5 (*GAS5*) RNA, showing the glucocorticoid receptor element (GRE)-mimic sequence (a), along with the 3D structure of the GRE-mimic sequence (b) (the 3D structure of the GRE-mimic was obtained from the Protein Data Bank with reference no. 4MCE [18]).

The regulation of the GR function is a complicated process, still unknown to a great extent. The role of *GAS5* was previously reported in childhood obesity, where it was shown to act as a regulatory repressor element of the GR [13]. In addition, *GAS5* was shown to play a role in metabolic processes such as obesity, anorexia or overweight situations. Interestingly, its expression in *in vivo* samples was influenced by methylation differences on its promoter region. If metabolic disorders are affected by gene expression and regulation, then, in the short-term, this should be evident in GR-related genes and regulatory elements such as *GAS5*. On the other hand, in the long-term perspective, metabolic disorders could be reflected in premature events, and be marked on the genome as methylated genes, as for example in infancy. These findings indicate that *GAS5* manifests multifaceted roles in various physiological processes, including tumor ontogenesis [19]. *GAS5* acts as a gene regulatory element through three basic modes of action. The first refers to its direct connection to its target gene, post-transcriptionally (Figure 2a). The second includes the indirect mechanism, which involves the binding of *GAS5* with a regulatory protein (e.g., GR) and the subsequent regulation of gene transcription (Figure 2b). There is also a secondary, indirect mode of action, which involves the formation of a *GAS5*/protein complex further acting as a regulatory element for the transcribed gene (Figure 2c).

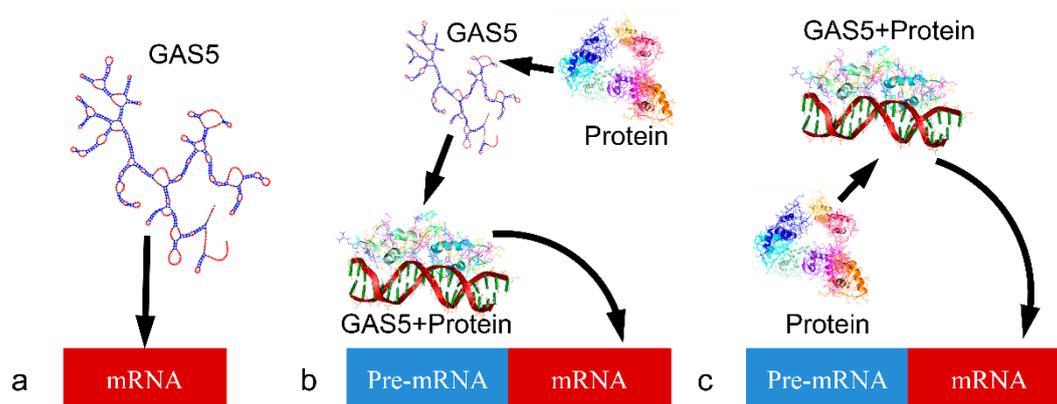


Figure 2. The three modes of action of *GAS5*. (b) *GAS5* acts directly on gene expression by regulating translation. (a) *GAS5* acts indirectly on gene expression. This includes the formation of a *GAS5*/protein complex regulating gene expression on the transcription level and finally again (c) indirectly through the formation of a *GAS5*/protein complex, which acts as a post-transcriptional regulatory mechanism. (The diagram was adopted from [20]. Exemplary molecules presented include the crystal structure of the DNA-free Glucocorticoid Receptor DNA Binding Domain with reference no. 6CFN [21], which is depicted as “Protein” and the *GAS5*/Protein complex is the crystallographic analysis of the interaction of the glucocorticoid receptor with DNA with reference no. 1R4R [22].)

The region of *GAS5* being responsible for binding to the GR and hence, crucial for its transcriptional repression, is enclosed between nucleotides 400 and 598. This portion of human *GAS5* contains two glucocorticoid response elements (GREs) at nucleotides 539–544 (GRE-1) and 553–559 (GRE-2), which double back and complement each other with a hairpin structure [23,24] (Figure 1a). These same GRE elements are preserved in mouse *Gas5*, which is the only other *Gas5* sequence available, although they share ~70% nucleotide homology in their exonic sequences [15,16]. In addition, a “mineralocorticoid response element” (MRE) is found at nucleotides 473–478, but it does not have a perfectly complementing sequence and, therefore, cannot form double-stranded RNA.

GAS5 shares common responsive sequences with other steroid receptors, such as the mineralocorticoid (MR), progesterone (PR) and androgen (AR) receptors, and can thus suppress their transcriptional activity in a ligand-dependent fashion. On the other hand, *GAS5* does not affect the transcriptional activity of the peroxisome proliferator activating receptor δ (PPAR δ) or p53. Likewise, *GAS5* functions as a general co-repressor of some steroid hormone receptors, repressing their transcriptional activity by binding to their DNA binding domain, as in the case of the transactivation domain (TAD) of VP16 fused with the DNA binding domain of GAL4 [13,25]. However, there is no single answer to the molecular mechanism of *GAS5* action, as differences have been found between rodent *Gas5* and human *GAS5* functions [26].

3. *GAS5* in Tumor Therapy-Related Resistance

The role of *GAS5* in cancer ontogenesis and progression is a relatively new subject of investigation. Yet, the role of *GAS5* in therapy-induced resistance observed across different types of tumors is important and needs to be further understood [13,16,27–29].

3.1. *GAS5* in Leukemia

GAS5 expression was recently found to be tightly linked to therapy progression in acute lymphoblastic leukemia (ALL) [30]. In the study of Gasic et al. [30], *GAS5* expression was reduced at day 33 of the induction therapy as compared to day 15, yet with still higher levels, compared to the time of diagnosis. This report suggested two interesting findings. The first was that *GAS5* expression was elevated due to treatment and the second, *GAS5* expression was low at diagnosis. At the same time, a recently discovered polymorphism in *GAS5* was found to be linked with poor prognosis in

acute myeloid leukemia (AML) patients [31]. The interesting finding was that *GAS5* molecules without the polymorphism, rs55829688 CC, were found to manifest higher expression levels in peripheral blood cells, compared to those that bared the polymorphism, rs55829688T [31]. However, in another report, it was shown that the down-regulation of *GAS5* led to the rescue of primary and malignant T-lymphocytes from the inhibition of the mammalian target of rapamycin (mTOR) [32]. In particular, this study showed that *GAS5* has tumor suppressor activity since it could suppress tumor growth, while, when silenced, tumor cells recovered and increased their proliferation rate [32] (Figure 3).

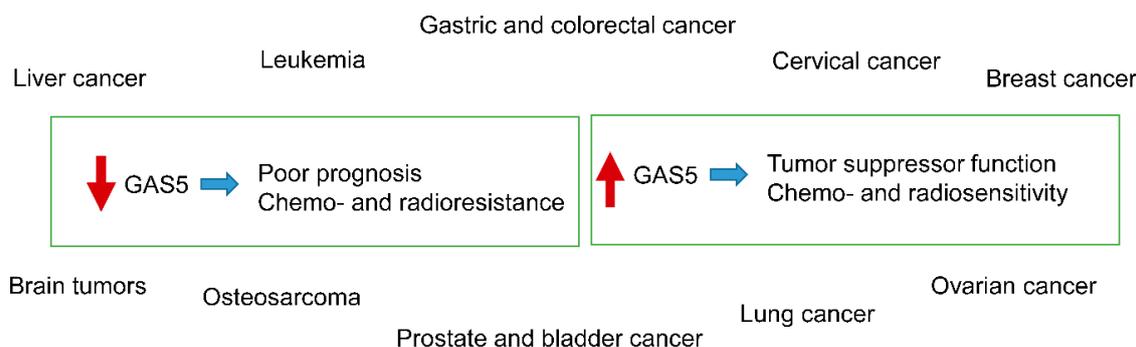


Figure 3. The action of *GAS5* on different tumor types. *GAS5* down-regulation or mutations are related to poor prognosis, as well as chemo- and radioresistance. On the other hand, the upregulated *GAS5* functions as a tumor suppressor and is related to chemo- and radiosensitivity.

3.2. *GAS5* in Cervical Cancer

Cervical cancer ranks second in women and is the fourth leading cause of deaths related to cancers. It can be very aggressive and, as such, it is still the subject of intense research. There are few reports regarding *GAS5* in cervical cancer. A recent study found that *GAS5* interacts with miR-106b and this complex inhibits the expression of the immediate early response 3 gene (*IER3*), leading to sensitivity to radiation therapy [33]. In another report, *GAS5* over-expression was shown to be connected to the down-regulation of miR-21 and the subsequent phosphorylation of *STAT3* and *E2F3* [33]. *GAS5* over-expression can also reduce the expression of two miR-21 targets: *TIMP3* and *PDCD4* [33]. All these events have been observed to lead to a G_0/G_1 arrest and enhancement of cisplatin-induced apoptosis [33]. Similarly, another study was in agreement with the findings by Gao et al. (2019), suggesting that *GAS5* negatively regulates miR-21 and upheaves cisplatin resistance [34] (Figure 3).

3.3. *GAS5* in Breast Cancer

Breast cancer is the most common type of malignancy in women and a leading cause of death. It is a complex, heterogeneous disease classified into hormone-receptor-positive, human epidermal growth factor receptor-2 overexpressing (HER2+) and triple-negative breast cancer (TNBC) based on histological features [35]. Although early diagnosis is of paramount importance for the treatment and prognosis of this tumor, there is still a lot to understand on the mechanisms of action of *GAS5* in it [36]. Overall, *GAS5* is also considered to function as a tumor suppressor in breast cancer [27,37–39]. This was also reported in a recent work, which indicated that *GAS5* is down-regulated in breast cancer and that it negatively impacts disease prognosis [33]. A way to alleviate *GAS5* down-regulation was proposed via the inhibition of the mTOR signaling pathway [40]. Interestingly, the magnitude of cell death, in vitro, was directly proportional to *GAS5* expression levels [40]. Finally, *GAS5* was able to promote apoptosis in estrogen receptor (ER)-positive cells and in the case of *GAS5* silenced cells, the inhibition of the PI3K/mTOR signaling pathway was able to recover *GAS5* expression [40]. This finding was quite interesting, because it postulated that in the case of low levels of *GAS5* expression, the inhibition of the mTOR pathway could be a complementary therapeutic target in the treatment of breast cancer. Furthermore, *GAS5* has been linked to trastuzumab resistance, which is a main obstacle

in HER2-positive breast cancer cells [41]. Li et al. (2016), showed that the down-regulation of *GAS5* is partly responsible for trastuzumab and lapatinib resistance. Both drugs interrupt the HER2/neu and EGFR pathways. In agreement with previous studies, it becomes evident that *GAS5* acts as tumor suppressor by interacting with miR-21 [41]. Tamoxifen is one of the basic chemotherapeutic agents in the treatment of breast cancer. In a recent report, it was shown that the down-regulation of *GAS5* is related to tamoxifen resistance. In particular, it was found that *GAS5* functions as a sponge for miR-222 suppressing PTEN expression and thus, inhibiting tamoxifen resistance. On the contrary, *GAS5* down-regulation functions reversely and tamoxifen resistance is promoted [42]. These findings are in agreement with a recent report where it was shown that *GAS5* is down-regulated in breast cancer tissues and linked to chemotherapy resistance [43]. Consequently, it seems that *GAS5* can be considered as new player in cancer ontogenesis, progression and prognosis, as well as it may have prognostic and therapeutic applications in this disease. Several new drugs have been designed and synthesized for the treatment of breast cancer (Figure 3).

3.4. *GAS5* in Ovarian Cancer

Ovarian cancer is a grave gynecological tumor and there are not many studies concerning the role of *GAS5* in it or its association with chemoresistance in this tumor. However, similar to other gynecological tumors, it seems that all studies converge to the conclusion that *GAS5* acts as a tumor suppressor in ovarian cancer, as well [44–47]. In a recent study, *GAS5* was found to be down-regulated in this disease [41]. In particular, in a meta-analysis of 561 microarrays and 136 RNA-seq specimens, *GAS5* was down-regulated and manifested high sensitivity and specificity in predicting platinum-based chemoresistance [48]. *GAS5* was also found to be down-regulated in epithelial ovarian cancer in another study, where it was related to disease prognosis, in particular [49]. At the same time, *GAS5* was found to be down-regulated in cisplatin resistant tumors. On the contrary, its up-regulation had the opposite effect, which significantly enhanced the sensitivity of ovarian cancer cells to cisplatin, both in vivo and in vitro. Further on, the up-regulation of *GAS5* was found to increase both the ratio of G₀/G₁ arrest and apoptosis in ovarian cancer [42]. The same study reported that a probable mechanism for *GAS5* action was mediated through the regulation of *PARP1* by recruiting the transcription factor E2F4 to its promoter and subsequent MAPK pathway activity [50] (Figure 3).

3.5. *GAS5* in Prostate and Bladder Cancers

Both prostate [51] and bladder cancers [52] are considered to be two major tumor types and causes of cancer-related deaths. In prostate cancer, *GAS5* was found to be down-regulated, while it was down-regulated in radio-resistant prostate tumor cells [53]. This effect, was found to be alleviated by the addition of α-Solanine, which up-regulates *GAS5* and at the same time, confers sensitivity to radiotherapy [53]. Similarly, *GAS5* was found to be down-regulated in transitional cell carcinomas of the urinary bladder, and its down-regulation was found to be positively correlated with higher pathological grades of the tumor [54]. However, in an in vitro system, *GAS5* overexpression could reduce chemo-resistance to doxorubicin and promoted apoptosis [54] (Figure 3).

3.6. *GAS5* in Lung Cancer

Lung cancer is the most common cause of death from tumor-related diseases [55]. A recent study reported that *GAS5* is down-regulated in lung cancer cells and at the same time, its knockdown increased cis-platin IC₅₀ in an in vitro system, while its overexpression decreased it [56]. In the same study, it was found that *GAS5* knockdown resulted in decreased autophagy in vitro, and therefore, resistance to cis-platin [56]. Similarly, *GAS5* up-regulation was found to be a significant factor of inhibition of tumorigenesis and an enhancer of radiosensitivity [57]. In addition, the mechanism of enhancement of radiosensitivity was found to function via the suppression of miR-135b in non-small cell lung cancer cells [57]. Another recent study confirmed that *GAS5* plays a significant role in non-small cell lung cancer, participating in cis-platin resistance. Cao et al. reported that chemo-sensitivity is

modulated by the tumor suppressor PTEN [58]. In the same study, it was found that a significant low *GAS5* expression in non-small cell lung cancer patients was correlated with poorer prognosis. In an in vitro system, *GAS5* knockdown promoted cell viability and regulated chemo-sensitivity to cis-platin. The authors showed that *GAS5* competed with PTEN for miR-21 binding, indicating a strong evidence that *GAS5*/miR-21/PTEN interactions are significant in cis-platin sensitivity in non-small cell lung cancer cells [58]. Similarly, *GAS5* was found to bind miR-21 and miR-23a, at the same time up-regulating PTEN and inhibiting PI3K/Akt phosphorylation [59]. This mechanism was found to function as an angiogenesis inhibitor, signifying that *GAS5* could be targeted therapeutically in order to inhibit angiogenesis in non-small cell lung cancer [59] (Figure 3).

3.7. *GAS5* in Gastric and Colorectal Cancers

Gastric cancer is the fourth most common malignancy, and the second most common cause of cancer-related deaths in the world [60]. *GAS5* also plays a significant role as a tumor suppressor in gastric cancer [61–64]. A recent study highlighted the fact that *GAS5* expression was significantly down-regulated in gastric cancer tissues, and that it was down-regulated in adriamycin-resistant cells [65]. *GAS5* was also found to have higher levels of promoter methylation in SGC-7901 cells, conferring resistance to chemotherapy [65]. There are no reports on the role of *GAS5* in chemoresistance in colorectal cancer; yet, reports suggest that *GAS5* is responsible for tumor suppression, inhibition of proliferation, metastasis and invasion [66]. In addition, a recent study indicated that *GAS5* inhibits angiogenesis and metastasis in colorectal cancer by suppressing the Wnt/beta-catenin signaling pathway, which is dedicated to promoting cell invasion and migration in this type of tumor (Figure 3) [67]. Recently, a 5-bp indel polymorphism (rs145204276) was found in the *GAS5* promoter region and proposed to have a carcinogenic effect [68].

3.8. *GAS5* in Liver Cancer

Liver cancer, in particular, hepatocellular carcinoma, is predominately present in eastern Asia and its rates are increasing in the northern hemisphere [69]. Liver cancer has a very fast progressing time span posing a significant threat to life. There are no reports on the role of *GAS5* in chemosensitivity or chemoresistance in liver cancer. Yet, there are some interesting reports suggesting that *GAS5* plays a synergistic role in the anti-tumor action of flavonoids and phytochemicals. In particular, phytochemicals, such as curcumin, resveratrol, sulforaphane, berberine and gambogic acid, have all been examined for their connection with non-coding RNAs. *GAS5* was reported as one of the ncRNAs that is regulated by phytochemicals, which can synergistically affect tumor development and progression. When phytochemicals were administered in combination with chemotherapeutics, they were found to have an additive effect on the overexpression of *GAS5* and the sensitization of cancer cells to chemotherapy. Finally, a recent study showed that corylin, a flavonoid extracted from the plant *Psoralea corylifolia* L. (Fabaceae), suppresses tumor growth and progression [70]. The interesting finding was that corylin was found to exert such effects on tumor growth through activation of *GAS5* [70] (Figure 3).

3.9. *GAS5* in Brain Tumors

Brain tumors, or tumors of the central nervous system, along with their extreme diversity, present a special case of malignancy due to the anatomical position in which they are diagnosed. This point is further strengthened by the fact that in several tumors, either benign or extremely aggressive, surgical excision is a drastic solution towards therapy, while in the case of brain tumors, this is not always the case, or it is less feasible. There are no studies connecting *GAS5* to chemoresistance. In fact, there are very few studies on the role of *GAS5* in brain tumors, in general. Yet, all the present studies agree that *GAS5* functions as a tumor suppressor and inhibits tumor proliferation, invasion, metastasis and migration [71–75] (Figure 3).

3.10. *GAS5* in Osteosarcoma

Another tumor type that we investigated is osteosarcoma. Osteosarcoma is a rare malignancy of the childhood with an incidence of 4–5 new cases per million per year [76]. It is an aggressive malignant neoplasm that arises from primitive transformed cells of mesenchymal origin, exhibits osteoblastic differentiation and produces malignant osteoid [77]. There are no reports concerning the role of *GAS5* in osteosarcoma with respect to chemotherapy-related resistance. However, there are some reports referring to *GAS5* as a significant gene in the tumor's progression. In particular, miR-663a and *ZBTB7A* were found to protect osteosarcoma from endoplasmic reticulum stress-induced apoptosis, through the down-regulation of *GAS5* [78], while in a similar study the CtBP1-HDAC1/2-IRF1 transcriptional complex was also found to be down-regulated in osteosarcoma cells [79]. In addition, it was found that *GAS5* sponges miR-203a [80] and miR-221 [81], thus suppressing tumor growth and inhibiting tumor invasion (Figure 3).

4. The Special Case of *GAS5* and miRNAs

4.1. *GAS5* and miRNAs in Leukemia

The topic of *GAS5* and miRNAs could not escape the attention of the present work. There are very few reports on the connection between *GAS5* and miRNAs. In the case of leukemia, there is one report suggesting the interaction of *GAS5* with miR-222, since their expression is negatively correlated [82]. In this study, it was also found that *GAS5* over-expression was related to the inhibition of leukemic cells proliferation, the enhancement of leukemic cell apoptosis and the inhibition of tumor cell invasion [82] (Table 1).

4.2. *GAS5* and miRNAs in Cervical Cancer

Similarly, few reports are available on the connection of *GAS5* and miRNAs in cervical cancer. Yet, all studies agree that *GAS5* acts as a suppressor or “sponge” for oncogenic miRNAs, whereas its overexpression is closely related to tumor suppression and induction of therapy-related sensitivity. In particular, previous studies indicated that *GAS5* interacts with miR-222 [82], miR-106b [33], miR-135a [83], miR-21 [84] and miR-205 [34,85], conferring tumor suppressor properties and induction of sensitivity to chemo- and radiotherapy (Table 1).

4.3. *GAS5* and miRNAs in Breast Cancer

The association between *GAS5* and miRNAs has been widely studied in breast cancer. A recent study highlighted the role of *GAS5* in breast cancer and adriamycin resistance, through the gene's interaction with miR-221-3p [86]. Another report showed that *GAS5* manifested tumor suppressor effects and induced chemosensitivity to breast cancer cells by indirectly targeting the miR-378-5p/SUFU signaling pathway [87], as well as by competitively binding miR-196a-5p [87]. Additionally, *GAS5* appeared to be a direct target of miR-221/222, suppressing tumor proliferation and enhancing tumor cell apoptosis [88]. In another report, it was shown that *GAS5* stimulates autophagy through the miR-23a/ATG3 axis, where it acts as a miRNA sponge [89]. Interestingly, miR-21 also has an oncogenic role in breast cancer, where it induces chemo- and radiosensitivity [41] (Table 1).

Table 1. Summary of the relation of GAS5 and miRNAs in several tumors.

Tumor	miRNA	Relation between GAS5 and miRNA	GAS5 Effect on Tumor	Effect on Therapy-Related Resistance	Citation
Leukemia	miR-222	Direct Suppression	Tumor suppressor	Unknown	Jing et al. (2019) [82]
Cervical Cancer	miR-106b	Direct Suppression/Sponge	Tumor suppressor	Induces chemo- and radiosensitivity	Gao et al. (2019) [33]
	miR-135a	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Yan et al. (2020) [83]
	miR-21	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Yao et al. (2019), Li (2016) [41,84]
	miR-205	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Yang et al. (2017), Wen et al. (2017) [34,85]
	miR-221-3p	Direct Suppression/Sponge	Tumor suppressor	Induces chemo- and radiosensitivity	Chen et al. (2020) [86]
Breast Cancer	miR-378-5p	Indirect Suppression/Sponge	Tumor suppressor	Induces chemo- and radiosensitivity	Zheng et al. (2020) [87]
	miR-221/222	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Zong et al. (2019), Gu et al. (2018) [42,88]
	miR-23a	Direct Suppression/sponge	Tumor suppressor/induces autophagy	Unknown	Gu et al. (2018) [89]
	miR-196a-5p	Direct Suppression/sponge	Induces autophagy	Unknown	Li et al. (2018) [90]
	miR-196a-5p	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Zheng et al. (2020) [87]
Ovarian Cancer	miR-21	Direct Suppression/Sponge	Tumor suppressor	Induces chemo- and radiosensitivity	Li (2016) [41]
	miR-196a-5p	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Zhao et al. (2018) [46]
Prostate Cancer	miR-940	Indirect Suppression	Tumor suppressor	Unknown	Chen et al. (2017) [91]
	miR-18a	Indirect Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Yang et al. (2019) [53]

Table 1. Cont.

Tumor	miRNA	Relation between GAS5 and miRNA	GAS5 Effect on Tumor	Effect on Therapy-Related Resistance	Citation
Lung Cancer	miR-21	Indirect Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Chen et al. (2020) [92]
	miR-29-3p	Indirect Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Cheng et al. (2019) [59]
	miR-21	Indirect Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Cao et al. (2017) [58]
	miR-205	Direct Suppression	Tumor suppressor	Unknown	Dong et al. (2019) [93]
	miR-135b	Direct Suppression	Tumor suppressor	Induces chemo- and radiosensitivity	Xue et al. (2017) [57]
	miR-23a	Indirect Suppression	Tumor suppressor	Unknown	Mei et al. (2017) [94]
Gastric Cancer	miR-18a	Direct Suppression	Tumor suppressor	Unknown	Wei et al. (2020) [95]
	miR-106a-5p	Indirect Suppression	Tumor suppressor	Unknown	Dong et al. (2019) [63]
	miR-222	Direct Suppression/sponge	Tumor suppressor	Unknown	Li et al. (2017) [96]
Colorectal Cancer	miR-182-5p	Direct Suppression/sponge	Tumor suppressor	Induces chemo- and radiosensitivity	Cheng et al. (2018) [97]
	miR-221	Indirect Suppression	Tumor suppressor	Unknown	Liu et al. (2018) [98]
Liver Cancer	miR-222	Direct Suppression/sponge	Tumor suppressor	Induces chemo- and radiosensitivity	Zhao et al. (2020) [99]
	miR-21	Direct Suppression/sponge	Tumor suppressor	Unknown	Wang et al. (2018), Hu et al. (2016) [100,101]
	miR-544	Indirect Suppression	Tumor suppressor	Unknown	Fang et al. (2019) [102]
	miR-135b	Indirect Suppression	Tumor suppressor	Unknown	Yang et al. (2019) [103]
	miR-34a	Indirect Suppression	Tumor suppressor/sponge	Unknown	Toraih et al. (2018) [104]
Glioma	miR-106b	Indirect Suppression	Tumor suppressor/sponge	Unknown	Huang et al. (2020) [105]
	miR-18a-3p	Indirect Suppression	Tumor suppressor/sponge	Unknown	Liu et al. (2018) [73]
Osteo-sarcoma	miR-663a	Indirect Suppression	Tumor suppressor/sponge	Unknown	Zhang et al. (2019) [79]
	miR-203a	Indirect Suppression	Tumor suppressor/sponge	Unknown	Wang et al. (2018) [80]
	miR-221	Direct Suppression	Tumor suppressor/sponge	Unknown	Ye et al. (2017) [81]

4.4. *GAS5 and miRNAs in Ovarian Cancer*

Although ovarian cancer is very common in women, there is only one report investigating the association between miRNAs and *GAS5* in it. In this, the role of miR-196-5p in relation to *GAS5* was reported. *GAS5* down-regulation was found to be related to high miR-196-5p expression, which induced tumor cell proliferation and progression. Thus, *GAS5* up-regulation confers tumor cell proliferation inhibition [46] (Table 1).

4.5. *GAS5 and miRNAs in Prostate and Bladder Cancers*

In the case of prostate and bladder cancers, two reports highlighted the connection of *GAS5* with miRNAs. In particular, it was reported that *GAS5* is down-regulated due to its targeting from miR-940 [91]. The relation between *GAS5* and miR-940 was reported to be a possible prognostic factor. Finally, a recent study indicated that *GAS5* negatively regulates miR-18a and, thus, confers radiosensitivity in human prostate cells [53] (Table 1).

4.6. *GAS5 and miRNAs in Lung Cancer*

Several reports have also identified the connection between *GAS5* and miRNAs. A recent report showed that *GAS5* probably indirectly regulates miR-21, whereas its over-expression suppresses miR-21 expression and, hence, increases radiosensitivity of lung tumor cells [92]. Recently, it was also shown that miR-29-3p antagonizes *GAS5* for binding PTEN [59]. It was also reported that *GAS5* exosomes are the basic vehicle of transmission conferring tumor inhibition [59]. The connection of *GAS5*/PTEN and miRNAs is also stated to be of significance through the competitive binding with miR-21 [58]. Similarly, the role of *GAS5*/PTEN is also shown to be of significance in lung cell proliferation and metastasis in connection to miR-205 [93]. Another recent study suggested that *GAS5* directly binds and suppresses miR-135b, enhancing radiosensitivity [57]. Finally, a connection between *GAS5* and miR-23a has been reported, where miR-23a was found to suppress *GAS5* expression and enhance tumor cell proliferation and tumorigenesis [94] (Table 1).

4.7. *GAS5 and miRNAs in Gastric and Colorectal Cancers*

In gastric cancer, three miRNAs have been reported to relate to *GAS5*, miR-18a [95], miR-106a-5p [63] and miR-222 [96]. In the case of miR-18a, it was reported that *GAS5* directly binds to it, inhibiting tumor growth via the stimulation of the activity of natural killer (NK) cells [95]. On the other hand, *GAS5* functions as sponge for miR-106a-5p, inactivating the Akt/mTOR pathway [63]. Finally, miR-222 was reported to directly bind to *GAS5* similarly, as in all previous cases, suppressing tumor cell proliferation [96]. In colorectal cancer, two different miRNAs were reported, miR-182-5p [97] and miR-221 [98]. *GAS5* could directly bind to miR-182-5p and inhibit tumor cell proliferation through the miR-182-5p/FOXO3a axis [97]. Similarly, miR-221 is negatively regulated to *GAS5* expression. If overexpressed, *GAS5* can suppress miR-221 expression and subsequently inhibit tumor cell proliferation in colorectal cancer [98] (Table 1).

4.8. *GAS5 and miRNAs in Liver Cancer*

In the case of gastric cancer, five miRNAs have been reported to be related to *GAS5*, miR-222 [99], miR-21 [100,101], miR-544 [102], miR-135b [103] and miR-34a [104]. *GAS5* was shown to sensitize hepatocellular cancer cells to chemotherapy by sponging miR-222 [99]. Similarly, *GAS5* directly acts as a sponge for miR-21, suppressing its expression and subsequently inhibiting hepatocellular carcinoma proliferation [100,101]. In the case of miR-544, *GAS5* negatively regulates its expression, inhibiting tumor cell proliferation [102]. *GAS5* inhibits cell proliferation also through the miR-544/RUNX3 pathway [102], where it stimulates NK cell activity and inhibits tumor growth [95]. In addition, *GAS5* and miR-135b reversely correlated and as reported in other tumors, *GAS5* over-expression reduces miR-135b expression and, thus, inhibits tumor cell proliferation [103]. Finally, miR-34a

manifested a different mode of action with respect to *GAS5*. It appeared that *GAS5* and miR-34a were positively correlated in three types of tumors; in hepatocellular carcinoma, glioblastoma and renal cell carcinoma [104]. *GAS5* under-expression was also related to tumor progression and proliferation (Table 1).

4.9. *GAS5* and miRNAs in Brain Tumors

Since brain tumors are not easily manageable, there are not many reports on the connection of *GAS5* and miRNAs. The existent studies are concerned with reports on glioma. In particular, two miRNAs are found to be related to *GAS5*. The miR-106b-5p [105] and miR-18a-5p [73]. Both of them were found to be significantly up-regulated in glioma cells, while *GAS5* was down-regulated. Additionally, it was found that *GAS5* over-expression results in miRNA down-regulation (Table 1).

4.10. *GAS5* and miRNAs in Osteosarcoma

Three miRNAs are related to *GAS5* in osteosarcoma: miR-663a [79], miR-203a [80] and miR-221 [81]. The miR-663a indirectly suppresses *GAS5* through the inhibition of its target, *ZBTB7A* [79]. Moreover, miR-203a suppresses *GAS5*, deactivates *TIMP2*, but activates the *PI3K/AKT/GSK2 β* pathway with simultaneous inhibition of the *NF- κ B* signaling cascade [80]. Therefore, *GAS5* indirectly regulates miR-203a, as also supported by their reverse-correlated expression. Finally, *GAS5* can directly suppress miR-221 through the miR-221/*ARH1* pathway [81] (Table 1).

5. Discussion

Functional ncRNAs affect every aspect of the biology in many organisms, from bacteria to higher eukaryotes. Specifically, they affect all stages of the coding sequence, including mRNA transcription, degradation and translation, and/or the nuclear translocation of proteins [5,106,107]. Among them, *GAS5* is mechanistically related to the bacterial 6S RNA, which binds the RNA polymerase and inhibits transcription [107,108]. Regarding nuclear receptor related ncRNAs, the ncRNA coactivator steroid receptor RNA activator (SRA) enhances nuclear receptor-induced transcriptional activity by associating with cofactor proteins, its stem-loop interacting protein, called SLIRP, and a pseudo-uridine synthase Pus1p [109,110]. *GAS5* is distinct from SRA in its activity and mode of action, while, similarly to SRA and other ncRNAs, its interaction with regulatory proteins might be critical for *Gas5*-mediated suppression of GR-induced transcriptional activity. Indeed, in relation to complex transcriptional regulation of endogenous, chromatin-associated genes [49], it would be interesting to investigate if *GAS5* can mimic the conformation of chromatin-integrated DNA interaction with histone-bound proteins and/or other chromatin components, with which the GR normally interacts to stimulate the transcription of endogenous, glucocorticoid-responsive genes.

In the present study, we explored the expression of *GAS5* along with that of various miRNAs across different tumor types and focused on its role in therapy-related sensitivity to these cells. The main conclusion is that *GAS5* seems to exert a tumor-suppressive role in the process of carcinogenesis across all tumor types. It does so, by interacting with or modulating the expression of various gene targets. As such, *GAS5* participates in tumor growth, proliferation, invasion, metastasis inhibition, as well as the induction of apoptosis. However, it seems that *GAS5* is also involved in the therapeutic response of cancer patients. Here, we review both in vitro and in vivo studies showing that *GAS5* contributes to the sensitization of cancer cells to chemotherapy and radiotherapy [33]. The tumor suppressive role of *GAS5* was recently supported by others as well [111–113], and all clues suggest that *GAS5* could be used a promising biomarker for disease diagnosis, tumor progression, or even as a therapeutic marker. However, there are a few studies investigating in-depth the role of *GAS5* in human tumors. Apart from differential expression, diverse genetic variants within *GAS5* have also been proposed to affect drug response, and could, thus, facilitate the categorization and dose adjustment [111].

6. Conclusions

Several studies highlight that *GAS5* plays an important role in various pathological and physiological conditions. Overall, *GAS5* acts as a tumor suppressor, whose down-regulation is directly connected to tumor progression, tumor cell proliferation and therapy-related resistance across different types of tumors. The agreement of different studies on the role of *GAS5* makes it a new attractive target for the prognosis and therapy of different cancer types.

Author Contributions: G.I.L.: conceptualization, methodology, investigation, data curation, writing—original draft preparation, visualization, supervision, project administration; K.H.: investigation, data curation, writing—review and editing; A.Z.: methodology, investigation, data curation, writing—review and editing, visualization, supervision, funding acquisition. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

ALL	Acute Lymphoblastic Leukemia
AML	Acute Myeloid Leukemia
AR	Androgen Receptor
<i>GAS5</i>	Growth Arrest Specific 5
GR	Glucocorticoid Receptor
GRE	Glucocorticoid Response Element
lncRNAs	long non-coding RNAs
miRNA	microRNA
MR	Mineralocorticoid Receptor
MRE	Mineralocorticoid Response Element
ncRNA	non-coding RNA
NK	Natural Killer cells
piRNAs	piwi-interacting
PR	Progesterone Receptor
snoRNAs	small non-coding nucleolar RNAs
SRA	Steroid Receptor RNA Activator
TAD	Transactivation Domain

References

- Lockhart, D.J.; Winzler, E.A. Genomics, gene expression and DNA arrays. *Nature* **2000**, *405*, 827–836. [[CrossRef](#)] [[PubMed](#)]
- Sherafatian, M.; Mowla, S.J. The origins and evolutionary history of human non-coding rna regulatory networks. *J. Bioinform. Comput. Biol.* **2017**, *15*, 1750005. [[CrossRef](#)] [[PubMed](#)]
- Jarroux, J.; Morillon, A.; Pinskaya, M. History, discovery, and classification of lncrnas. *Adv. Exp. Med. Biol.* **2017**, *1008*, 1–46. [[PubMed](#)]
- Eisen, M.B.; Spellman, P.T.; Brown, P.O.; Botstein, D. Cluster analysis and display of genome-wide expression patterns. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 14863–14868. [[CrossRef](#)] [[PubMed](#)]
- Mattick, J.S. The functional genomics of noncoding rna. *Science* **2005**, *309*, 1527–1528. [[CrossRef](#)] [[PubMed](#)]
- Pang, K.C.; Frith, M.C.; Mattick, J.S. Rapid evolution of noncoding rnas: Lack of conservation does not mean lack of function. *Trends Genet* **2006**, *22*, 1–5. [[CrossRef](#)]
- Taft, R.J.; Pang, K.C.; Mercer, T.R.; Dinger, M.; Mattick, J.S. Non-coding rnas: Regulators of disease. *J. Pathol.* **2010**, *220*, 126–139. [[CrossRef](#)]
- Esteller, M. Non-coding rnas in human disease. *Nat. Rev. Genet.* **2011**, *12*, 861–874. [[CrossRef](#)]
- Stefani, G.; Slack, F.J. Small non-coding rnas in animal development. *Nat. Rev. Mol. Cell Biol.* **2008**, *9*, 219–230. [[CrossRef](#)]
- Mercer, T.R.; Dinger, M.E.; Mattick, J.S. Long non-coding rnas: Insights into functions. *Nat. Rev. Genet.* **2009**, *10*, 155–159. [[CrossRef](#)]

11. Katayama, S.; Tomaru, Y.; Kasukawa, T.; Waki, K.; Nakanishi, M.; Nakamura, M.; Nishida, H.; Yap, C.C.; Suzuki, M.; Kawai, J.; et al. Antisense transcription in the mammalian transcriptome. *Science* **2005**, *309*, 1564–1566. [[PubMed](#)]
12. Schneider, C.; King, R.M.; Philipson, L. Genes specifically expressed at growth arrest of mammalian cells. *Cell* **1988**, *54*, 787–793. [[CrossRef](#)]
13. Kino, T.; Hurt, D.E.; Ichijo, T.; Nader, N.; Chrousos, G.P. Noncoding rna gas5 is a growth arrest and starvation-associated repressor of the glucocorticoid receptor. *Sci. Signal.* **2010**, *3*, ra8. [[CrossRef](#)] [[PubMed](#)]
14. Garabedian, M.J.; Logan, S.K. Glucocorticoid receptor DNA binding decoy is a gas. *Sci. Signal.* **2010**, *3*, pe5. [[CrossRef](#)]
15. Coccia, E.M.; Cicala, C.; Charlesworth, A.; Ciccarelli, C.; Rossi, G.B.; Philipson, L.; Sorrentino, V. Regulation and expression of a growth arrest-specific gene (gas5) during growth, differentiation, and development. *Mol. Cell. Biol.* **1992**, *12*, 3514–3521. [[CrossRef](#)]
16. Smith, C.M.; Steitz, J.A. Classification of gas5 as a multi-small-nucleolar-rna (snorna) host gene and a member of the 5'-terminal oligopyrimidine gene family reveals common features of snorna host genes. *Mol. Cell. Biol.* **1998**, *18*, 6897–6909. [[CrossRef](#)]
17. Amaldi, F.; Pierandrei-Amaldi, P. Top genes: A translationally controlled class of genes including those coding for ribosomal proteins. *Prog. Mol. Subcell. Biol.* **1997**, *18*, 1–17.
18. Hudson, W.H.; Pickard, M.R.; de Vera, I.M.; Kuiper, E.G.; Mourtada-Maarabouni, M.; Conn, G.L.; Kojetin, D.J.; Williams, G.T.; Ortlund, E.A. Conserved sequence-specific lincrna-steroid receptor interactions drive transcriptional repression and direct cell fate. *Nat. Commun.* **2014**, *5*, 5395. [[CrossRef](#)]
19. Yang, X.; Xie, Z.; Lei, X.; Gan, R. Long non-coding rna gas5 in human cancer. *Oncol. Lett.* **2020**, *20*, 2587–2594. [[CrossRef](#)]
20. Zhou, Y.; Chen, B. Gas5mediated regulation of cell signaling (review). *Mol. Med. Rep.* **2020**, *22*, 3049–3056. [[PubMed](#)]
21. Frank, F.; Okafor, C.D.; Ortlund, E.A. The first crystal structure of a DNA-free nuclear receptor DNA binding domain sheds light on DNA-driven allostery in the glucocorticoid receptor. *Sci. Rep.* **2018**, *8*, 13497. [[CrossRef](#)]
22. Luisi, B.F.; Xu, W.X.; Otwinowski, Z.; Freedman, L.P.; Yamamoto, K.R.; Sigler, P.B. Crystallographic analysis of the interaction of the glucocorticoid receptor with DNA. *Nature* **1991**, *352*, 497–505. [[CrossRef](#)] [[PubMed](#)]
23. Varani, G.; McClain, W.H. The g x u wobble base pair. A fundamental building block of rna structure crucial to rna function in diverse biological systems. *EMBO Rep.* **2000**, *1*, 18–23. [[CrossRef](#)] [[PubMed](#)]
24. Dimitrov, R.A.; Zuker, M. Prediction of hybridization and melting for double-stranded nucleic acids. *Biophys. J.* **2004**, *87*, 215–226. [[CrossRef](#)] [[PubMed](#)]
25. Mayama, T.; Marr, A.K.; Kino, T. Differential Expression of Glucocorticoid Receptor Noncoding RNA Repressor Gas5 in Autoimmune and Inflammatory Diseases. *Horm. Metab. Res.* **2016**, *48*, 550–557. [[CrossRef](#)] [[PubMed](#)]
26. Goustin, A.S.; Thepsuwan, P.; Kosir, M.A.; Lipovich, L. The growth-arrest-specific (gas)-5 long non-coding rna: A fascinating lincrna widely expressed in cancers. *Non-Coding RNA* **2019**, *5*, 46. [[CrossRef](#)]
27. Mourtada-Maarabouni, M.; Pickard, M.R.; Hedge, V.L.; Farzaneh, F.; Williams, G.T. Gas5, a non-protein-coding rna, controls apoptosis and is downregulated in breast cancer. *Oncogene* **2009**, *28*, 195–208. [[CrossRef](#)]
28. Nakamura, Y.; Takahashi, N.; Kakegawa, E.; Yoshida, K.; Ito, Y.; Kayano, H.; Niitsu, N.; Jinnai, I.; Bessho, M. The gas5 (growth arrest-specific transcript 5) gene fuses to bcl6 as a result of t(1;3)(q25;q27) in a patient with b-cell lymphoma. *Cancer Genet. Cytogenet.* **2008**, *182*, 144–149. [[CrossRef](#)]
29. Mourtada-Maarabouni, M.; Hedge, V.L.; Kirkham, L.; Farzaneh, F.; Williams, G.T. Growth arrest in human t-cells is controlled by the non-coding rna growth-arrest-specific transcript 5 (gas5). *J. Cell Sci.* **2008**, *121*, 939–946. [[CrossRef](#)]
30. Gasic, V.; Stankovic, B.; Zukic, B.; Janic, D.; Dokmanovic, L.; Krstovski, N.; Lazic, J.; Milosevic, G.; Lucafo, M.; Stocco, G.; et al. Expression pattern of long non-coding rna growth arrest-specific 5 in the remission induction therapy in childhood acute lymphoblastic leukemia. *J. Med. Biochem.* **2019**, *38*, 292–298. [[CrossRef](#)]
31. Yan, H.; Zhang, D.Y.; Li, X.; Yuan, X.Q.; Yang, Y.L.; Zhu, K.W.; Zeng, H.; Li, X.L.; Cao, S.; Zhou, H.H.; et al. Long non-coding rna gas5 polymorphism predicts a poor prognosis of acute myeloid leukemia in chinese patients via affecting hematopoietic reconstitution. *Leuk. Lymphoma* **2017**, *58*, 1948–1957. [[CrossRef](#)] [[PubMed](#)]

32. Mourtada-Maarabouni, M.; Hasan, A.M.; Farzaneh, F.; Williams, G.T. Inhibition of human t-cell proliferation by mammalian target of rapamycin (mTOR) antagonists requires noncoding rna growth-arrest-specific transcript 5 (gas5). *Mol. Pharmacol.* **2010**, *78*, 19–28. [[CrossRef](#)]
33. Gao, J.; Liu, L.; Li, G.; Cai, M.; Tan, C.; Han, X.; Han, L. Lncrna gas5 confers the radio sensitivity of cervical cancer cells via regulating mir-106b/ier3 axis. *Int. J. Biol. Macromol.* **2019**, *126*, 994–1001. [[CrossRef](#)] [[PubMed](#)]
34. Wen, Q.; Liu, Y.; Lyu, H.; Xu, X.; Wu, Q.; Liu, N.; Yin, Q.; Li, J.; Sheng, X. Long noncoding rna gas5, which acts as a tumor suppressor via microRNA 21, regulates cisplatin resistance expression in cervical cancer. *Int. J. Gynecol. Cancer Off. J. Int. Gynecol. Cancer Soc.* **2017**, *27*, 1096–1108. [[CrossRef](#)]
35. Nagini, S. Breast cancer: Current molecular therapeutic targets and new players. *Anti-Cancer Agents Med. Chem.* **2017**, *17*, 152–163. [[CrossRef](#)] [[PubMed](#)]
36. Momenimovahed, Z.; Salehiniya, H. Epidemiological characteristics of and risk factors for breast cancer in the world. *Breast Cancer Targets Ther.* **2019**, *11*, 151–164. [[CrossRef](#)] [[PubMed](#)]
37. Arshi, A.; Sharifi, F.S.; Khorrarnian Ghahfarokhi, M.; Faghieh, Z.; Doosti, A.; Ostovari, S.; Mahmoudi Maymand, E.; Ghahramani Seno, M.M. Expression analysis of malat1, gas5, sra, and neat1 lncrnas in breast cancer tissues from young women and women over 45 years of age. *Mol. Ther. Nucleic Acids* **2018**, *12*, 751–757. [[CrossRef](#)]
38. Gee, H.E.; Buffa, F.M.; Camps, C.; Ramachandran, A.; Leek, R.; Taylor, M.; Patil, M.; Sheldon, H.; Betts, G.; Homer, J.; et al. The small-nucleolar rnas commonly used for microRNA normalisation correlate with tumour pathology and prognosis. *Br. J. Cancer* **2011**, *104*, 1168–1177. [[CrossRef](#)]
39. Zhang, Z.; Zhu, Z.; Watabe, K.; Zhang, X.; Bai, C.; Xu, M.; Wu, F.; Mo, Y.Y. Negative regulation of lncrna gas5 by mir-21. *Cell Death Differ.* **2013**, *20*, 1558–1568. [[CrossRef](#)]
40. Pickard, M.R.; Williams, G.T. Regulation of apoptosis by long non-coding rna gas5 in breast cancer cells: Implications for chemotherapy. *Breast Cancer Res. Treat.* **2014**, *145*, 359–370. [[CrossRef](#)]
41. Li, W.; Zhai, L.; Wang, H.; Liu, C.; Zhang, J.; Chen, W.; Wei, Q. Downregulation of lncrna gas5 causes trastuzumab resistance in breast cancer. *Oncotarget* **2016**, *7*, 27778–27786. [[CrossRef](#)] [[PubMed](#)]
42. Gu, J.; Wang, Y.; Wang, X.; Zhou, D.; Shao, C.; Zhou, M.; He, Z. Downregulation of lncrna gas5 confers tamoxifen resistance by activating mir-222 in breast cancer. *Cancer Lett.* **2018**, *434*, 1–10. [[CrossRef](#)] [[PubMed](#)]
43. Li, J.; Li, L.; Yuan, H.; Huang, X.W.; Xiang, T.; Dai, S. Up-regulated lncrna gas5 promotes chemosensitivity and apoptosis of triple-negative breast cancer cells. *Cell Cycle* **2019**, *18*, 1965–1975. [[CrossRef](#)]
44. Guo, L.L.; Wang, S.F. Downregulated long noncoding rna gas5 fails to function as decoy of cebpb, resulting in increased gdf15 expression and rapid ovarian cancer cell proliferation. *Cancer Biother. Radiopharm.* **2019**, *34*, 537–546. [[CrossRef](#)]
45. Ma, C.; Wang, W.; Li, P. Lncrna gas5 overexpression downregulates il-18 and induces the apoptosis of fibroblast-like synoviocytes. *Clin. Rheumatol.* **2019**, *38*, 3275–3280. [[CrossRef](#)]
46. Zhao, H.; Yu, H.; Zheng, J.; Ning, N.; Tang, F.; Yang, Y.; Wang, Y. Lowly-expressed lncrna gas5 facilitates progression of ovarian cancer through targeting mir-196-5p and thereby regulating hoxa5. *Gynecol. Oncol.* **2018**, *151*, 345–355. [[CrossRef](#)]
47. Zhao, Q.; Fan, C. A novel risk score system for assessment of ovarian cancer based on co-expression network analysis and expression level of five lncrnas. *BMC Med. Genet.* **2019**, *20*, 103. [[CrossRef](#)]
48. Song, J.; Zhang, W.; Wang, S.; Liu, K.; Song, F.; Ran, L. A panel of 7 prognosis-related long non-coding rnas to improve platinum-based chemoresistance prediction in ovarian cancer. *Int. J. Oncol.* **2018**, *53*, 866–876. [[CrossRef](#)]
49. Chrousos, G.P.; Kino, T. Intracellular glucocorticoid signaling: A formerly simple system turns stochastic. *Sci. STKE* **2005**, *2005*, pe48. [[CrossRef](#)]
50. Long, X.; Song, K.; Hu, H.; Tian, Q.; Wang, W.; Dong, Q.; Yin, X.; Di, W. Long non-coding rna gas5 inhibits ddp-resistance and tumor progression of epithelial ovarian cancer via gas5-e2f4-parp1-mapk axis. *J. Exp. Clin. Cancer Res. CR* **2019**, *38*, 345. [[CrossRef](#)]
51. Pernar, C.H.; Ebot, E.M.; Wilson, K.M.; Mucci, L.A. The epidemiology of prostate cancer. *Cold Spring Harb. Perspect. Med.* **2018**, *8*, a030361. [[CrossRef](#)]
52. Wong, M.C.S.; Fung, F.D.H.; Leung, C.; Cheung, W.W.L.; Goggins, W.B.; Ng, C.F. The global epidemiology of bladder cancer: A joinpoint regression analysis of its incidence and mortality trends and projection. *Sci. Rep.* **2018**, *8*, 1129. [[CrossRef](#)] [[PubMed](#)]

53. Yang, J.; Hao, T.; Sun, J.; Wei, P.; Zhang, H. Long noncoding RNA GAS5 modulates α -Solanine-induced radiosensitivity by negatively regulating miR-18a in human prostate cancer cells. *Biomed. Pharmacother.* **2019**, *112*, 108656. [[CrossRef](#)] [[PubMed](#)]
54. Zhang, H.; Guo, Y.; Song, Y.; Shang, C. Long noncoding rna gas5 inhibits malignant proliferation and chemotherapy resistance to doxorubicin in bladder transitional cell carcinoma. *Cancer Chemother. Pharmacol.* **2017**, *79*, 49–55. [[CrossRef](#)]
55. De Groot, P.M.; Wu, C.C.; Carter, B.W.; Munden, R.F. The epidemiology of lung cancer. *Transl. Lung Cancer Res.* **2018**, *7*, 220–233. [[CrossRef](#)] [[PubMed](#)]
56. Zhang, N.; Yang, G.Q.; Shao, X.M.; Wei, L. Gas5 modulated autophagy is a mechanism modulating cisplatin sensitivity in nsclc cells. *Eur. Rev. Med. Pharmacol. Sci.* **2016**, *20*, 2271–2277. [[PubMed](#)]
57. Xue, Y.; Ni, T.; Jiang, Y.; Li, Y. Long noncoding rna gas5 inhibits tumorigenesis and enhances radiosensitivity by suppressing mir-135b expression in non-small cell lung cancer. *Oncol. Res.* **2017**, *25*, 1305–1316. [[CrossRef](#)] [[PubMed](#)]
58. Cao, L.; Chen, J.; Ou, B.; Liu, C.; Zou, Y.; Chen, Q. Gas5 knockdown reduces the chemo-sensitivity of non-small cell lung cancer (nsclc) cell to cisplatin (ddp) through regulating mir-21/pten axis. *Biomed. Pharmacother.* **2017**, *93*, 570–579. [[CrossRef](#)]
59. Cheng, Y.; Dai, X.; Yang, T.; Zhang, N.; Liu, Z.; Jiang, Y. Low long noncoding rna growth arrest-specific transcript 5 expression in the exosomes of lung cancer cells promotes tumor angiogenesis. *J. Oncol.* **2019**, *2019*, 2476175. [[CrossRef](#)] [[PubMed](#)]
60. Sitarz, R.; Skierucha, M.; Mielko, J.; Offerhaus, G.J.A.; Maciejewski, R.; Polkowski, W.P. Gastric cancer: Epidemiology, prevention, classification, and treatment. *Cancer Manag. Res.* **2018**, *10*, 239–248. [[CrossRef](#)]
61. Aminian, K.; Mashayekhi, F.; Mirzanejad, L.; Salehi, Z. A functional genetic variant in gas5 lncrna (rs145204276) modulates p27(kip1) expression and confers risk for gastric cancer. *Br. J. Biomed. Sci.* **2019**, *76*, 83–85. [[CrossRef](#)] [[PubMed](#)]
62. Cheng, C.; Wang, Q.; Zhu, M.; Liu, K.; Zhang, Z. Integrated analysis reveals potential long non-coding rna biomarkers and their potential biological functions for disease free survival in gastric cancer patients. *Cancer Cell Int.* **2019**, *19*, 123. [[CrossRef](#)] [[PubMed](#)]
63. Dong, S.; Zhang, X.; Liu, D. Overexpression of long noncoding rna gas5 suppresses tumorigenesis and development of gastric cancer by sponging mir-106a-5p through the akt/mtor pathway. *Biol. Open* **2019**, *8*, bio.041343. [[CrossRef](#)] [[PubMed](#)]
64. Liu, Y.; Yin, L.; Chen, C.; Zhang, X.; Wang, S. Long non-coding RNA GAS5 inhibits migration and invasion in gastric cancer via interacting with p53 protein. *Dig. Liver Dis.* **2020**, *52*, 331–338. [[CrossRef](#)] [[PubMed](#)]
65. Zhang, N.; Wang, A.-Y.; Wang, X.; Sun, X.-M.; Xue, H.-Z. Gas5 is downregulated in gastric cancer cells by promoter hypermethylation and regulates adriamycin sensitivity. *Eur. Rev. Med. Pharmacol. Sci.* **2016**, *20*, 3199–3205. [[PubMed](#)]
66. Abel, G.A.; Wochnik, G.M.; Ruegg, J.; Rouyer, A.; Holsboer, F.; Rein, T. Activity of the gr in g2 and mitosis. *Mol. Endocrinol.* **2002**, *16*, 1352–1366. [[CrossRef](#)] [[PubMed](#)]
67. Song, J.; Shu, H.; Zhang, L.; Xiong, J. Long noncoding RNA GAS5 inhibits angiogenesis and metastasis of colorectal cancer through the Wnt/ β -catenin signaling pathway. *J. Cell. Biochem.* **2019**, *120*, 6937–6951. [[CrossRef](#)] [[PubMed](#)]
68. Lu, S.; Su, Z.; Fu, W.; Cui, Z.; Jiang, X.; Tai, S. Altered expression of long non-coding RNA GAS5 in digestive tumors. *Biosci. Rep.* **2019**, *39*, 39. [[CrossRef](#)]
69. Rowe, J.H.; Ghouri, Y.A.; Mian, I. Review of hepatocellular carcinoma: Epidemiology, etiology, and carcinogenesis. *J. Carcinog.* **2017**, *16*, 1. [[CrossRef](#)]
70. Mishra, S.; Verma, S.S.; Rai, V.; Awasthee, N.; Chava, S.; Hui, K.M.; Kumar, A.P.; Challagundla, K.B.; Sethi, G.; Gupta, S.C. Long non-coding rnas are emerging targets of phytochemicals for cancer and other chronic diseases. *Cell Mol. Life Sci.* **2019**, *76*, 1947–1966. [[CrossRef](#)]
71. Chen, L.; Han, L.; Wei, J.; Zhang, K.; Shi, Z.; Duan, R.; Li, S.; Zhou, X.; Pu, P.; Zhang, J.; et al. Snord76, a box c/d snorna, acts as a tumor suppressor in glioblastoma. *Sci. Rep.* **2015**, *5*, 8588. [[CrossRef](#)]
72. Huo, J.F.; Chen, X.B. Long noncoding rna growth arrest-specific 5 facilitates glioma cell sensitivity to cisplatin by suppressing excessive autophagy in an mtor-dependent manner. *J. Cell. Biochem.* **2019**, *120*, 6127–6136. [[CrossRef](#)]

73. Liu, Q.; Yu, W.; Zhu, S.; Cheng, K.; Xu, H.; Lv, Y.; Long, X.; Ma, L.; Huang, J.; Sun, S.; et al. Long noncoding rna gas5 regulates the proliferation, migration, and invasion of glioma cells by negatively regulating mir-18a-5p. *J. Cell. Physiol.* **2018**, *234*, 757–768. [[CrossRef](#)] [[PubMed](#)]
74. Shen, J.; Hodges, T.R.; Song, R.; Gong, Y.; Calin, G.A.; Heimberger, A.B.; Zhao, H. Serum hotair and gas5 levels as predictors of survival in patients with glioblastoma. *Mol. Carcinog.* **2018**, *57*, 137–141. [[CrossRef](#)] [[PubMed](#)]
75. Wang, Y.; Xin, S.; Zhang, K.; Shi, R.; Bao, X. Low gas5 levels as a predictor of poor survival in patients with lower-grade gliomas. *J. Oncol.* **2019**, *2019*, 1785042. [[CrossRef](#)] [[PubMed](#)]
76. Ottaviani, G.; Jaffe, N. The epidemiology of osteosarcoma. *Cancer Treat. Res.* **2009**, *152*, 3–13. [[PubMed](#)]
77. Luetke, A.; Meyers, P.A.; Lewis, I.; Juergens, H. Osteosarcoma treatment—Where do we stand? A state of the art review. *Cancer Treat. Rev.* **2014**, *40*, 523–532. [[CrossRef](#)]
78. Zhang, X.; Du, K.; Lou, Z.; Ding, K.; Zhang, F.; Zhu, J.; Chang, Z. The ctbp1-hdac1/2-irf1 transcriptional complex represses the expression of the long noncoding rna gas5 in human osteosarcoma cells. *Int. J. Biol. Sci.* **2019**, *15*, 1460–1471. [[CrossRef](#)]
79. Zhang, L.; Wang, Y.; Zhang, L.; Xia, X.; Chao, Y.; He, R.; Han, C.; Zhao, W. Zbtb7a, a mir-663a target gene, protects osteosarcoma from endoplasmic reticulum stress-induced apoptosis by suppressing lncrna gas5 expression. *Cancer Lett.* **2019**, *448*, 105–116. [[CrossRef](#)]
80. Wang, Y.; Kong, D. Lncrna gas5 represses osteosarcoma cells growth and metastasis via sponging mir-203a. *Cell. Physiol. Biochem. Int. J. Exp. Cell. Physiol. Biochem. Pharmacol.* **2018**, *45*, 844–855. [[CrossRef](#)]
81. Ye, K.; Wang, S.; Zhang, H.; Han, H.; Ma, B.; Nan, W. Long noncoding rna gas5 suppresses cell growth and epithelial-mesenchymal transition in osteosarcoma by regulating the mir-221/arhi pathway. *J. Cell. Biochem.* **2017**, *118*, 4772–4781. [[CrossRef](#)] [[PubMed](#)]
82. Jing, Z.; Gao, L.; Wang, H.; Chen, J.; Nie, B.; Hong, Q. Long non-coding rna gas5 regulates human b lymphocytic leukaemia tumorigenesis and metastasis by sponging mir-222. *Cancer Biomark. Sect. A Dis. Mark.* **2019**, *26*, 385–392. [[CrossRef](#)] [[PubMed](#)]
83. Yan, Z.; Ruoyu, L.; Xing, L.; Hua, L.; Jun, Z.; Yaqin, P.; Lu, W.; Aili, T.; Yuzi, Z.; Lin, M.; et al. Long non-coding rna gas5 regulates the growth and metastasis of human cervical cancer cells via induction of apoptosis and cell cycle arrest. *Arch. Biochem. Biophys.* **2020**, *684*, 108320. [[CrossRef](#)] [[PubMed](#)]
84. Yao, T.; Lu, R.; Zhang, J.; Fang, X.; Fan, L.; Huang, C.; Lin, R.; Lin, Z. Growth arrest-specific 5 attenuates cisplatin-induced apoptosis in cervical cancer by regulating stat3 signaling via mir-21. *J. Cell. Physiol.* **2019**, *234*, 9605–9615. [[CrossRef](#)] [[PubMed](#)]
85. Yang, W.; Hong, L.; Xu, X.; Wang, Q.; Huang, J.; Jiang, L. Lncrna gas5 suppresses the tumorigenesis of cervical cancer by downregulating mir-196a and mir-205. *Tumour Biol. J. Int. Soc. Oncodev. Biol. Med.* **2017**, *39*, 1010428317711315. [[CrossRef](#)] [[PubMed](#)]
86. Chen, Z.; Pan, T.; Jiang, D.; Jin, L.; Geng, Y.; Feng, X.; Shen, A.; Zhang, L. The lncrna-gas5/mir-221-3p/dkk2 axis modulates abcb1-mediated adriamycin resistance of breast cancer via the wnt/beta-catenin signaling pathway. *Mol. Ther. Nucleic Acids* **2020**, *19*, 1434–1448. [[CrossRef](#)]
87. Zheng, S.; Li, M.; Miao, K.; Xu, H. Lncrna gas5-promoted apoptosis in triple-negative breast cancer by targeting mir-378a-5p/sufu signaling. *J. Cell. Biochem.* **2020**, *121*, 2225–2235. [[CrossRef](#)]
88. Zong, Y.; Zhang, Y.; Sun, X.; Xu, T.; Cheng, X.; Qin, Y. Mir-221/222 promote tumor growth and suppress apoptosis by targeting lncrna gas5 in breast cancer. *Biosci. Rep.* **2019**, *39*, BSR20181859. [[CrossRef](#)]
89. Gu, J.; Wang, Y.; Wang, X.; Zhou, D.; Wang, X.; Zhou, M.; He, Z. Effect of the LncRNA GAS5-MiR-23a-ATG3 Axis in Regulating Autophagy in Patients with Breast Cancer. *Cell. Physiol. Biochem.* **2018**, *48*, 194–207. [[CrossRef](#)]
90. Li, S.; Zhou, J.; Wang, Z.; Wang, P.; Gao, X.; Wang, Y. Long noncoding RNA GAS5 suppresses triple negative breast cancer progression through inhibition of proliferation and invasion by competitively binding miR-196a-5p. *Biomed. Pharmacother.* **2018**, *104*, 451–457. [[CrossRef](#)]
91. Chen, X.; Yang, C.; Xie, S.; Cheung, E. Long non-coding rna gas5 and zfas1 are prognostic markers involved in translation targeted by mir-940 in prostate cancer. *Oncotarget* **2018**, *9*, 1048–1062. [[CrossRef](#)]
92. Chen, L.; Ren, P.; Zhang, Y.; Gong, B.; Yu, D.; Sun, X. Long noncoding rna gas5 increases the radiosensitivity of a549 cells through interaction with the mir21/pten/akt axis. *Oncol. Rep.* **2020**, *43*, 897–907. [[PubMed](#)]

93. Dong, L.; Li, G.; Li, Y.; Zhu, Z. Upregulation of long noncoding rna gas5 inhibits lung cancer cell proliferation and metastasis via mir-205/pten axis. *Med. Sci. Monit. Int. Med. J. Exp. Clin. Res.* **2019**, *25*, 2311–2319. [[CrossRef](#)] [[PubMed](#)]
94. Mei, Y.; Si, J.; Wang, Y.; Huang, Z.; Zhu, H.; Feng, S.; Wu, X.; Wu, L. Long noncoding rna gas5 suppresses tumorigenesis by inhibiting mir-23a expression in non-small cell lung cancer. *Oncol. Res.* **2017**, *25*, 1027–1037. [[CrossRef](#)] [[PubMed](#)]
95. Wei, M.F.; Gu, Z.S.; Zheng, L.L.; Zhao, M.X.; Wang, X.J. Long non-coding rna gas5 promotes natural killer cell cytotoxicity against gastric cancer by regulating mir-18a. *Neoplasma* **2020**. [[CrossRef](#)] [[PubMed](#)]
96. Li, Y.; Gu, J.; Lu, H. The gas5/mir-222 axis regulates proliferation of gastric cancer cells through the pten/akt/mTOR pathway. *Dig. Dis. Sci.* **2017**, *62*, 3426–3437. [[CrossRef](#)] [[PubMed](#)]
97. Cheng, K.; Zhao, Z.; Wang, G.; Wang, J.; Zhu, W. Lncrna gas5 inhibits colorectal cancer cell proliferation via the mir1825p/foxo3a axis. *Oncol. Rep.* **2018**, *40*, 2371–2380.
98. Liu, L.; Meng, T.; Yang, X.H.; Sayim, P.; Lei, C.; Jin, B.; Ge, L.; Wang, H.J. Prognostic and predictive value of long non-coding rna gas5 and microRNA-221 in colorectal cancer and their effects on colorectal cancer cell proliferation, migration and invasion. *Cancer Biomark. Sect. A Dis. Markers* **2018**, *22*, 283–299. [[CrossRef](#)]
99. Zhao, P.; Cui, X.; Zhao, L.; Liu, L.; Wang, D. Overexpression of growth-arrest-specific transcript 5 improved cisplatin sensitivity in hepatocellular carcinoma through sponging mir-222. *DNA Cell Biol.* **2020**, *39*, 724–732. [[CrossRef](#)]
100. Wang, C.; Ke, S.; Li, M.; Lin, C.; Liu, X.; Pan, Q. Downregulation of lncrna gas5 promotes liver cancer proliferation and drug resistance by decreasing pten expression. *Mol. Genet. Genom.* **2020**, *295*, 251–260. [[CrossRef](#)]
101. Hu, L.; Ye, H.; Huang, G.; Luo, F.; Liu, Y.; Liu, Y.; Yang, X.; Shen, J.; Liu, Q.; Zhang, J. Long noncoding RNA GAS5 suppresses the migration and invasion of hepatocellular carcinoma cells via miR-21. *Tumor Biol.* **2015**, *37*, 2691–2702. [[CrossRef](#)] [[PubMed](#)]
102. Fang, P.; Xiang, L.; Chen, W.; Li, S.; Huang, S.; Li, J.; Zhuge, L.; Jin, L.; Feng, W.; Chen, Y.; et al. Lncrna gas5 enhanced the killing effect of nk cell on liver cancer through regulating mir-544/runx3. *Innate Immun.* **2019**, *25*, 99–109. [[CrossRef](#)] [[PubMed](#)]
103. Yang, L.; Jiang, J. Gas5 regulates reck expression and inhibits invasion potential of hcc cells by sponging mir-135b. *BioMed Res. Int.* **2019**, *2019*, 2973289. [[CrossRef](#)] [[PubMed](#)]
104. Toraih, E.A.; Alghamdi, S.A.; El-Wazir, A.; Hosny, M.M.; Hussein, M.H.; Khashana, M.S.; Fawzy, M.S. Dual biomarkers long non-coding rna gas5 and microRNA-34a co-expression signature in common solid tumors. *PLoS ONE* **2018**, *13*, e0198231. [[CrossRef](#)]
105. Huang, W.; Shi, Y.; Han, B.; Wang, Q.; Zhang, B.; Qi, C.; Liu, F. Lncrna gas5-as1 inhibits glioma proliferation, migration, and invasion via mir-106b-5p/tusc2 axis. *Hum. Cell* **2020**, *33*, 416–426. [[CrossRef](#)]
106. Willingham, A.T.; Orth, A.P.; Batalov, S.; Peters, E.C.; Wen, B.G.; Aza-Blanc, P.; Hogenesch, J.B.; Schultz, P.G. A strategy for probing the function of noncoding RNAs finds a repressor of nfat. *Science* **2005**, *309*, 1570–1573. [[CrossRef](#)]
107. Barrandon, C.; Spiluttini, B.; Bensaude, O. Non-coding RNAs regulating the transcriptional machinery. *Biol. Cell* **2008**, *100*, 83–95. [[CrossRef](#)]
108. Willkomm, D.K.; Hartmann, R.K. 6s rna—An ancient regulator of bacterial rna polymerase rediscovered. *Biol. Chem.* **2005**, *386*, 1273–1277. [[CrossRef](#)]
109. Lanz, R.B.; McKenna, N.J.; Onate, S.A.; Albrecht, U.; Wong, J.; Tsai, S.Y.; Tsai, M.J.; O'Malley, B.W. A steroid receptor coactivator, sra, functions as an rna and is present in an src-1 complex. *Cell* **1999**, *97*, 17–27. [[CrossRef](#)]
110. Leygue, E. Steroid Receptor RNA Activator (SRA1): Unusual Bifaceted Gene Products with Suspected Relevance to Breast Cancer. *Nucl. Recept. Signal.* **2007**, *5*, e006. [[CrossRef](#)]
111. Ghaforui-Fard, S.; Taheri, M. Growth arrest specific transcript 5 in tumorigenesis process: An update on the expression pattern and genomic variants. *Biomed. Pharmacother.* **2019**, *112*, 108723. [[CrossRef](#)]

112. Ma, C.; Shi, X.; Zhu, Q.; Li, Q.; Liu, Y.; Yao, Y.; Song, Y. The growth arrest-specific transcript 5 (GAS5): A pivotal tumor suppressor long noncoding RNA in human cancers. *Tumor Biol.* **2015**, *37*, 1437–1444. [[CrossRef](#)] [[PubMed](#)]
113. Pickard, M.R.; Williams, G.T. Molecular and cellular mechanisms of action of tumour suppressor gas5 lncrna. *Genes* **2015**, *6*, 484–499. [[CrossRef](#)] [[PubMed](#)]

Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).