

Supplementary Materials

Linking peptidoglycan synthesis protein complex with asymmetric cell division during *Bacillus subtilis* sporulation

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Table S1. Bacterial strains

Strain	Genotype	Source/reference
<i>B. subtilis</i>		
PY79	Prototrophic derivative of <i>B. subtilis</i> 168	[43]
PY180	<i>spoIIIE::Tn917ΩHU7</i>	[8]
JR46	<i>gpsB::kan</i>	[20]
KM1202	<i>p_{gpsB} gpsB-ypet cat</i>	this work
KM1309	<i>p_{gpsB} gpsB-mneongreen cat</i>	this work
KM1322	<i>p_{gpsB} gpsB-mscarlet kan</i>	this work
KM1324	<i>p_{spoIIIE} spoIIIE-ypet cat p_{gpsB} gpsB-mscarlet kan</i>	this work
KM1325	<i>p_{gpsB} gpsB-ypet cat spoIIIE::Tn917ΩHU7</i>	this work
KM1327	<i>p_{spoIIIE} spoIIIE-ypet cat gpsB::kan</i>	this work
IB1537	<i>p_{spoIIIE} spoIIIE-ypet cat</i>	[15]
<i>E. coli</i>		
MM294	<i>F⁻ endA-1 hsdR-1, (rk⁻, mk) supE44 thi-1 recA1</i>	[44]
DH5α	<i>F⁻ Φ80lacZΔM15 Δ(lacZYA-argF) U169 recA1 endA1 hsdR17 (rK⁻, mK⁺) phoA supE44 λ⁻ thi-1 gyrA96 relA1</i>	Invitrogen
BL21(DE3)	<i>hsdS gal (λcts857 indt Sam7 nin5 lacUV5-T7gene</i>	Novagen

BTH101	<i>F⁻cya-99 araD139 galE15 galK16 rpsL1(Str^r)hsdR2 mcrA1 mcrB1</i>	[48]
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Table S2. Plasmids

Plasmid	Description	Reference
pSG1151	<i>bla cat gfpmut1</i>	[45]
pUK19	<i>bla kan</i>	[46]
pET15 b(+)	expression vector used for protein expression, <i>bla lacI T7 promoter</i>	Novagen
pUC19	vector with a multiple cloning site (MCS) <i>bla</i>	[47]
pSGIIE-Ypet	<i>bla cat spoIIE (724-827aa)-ypet</i>	[15]
pSGgpsB-ypet	<i>bla cat gpsB (31-98aa)-ypet</i>	this work
pSGgpsB-mneongreen	<i>bla cat gpsB (31-98aa)-mneongreen</i>	this work
pUCkangpsB-mscarlet	<i>bla kan gpsB (31-98aa)-mscarlet</i>	this work
pETgpsB	<i>bla lacI pT7 his-gpsB</i>	this work
pKT25	enable fusion to C-terminal end of adenylate cyclase T25 fragment	[48]
pKNT25	enable fusion to N-terminal end of adenylate cyclase T25 fragment	[48]
pUT18	enable fusion to N-terminal end of adenylate cyclase T18 fragment	[48]
pUTC18	enable fusion to C-terminal end of adenylate cyclase T18 fragment	[48]
pKTgpsB	<i>p_{lac}-T25-gpsB-kan</i>	this work
pKNTgpsB	<i>p_{lac}-gpsB-T25 kan</i>	this work

pUTgpsB	<i>p_{lac}-gpsB-T18 bla</i>	this work
pUTCgpsB	<i>p_{lac}-T18-gpsB bla</i>	this work
pKTIIE	<i>p_{lac}-T25-spoIIIE kan</i>	[15]
pKNTIIE	<i>p_{lac}-spoIIIE-T25 kan</i>	[15]
pUTIIE	<i>p_{lac}-spoIIIE-T18 bla</i>	[15]
pUTCIIIE	<i>p_{lac}-T18-spoIIIE bla</i>	[15]
pKTnctIIE	<i>p_{lac}-T25- spoIIIE domain I,II kan</i>	[15]
pKNTnctIIE	<i>p_{lac}-spoIIIE domain I,II-T25 kan</i>	[15]
pUTCnctIIE	<i>p_{lac}-T18-spoIIIE domain I,II bla</i>	[15]
pUTnctIIE	<i>p_{lac}-spoIIIE domain I,II T18 bla</i>	[15]
pKTctIIE	<i>p_{lac}-T25- spoIIIE domain II kan</i>	[15]
pKNTctIIE	<i>p_{lac}-spoIIIE domain II-T25 kan</i>	[15]
pUTCctIIE	<i>p_{lac}-T18-spoIIIE domain II bla</i>	[15]
pUTctIIE	<i>p_{lac}-T18-spoIIIE domain II bla</i>	[15]
pKNTezrA	<i>p_{lac}-ezrA-T25 kan</i>	this work
pUTEzrA	<i>p_{lac}-ezrA-T18 bla</i>	this work
pKTponA	<i>p_{lac}-T25-ponA kan</i>	this work
pUTCponA	<i>p_{lac}-T18-ponA bla</i>	this work

Table S3. Oligonucleotides used in this work

Primer	Sequence 5' - 3'
gpsBSKpnI	GCTGAATATGGTACCGACAAATTTTAGATATGATTATTAAGGATTATG
gpsBEKpnI	GATGATGATGGTACCCCAATCATAAAGCTTGCTGCCAAAAAC
gpsBBamF4	GTAGTAGTAGGATCCCATGCTTGCTGATAAAGTAAAGCTTTCTG
gpsBEcoR	GTAGTAGTAGAATTCTGAATCATAAAGCTTGCTGCCAAAAAC
gpsBKTEcoR	GTAGTAGTAGAATTCTCAATCATAAAGCTTGCTGCCAAAAAC
ezrABamF2	GATGATGATGGATCCCATGGAGTTTGTTCATTGGATTATTAATTG

ezrAEcoR	GATGATGATGAATTCGAAGCGGATATGTCAGCTTTGATTTTTTC
ezrAKTEcoR	GATGATGATGAATTCCTAAGCGGATATGTCAGCTTTGATTTTTTC
ponABamF	CTACTACTAGGATCCCATGTCAGATCAATTTAACAGCCGTG
ponAEcoR2	CTACTACTAGAATTCGAATTTGTTTTTTCAATGGATGATGAG
ponAKTEcoR	CTACTACTAGAATTCCTAATTTGTTTTTTCAATGGATGATGAG
gpsBSNd	CATGCGTCCCATATGCTTGCTGATAAAGTAAAGC
gpsBEBam	GCACCGGATGGATCCTCAATCATAAAGCTTGCTGCCAAAAAC
mscarletSKpn	GCGAACGCAGGTACCATGGTCTCCAAAGGAGAG
mscarletEPst	GCGAACGCACCTGCAGTTATTTATACAGCTCATCCATAC
mneongreenSKpn	GCGAACGCAGGTACCATGGTGAGCAAGGGCGA
mneongreenEPst	GCGAACGCACCTGCAGCTACTTGTACAGCTCGTC
kanSHind	GATGATGATAAGCTTTCGCCGTATGTAAGGATTTC
kanEHind	GATGATGCTAAGCTTCTAAAACAATTCATCCAGTAAAATATA