

Figure S1. *CXCR7* expression in DLBCL: (a) Expression analysis of *CXCR7* in non-neoplastic control germinal center B cells (GC-B) and diffuse large B cell lymphoma cells (DLBCL) consisting of DLBCL-NGCB and DLCBL-GCB by RQ-PCR. GCB-DLBCL were further subdivided into primary (DLBCL-pGCB) and transformed DLBCL (DLBCL-pGCB) originating from follicular lymphoma. (b) Expression analysis of *CXCR7* in DLBCL samples with early (stage 1) and advanced stage (stage 2–4) (left graph) and DLBCL samples with and without bone marrow infiltration (right graph) by RQ-PCR. mRNA expression levels were calculated as the relative expression in comparison to GC-B cells. The comparison of the expression levels was performed by using the Mann–Whitney U-test or Student's t-test.



Figure S2. Survival analysis based on expression levels of *CXCR4*, *CXCL12*, and *CXCR7*. (a) Probability of 5-yearsurvival in de novo DLBCL patients based on CXCR4 expression. (b,c) Probability of 5-year-survival in all (b) and de novo (c) DLBCL patients based on *CXCL12* expression. (d,e) Probability of 5-year-survival in all (d) and de novo (e) DLBCL patients based on *CXCR7* expression.



Figure S3. CXCR4 expression and CXCL12^{AF647} binding: (**a**) Surface expression of CXCR4 on SuDHL4, RI-1, U2932, and BL2 cell lines as determined by flow cytometry using a Phycoerythrin (PE) labeled antibody targeting CXCR4 (clone 2B11). (**b**) Percentage of CXCL12^{AF647} binding in the presence and absence of blocking antibodies against CXCR4 and CXCR7 as determined by flow cytometry. CXCL12 is bound by CXCR4 in all cell lines examined, while it is also bound by CXCR7 on RI-1 cells. IT denotes isotype control. * indicates significant reduction of CXCL12^{AF647} binding in the presence of antibodies against CXCR4 or CXCR7 compared to the binding isotype controls (p < 0.01).



Figure S4. CXCR4 antagonists and CXCL12^{AF647} binding. Percentage of CXCL12^{AF647} binding in the presence of AMD3100, AMD070, and its niacin derivative WK1 on BL2 cells. CXCL12^{AF647} binding to CXCR4 is inhibited by all three antagonists tested. * indicates significant differences of CXCL12^{AF647} binding in the presence of AMD070 and WK1, respectively, in comparison to AMD3100 (p < 0.01). ° indicates significant differences of CXCL12^{AF647} binding in the presence of CXCL12^{AF647} binding in the presence of CXCL12^{AF647} binding in the presence of AMD070 and WK1, respectively, in comparison to AMD3100 (p < 0.01). ° indicates significant differences of CXCL12^{AF647} binding in the presence of WK1 in comparison to AMD070 (p < 0.05).



Figure S5. CXCR4 antagonists and transmigration. Percentage of transmigration of BL2 (**left**) and U2932 (**right**) cells in the presence of AMD070 and its niacin derivative WK1, respectively, in comparison to transmigration with DMSO as control. Transmigration was inhibited in both cell lines using AMD070 while inhibition by WK1 was not that efficient. ° indicates significant inhibition of transmigration compared to the DMSO control (p < 0.05). ‡ indicates significantly lower inhibition of WK1 compared to AMD070 (p < 0.05).



Figure S6. CXCR4 antagonists and cell viability. Percentage of Annexin V/7AAD positive SuDHL4, RI-1, U2932 and BL2 cell lines in the presence of increasing concentrations of AMD3100, AMD070, and its niacin derivative WK1 as determined by flow cytometry and compared to DMSO treated control cells. Only treatment of BL2 and SUDHL4 cells with 10 μ M, 20 μ M, and 40 μ M WK1 led to a decrease in viable, Annexin V/7AAD negative cells, while all other cell lines or antagonists showed no effect. * indicates significantly reduced viability compared to AMD3100 (p < 0.05). ° indicates significantly reduced viability compared to AMD070 (p < 0.05).



Figure S7. Gene expression of pro- and anti-apoptotic BCL2 members upon treatment with AMD070 and WK1. (a) BL2 (as Burkitt model) and (b) U2932 (as NGCB-DLBCL model) were treated with 40μM AMD070 or its niacin derivative WK1 and after 1 h, 3 h, 6 h, and 12 h gene expression levels of (I) pro-apoptotic and (II) anti-apoptotic BCL2 family members were determined in comparison to the DMSO treated control cells by RQ PCR. mRNA expression levels were calculated as the relative expression in comparison to DMSO controls. Each bar represents the mean values of expression levels ± standard error of the mean (SEM). The comparison of the expression levels

was performed by using the Mann–Whitney U-test or Student's t-test. * Expression is significant to the DMSO control ($p \le 0.05$). ° Expression is significantly different compared to the AMD070 and WK1 treatment ($p \le 0.05$).



Figure S8. Gene expression of pro- and anti-apoptotic BCL2 members upon treatment with AMD3100. (a) BL2 and (b) U2932 cells were treated with 40 μ M AMD3100 and after 1 h, 3 h, 6 h, and 12 h gene expression levels of (I) pro-apoptotic and (II) anti-apoptotic BCL2 members were determined by RQ PCR. mRNA expression levels

were calculated as the relative expression in comparison to DMSO treated controls. Assays were carried out in duplicate. Each bar represents the mean values of expression levels \pm standard error of the mean (SEM). The comparison of the expression levels was performed by using the Mann–Whitney U-test or Student's t-test. * Expression is significant to the DMSO control ($p \le 0.05$).



Figure S9: Gene expression of JNK-, ERK1/2-, and NF- κ B/ BCR-target genes upon treatment with AMD070 and WK1. (a) BL2 (as Burkitt model) and (b) U2932 (as NGCB-DLBCL model) were treated with 40 μ M AMD070 or its niacin derivative WK1 and after 24 h gene expression levels of CCR7, IL-10, CFLAR, ADARB, CCL22, and FN as JNK-targets; cFOS, BUB1, MXD1, JUNB, cJUN, ETV5 and DUSP1 as ERK1/2; and RGS1, KLF10, TNF, BCL2A1, OAS1, and CCL4 as NF- κ B/ BCR targets were determined in comparison to the DMSO treated control cells by RQ PCR. mRNA expression levels were calculated as the relative expression in comparison to the DMSO controls. Each bar represents the mean values of the expression levels \pm standard error of the mean (SEM). The comparison of the expression levels was performed by using the Mann–Whitney U-test or Student's t-test. * Expression is significant to the DMSO control ($p \le 0.05$). ° Expression is significantly different when comparing the AMD070 and WK1 treatment ($p \le 0.05$).







Figure S11. Negative (I & III) and positive (II & IV) controls for (**a**) CXCR4 and (**b**) CXCR7. Reactive tonsils were used for testing the two antibodies. All images (magnification 20×) were captured using an Olympus BX51 microscope and an Olympus E-330 camera.

Lymphoma	Disease	BCL2 status	References	
cell lines				
SUDHL4	GCB-DLBCL	BCL-2-positive; major rearrangement in the BCL-2 gene	Klanova et al. 2015 Clinical Cancer Research [1]	Masir et al. 2010 Pathology [2]
RI-1	NGCB- DLBCL	BCL-2-positive; amplification of BCL-2	Klanova et al. 2015 Clinical Cancer Research [1]	Masir et al. 2010 Pathology [2]
U2932	NCGB- DLBCL	BCL-2-positive; overexpress BCL-2	Amini et al. 2002 Leukemia & Lymphoma [3]	Klanova et al. 2015 Clinical Cancer Research [1]
BL-2	Burkitt's Lymphoma	express BCL-2	Finke et al. 1992 Blood [4]	
Raji	Burkitt's Lymphoma	express BCL-2	Finke et al. 1992 Blood [4]	

Table S1. BCL2 status of the used lymphoma cell lines with type of DLBCL and references.

GAPDH3428-f	AAG GTC GGA GTC AAC GGA TTT
CAPDH3428-r	
HPRT1_f	
HPRT1 r	
	CAC CAC AIG CIT GCC AIC C
	Qiagen #Q100089817
CCR/	Qiagen #Q101666686
CD44	Qiagen #Q1000/3549
	Qiagen #Q100041685
MMP2	Qiagen #Q100088396
FNI	Qiagen #Q100038024
COLIA	Qiagen #Q100037793
CFLAR	Qiagen #Q100064554
ADARB	Qiagen #QT00081655
EGR3	Qiagen #QT00246498
cFOS	Qiagen #QT00007070
BUB1	Qiagen #QT00082929
MXD1	Qiagen #QT00082915
JUNB	Qiagen #QT00201341
cJUN	Qiagen #QT00242956
ETV5	Qiagen #QT00009485
DUSP1	Qiagen #QT00036638
CCL3-f	CTC CAA GCC CGG TGT CAT CT
CCL3-r	TTC TGG ACC CAC TCC TCA CT GG
CCL4-f	TAT GAG ACC AGC AGC CTC TG
CCL4-r	GCT TCT TTT GGT TTG GAA TAC C
KLF10-f	ATG CTC AAC TTC GGT GCC T
KLF10-r	TTC CAT TCT TTC CTC CGC
OAS3-f	CTG GTG TCC ACA GCC CTG AA
OAS3-r	TGC CAG AAC TGA GCT GCC C
RGS1-f	AAC TTC TTG CCA ACC AAA CTG
RGS1-r	CAA GCC AGC CAG AAC TCA
TNF-f	AAG CCT GTA GCC CAT GTT GTA G
TNF-r	AGA TGA GGT ACA GGC CCT CTG A
BCI 2A1_f	
BCI 2A1_r	
BAD-f	
BAD r	
PLIMA f	
PAN F	
DAA-F	GAG GEE GIE CEA ACE AC
BCL-AL-I	
BCL-XL-r	
BCL-2-f	GGA GGA HIG IGG CCI ICI HIG
BCL-2-r	GCC GGT ICA GGT ACT CAG ICA I
MCL-1-t	CCA AGG ACA CAA AGC CAA TG
MCL-1-r	AAG AAC TCC ACA AAC CCA TCC
BIK-f	CTT GAT GGA GAC CCT CCT GTA TG
BIK-r	AGG GTC CAG GTC CTC TTC AGA
BAK-f	ATGGTCACCTTACCTCTGCAA
BAK-r	TCATAGCGTCGGTTGATGTCG
BIM Isoform 9-f	AAC CAC TAT CTC AGT GCA ATG G

Table S2. Oligonucleotide sequences of used primers.

BIM Isoform 9-r	TTG ACT ATG GTG GTG GCC A	
BID-f	GGA ACC GTT GTT GAC CTC AC	
BID-r	GAG GAG CAC AGT GCG GAT	
BMF-f	TTC AAA GCA AGG TTG TGC AG	
BMF-r	TTG TGG GGT GAC TGA GGA AC	
NOXA-f	AGC TGG AAG TCG AGT GTG CT	
NOXA-r	TCC TGA GCA GAA GAG TTT GGA	
CXCR4	Qiagen #QT00223188	
CXCR7	Qiagen #QT00069650	
CXCL12	Qiagen #QT01008133	
CXCR4-ex1 seq fw	CAGCAGGTAGCAAAGTGAC	
CXCR4-ex1 seq rev	TCAAGAAAACTCCTTTCGGTG	
CXCR4-ex2 seq fw	ATGGGAAAAGATGGGGAGG	
CXCR4-ex2 seq rev	AGACTCAGACTCAGTGGAAAC	

References

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