

Antiproliferative activity of Pt(IV) conjugates containing the Non-Steroidal Anti-Inflammatory Drugs (NSAIDs) ketoprofen and naproxen

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SUPPLEMENTARY MATERIAL

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Figure S4-Figure S5. NMR characterisation of 2-(6-metoxynaphtalen-2-yl)propanoyl chloride

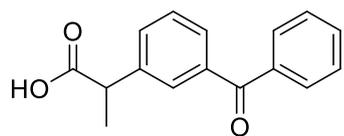
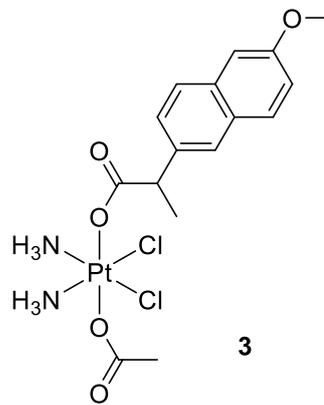
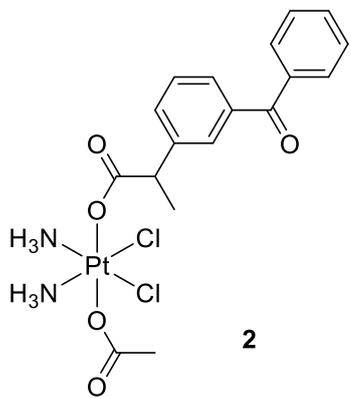
Figure S6-Figure S9. MS and NMR characterisation of complex **2**

Figure S10-Figure S13. MS and NMR characterisation of complex **3**

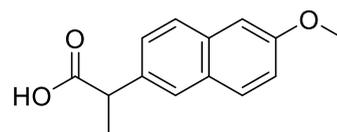
Figure S14-Figure S17. Residual viability (resazurin reduction assay) data of HCT 116 and HT-29 cells after treatment with cisplatin, NSAIDs and a cisplatin : NSAIDs 1 : 500 mixture. These data were used to obtain the Combination Index (CI) value with the method of Chou and Talalay.

Figure S18-Figure S19. Representative pictures of HCT 116 and A-549 cells 24 h after a treatment with **2** and **3**.

Table S1. Genes analyzed by means of Quantitative Reverse Transcription PCR (RT-qPCR).



ketoprofen



naproxen

Figure S1. Sketch of the complexes under investigation.

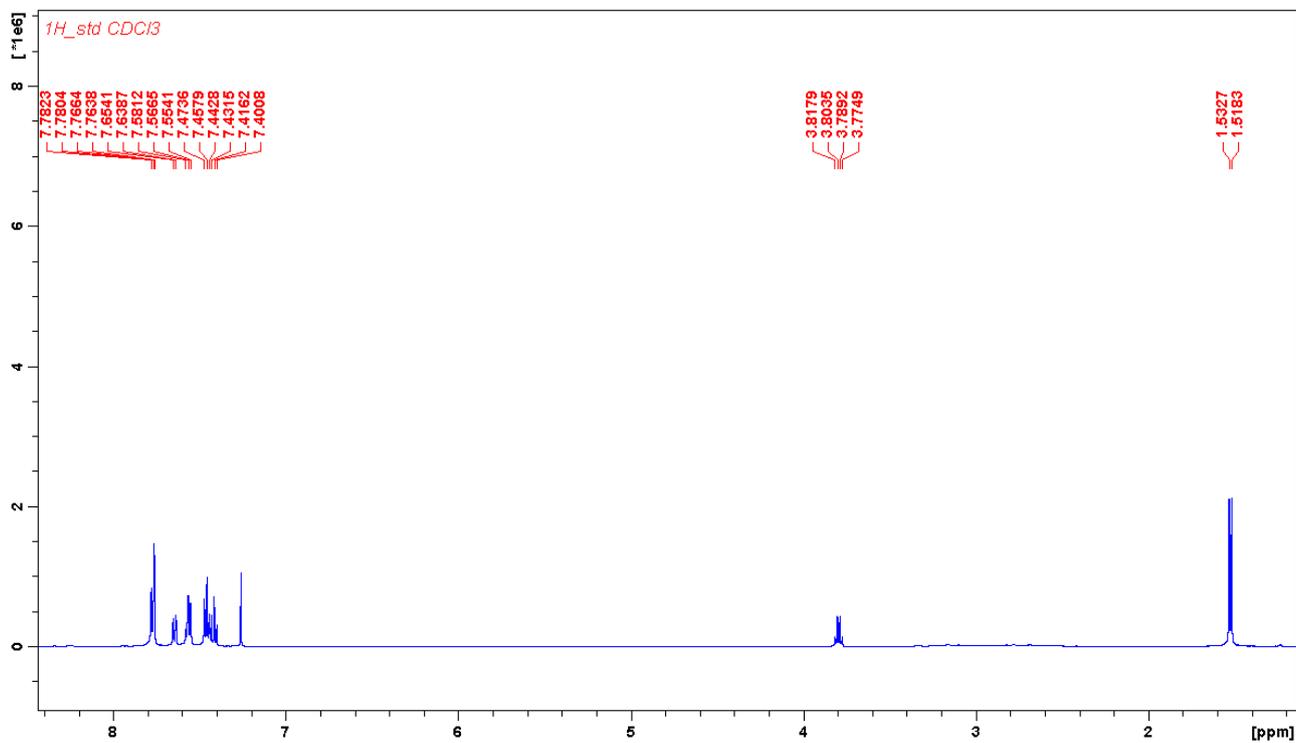
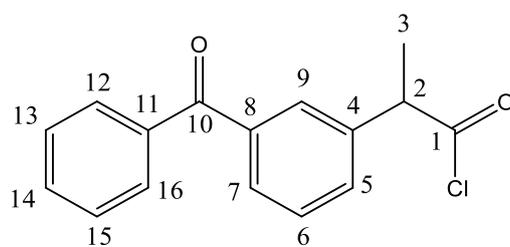


Figure S2. ¹H-NMR spectrum of (RS)-2-[3-(benzoyl)phenyl]propanoyl chloride in CDCl₃.

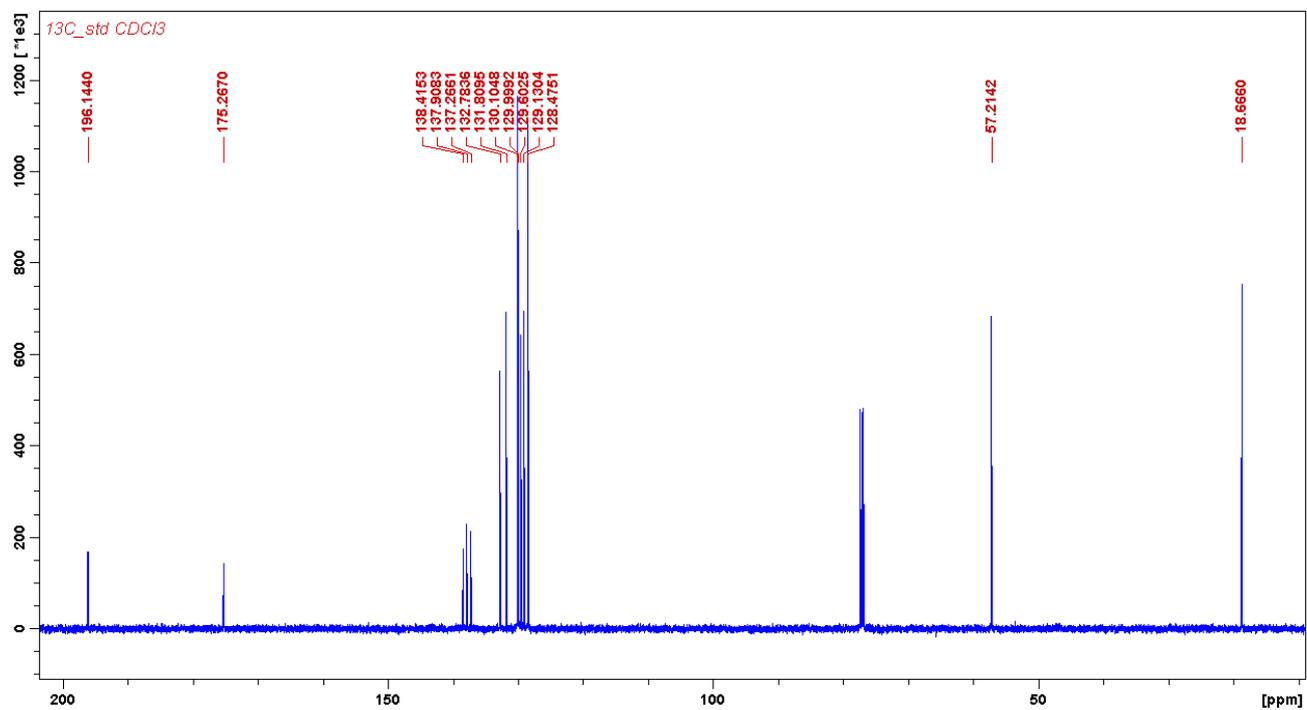


Figure S3. ¹³C-NMR spectrum of (RS)-2-[3-(benzoyl)phenyl]propanoyl chloride in CDCl₃.

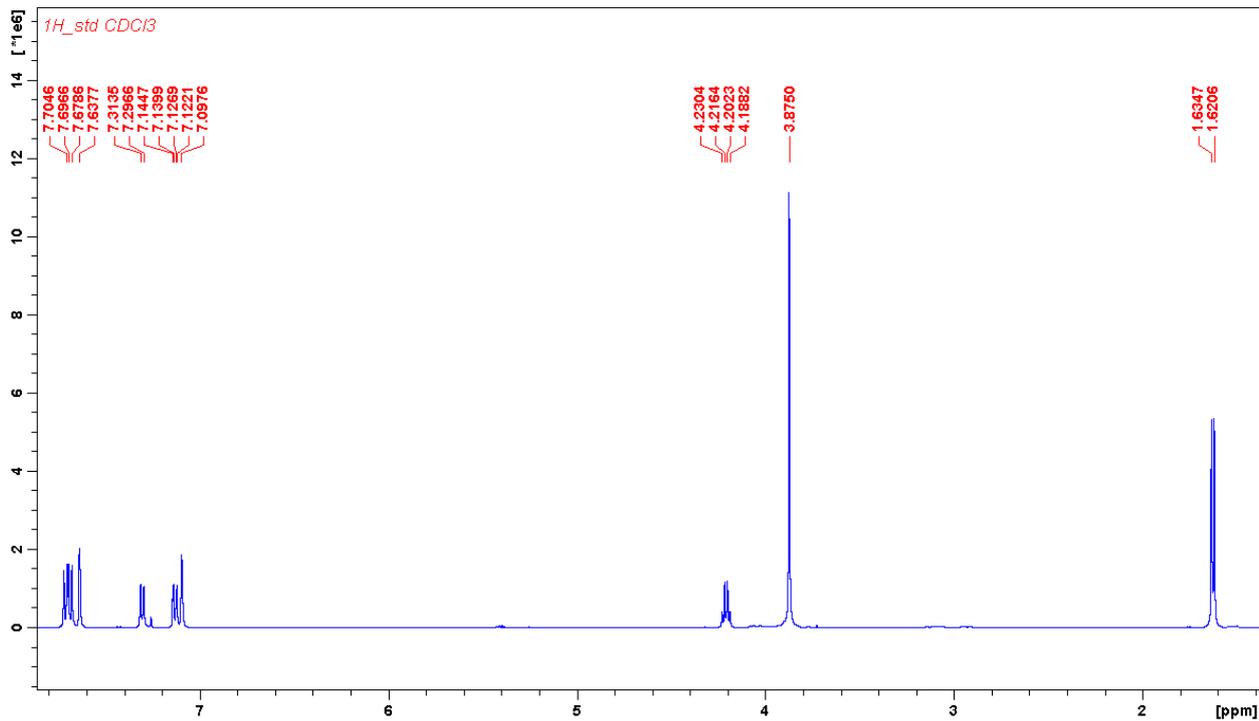
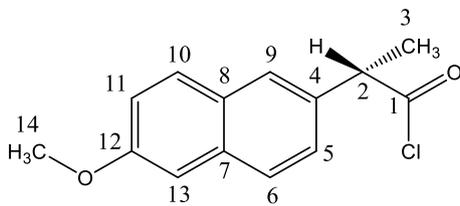


Figure S4. ¹H-NMR spectrum of 2-(6-methoxynaphthalen-2-yl)propanoyl chloride in CDCl₃.

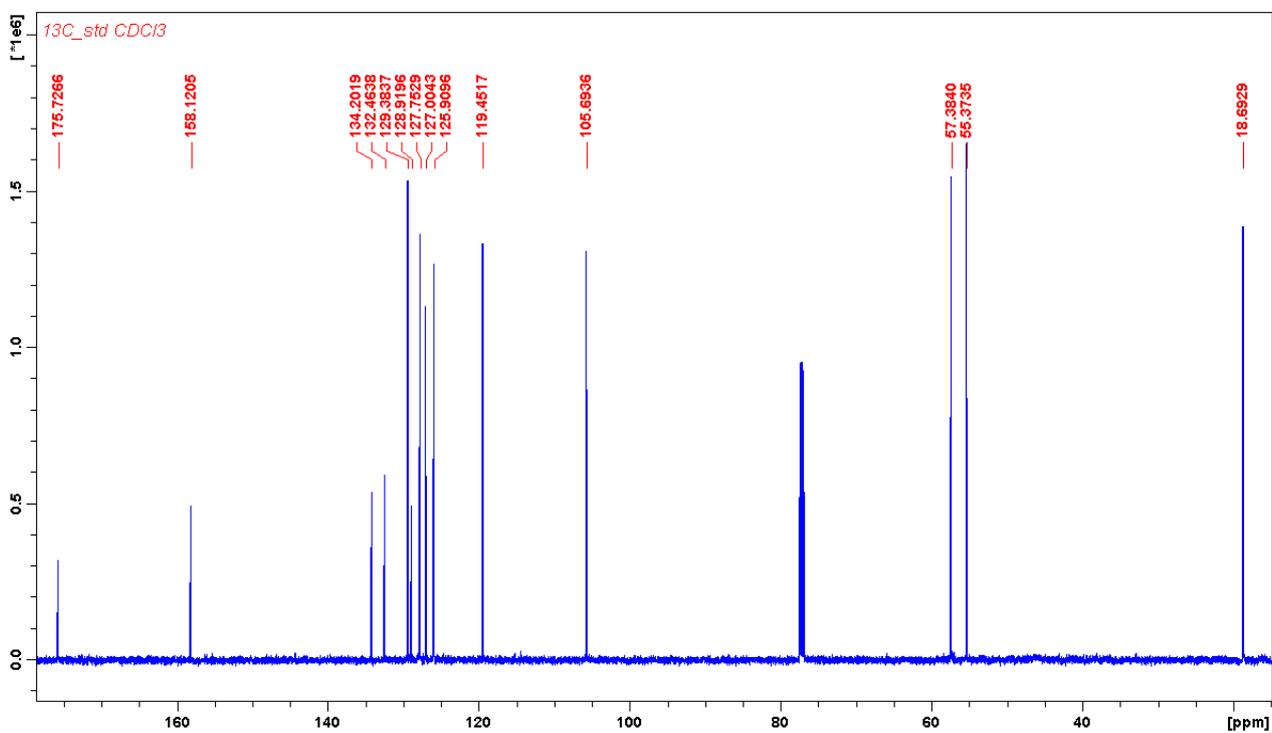


Figure S5. ¹³C-NMR spectrum of 2-(6-methoxynaphthalen-2-yl)propanoyl chloride in CDCl₃.

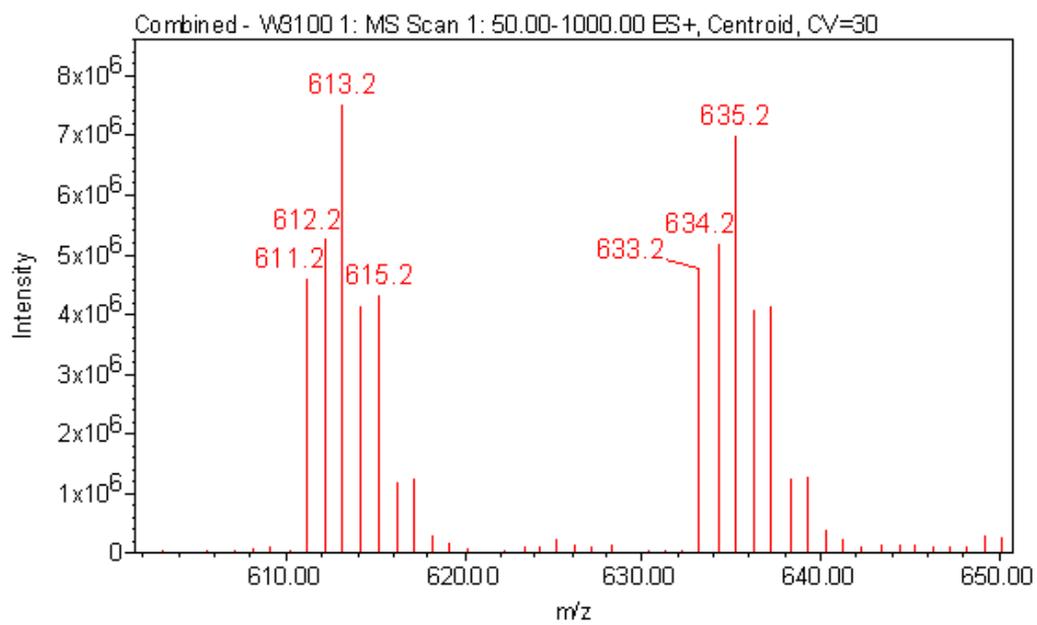


Figure S6. ESI-MS spectrum of complex **2**

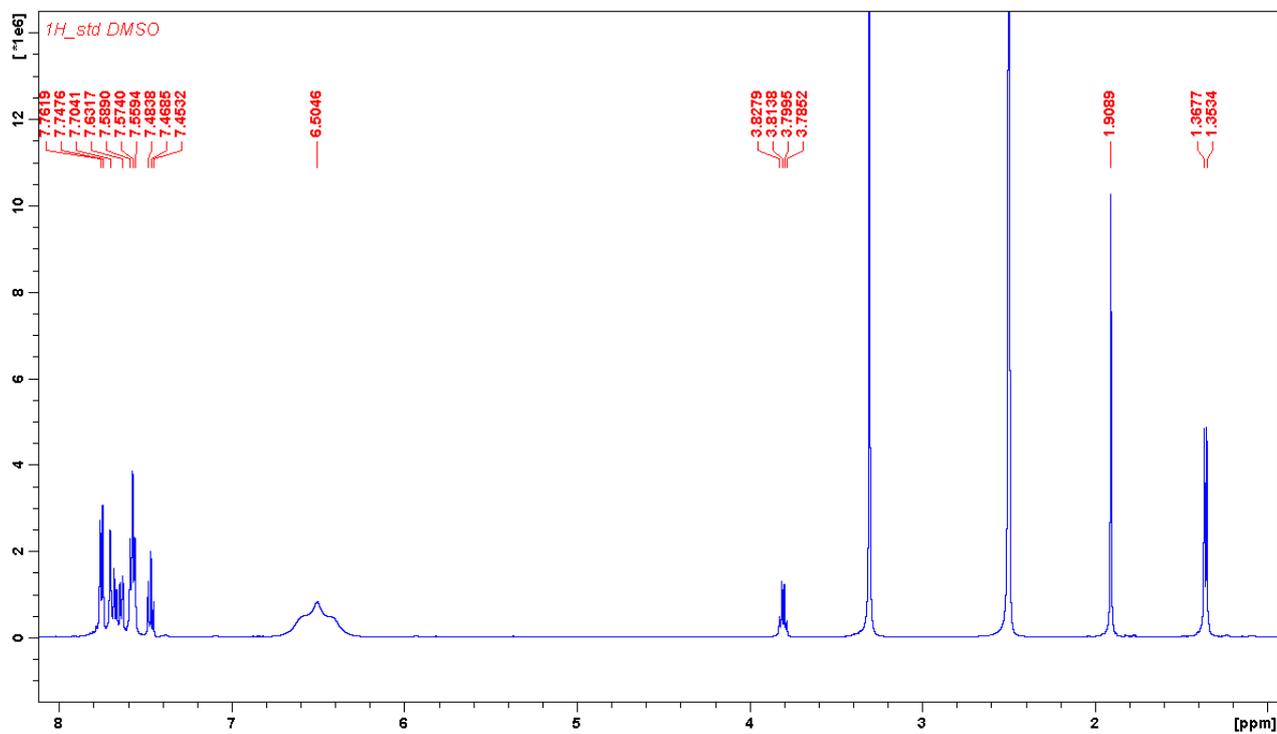
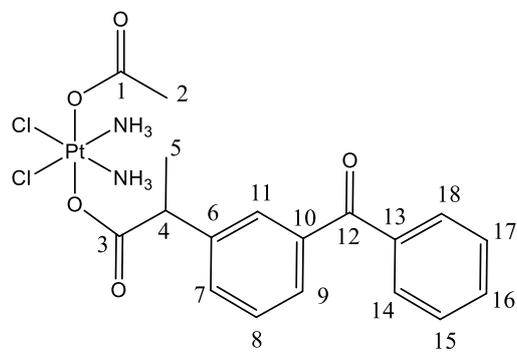


Figure S7. ^1H -NMR spectrum of complex **2** in DMSO-d_6 .

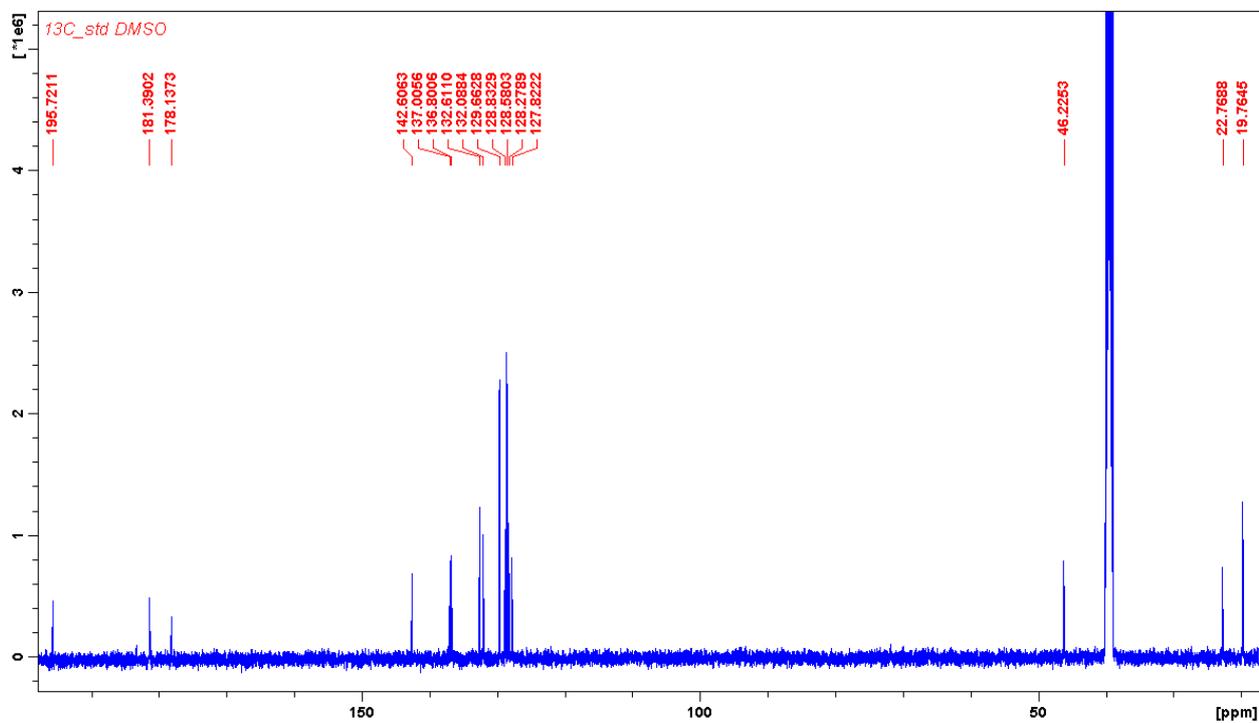


Figure S8. ^{13}C -NMR spectrum of complex **2** in DMSO- d_6

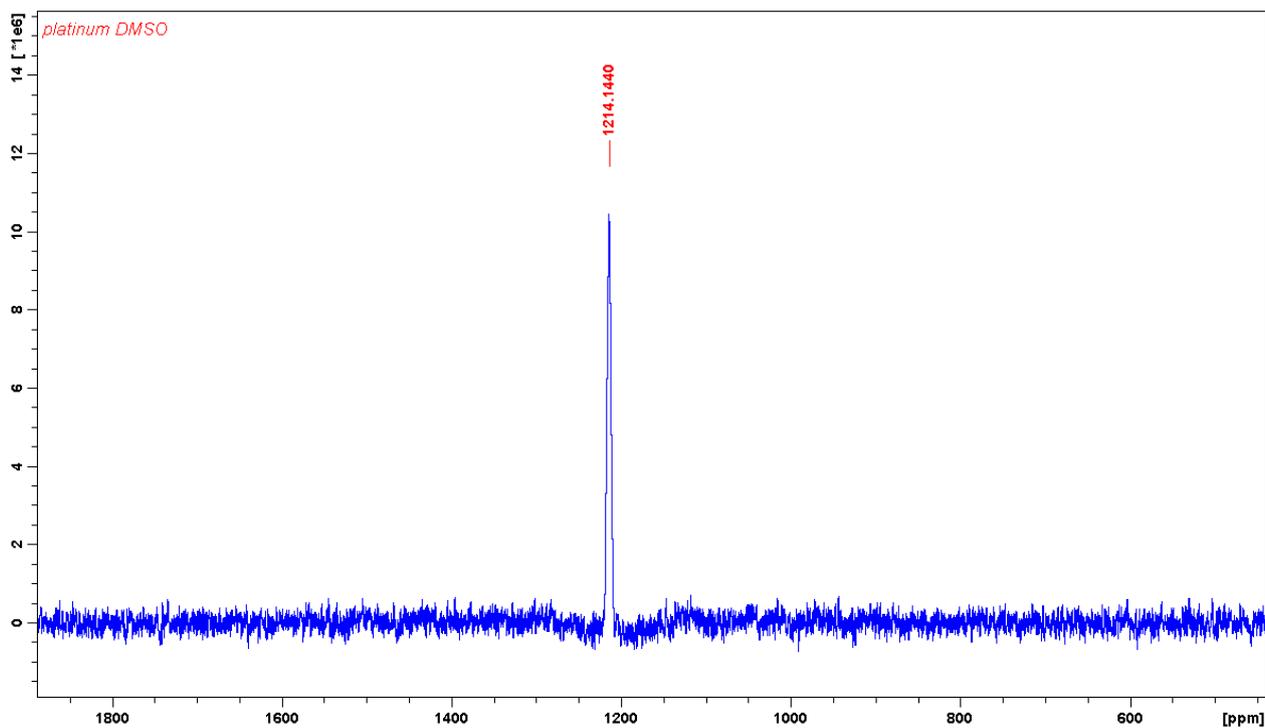


Figure S9. ^{195}Pt -NMR spectrum of complex **2** in DMSO- d_6

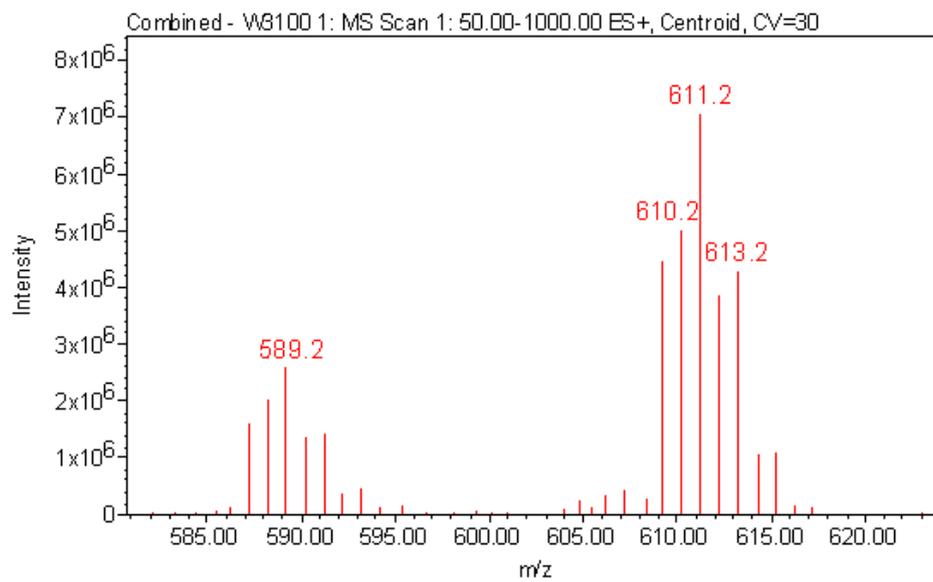


Figure S10. ESI-MS spectrum of complex **3**

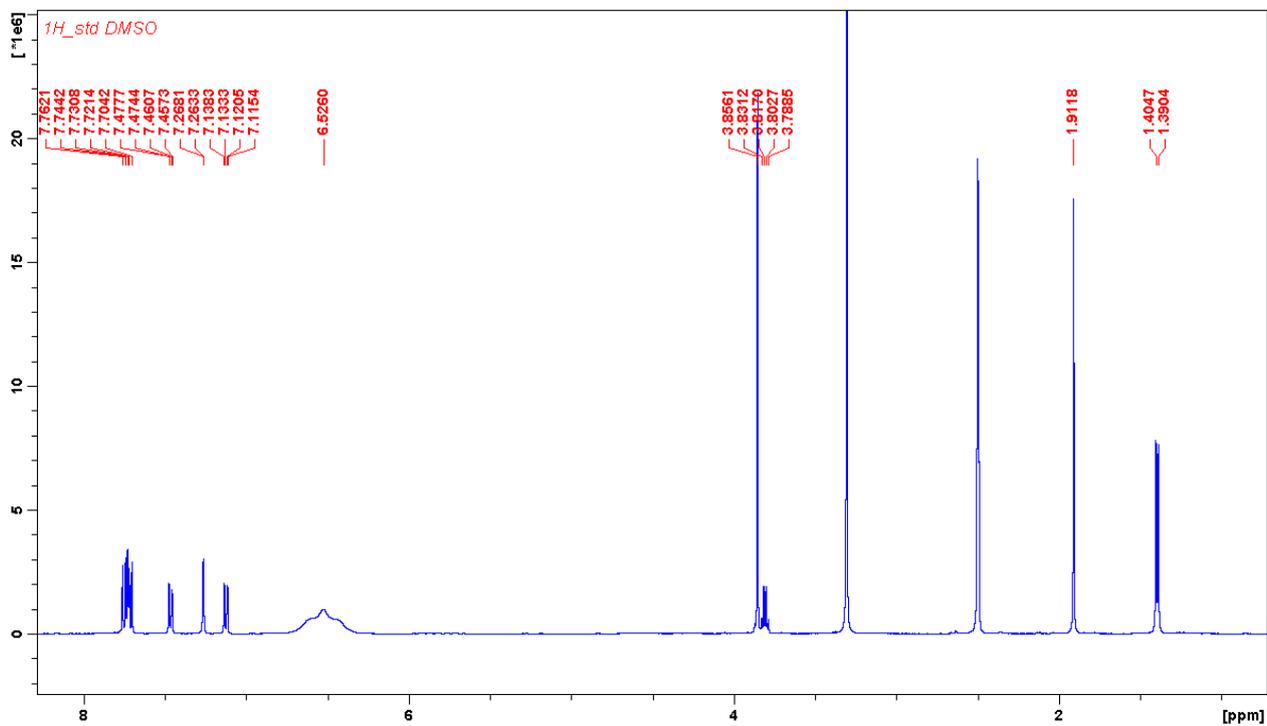
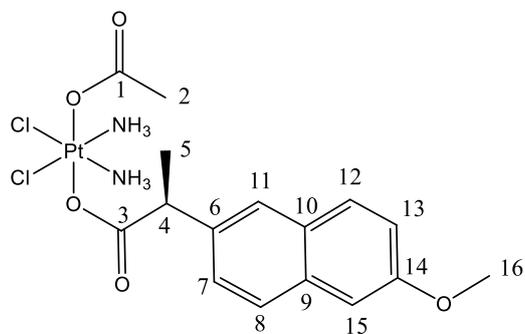


Figure S11. ^1H -NMR spectrum of complex **3** in DMSO-d_6 .

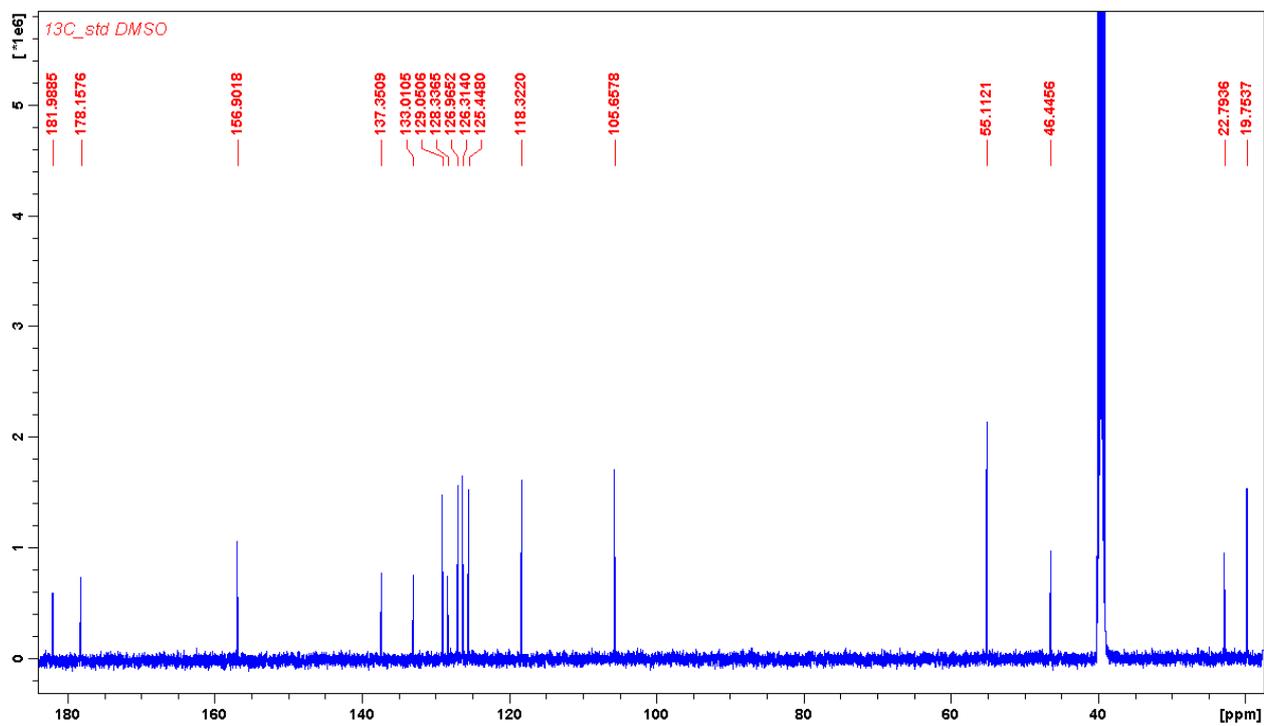


Figure S12. ^{13}C -NMR spectrum of complex **3** in DMSO- d_6 .

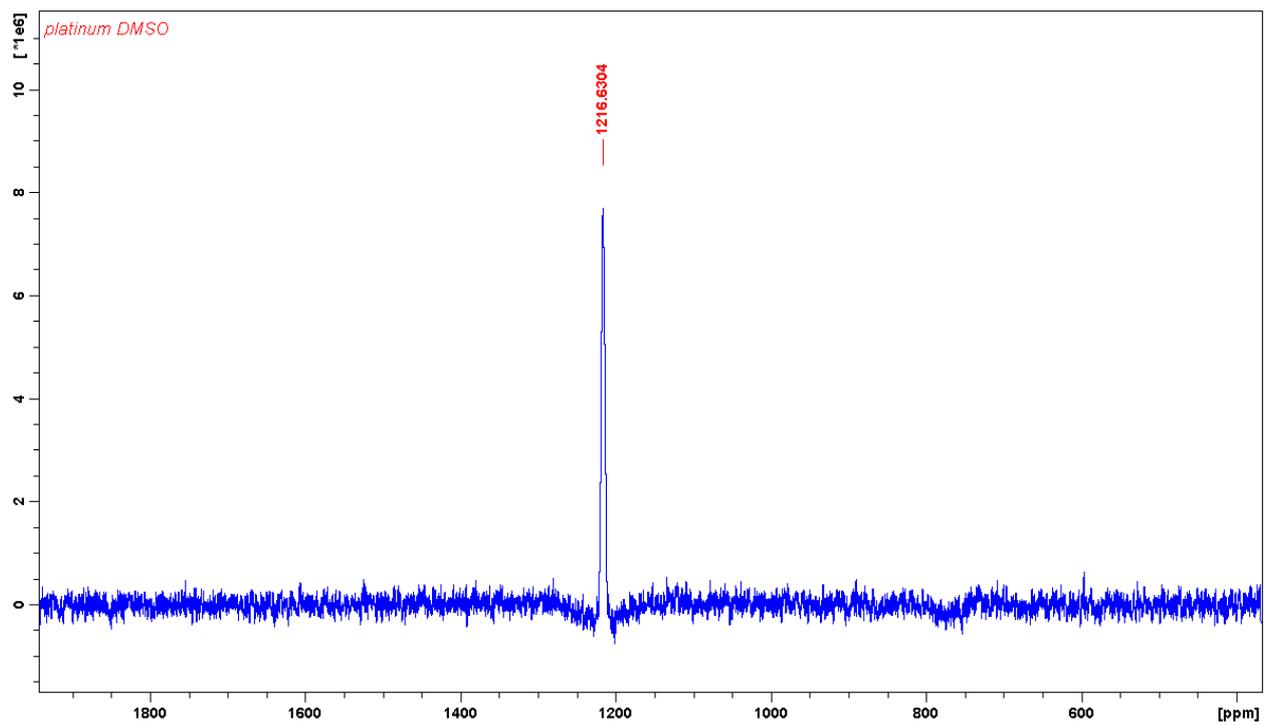


Figure S13. ^{195}Pt -NMR spectrum of complex **3** in DMSO- d_6 .

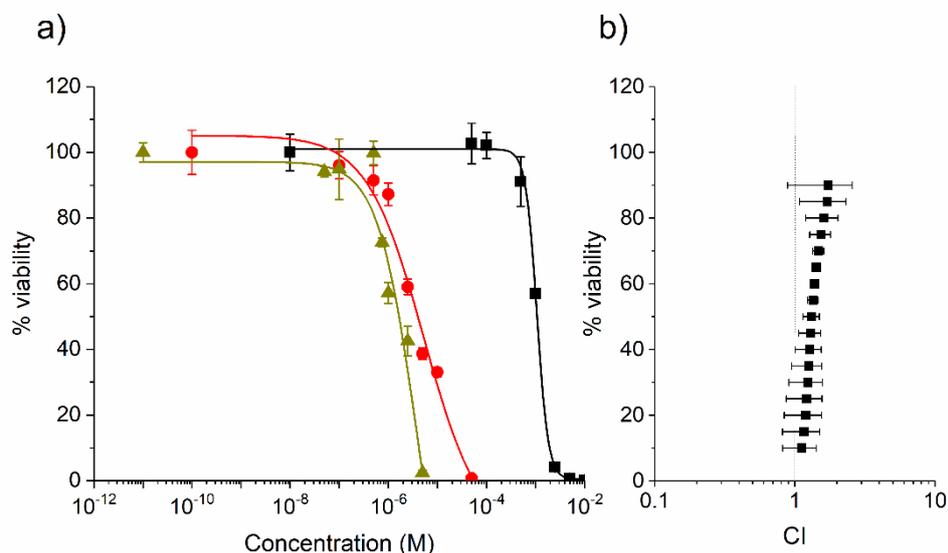


Figure S14. HCT116 cells were treated for 72 h with cisplatin (red dots), ketoprofen (black squares) or a cisplatin : ketoprofen mixture (in a fixed 1:500 ratio, according to their respective IC_{50} values, green triangles). a) Residual viability was assessed by means of the resazurin reduction assay and data were fitted with a four-parameter function. b) Residual viability data of the 1:500 mixture were compared to obtain the Combination Index (CI) value with the method of Chou and Talalay (equation for non-mutually exclusive drugs).

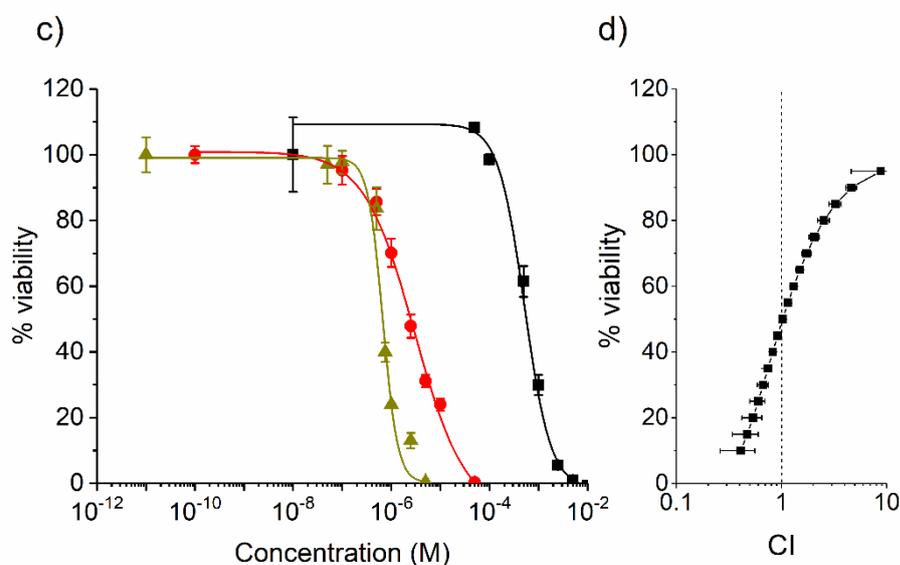


Figure S15. HCT116 cells were treated for 72 h with cisplatin (red dots), naproxen (black squares) or a cisplatin : naproxen mixture (in a fixed 1:500 ratio, according to their respective IC_{50} values, green triangles). c) Residual viability was assessed by means of the resazurin reduction assay and data were fitted with a four-parameter function. d) Residual viability data of the 1:500 mixture were compared to obtain the Combination Index (CI) value with the method of Chou and Talalay (equation for non-mutually exclusive drugs).

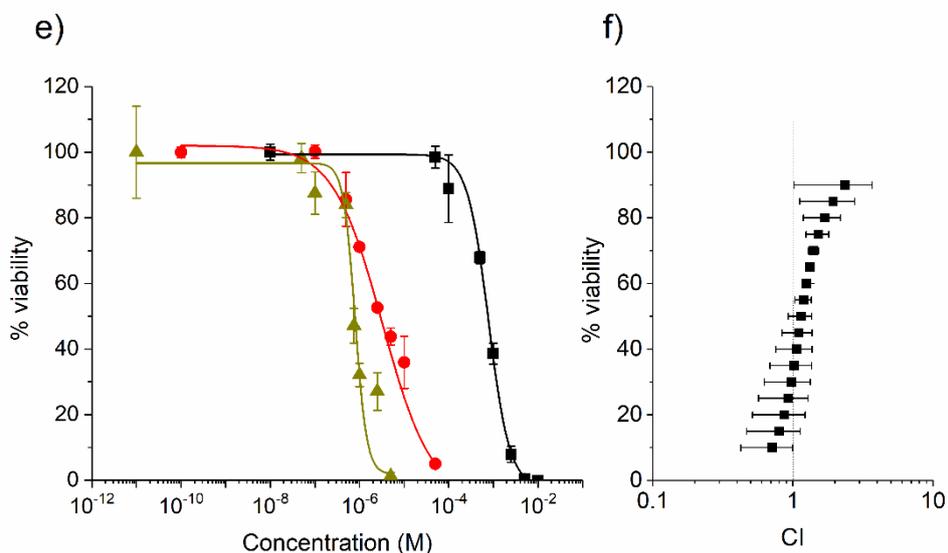


Figure S16. HT-29 cells were treated for 72 h with cisplatin (red dots), ketoprofen (black squares) or a cisplatin : ketoprofen mixture (in a fixed 1:500 ratio, according to their respective IC_{50} values, green triangles). e) Residual viability was assessed by means of the resazurin reduction assay and data were fitted with a four-parameter function. f) Residual viability data of the 1:500 mixture were compared to obtain the Combination Index (CI) value with the method of Chou and Talalay (equation for non-mutually exclusive drugs).

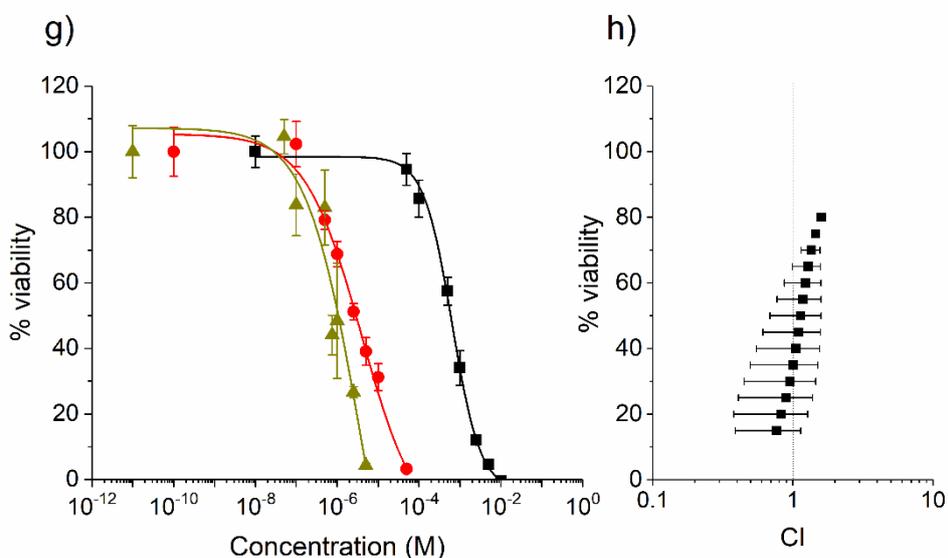


Figure S17. HT-29 cells were treated for 72 h with cisplatin (red dots), naproxen (black squares) or a cisplatin : naproxen mixture (in a fixed 1:500 ratio, according to their respective IC_{50} values, green triangles). g) Residual viability was assessed by means of the resazurin reduction assay and data were fitted with a four-parameter function. h) Residual viability data of the 1:500 mixture were compared to obtain the Combination Index (CI) value with the method of Chou and Talalay (equation for non-mutually exclusive drugs).

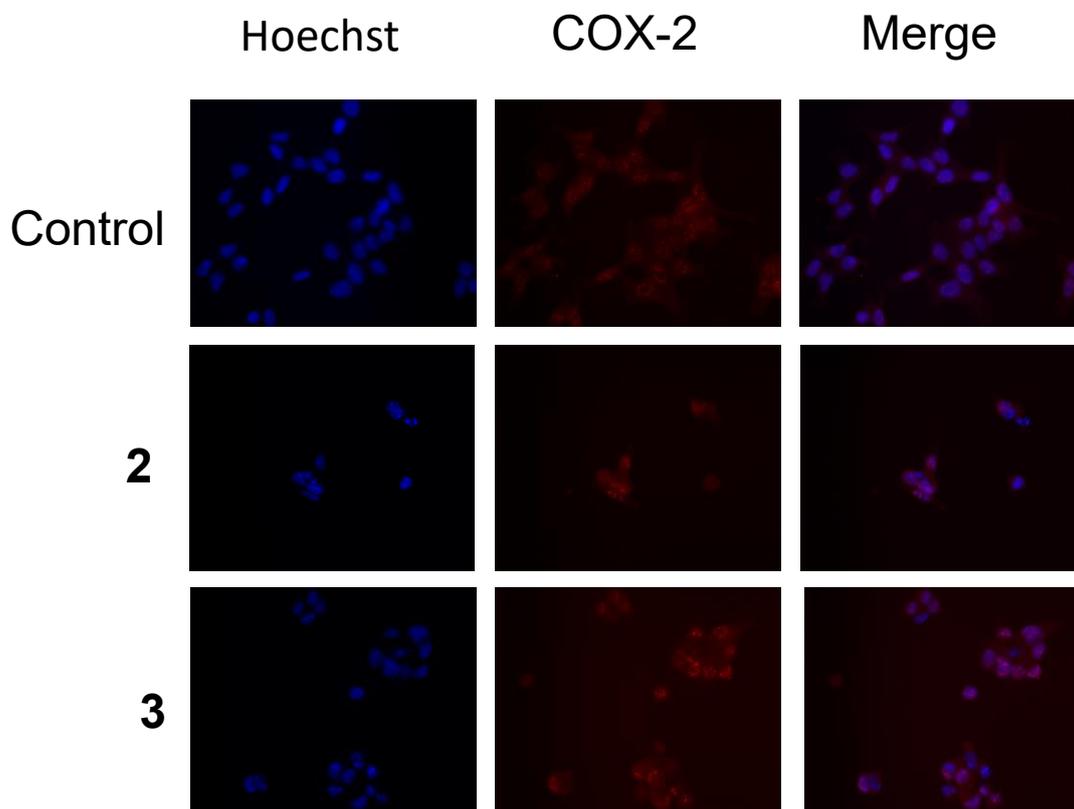


Figure S18. Representative pictures of HCT 116 cells 24 h after a 3 μ M treatment with **2** and **3** (from top to bottom: control, **2**, and **3**). Nuclei were stained with Hoechst 33258 (left), COX-2 was revealed with an Alexa 594-conjugated secondary antibody (middle), and images were merged (right). Pictures taken by means of Olympus BX51 microscope at 40 \times magnification. (For the description of the experiments see note after Figure S19.)

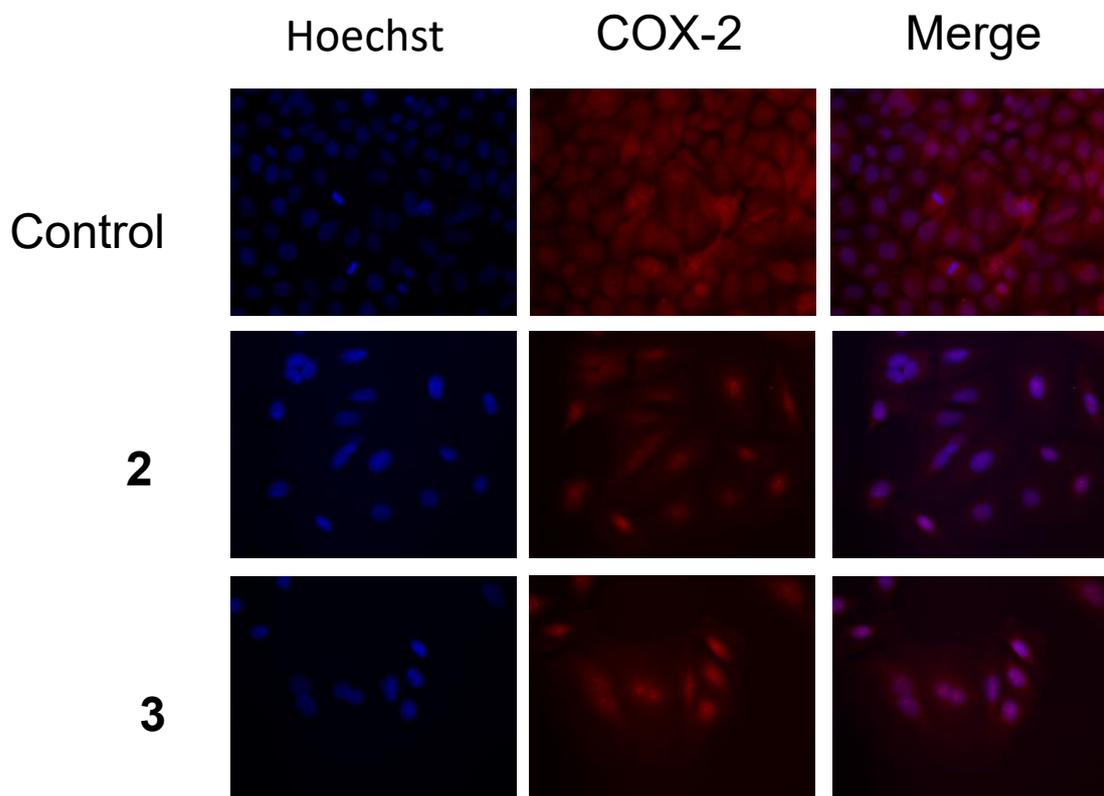


Figure S19. Representative pictures of A-549 cells 24 h after a 5 μM treatment with **2** and **3** (from top to bottom: control, **2**, and **3**). Nuclei were stained with Hoechst 33258 (left), COX-2 was revealed with an Alexa 594-conjugated secondary antibody (middle), and images were merged (right). Pictures taken by means of Olympus BX51 microscope at 40 \times magnification.

Immunochemical analysis. HCT 116 and A-549 cells (5×10^5) were seeded on coverslips and allowed to grow for 24 h. The cells were treated with equitoxic concentrations of the compounds under investigation (*i.e.*, HCT 116: 3 μM for both complexes **2** and **3**; A-549: 5 μM for both complexes **2** and **3**). After 24 h, the samples were fixed with 4% formalin and post-fixed with 70% ethanol at -20 $^{\circ}\text{C}$ for at least 24 h. After rehydration in PBS for 10 minutes, the cells were incubated with a primary antibody against COX-2 (Santa Cruz Biotechnology), at 1:200 dilution, for 60 min. Then, the coverslips were washed three times with PBS and incubated with Alexa 594-conjugated anti-goat secondary antibody (Molecular Probes) at 1:200 dilution. All the incubations were performed in the dark at room temperature. Finally, the sections were counterstained for DNA with 0.1 $\mu\text{g mL}^{-1}$ Hoechst 33258 (Sigma-Aldrich, Milano, Italy) for 10 minutes, washed with PBS, and mounted in a drop of Mowiol (Calbiochem, Inalco, Italy) for fluorescence microscopy analysis. An Olympus BX51 microscope equipped with a 100-W mercury lamp was used under the following conditions:

- for Hoechst 33258: 330–385 nm excitation filter, 400 nm dichroic mirror, and 420 nm barrier filter
- for Alexa 594: 540 nm excitation filter, 580 nm dichroic mirror, and 620 nm barrier filter.

Images were recorded with an Olympus MagniFire camera system and processed with the Olympus Cell F software.

Table S1. Genes analyzed by means of Quantitative Reverse Transcription PCR (RT-qPCR). The NCBI accession number is reported along with the 5'-3' sequence of the forward and reverse primer and the expected product length.

Gene	Accession no.	Forward	Reverse	Product length (bp)
COX-2	M90100.1	CCCTGAGCATCTACGGTTTG	CATCGCATACTCTGTTGTGTTC	107
GAPDH	NG_007073.2	ATCCCTGAGCTGAACGGGAA	GGCAGGTTTTTCTAGACGGC	99
HPRT1	NM_000194.2	TTGCTTTCCTTGGTCAGGCA	ATCCAACACTTCGTGGGGTC	85
RNA18SN1	NR_145820.1	CGTCTGCCCTATCAACTTTCG	TGCCTTCCTTGGATGTGGTAG	124
BAX	NM_001291428.1	GACCATCTTTGTGGCGGGAG	GAGGAAAAACACAGTCCAAGGC	94
BAD	NM_004322.3	GAGACCTGTGCGCCGTCA	AGGACCTCAGTCTCCCCTCAG	74
Bcl2a	NM_000633.2	CTTTGAGTTCGGTGGGGTCA	GGGCCGTACAGTTCCACAAA	162
NAG-1/GDF15	NM_004864.3	TTGCGGAAACGCTACGAGG	GCACTTCTGGCGTGAGTATCC	115

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