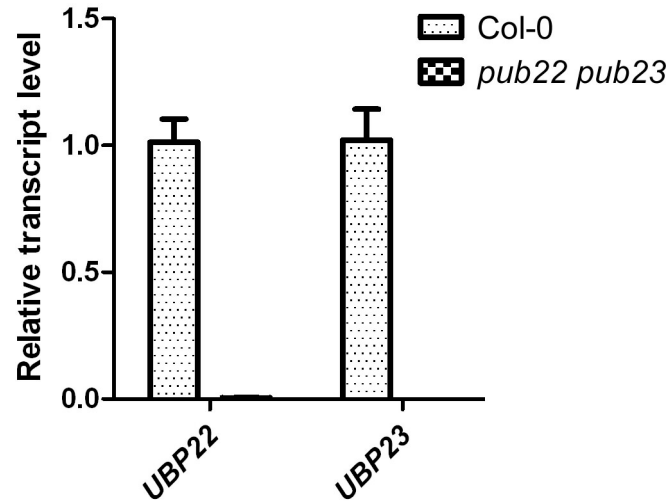
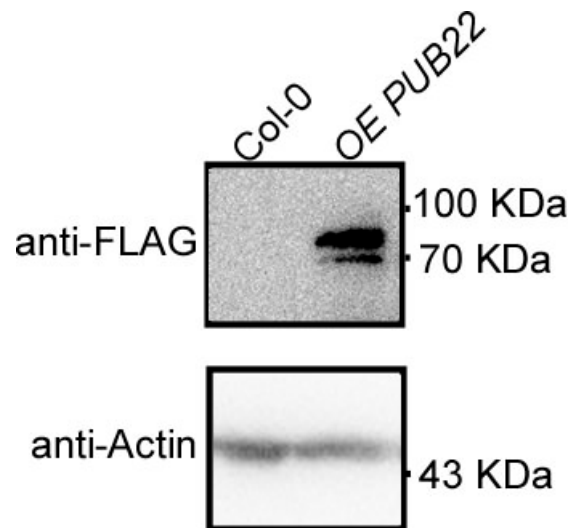
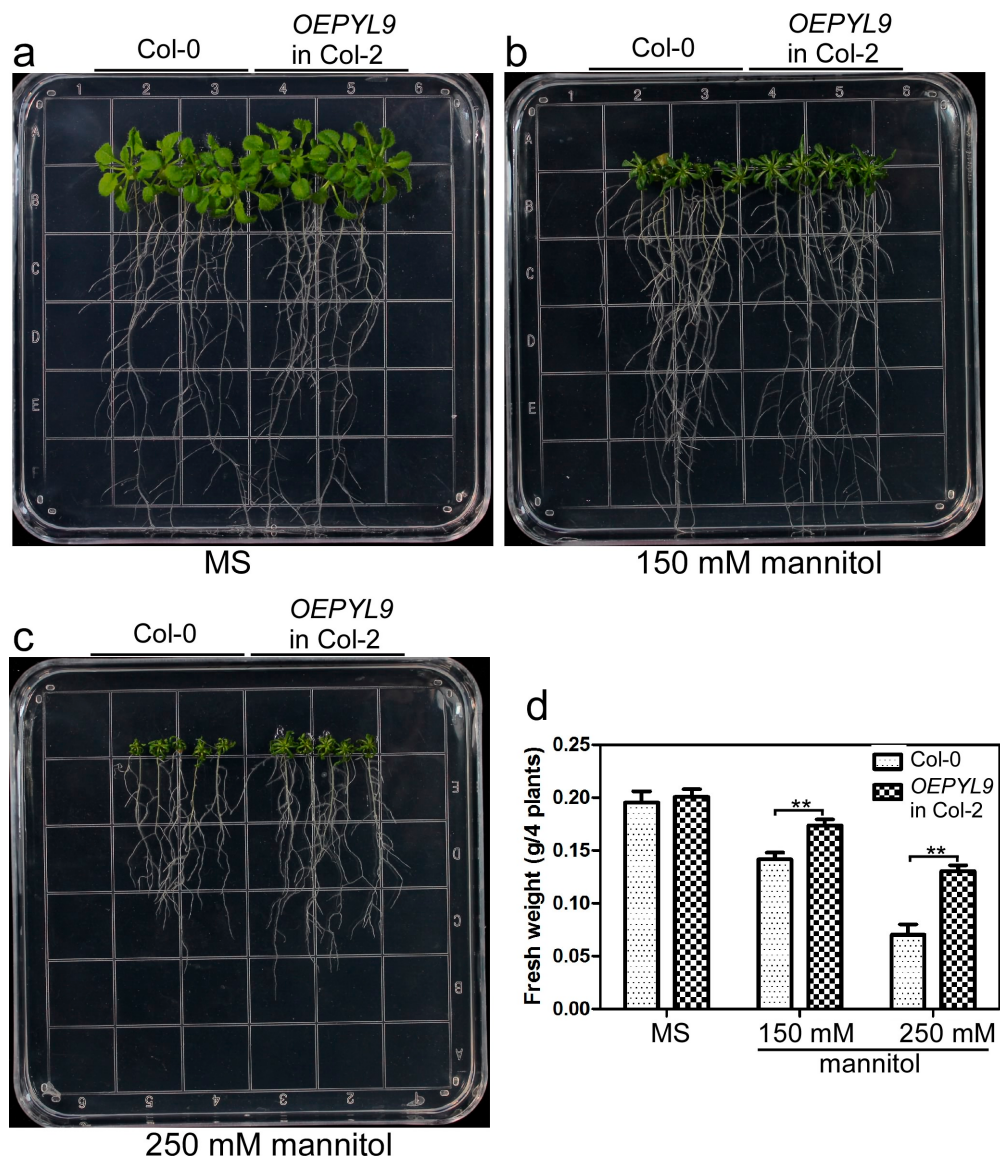




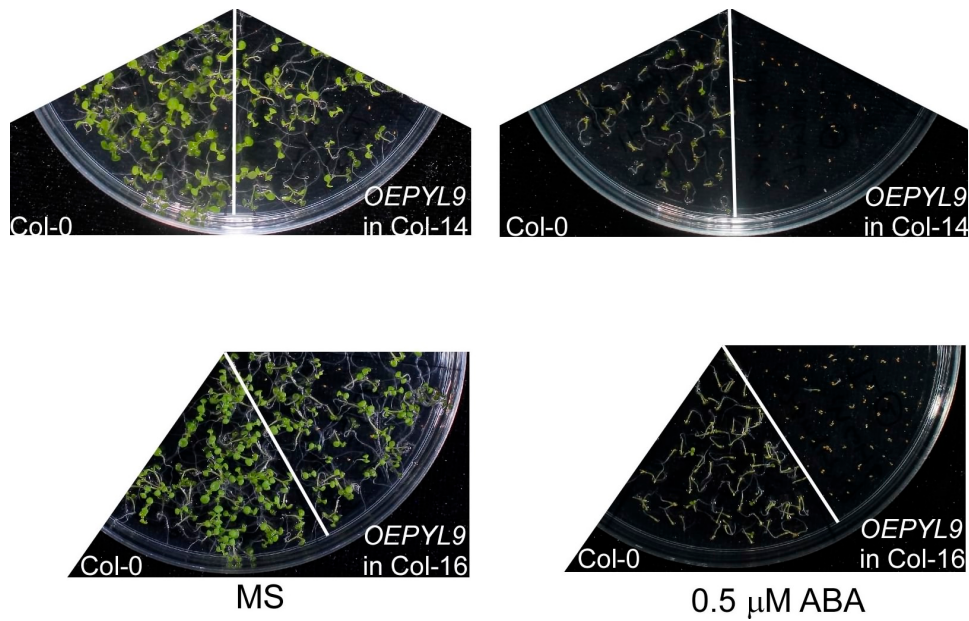
Supplementary Figure S1. Expression of *PUB22* and *PUB23* in the *pub22 pub23* double mutant. Quantitative real-time PCR determined the expression of *PUB22* and *PUB23* in *pub22 pub23* double mutant. The *pub22* (SALK_072621) and *pub23* (SALK_063470) single mutants[35]. Data are the means $\pm$ standard errors ($n = 3$).



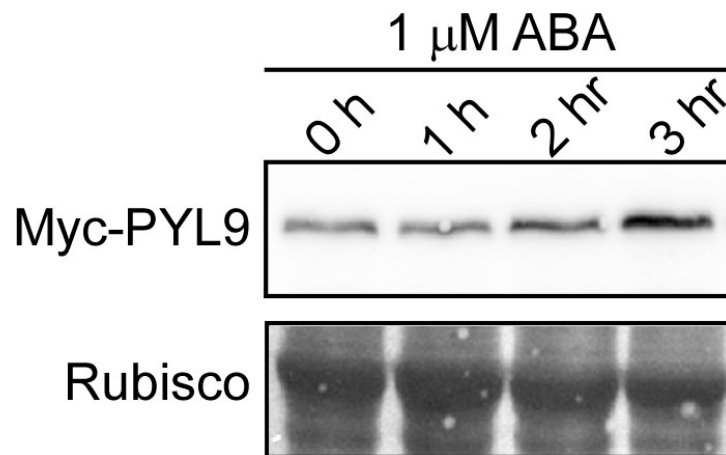
Supplementary Figure S2. The expression of *FLAG-PUB22* in transgenic plants. The expression of *FLAG-PUB22* in transgenic plants was analyzed by immunoblotting with anti-FLAG antibody.



Supplementary Figure S3. The seedling growth of the wild-type and the transgenic seedlings harboring *Myc-PYL9* on MS with or without mannitol. (a), (b) and (c). Seven-day-old seedlings grown on MS medium were transferred to the medium with different concentration of mannitol for 12 days. (d) Quantitative evaluation of the fresh weight of seedlings in (a), (b) and (c). Asterisks indicate significant differences ($p < 0.05$).



Supplementary Figure S4. The seeds germination of the wild-type and the independent transgenic seeds with *Myc-PYL9* on MS with or without 0.5 μM ABA. The seeds were vernalized at 4 $^{\circ}\text{C}$ for 3 days and grown on the chamber at 22 $^{\circ}\text{C}$ for 9 days.



Supplementary Figure S5. Accumulation of the Myc-PYL9 protein induced by ABA. The transgenic plants containing *Myc-PYL9* in Col-0 background were treated by 1 μM ABA and the samples were harvested at the indicated time points. Coomassie brilliant blue (CBB) staining of Rubisco was used as loading control.

Supplementary Table 1. Primers used for plasmids construction.

Plasmids	Forward primer (5'-3')	Reverse primer (5'-3')
<i>pCambia1307-Myc-PYL9</i>	ACGCGTCGACATGATGGACGGCGTTGAAGGC	GGGGTACCTCACTGAGTAATGTCCTGAG
<i>pCambia1307-FLAG-PUB22</i>	ACGCGTCGACATGATGGATCAAGAGATAGAGATTC	GGGGTACCTCAAGCAGGATACGAATCATAC
<i>BD-PYL1</i>	CATGGAGGCCGAATTCATGCCTTCGGAGTTAACACC	GCAGGTCGACGGATCCTCACGTCACCTGAGAACCAC
<i>BD-PYL1</i>	CATGGAGGCCGAATTCATGGCGAATTCAGAGTCCTCC	GCAGGTCGACGGATCCTTACCTAACCTGAGAAGAGTTG
<i>BD-PYL2</i>	CATGGAGGCCGAATTCATGAGCTCATCCCCGGCCGTG	GCAGGTCGACGGATCCTTATTCATCATCATGCATAGGT
<i>BD-PYL3</i>	CATGGAGGCCGAATTCATGAATCTTGCTCCAATCCATG	GCAGGTCGACGGATCCTCAGGTCGGAGAAGCCGTG
<i>BD-PYL4</i>	CATGGAGGCCGAATTCATGCTTGCCGTTACCCGTCC	GCAGGTCGACGGATCCTCACAGAGACATCTTCTTCTTG
<i>BD-PYL5</i>	ACGAGGTCGGAATTCTCGCTGAGG	CGCGTCGACATAACTAATCATCAATTTGCC
<i>BD-PYL6</i>	CATGGAGGCCGAATTCATGCCAACGTCGATACAGTTTC	GCAGGTCGACGGATCCTTACGAGAATTTAGAAGTGTCTC
<i>BD-PYL7</i>	CATGGAGGCCGAATTCATGGAGATGATCGGAGGAGAC	GCAGGTCGACGGATCCTCAAAGGTTGGTTTCTGTATG
<i>BD-PYL8</i>	CATGGAGGCCGAATTCATGGAAGCTAACGGGATTGAG	GCAGGTCGACGGATCCTTAGACTCTCGATTCTGTCGTG
<i>BD-PYL9</i>	CATGGAGGCCGAATTCATGGACGGCGTTGAAGGC	GCAGGTCGACGGATCC TCACTGAGTAATGTCCTGAG
<i>BD-PYL10</i>	CATGGAGGCCGAATTCATGAACGGTGACGAAACAAAGAAG	GCAGGTCGACGGATCCTCATATCTTCTTCTCCATAGATTC
<i>BD-PYL13</i>	CATGGAGGCCGAATTCATGGAAAGTTCTAAGCAAAAACG	GCAGGTCGACGGATCCTTACTTCATCATTTTTCTTTGTGAGC
<i>AD-PUB18</i>	GGAGGCCAGTGAATTCAGTCATAGCAGCATGATCCATACG	CGAGCTCGATGGATCCCCCGAGCTAAATATACAAACA

<i>AD-PUB22</i>	GGAGGCCAGTGAATTCATGGATCAAGAGATAGAGATT	CGAGCTCGATGGATCCTCAAGCAGGATACGAATCATAC
<i>AD-PUB23</i>	GGAGGCCAGTGAATTCATGTCCGGAGGAATAATGGATG	CGAGCTCGATGGATCCTCAGCAGGGATATGCAAGAATC
<i>pCold-GST-PYL5</i>	CGAGGGATCCGAATTCATGAGGTCACCGGTGCAACTC	TAGACTGCAGGTCGACTTATTGCCGGTTGGTACTTCG
<i>pCold-GST-PYL7</i>	CGAGGGATCCGAATTCATGGAGATGATCGGAGGAGAC	TAGACTGCAGGTCGACTCAAAGGTTGGTTTCTGTATG
<i>pCold-GST-PYL8</i>	CGAGGGATCCGAATTCATGGAAGCTAACGGGATTGAG	TAGACTGCAGGTCGACTTAGACTCTCGATTCTGTCTGTG
<i>pCold-GST-PYL9</i>	CGAGGGATCCGAATTCATGATGGACGGCGTTGAAGGC	TAGACTGCAGGTCGACTCACTGAGTAATGTCCTGAG
<i>pCold-GST-PYL10</i>	CGAGGGATCCGAATTCATGAACGGTGACGAAACAAAGAAG	TAGACTGCAGGTCGACTCATATCTTCTTCTCCATAGATTC
<i>pCold-MBP-PUB22</i>	CGAGGGATCCGAATTCATGGATCAAGAGATAGAGATTC	TAGACTGCAGGTCGACTCAAGCAGGATACGAATCATAC
<i>pCold-MBP-PUB22C13A</i>	CTTCCTTCTTCTTGCTCCAATCTCTCTAG	CTAGAGAGATTGGAGCAAGGAAGAAGGAAG
<i>pCold-MBP-PUB23</i>	CGAGGGATCCGAATTCATGTCCGGAGGAATAATGGATG	TAGACTGCAGGTCGACTCAGCAGGGATATGCAAGAATC
<i>pCold-MBP-PUB23C18A</i>	CCTCCGTTCTTCTTGCTCCTATCTCTTTGG	CCAAAGAGATAGGAGCAAGGAAGAACGGAGG
<i>pCAMBIA1300-PUB22C13A-NUC</i>	AGCTCGAGTAGTCGACATGGATCAAGAGATAGAG	ACGAGATCTGGTCGACAGCAGGATACGAATCATAC
<i>pCAMBIA1300-CLUC-PYL9</i>	ACGCGTCCCGGGGCGGTACCATGATGGACGGCGTTGAAGGC	GCCCTCTAGAGGATCCTCACTGAGTAATGTCCTGAG
<i>pCAMBIA1300-CLUC-OsNAC2</i>	TCCCGGGGCGGTACCATGGAAGTTGCCCCCTGGC	GCTCTGCAGGTCGACTTAGTAGCCCCATAGCGC
<i>pSPYCE (M)-PUB22C13A</i>	CGCCACTAGTGGATCCATCAAGAGATAGAGATT	TACCCTCGAGGTCGACAGCAGGATACGAATCATAC
<i>pSPYNE173-PYL9</i>	GCCTACTAGTGGATCCATGATGGACGGCGTTGAAGGC	TACCCTCGAGGTCGACTCACTGAGTAATGTCCTGAG

Supplementary Table 2. Primers used for the quantitative real-time PCR.

Plasmids	Forward primer (5'-3')	Reverse primer (5'-3')
<i>ACTIN</i>	ACTCTCCCGCTATGTATGTC	GATGGAAGAGCTGGTCTTTG
<i>PYL9</i>	AGCTCCTCTTCATCTCGTTTGG	AGACTGCCGATTTCAAGGATCAC
<i>Myc-PYL9</i>	ATGGAGAGCTTGGGCGACCTCAC	ATCACCATCGTTCCTGCTCTTC
<i>PUB22</i>	TTCGGTTGGGAGGTTCTG	CAAACCCTAGCGTGAAGC
<i>PUB23</i>	GGCCGTGGAAGCTGGAGTAATC	CCCTAACCGCTCTATCGCTTGC
<i>PUB24</i>	TAGAGCCGAGATTCTTGC	ACCGTCCCAACATTAACC
<i>NLUC</i>	TTCTATCCGCTGGAAGATGGAACC	TTCATAGCTTCTGCCAACCGAACG
<i>CLUC</i>	TGGATGGCTACATTCTGGAGAC	GGTGTTGGAGCAAGATGGATTC