Supporting Information

Spatiotemporal control of doxorubicin delivery from 'stealth'like prodrug micelles

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References

Synthesis of 1



Synthetic scheme to 1. (i) 4-nitrophenylchloroformate, Et₃N, CH₂Cl₂. (ii) Doxorubicin.HCl, Et₃N, DMF.

MethoxyPEG₂₀₀₀ 4-(4-(1-hydroxyethyl)-2-methoxy-5-nitrophenoxy)butanoate (2) was synthesized as previously reported ^[1].

MethoxyPEG₂₀₀₀ 4-(2-methoxy-5-nitro-4-(1-(((4-nitrophenoxy)carbonyl)oxy)ethyl) phenoxy)butanoate, 3

To a stirred solution of **2** (500 mg, 0.22 mmol) and 4-nitrophenyl chloroformate (265 mg, 1.31 mmol, 6eq.) in CH₂Cl₂ (20mL) was added Et₃N (305 μ L, 2.19 mmol, 10eq.). The reaction mixture was stirred at room temperature in the dark overnight. Following solvent removal *in vacuo*, purification by column chromatography (*Gradient*: CH₂Cl₂ to 15% MeOH in CH₂Cl₂) afforded **3** (278 mg, 0.11 mmol, 52%) as a yellow powder.

R_f: 0.30 (CH₂Cl₂:MeOH; 12:1)

¹**H** NMR (CDCl₃, 400 MHz): 8.26 (d, J = 8 Hz, ArH-o-NO₂, 2H); 7.61 (s, ArH-o-NO₂, 1H); 7.35 (d, J = 8Hz, ArH-m-NO₂, 2H); 7.11 (s, ArH-m-NO₂, 1H); 6.52 (q, J = 8 Hz, CH(CH₃)OCOO, 1H); 4.26 (m,COOC H_2 CH₂O, 2H); 4.14 (t, J = 8 Hz, OOCCH₂CH₂CH₂O, 2H); 4.00 (s, C H_3 O, 3H); 3.45-3.95 (m, OCC H_2 CH₂, 196H); 3.32 (s, C H_3 OCH₂CH₂O, 3H); 2.59 (m, OOCCH₂CH₂CH₂O, 2H); 1.78 (d, J = 8 Hz, CH(C H_3) OCOO, 3H).

MethoxyPEG₂₀₀₀ 4-(4-(1-(((3-hydroxy-2-methyl-6-(((1S,3S)-3,5,12-trihydroxy-3-(2hydroxyacetyl)-10-methoxy-6,11-dioxo-1,2,3,4,6,11-hexahydrotetracen-1-yl)oxy)tetrahydro-2H-pyran-4-yl)carbamoyl)oxy)ethyl)-2-methoxy-5-nitrophenoxy)butanoate, 1

To a stirred solution of **3** (86 mg, 0.034 mmol) and doxorubicin.HCl (20 mg, 0.037 μ mol) in DMF (500 μ L) was added Et₃N (47.2 μ L, 0.34 mmol, 10eq.). The reaction mixture was stirred at RT in the dark overnight. CH₂Cl₂ (20 mL) was then added to the reaction mixture and the solution washed with brine (15 mL). The organic fraction was dried (Na₂SO₄) and solvent removed *in vacuo*. Column chromatography (*Gradient*: CH₂Cl₂ to 2% MeOH in CH₂Cl₂ to 10% MeOH in CH₂Cl₂) yielded **1** (58.1 mg, 61%) as a red powder.

R_f: 0.20 (CH₂Cl₂:MeOH; 12:1)

¹**H NMR** (CDCl₃, 400 MHz): Partial peak assignment annotated in Figure S3. ¹H NMR of DOX with partial peak assignment included in Figure S2.

MS – despite numerous attempts to characterize this compound (MALDI, ESI), MS data was inconclusive – most likely due to compound instability and/or poor ionization of this compound during mass spec analysis. Following UV irradiation however, the MS of the photolysis products could be clearly detected (Figure S5 and S6). These products – nitroso-PEG and DOX – can only arise from the photolysis of **1**.



Figure S1. ¹H-NMR of **3**.



Figure S2. ¹H-NMR of **Doxorubicin**.



Figure S3. ¹H-NMR of **1**.



Figure S4. HPLC trace of 1. Retention time – 17.8 min. UV detection – 214 nm.



Figure S5. ESI-MS spectra (raw data) following photolysis of **1** and showing the expected photoproducts – DOX and nitroso-benzyl-PEG₂₀₀₀ – as the only significant species present. The presence of DOX clusters – $[2DOX]^+$ and $[3DOX]^+$ - in the raw spectra arise from 'soft' electrospray ionization techniques.



Figure S6. Deconvoluted (software: MaxEnt1) mass spectra of nitroso-PEG envelope signals.



Figure S7. *(left)* Time course DLS size distributions of **1** (300µM in PBS) diluted (1:1) in DMEM+FCS. *(right)* DLS size distributions of **1** (varying concentrations) in PBS.



Figure S8. Cells (bright field) irradiated for varying times (UV-A, 365nm, 15-17 mWcm⁻²) and imaged immediately. As UV-A irradiation times increase cells become smaller (shrinkage) and more rounded, hallmarks of the onset of UV-A induced apoptosis.²



Figure S9. FACS analysis showing increased uptake of DOX (released from a solution of 1 (300μ M in PBS)) by HeLa cells with increasing irradiation times. A) Dot plots of HeLa cells after *t*=0 ,5, 10 and 20 mins of irradiation; cell population was gated based on FSC-A vs SSC-A (cell doublets were gated out using FSC-A vs FSC-H). B) Histograms of HeLa cells after *t*=0 mins (pink) *t*=5 mins (blue), *t*=10 mins (orange) and *t*=20 mins (green) irradiation. C) Mean Fluorescence Intensity (MFI) of HeLa cells after different irradiation times. Error bars ± SD.

References

[1] Kong, L.; Askes, S. H. C.; Bonnet, S.; Kros, A.; Campbell, F. *Angew. Chem. Int. Ed.* **2016**, 55, 1396–1400.

[2] Elmore, S. Toxicol. Pathol. 2007, 35, 495-516.