

## Supplementary Information

### Determining the Value of $E_{PE}^{(50S)}$

From Figure 2 for the translation through the single-stranded mRNA with  $P_E = 1$ , the mean time of an elongation cycle under saturating concentrations of EF-G·GTP and the aminoacyl-tRNA·EF-Tu·GTP ternary complex is calculated by

$$\tau = \frac{k_1 + k_2 + k_3}{k_1 k_3} + \frac{1}{k_4} + \frac{1}{k_5} + \frac{k_7 + k_{-7} + k_8}{k_7 k_8} + \frac{1}{k_9} + \frac{1}{k_{10}} + \frac{1}{k_{11}} \quad (S1)$$

The translation rate is calculated by

$$v_0 = \frac{1}{\tau} \quad (S2)$$

Then, consider the translation through the mRNA duplex, with  $P_E < 1$ . Based on Figure 2, for the mRNA duplex with  $n$  base pairs next to the mRNA-entry channel of the ribosome being open spontaneously the translation rate under saturating concentrations of EF-G·GTP and the ternary complex is calculated by

$$v^{(n)} = \left[ \tau P_E^{(n)} + \tau P_E^{(n)} (1 - P_E^{(n)}) + \tau P_E^{(n)} (1 - P_E^{(n)})^2 + \dots + \tau_F (1 - P_E^{(n)}) + \tau_F (1 - P_E^{(n)})^2 + \dots \right]^{-1} \quad (S3)$$

where  $P_E^{(n)}$  is calculated by Equations (1)–(4),  $\tau$  is calculated by Equation (S1) and  $\tau_F$  is calculated by

$$\tau_F = \frac{k_1 + k_2 + k_3}{k_1 k_3} + \frac{1}{k_4} + \frac{1}{k_t} \quad (S4)$$

Equation (S3) can be rewritten as

$$v^{(n)} = \frac{1}{\tau + \tau_F (1 - P_E^{(n)}) / P_E^{(n)}} \quad (S5)$$

The mean translation rate is then calculated by

$$v = \sum_{n=0}^{\infty} (v^{(n)} f_O^{(n)}) \quad (S6)$$

Since  $v^{(n \geq 3)} = v^{(3)}$  and  $\sum_{n=0}^{\infty} f_O^{(n)} = 1$ , Equation (S6) can be rewritten as

$$v = \sum_{n=0}^2 (v^{(n)} f_O^{(n)}) + v^{(3)} \left( 1 - \sum_{n=0}^2 f_O^{(n)} \right) \quad (S7)$$

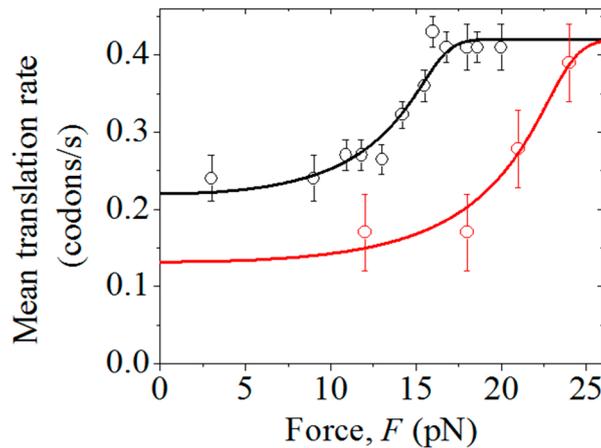
It is seen that when  $P_E^{(n)} = 1$ , Equations (S5) and (S6) or (S7) are reduced to Equation (S2). It is also noted that if the time of slow EF-G·GDP release from the pre-translocation State F is close to the time of fast EF-G·GDP release from the post-translocation state plus the time from the binding of the ternary complex through the peptidyl transfer, *i.e.*,  $\tau \approx \tau_F$ , Equations (S5) and (S6) or (S7) are reduced

to  $v \approx \sum_{n=0}^{\infty} (v_0 P_E^{(n)} f_O^{(n)})$ , *i.e.*,

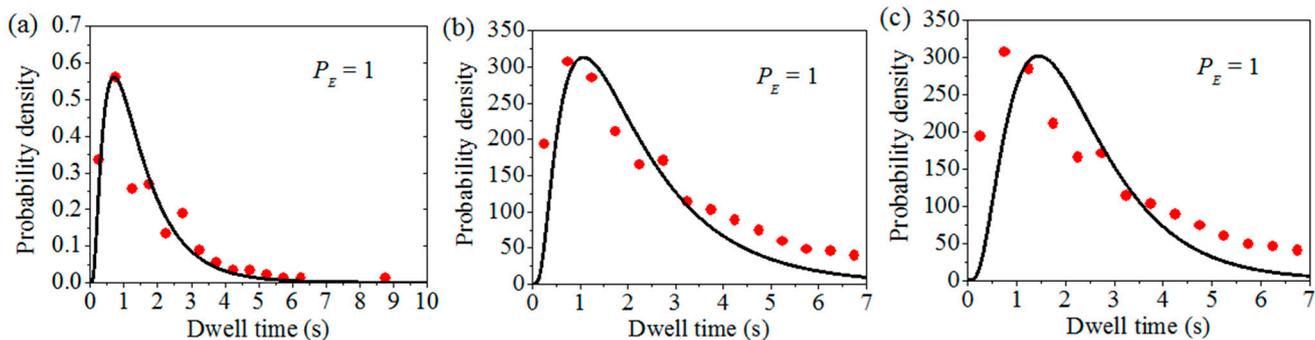
$$v(F) \approx v_0 \left\{ \sum_{n=0}^2 \frac{1}{1 + \exp\left[-\left(E_{PE}^{(50S)} - (3-n)E_{bp}\right)/k_B T\right]} f_O^{(n)} + \sum_{n=3}^{\infty} \frac{1}{1 + \exp\left(-E_{PE}^{(50S)}/k_B T\right)} f_O^{(n)} \right\} \quad (\text{S8})$$

Equation (S8) is the same as that used in the previous work [13]. It is noted here that with values of the rate constants given in Table 1, from Figure 2 it is calculated that the mean time of transition from State POST to State C at saturating ternary complex,  $\tau_{POST \rightarrow C} = 1/k_5 + (k_7 + k_{-7} + k_8)/(k_7 k_8) + 1/k_9 + 1/k_{10} + 1/k_{11}$ , is comparable to the time  $\tau_r = 1/k_r$ . Thus, as seen from Equations (S1) and (S4), the approximation  $\tau \approx \tau_F$  is valid.

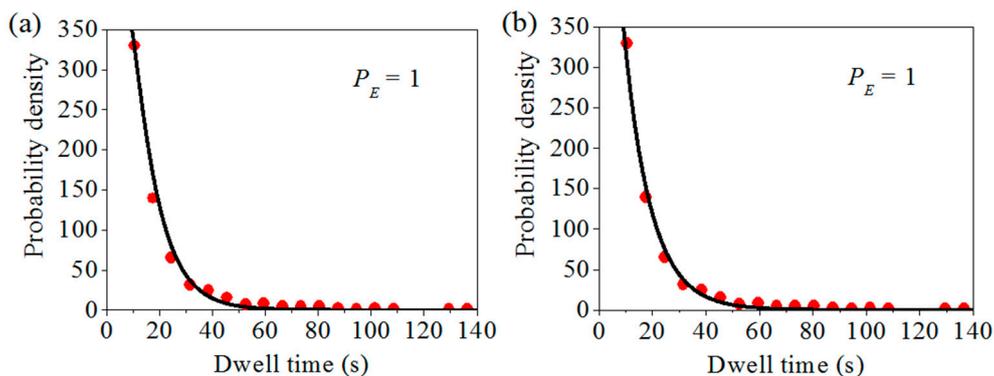
As done in the previous work [13], for fixed  $Z_{\max} = 0.58$  nm/base,  $E_{bp} = 3k_B T$  and  $b = 1.85$  nm for mRNA duplex hpVal<sub>GC50</sub>, using Equation (S8) the experimental data on the rate of the ribosome translation through mRNA duplex hpVal<sub>GC50</sub> vs. the pulling force  $F$  [7] can be fitted well, as shown in Figure S1, where the parameters for the ribosome are  $E_{PE}^{(50S)} = 9k_B T$  and  $v_0 = 0.42$  codons/s. Similarly, for  $Z_{\max} = 0.58$  nm/base,  $E_{bp} = 3.3k_B T$  and  $b = 0.72$  nm for mRNA duplex hpVal<sub>GC100</sub>, and  $E_{PE}^{(50S)} = 9k_B T$  and  $v_0 = 0.42$  codons/s for the ribosome, using Equation (S8) the theoretical data on the rate of the ribosome translation through mRNA duplex hpVal<sub>GC100</sub> are also in good agreement with the experimental data [7] (Figure S1). It is noted here that the base-pairing energies  $E_{bp} = 3k_B T$  for hpVal<sub>GC50</sub> and  $E_{bp} = 3.3k_B T$  for hpVal<sub>GC100</sub> are consistent with those estimated by using the nearest-neighboring thermodynamic model for the RNA duplex stability [51]. The maximum extension of the single-stranded mRNA containing one base under the pulling force,  $Z_{\max} = 0.58$  nm/base, is the same as that used in the literature (see main text). The Kuhn length  $b = 1.85$  nm for hpVal<sub>GC50</sub> and  $b = 0.72$  nm for hpVal<sub>GC100</sub> are consistent with those ( $b = 0.6$ – $2.5$  nm) estimated in the literature (see main text). Moreover, since the Kuhn length is dependent on the persistence length that is sequence dependent [52], it is understandable that the two duplexes hpVal<sub>GC50</sub> and hpVal<sub>GC100</sub> have different Kuhn lengths.



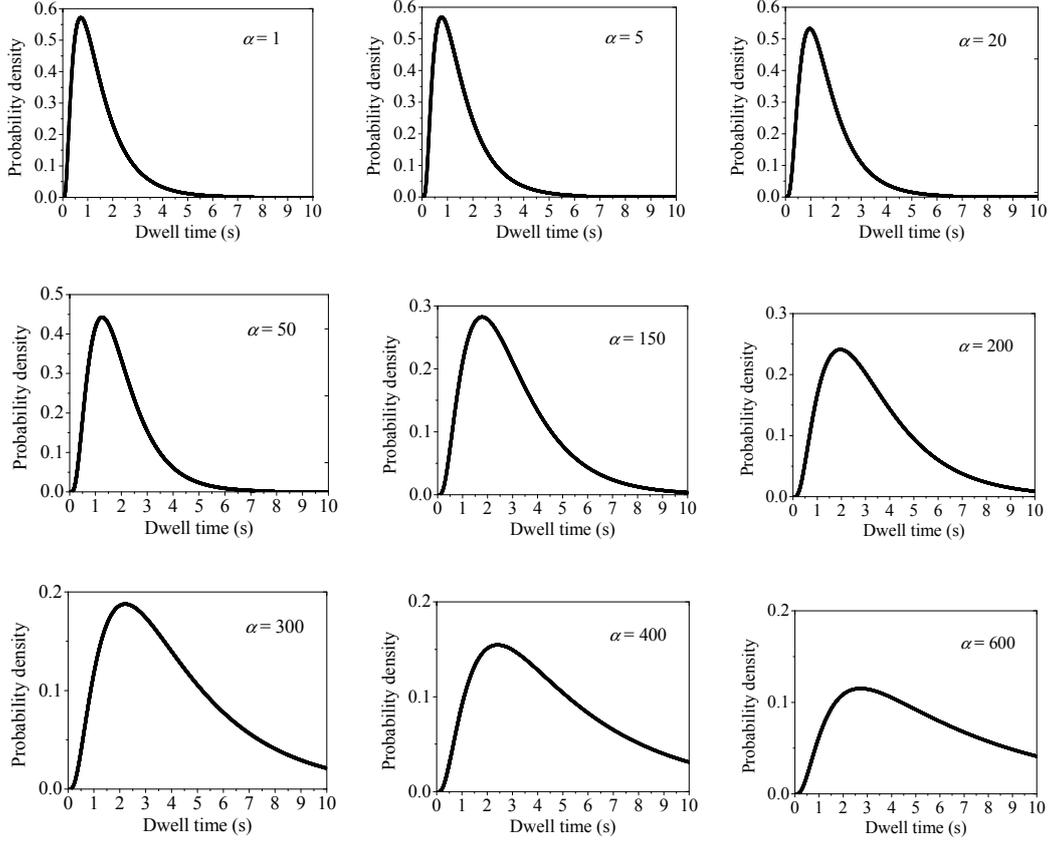
**Figure S1.** Mean rate of translation through mRNA duplexes vs. the pulling force  $F$  to unzip the duplexes. Solid lines are theoretical results and symbols are experimental data taken from Qu *et al.* [7] (reproduced with permission from Nature). The black line and symbols are for mRNA duplex hpVal<sub>GC50</sub>, while the red line and symbols are for mRNA duplex hpVal<sub>GC100</sub>.



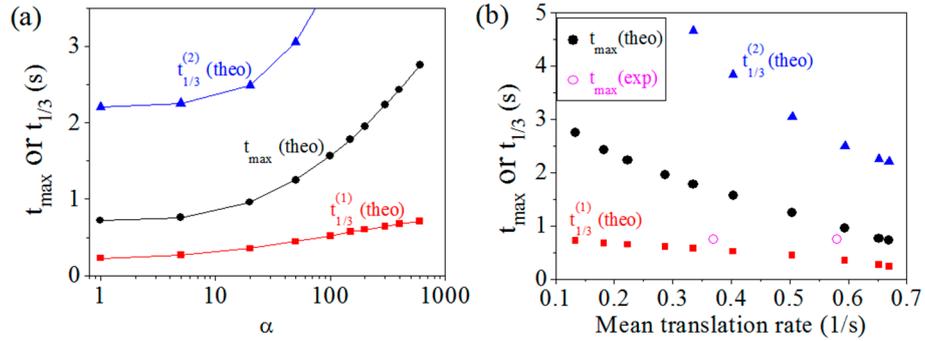
**Figure S2.** Distributions of dwell times calculated by taking  $P_E = 1$  (black lines), which corresponds to the translation through the single-stranded mRNA. In order to make a direct comparison with the experimental data, we multiply the calculated distribution of dwell times,  $h(t)$ , by a constant  $C$ . Red dots are experimental data taken from Wen *et al.* [6]. **(a)** With values of rate constants  $k_1$ – $k_{11}$  as given in Table 1. The experimental data are taken from Figure 4a in Wen *et al.* [6]; **(b)** With values of rate constants  $k_1$ – $k_{11}$  given in Table 1 being divided by 1.5. The experimental data are taken from Figure S5a in Wen *et al.* [6]; **(c)** With rate constant of the translocation step,  $k_4$ , given in Table 1 being divided by 80 while with values of other rate constants as given in Table 1. The experimental data are taken from Figure S5a in Wen *et al.* [6] (reproduced with permission from Nature).



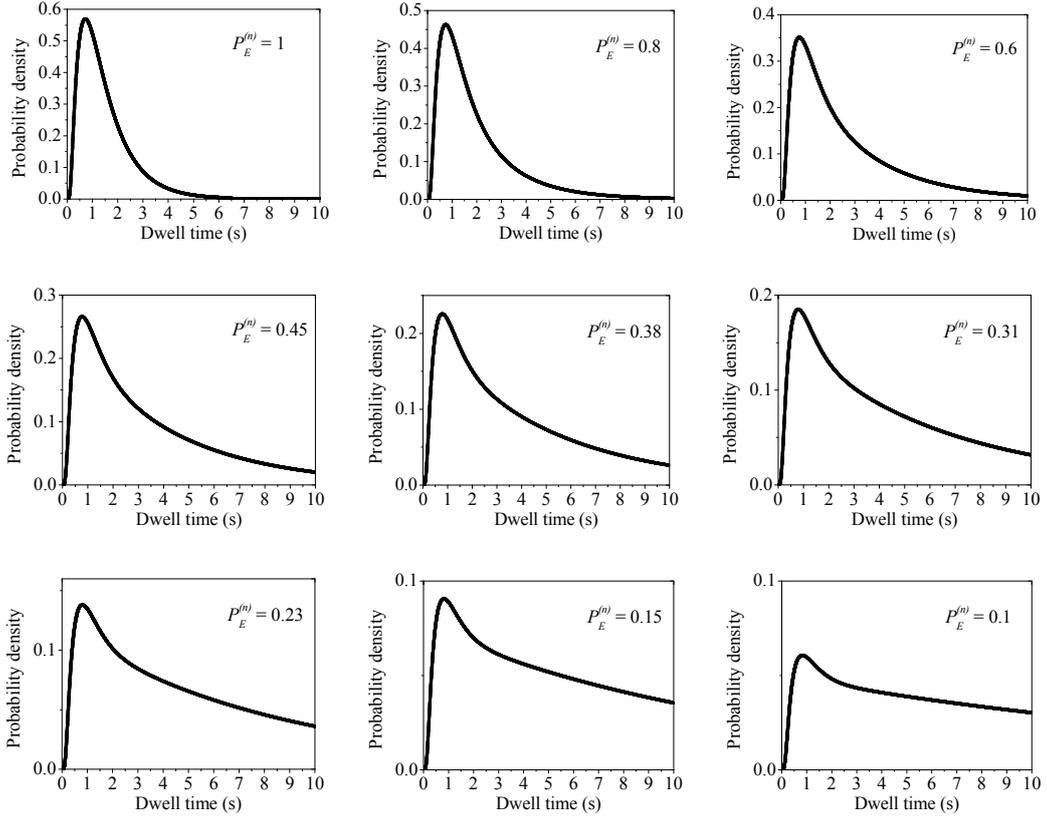
**Figure S3.** Distributions of dwell times with long durations ( $>7$  s) calculated by taking  $P_E = 1$  (black line), which corresponds to the translation through the single-stranded mRNA. In order to make a direct comparison with the experimental data, we multiply the calculated distribution of dwell times,  $h(t)$ , by a constant  $C$ . Red dots are experimental data taken from Figure S5b in Wen *et al.* [6] (reproduced with permission from Nature). **(a)** With values of rate constants  $k_1$ – $k_{11}$  given in Table 1 being divided by 9; and **(b)** With rate constant of the translocation step,  $k_4$ , given in Table 1 being divided by 1000 while with values of other rate constants as given in Table 1.



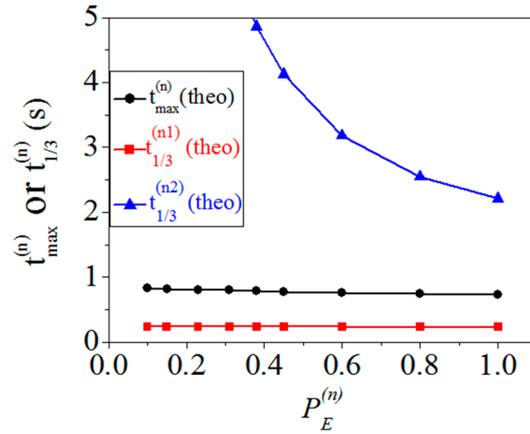
**Figure S4.** Normalized distributions of dwell times calculated by taking  $P_E = 1$ , which corresponds to the translation through the single-stranded mRNA, but with different values of  $\alpha$ , where  $k_4$  is replaced by  $k_4/\alpha$ .



**Figure S5.** Dwell time,  $t_{max}$ , at which the maximum value of the distribution occurs, dwell time,  $t_{1/3}^{(1)}$ , at which the distribution increases from zero to the value equal to one third of the maximal value, and dwell time,  $t_{1/3}^{(2)}$ , at which the distribution decreases from the maximal value to the value equal to one third of the maximal value, vs.  $\alpha$  (left panel, **a**), which are obtained from Figure S4. The right panel (**b**) corresponds to  $t_{max}$ ,  $t_{1/3}^{(1)}$  and  $t_{1/3}^{(2)}$  vs. the translation rate  $v_0$ , where the theoretical data for  $v_0$  are calculated by using Equations (S1) and (S2). The two experimental data for  $t_{max}$  are taken from and those for  $v$  are calculated from the two distributions of dwell times shown in Figure 4a and Figure S5a of Wen *et al.* [6].



**Figure S6.** Normalized distributions of dwell times,  $h^{(n)}(t)$  for different values of  $P_E^{(n)}$ .



**Figure S7.** Dwell time,  $t_{\max}^{(n)}$ , at which the maximum value of the distribution  $h^{(n)}(t)$  occurs, dwell time,  $t_{1/3}^{(n1)}$ , at which the distribution  $h^{(n)}(t)$  increases from zero to the value equal to one third of the maximal value, and dwell time,  $t_{1/3}^{(n2)}$ , at which the distribution  $h^{(n)}(t)$  decreases from the maximal value to the value equal to one third of the maximal value, vs. the effective translocation probability  $P_E^{(n)}$ , which are obtained from Figure S6.