## **Supplementary Information**

Species	Genome Database
Arabidopsis lyrata	http://www.phytozome.net/
Arabidopsis thaliana	http://www.phytozome.net/
Aspergillus nidulans FGSC A4	http://www.broad.mit.edu/annotation/fgi
Chlamydomonas reinhardtii	http://www.ncbi.nlm.nih.gov/
Culex quinquefasciatus	http://www.vectorbase.org/Culex_quinquefasciatus/Info/Index
Escherichia coli str. K-12 substr. W3110	http://www.broad.mit.edu
Glycine max	http://www.phytozome.net/
Larix gmelinii	http://dendrome.ucdavis.edu/
Lonicera japonica Thunb	Database in our group
Lonicera japonica Thunb. var. chinensis (Wats.)	Database in our group
Oryza sativa	http://www.phytozome.net/
Penicillium marneffei ATCC 18224	http://fungalgenomes.org/data/PEP/
Pinus pinaster	http://dendrome.ucdavis.edu/
Pinus taeda	http://dendrome.ucdavis.edu/
Populus trichocarpa	http://www.phytozome.net/
Pseudotsuga menziesii	http://dendrome.ucdavis.edu/
Selaginella moellendorffii	http://www.phytozome.net/
Sorghum bicolor	http://www.phytozome.net/
Vitis vinifera	http://www.phytozome.net/
Volvox carteri	http://www.ncbi.nlm.nih.gov/
Zea mays	http://www.phytozome.net/

Table S1. Sequences from twenty-one species in this study.

LJACS1	LJAAE	LJ4CL	LJACS2
T (151,2) #	T (169, 3)	S (199,1)	M (228, 1)
S (152,2)	S (170,2)	S (200,1)	T (230, 2)
	G (171,1)		
	T (172,1)		
	V (179,1)	V (209,1)	
N (171,2)	S (189,1)	S (219,2)	S (231, 2)
	I (193,1)	Q (223,1)	
L (191,10)	L (209,1)	L (243,1)	L (250, 1)
P (192,10)	P (210,1)	P (244,1)	
A (194,1)			A (278, 1)
H (195,7)	H (213,7)	H (247,5)	H (279, 5)
I (196,5)	V (214,7)	** I (248,6)	I (280, 5)
Y (197,3)	H (215,7)	** Y (249,6)	F (281, 3)
E (198,1)		S (250,1)	
R (199,1)			R (283, 1)
A (200,3)			V (284, 2)
N (201,1)	A (219,6)	I (253,6)	I (285, 1)
C (235,2)	T (257,2)	P (288,2)	C (319, 2)
V (237,1)	V (259,1)	V (290,1)	A (320, 1)

**Table S2.** Putative binding-site residues in the model Lonicera japonica.

LJACS1	LJAAE	LJ4CL	LJACS2
V (304,1)	R (285,1)	M (315,3)	L (391, 2)
S (305,3)	S (286,3)	S (316,3)	S (392, 3)
G (306,6)	C (287,7)	* G (317,6)	G (393, 6)
A (307,8)	S (288,8)	A (318,7)	A (394, 7)
S (308,5)	A (289,6)	A (319,4)	A (395, 5)
P (309,4)	S (290,5)	P (320,3)	P (396, 5)
V (327,1)	L (308,1)	G (339,2)	L (414, 1)
E (328,4)	E (309,4)	Q (340,5)	Q (415, 6)
G (329,7)	A (310,8)	* G (341,6)	G (416, 6)
Y (330,5)	Y (311,7)	Y (342,4)	Y (417, 5)
G (331,9)	A (312,8)	* G (343,8)	G (418, 8)
M (332,8)	M (313,10)	M (344,7)	L (419, 7)
T (333,8)	T (314,9)	T (345,7)	T (420, 7)
E (334,1)	E (315,2)	E (346,1)	E (421, 1)
S (336,4)	T (317,3)	M (348,4)	C (423, 3)
C (337,4)	H (318,4)	* P (349,4)	A (424, 3)
I (338,3)	L (319,3)	* V (350,6)	G (425, 4)
I (339,1)	M (320,1)	* L (351,1)	T (426, 1)
		M (353,1)	
V (352,2)	V (336,4)		V (440, 2)
		C (369,1)	
S (413,1)	T (391,4)		
D (415,4)	D (393,5)		D (501, 5)
I (427, 4)	L (405,5)		I (513, 5)
I (428, 1)	V (406,1)		
D (429,2)	G (407,1)		D (515, 1)
R (430,2)	R (408,6)		R (516, 2)

Table S2. Cont.

# (a,b), a, the site of putative binding-site residues; b, numbers in the 10 models; \* Related with 4-coumaric acid; \*\* Related with ferulic acid.

**Figure S1.** Chemical structures of the six naturally occurring 4CL substrates, sinapic acid, 5-hydroxyferulate, ferulic acid, caffeic acid, 4-coumarate and trans-cinnamic acid. Acetate and propanoate was toward to acetyl-CoA and propanoyl-CoA by ACS. Succinylbenzoate was toward to Succinylbenzoyl-CoA by AAE.



**Figure S2.** Phylogeny tree of AMP-binding enzyme (PF00501). A neighbor-joining tree containing 1207 sequences was generated based on the AMP-dependent synthetase/ligase domain sequences by Mega 5.02. A bootstrap value of 1000 replications was applied and all of sequences were classified into three clusters and 4 subgroups in the first cluster. Gene function of Arabidopsis thaliana in three clusters follow as: **1-1**, long chain acyl-CoA synthase; **1-2**, Acyl-acting anzyme/o-succinylbenzoate-CoA ligase/benzoate-CoA ligase; **1-3**, 4-coumarate:CoA ligase; **1-4**, acyl-CoA synthase/malony-CoA synthase; **2**, Acyl-acting anzyme/acetate-CoA ligase/AA-adenyl-dom(amino acid adenylation domain) protein; **3**, unknown.



**Figure S3.** HPLC-MS analysis in bud of *Lonicera japonica* Thunb. (**A**,**B**) Chlorogenic acid(Retention time (RT) 13.20 min); (**C**) ferulic acid (RT 24.24 min); (**D**) Rutin(RT 24.24 min); (**E**) Hyperoside (RT 25.53 min), Isoquercitrin (RT 25.81 min); (**F**) Luteoloside (RT 25.39 min), quercitrin (RT 28.78 min); (**G**) luteolin (RT 37.78 min); (**H**) Quercetin (RT 38.19 min); (**I**) Apigenin(RT 43.19 min).



**Figure S4.** Phylogeny and expression of unknown protein sequences in Cluster2. A neighbor-joining tree containing 20 sequences was generated based on the AMP-dependent synthetase/ligase domain sequences. A bootstrap value of 1000 replications was applied. The rpkm value of sequences in flowers of Lonicera japonica Thunb is shown.



**Figure S5.** Phylogeny and expression of ACS sequences in Cluster1. A neighbor-joining tree containing 37 sequences was generated based on the AMP-dependent synthetase/ligase domain sequences. A bootstrap value of 1000 replications was applied. The rpkm value of sequences in flowers of Lonicera japonica Thunb is shown.



**Figure S6.** Phylogeny and expression of ACS sequences in Cluster4. A neighbor-joining tree containing 50 sequences was generated based on the AMP-dependent synthetase/ligase domain sequences. A bootstrap value of 1000 replications was applied. The rpkm value of sequences in flowers of Lonicera japonica Thunb is shown.





## Figure S7. Protein structure of LJACS1, LJ4CL, LJAAE and LJACS2.

LJAAE (Cluster2)



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