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Review

βArrestins in Cardiac G Protein-Coupled Receptor Signaling and Function: Partners in Crime or "Good Cop, Bad Cop"?

Anastasios Lymperopoulos * and Shmuel Negussie

The Laboratory for the Study of Neurohormonal Control of the Circulation, Department of Pharmaceutical Sciences, Nova Southeastern University College of Pharmacy, Fort Lauderdale, FL 33328, USA

* Author to whom correspondence should be addressed; E-Mail: al806@nova.edu; Tel.: +1-954-262-1338; Fax: +1-954-262-2278.

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Abstract: βarrestin (βarr)-1 and -2 (βarrs) (or Arrestin-2 and -3, respectively) are universal G protein-coupled receptor (GPCR) adapter proteins expressed abundantly in extra-retinal tissues, including the myocardium. Both were discovered in the lab of the 2012 Nobel Prize in Chemistry co-laureate Robert Lefkowitz, initially as terminators of signaling from the β -adrenergic receptor (β AR), a process known as functional desensitization. They are now known to switch GPCR signaling from G protein-dependent to G protein-independent, which, in the case of βARs and angiotensin II type 1 receptor (AT₁R), might be beneficial, e.g., anti-apoptotic, for the heart. However, the specific role(s) of each βarr isoform in cardiac GPCR signaling and function (or dysfunction in disease), remain unknown. The current consensus is that, whereas both Barr isoforms can desensitize and internalize cardiac GPCRs, they play quite different (even opposing in certain instances) roles in the G protein-independent signaling pathways they initiate in the cardiovascular system, including in the myocardium. The present review will discuss the current knowledge in the field of βarrs and their roles in GPCR signaling and function in the heart, focusing on the three most important, for cardiac physiology, GPCR types (β_1AR , β_2AR & AT_1R), and will also highlight important questions that currently remain unanswered.

Keywords: βarrestins; signal transduction; GPCR; heart; adrenergic receptors; angiotensin II receptors; desensitization

1. General Considerations of Barrs

The G protein-coupled receptors (GPCRs) or heptahelical or seven-transmembrane spanning receptors are by far the largest and most diverse superfamily of cell surface receptors. Approximately 600 distinct genes encoding non-olfactory GPCRs make up greater than 1% of the human genome [1]. Such evolutionary diversity enables GPCRs to detect an extraordinary array of extracellular stimuli. GPCRs function in neurotransmission, neuroendocrine control of physiologic homeostasis and reproduction, and regulation of hemodynamics and intermediary metabolism, and they control the growth, proliferation, differentiation, and death of multiple cell types. It is estimated that more than half of all drugs in clinical use target GPCRs, acting either to mimic endogenous GPCR ligands, to block ligand access to the receptor, or to modulate ligand production [1]. The importance of this receptor superfamily was highlighted last year by the awarding of the two pioneers of the field, Bob Lefkowitz and Brian Kobilka, with the Nobel Prize in Chemistry [2]. Among the various organ systems, whose physiology and homeostasis GPCRs regulate, perhaps the most prominent is the cardiovascular system, including the heart per se. For instance, cardiac function (contractility) is tightly controlled by the activity of β -adrenergic receptors (β_1 - and β_2 ARs) located in the membranes of cardiac myocytes [3]. Cardiac structure and morphology are regulated by angiotensin II (AngII) type 1 receptors (AT₁Rs) present (mainly) in cardiac fibroblast and endothelial cell membranes, but also, to a lesser extent, in cardiomyocyte membranes [4]. Moreover, the neurohormonal control of the circulation, e.g., catecholamine and corticosteroid release by the adrenal glands or activation of the renin-angiotensin-aldosterone system (RAAS) by the juxtagomerular apparatus of the kidneys, is also under tight regulation by various GPCRs [5].

Agonist binding of all these receptors promotes their interaction with heterotrimeric G proteins, which initiates the classical intracellular signaling of these receptors that ultimately leads to a variety of cellular responses/physiological effects. At the same time though, agonist binding promotes the phenomenon of homologous or agonist-dependent receptor desensitization, which is the molecular basis of the waning of the cellular responsiveness to persistent receptor stimulation and constitutes a major classical homeostatic mechanism of cellular physiology [6]. This agonist-dependent desensitization is conferred, at the molecular level, by phosphorylation of the receptor by the family of kinases known as G protein-coupled receptor kinases (GRKs). GRK2 is the most prominent member of this family, binding to and phosphorylating a vast majority of GPCRs, including cardiac βARs and AT₁Rs [7]. It is also the most abundant GRK isoform in the heart and in the cardiovascular system in general [7]. GRK-dependent phosphorylation enhances the affinity of the receptor for binding to the ubiquitous receptor adapter proteins βarrestins (βarrs), which comprise two isoforms in mammals, βarr1 and -2, also known as arrestin (Arr)-2 and -3, respectively [6]. The βarrs are also abundantly expressed in the heart and in the vasculature, as well as in several other tissues [1,6]. Barr1 and -2 bind directly to GRK-phosphorylated GPCRs, forming a stoichiometric complex that is stereochemically incapable of further G protein coupling (a phenomenon referred to as "functional desensitization" of a GPCR). Barr activation occurs when the polar core located in the hinge region between the two globular domains of the βarr molecule interacts with GRK-phosphorylated residues on the receptor, displacing the βarr C-terminus and exposing the concave surface of the globular domains to interact with the receptor [8]. Receptor binding produces significant conformational changes in the βarr

molecule [9], whereas, conversely, βarr binding stabilizes a high-agonist affinity state of the receptor, prompting some authors to characterize the receptor-βarr complex as an "alternative ternary complex" analogous to the ternary complex existing between agonist-receptor-G protein in the absence of GTP [10]. Barrs (contrary to their retinal system-residing counterparts, the visual Arrs) further dampen G protein signaling by linking receptors to the clathrin-dependent endocytic machinery [1,11]. The C-terminus of βarrs directly binds clathrin heavy chain and the β2 adaptin subunit of the AP-2 complex, two intergral components of the endocytic machinery, as it gets displaced by the engagement of the receptor [12]. Clathrin/AP-2 binding causes βarr-bound GPCRs to cluster in clathrin-coated pits, which are pinched off the plasma membrane by the motor protein dynamin. This Barr-dependent endocytosis (receptor internalization or sequestration) removes receptors from the cell surface, rendering them less responsive to subsequent stimuli. From that point on, most GPCRs fall into one of two classes based on their affinity for the two βarr isoforms and the longevity of the receptor-βarr interaction [13]. One class exhibits higher affinity for Barr2 than Barr1 and forms transient receptor-ßarr complexes that dissociate soon after the receptor internalizes. These receptors (e.g., the β₂AR) rapidly recycle back to the plasma membrane ready to signal again upon the next encounter with agonist (receptor resensitization). The other class exhibits equivalent affinities for βarr1 or -2 and forms more stable receptor-βarr complexes that remain intact as the receptor undergoes endosomal These receptors (e.g., AT₁R & vasopressin V₂ receptor) are sequestered in endosomes and tend to recycle slowly or undergo lysosomal targeting for degradation (receptor downregulation, i.e., total cellular receptor number reduced).

Unlike the catalytic interaction of a GPCR with its cognate G protein, GPCRs form relatively stable complexes with βarrs that persist on a time scale of minutes to hours [14]. It was the discovery that Barrs serve as adapters not only in the context of GPCR endocytosis but also in linking activated receptors to other enzymatic effectors that ushered in a new paradigm shift in GPCR signal transduction [6,11]. It is nowadays well known that βarrs bind a number of catalytically active proteins and recruit them to agonist-occupied GPCRs, among them Src family tyrosine kinases, components of the ERK1/2 and c-Jun N-terminal kinase 3 (JNK3) mitogen-activated protein (MAP) kinase cascades, the E3 ubiquitin ligase Mdm2, the cAMP phosphodiesterases (PDE) PDE4D3/5, diacylglycerol kinase (DGK), the inhibitor of nuclear factor (NF)-κΒ ΙκΒα, and the Ser/Thr protein phosphatase (PP) PP2A [6,11]. It is via these interactions that βarr binding to the receptor initiates secondary waves of signal transduction independently of G proteins, usually as the GPCR-Barr complex travels through endosomal compartments during its endocytosis [6]. Thus, in addition to GPCR desensitization (G protein uncoupling) and internalization, βarrs perform a third very important biological function in cells: Signal transduction from GPCRs, i.e., in essence switching of the signaling of the receptor from G protein-dependent to G protein-independent [11]. Of course, βarr-mediated signaling is qualitatively different from the G protein-mediated signal transduction; for instance, βarr signaling does not result in signal amplification, as it usually proceeds through a 1:1 stoichiometry [6]. Additionally, it exhibits specific subcellular localization, dictated by the location of the Barr-based molecular signaling scaffold, which significantly affects the ultimate signaling events produced, i.e., Barrs provide compartmentalization to the signaling cascade they induce, whereas G protein signaling proceeds ("diffuses") throughout the cell all the way down to the cell nucleus [6,11].

Given the importance of GPCRs in regulation of cardiovascular homeostasis touched upon above, it comes as no surprise that the two βarrs play very important roles in regulation of cardiovascular (and, in particular, of cardiac) physiology and homeostasis, as well [1]. The cardiovascular roles of βarrs have only recently started to become elucidated thanks to a combination of techniques and tools, such as the utilization of the (global) βarr1 and βarr2 knockout (KO) mice, usage of isoform-specific siRNA knockdown in *in vitro* systems, employment of *in vitro* and *in vivo* systems of artificially constructed GPCR mutants incapable of signaling through G proteins, and, finally, synthesis and development of signaling pathway-selective (biased) GPCR ligands that preferentially elicit βarr signaling over G protein signaling (or *vice versa*). The present review will focus on the roles the two mammalian βarrs play in GPCR signaling and function specifically in the heart. We will start with an overview of what is currently known about the physiology/pathophysiology of the two cardiac βarrs, and then discuss topics pertaining to cardiac βarrs that are currently under intense investigation, highlighting particular areas and questions that await elucidation and answers.

2. Cardiac β₁AR Signaling and βarrs

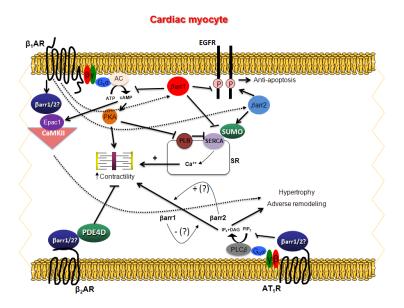
The principal role of βARs in the heart is regulation of cardiac rate and contractility in response to the catecholamines [3]. Out of the three known mammalian βAR subtypes, the $\beta_1 AR$ is the predominant one in cardiac myocytes, representing 75%–80% of total βAR density, followed by the β₂AR, which comprises about 15%–18% of total cardiomyocyte βARs and the remaining 2%–3% is β_3 ARs (under normal conditions) [5]. β_1 AR stimulation by catecholamines results in the dissociation of the stimulatory G protein alpha subunit ($G\alpha_s$) from $G_{\beta\gamma}$. $G\alpha_s$ stimulates adenylyl cyclase (AC) to produce 3'-5'-adenosine monophosphate (cyclic AMP, or cAMP), which in turn, by activating protein kinase A (PKA), regulates different intracellular, sarcolemmal and myofibrillar substrates, thus exerting the cellular effects of β_1AR activation on cardiac chronotropy, inotropy and lusitropy. In addition, $G_{\beta\gamma}$ can also activate downstream effectors that participate in cardiac signaling regulation [3]. β₂AR also mediates the effects of catecholamines on the heart, but in a qualitatively different manner from β_1AR , as it can also couple to the AC inhibitory G protein (G_i). It is now generally accepted that in the heart, $\beta_2 AR$ signals and functions in a substantially different way compared to $\beta_1 AR$ [15–17]. Importantly, whereas β_1AR activation enhances cardiomyocyte apoptosis, β_2AR exerts beneficial anti-apoptotic effects in the heart, purportedly through this G_i-mediated signaling [15–17]. Several studies using transgenic mice, β_2AR -selective stimulation and adenoviral-mediated β_2AR overexpression have established a consensus that β_2AR signaling is predominantly cardio-protective, improving cardiac function and decreasing apoptosis, whereas β_1AR -elicited signaling has detrimental effects in the heart [18,19]. Of note, the differences between these two predominant cardiac βARs in terms of signaling properties might take a quite different shape and have a much bigger bearing on pathophysiologic implications in the setting of heart failure (HF): for instance, β_1AR is selectively down-regulated (i.e., total cellular receptor number reduced) in HF, thus shifting the above mentioned stoichiometry of β₁AR:β₂AR towards 50:50 in the failing heart from 75:~25 in the normal, healthy heart [3]. However, β_2 AR is also non-functional and does not signal properly in the failing heart [3]. In addition, β₂AR signaling in the failing heart might differ substantially from the beneficial signaling pattern of the normal heart; it seems to be more "diffuse" (less "compartmentalized") and resembles more the pro-apoptotic cAMP signaling pattern of its β_1AR counterpart in HF [5]. Therefore, the aforementioned stoichiometric shift in favor of the "good" β_2AR in HF appears unable to help the heart improve its structure and function.

As co-factors of GRKs in cardiomyocyte β_1AR desensitization and downregulation, β_1AR desensitization and downregulation, β_1AR desensitization and downregulation. to the diminished inotropic and adrenergic reserves of the failing heart and their inhibition should theoretically be beneficial in acute HF, as it would enhance the Gas-AC-PKA-mediated pro-contractile signaling of cardiac βARs, which increases cardiac contractility [3]. Studies in (global) βarr1KO mice have confirmed that cardiac Barr1 diminishes inotropic and adrenergic reserves by means of desensitizing cardiomyocyte β_1 - and β_2 ARs, since contractility of β_1 - arr1KO mice in response to isoproterenol, a BAR agonist cardio-stimulant (positive inotrope), is significantly augmented compared to wild type mice (but basal contractility is unaffected) [20]. Of note, cardiac βarr2 does not appear to be up-regulated compensatively in these \(\beta \text{arr1KO} \) mice and to compensate for the loss of \(\beta \text{arr1} \) in the myocardium [20], strongly indicating that the two βarr isoforms are rarely (if at all) physiologically interchangeable. In HF, chronic catecholaminergic stimulation of the β₁AR promotes cardiac hypertrophy, decreases contractility, and increases myocyte apoptosis [21]. As a result, administration of β-blockers is currently part of standard care in the clinical management of congestive HF [22,23]. Barrs have been shown *in vitro* to mediate the mitogenic signaling of EGF (Epidermal Growth Factor) receptor (EGFR) transactivation by the β₁AR [24]. As inhibition of EGFR contributes to dilated cardiomyopathy, β_1AR signaling via β arr2 appears to be protective rather than deleterious for the heart and βarr2-dependent EGFR transactivation might exert a cardio-protective effect [25,26]. The physiologic relevance of EGFR transactivation by β₁AR-bound βarr2 has been demonstrated in transgenic mice overexpressing wild-type β_1ARs or mutant β_1ARs lacking GRK phosphorylation sites and thus unable to undergo GRK-mediated phosphorylation and bind Barrs [24]. Under conditions of chronic catecholamine stress, transgenic mice overexpressing the mutant β_1AR are incapable of transactivating cardiac EGFRs and show marked myocyte apoptosis and left ventricular dilatation compared to mice expressing wild-type β₁ARs [24]. Further substantiating a role for EGFR transactivation in the mechanism underlying these findings, pretreatment of mice overexpressing the wild-type β_1AR in their hearts with the known selective EGFR inhibitor (and anti-cancer drug) erlotinib prevents any improvement in cardiac function upon chronic stimulation with catecholamines [24].

Very recently, our laboratory was able to delineate the specific roles of cardiac β arr1 in β AR signaling and function during post-myocardial infarction (MI) HF progression by studying the β arr1KO mice after MI [27]. More specifically, we found that cardiac β arr1 (and not β arr2) is the β arr isoform responsible for the aforementioned β AR desensitization and downregulation (Figure 1), the molecular hallmark of HF, since β arr1KO mice have significantly better cardiac β -adrenergic and inotropic reserves, accompanied by elevated cardiac functional β AR numbers and cAMP pro-contractile signaling, compared to control, age-matched wild type mice, both in healthy conditions and, even more so, after MI. Notably, cardiac β arr2 did not compensate for these phenotypic effects of cardiac β arr1 absence [27]. In addition, the genetic deletion of β arr1 resulted in reduced cardiac apoptosis, inflammation, and adverse remodeling post-MI [27]. Importantly, and with regards to cardiac β arr-dependent signaling, we uncovered that β arr2 is the β arr isoform that mediates the beneficial, anti-apoptotic EGFR transactivation by the β 1AR in the heart mentioned above, and that cardiac β arr1 actually antagonizes β arr2 for this effect (Figure 1), since β arr1KO mouse hearts display

significantly elevated EGFR activity levels in response to β_1 AR-selective stimulation in vivo (treatment with isoproterenol in the presence of the β_2 AR-selective antagonist ICI 118,551), compared to control, wild type mouse hearts, both under healthy conditions and post-MI [27]. Another interesting finding from our study of βarr1KO mice was that cardiac sarco (endoplasmic) reticulum Ca²⁺-ATPase (SERCA)-2a activity of these mice was elevated, compared to control wild type mice, again both under healthy conditions and post-MI [27]. SERCA2a is an essential positive regulator of cardiac contractility activated by BARs (normally via PKA) [28]. Elevated activity of this cardiomyocyte calcium pump in post-MI βarr1KO mice was expected, since cardiac βarr1 desensitizes and downregulates β ARs in post-MI HF. The fact that it was found elevated also in normal, healthy Barr1KO mice however, whose cardiac β-adrenergic reserve is no different from that of control, wild type mice, raises the interesting possibility that cardiac βarr1 suppresses SERCA2a activity not only indirectly (by desensitizing \(\beta ARs \) but also directly [27]. Indeed, currently ongoing studies in our laboratory indicate that cardiac βarr2 is capable of interacting with SERCA2a, increasing its SUMO (small ubiquitin-like modifier)-vlation and activity [29], and cardiac βarr1 might normally antagonize this [30] (Figure 1). Taken together, all of the above studies seem to establish a consensus for cardiac β_1AR signaling, according to which: (a) β_1AR signaling is beneficial for the heart when it is mediated by Barrs (and specifically Barr2), and cardiotoxic (pro-apoptotic) when it is mediated by G proteins; and (b) β_1AR -bound β arr1 is detrimental for cardiac function (diminishes contractility, enhances apoptosis and inflammation, antagonizes βarr2-dependent beneficial signaling), whereas β₁AR-boundβarr2 is (mostly) beneficial for the heart, courtesy of its positive signaling effects (EGFR transactivation, SERCA2a potentiation) (Figure 1).

Figure 1. Schematic illustration of some important signaling pathways inside a cardiac myocyte affected by the two βarrs. SR, sarcoplasmic reticulum; PLB, phospholamban; ATP, adenosine triphosphate; PLCβ, phospholipase C-beta; IP₃, inositol (1,4,5)-trisphosphate; DAG, 2-diacylglycerol; PIP₂, phosphatidylinositol (4,5)-bisphosphate. See text for all other acronyms and details. "?" indicates that the suggested effect is currently under investigation, "βarr1/2?" indicates that which one of the two βarrs mediates the suggested effect (or if both βarrs do) is currently unknown.



However, this consensus was recently challenged by a study by Rockman and colleagues, in which cardiac βarrs were shown to be involved in cardiac β₁AR-induced activation of Ca²⁺/Calmodulin Kinase II (CaMKII) [31]. CaMKII is known to induce myocyte hypertrophy and apoptosis in the heart, thereby playing a detrimental role in HF pathophysiology [32]. Both βarrs were shown to orchestrate a multi-protein scaffold comprising β₁AR, CaMKII and Epac (Exchange protein directly activated by cAMP)-1 in mouse hearts in vivo, which leads to enhanced downstream cardiac CaMKII signaling [31] (Figure 1). This multi-molecular complex appears to form under basal conditions and β_1AR activation (e.g., with isoproterenol) enhances its formation [31]. Importantly, β₂AR does not seem capable of forming this complex, its formation is not affected by PKA and, finally, in contrast to EGFR transactivation (see above), both βarr1 and βarr2 appear capable of scaffolding this complex [31] (Figure 1). Certainly, these findings warrant further confirmation, but, if proven to hold true across species, they could potentially have enormous implications for cardiac β_1AR pathophysiology; since CaMKII signaling is generally damaging for the heart (pro-apoptotic, pro-arrhythmic, etc.), this appears to be a cardiotoxic effect of cardiac β₁AR βarr-dependent signaling, contrary to the EGFR transactivation by the same GPCR and via cardiac Barrs which is postulated to be beneficial in the heart (see above). On the other hand, if cardiac βarrs can mediate β₁AR-inducedEpac1 activation (Figure 1), then this would mean that cardiac βarrs cannot terminate the cAMP-mediated signaling of the cardiac β_1AR completely, despite the fact they uncouple the receptor from the G proteins; they merely terminate specifically its PKA-dependent signaling only.

3. Cardiac β₂AR Signaling and βarrs

Since β arrs can desensitize (uncouple from G proteins) the cardiac β ₂AR, as well, β arr binding to this receptor subtype in the heart should also be (predominantly) deleterious, as both the G_s protein-mediated, pro-contractile signaling and the G_{i/o} protein-mediated, anti-apoptotic signaling of the cardiac β_2 AR (see above) are inhibited [33]. However, β_2 Arr-dependent signaling stimulated by the β_2 AR can exert some beneficial effects in the cardiac myocyte, as it can be anti-apoptotic and also anti-inflammatory in its own right, e.g., by promoting ERK activation which increases cardiomyocyte survival and proliferation, and by blocking NF-κB activation which leads to pro-inflammatory cytokine production [34–36]. In contrast to the β_1AR , which forms a complex with PDE4D8 directly (without βarr involvement) when inactive and gets disassembled upon agonist binding, the β₂AR can form, thanks to the βarrs, a complex with another PDE variant, PDE4D5, upon its agonist activation [37] (Figure 1). ThisPDE4D5 recruitment to the β₂AR plays a crucial role in compartmentalizing the generated cAMP signal, posing a "brake" on the ability of cAMP to stimulate contractility [38] (Figure 1). Working in mouse hearts that lack β₁ARs but express the known GRK2 inhibitor mini-gene β ARKct (β_1 AR^{-/-}/ β ARKct), we found that this β arr-dependent PDE4D recruitment to the cardiac β₂AR is dependent on GRK2 and that it is, indeed, sufficient to prevent this βAR subtype from signaling to increased contractility in the heart in vivo [39]. GRK2 inhibition with βARKct in the hearts of β₁AR KO mice proved essential and capable of allowing these hearts to increase their contractile function in response to catecholaminergic stimulation, all the while diminishing the interaction of PDE4D with the cardiac β_2AR in vivo at the same time [39]. Thus, cardiac βarrs seem to impede β₂AR pro-contractile signaling in vivo, not only by uncoupling the

receptor from the G_s protein-cAMP signaling pathway, but also by recruiting PDE4D to the cardiac β_2AR (following its agonist-dependent phosphorylation by GRK2). Which one of the two β_2AR normally mediates this effect or whether both do remains an open question worth investigating in the future (Figure 1). In heterologous, transfected cell systems however, both β_2AR are capable of binding PDE4D and of recruiting it to the β_2AR [40].

Of note, several β_2AR ligands (the majority of them β -blockers, very useful in cardiovascular practice) have been tested at their ability to stimulate β_2AR (i.e., "bias" towards β_2AR (i.e., "bias" towards β_2AR (i.e., "bias" towards β_2AR (i.e., activation as a readout in transfected HEK293 cells, isoetharine and carvedilol have been identified as β_2AR (i.e., activate β_2AR (i.e., activate β_2AR signaling from the receptor without eliciting G protein activation) [42]. This finding has been postulated to be part of the mechanism of carvedilol's beneficial effects in HF [23]; however, it awaits confirmation with *in vivo* studies specifically on cardiac β_2AR , and, once confirmed, it would be interesting to delineate whether carvedilol preferentially stimulates the binding to the cardiac β_2AR s (and β_1AR s) of one β_2AR is equally induced.

4. Cardiac AT₁R Signaling and βarrs

In the heart, the AT₁R is (mainly) expressed in cardiac fibroblasts, where it stimulates cellular proliferation thus promoting fibrosis, and in cardiac myocytes, where it again stimulates growth thus promoting cardiac hypertrophy [43]. Whether it can also promote cardiomyocyte contractility however, is still a matter of debate [43]. Nonetheless, combined with other cellular effects leading to inflammation and oxidative stress development in the heart, cardiac AT₁R effects are clearly maladaptive and damaging for both the structure and function of the cardiac muscle, playing a pivotal role in the so-called adverse remodeling of the post-MI heart progressing to HF [4,43]. AT₁R is a classic G_{q/11}-coupled receptor that can also couple to G_{i/o} proteins [4]. With regards to their classical role as G protein-dependent signaling terminators (desensitizers), very little is known about βarrs and AT₁Rs and even less about cardiac βarrs and AT₁Rs. The AT₁R is a known GRK substrate, thus cardiac βarrs are bound to confer its desensitization in the heart secondary to its phosphorylation by GRKs, which has been demonstrated in vivo [4]. However, βarr-mediated AT₁R desensitization per se has never been directly investigated in vivo. Intriguingly, the AT₁R displays a somewhat peculiar behavior in terms of its desensitization. Not only is it subject to phosphorylation by other kinases (such as PKA and protein kinase C, PKC) in addition to GRKs [44], but also its phosphorylation is sometimes not even required for desensitization [45]. Thus, it apparently can desensitize through a plethora of different mechanisms and interactions with various other proteins, and what is more, some of the signaling pathways it elicits display different desensitization kinetics from others, e.g., Ca²⁺ transients induced by AT₁Rs readily and rapidly desensitize, whereas ERK activation and Janus kinase/Signal transducer and activator of transcription (JAK/STAT) signaling emanating from this receptor persist for longer periods of time [46]. Much more has come to light over the past several years about the physiological roles of cardiac Barrs when they mediate G protein-independent signal transduction by the AT₁Rs in the heart. The first such landmark study was conducted in 2005 and showed, remarkably, that an artificially constructed AT_{1A}R mutant (AT1-i2m), incapable of activating G proteins but able to interact with βarrs, led to significantly less myocardial apoptosis and fibrosis,

and enhanced cardiomyocyte hypertrophy, bradycardia, and fetal cardiac gene expression upon its exogenous overexpression in cardiomyocytes of transgenic mice in vivo, compared to wild type cardiac AT₁R expressed at similar receptor levels (B_{max} values) [47]. In primary cardiomyocytes, the AT₁R βarr-"biased" agonist AngII peptide analog SII [48] stimulates cardiomyocyte proliferation independently of G proteins [49], but not hypertrophy, which requires $G_{g/11}$ protein signaling [50,51] (Figure 1). In addition, this βarr agonist peptide produces positive inotropic and lusitropic effects in isolated murine cardiomyocytes through GRK6-mediated phosphorylation of the cardiomyocyte AT₁R and subsequent βarr2 activation [49] (Figure 1). Interestingly, GRK2-mediated phosphorylation of the AT₁R in cardiac myocytes leads to activation of the other βarr isoform (βarr1), and cardiac βarr1 seems to oppose these positive effects of βarr2 on AT₁R-elicited contractility and relaxation, i.e., βarr1-mediated signaling results in negative inotropy and lusitropy upon AT₁R activation in cardiac myocytes [49] (Figure 1). These findings are entirely consistent with specialized roles of the various GRK isoforms described in transfected systems [52], and also with the concept of GRK-induced receptor "barcoding", i.e., the phenomenon in which different GRK isoforms acting on the very same GPCR induce subsequent recruitment of different Barr isoforms resulting in different downstream signaling events and cellular responses, presumably by phosphorylating the same receptor at different sites/residues [53]. In contrast with isolated murine cardiac myocytes however, SII-activated AT₁R (i.e., AT₁R-bound βarrs) does not seem to produce any inotropic or chronotropic effects in isolated Langendorff-perfused cardiac preparations, despite the fact that ERK1/2, which presumably mediate the positive inotropic effects of βarr2 in isolated cardiac myocytes, are also activated by AT₁R-bound Barrs in Langendorff preparations [54]. Thus, it seems that these positive inotropic effects of cardiac Barr2 are strongly cell type- and experimental condition-dependent. Nevertheless, a consensus has emerged, according to which cardiomyocyte-residing AT₁Rs promote hypertrophy and cardiomyocyte proliferation via βarrs, as well as contractility via (at least) βarr2, whereas cardiac fibroblast-residing AT₁Rs promote fibrosis and cardiac adverse remodeling via the classical G_{q/11} protein-PKC-Ca²⁺ signaling pathway (Figure 1). Since βarr2 also terminates the G protein-mediated signaling of the AT₁R, stimulation of cardiac βarr2 activity and/or blockade of cardiac βarr1 activity at the AT₁Rs of the heart might be sought after for the treatment of post-MI HF and the cardiac hypertrophy and adverse remodeling that accompany this devastating disease. Indeed, a compound analogous to SII, i.e., a βarr-"biased" AT₁R peptide agonist that selectively activates βarrs while blocking G-protein signaling, TRV120027, has shown very promising results in canine models of acute HF, blocking the undesirable G protein-mediated AT₁R-induced vasoconstriction, thereby preserving renal function, while, at the same time, enhancing the desirable (in acute HF) βarr-dependent contractility of cardiac myocytes [55], and it is currently under development for the treatment of HF.

Another interesting example of cardiac AT_1R β arr-mediated signaling is that of the mechanical stretch-activated AT_1R . A recent study showed that simple mechanical stretch (in the absence of any ligand) can actually activate the $AT_{1A}R$ leading to selective β arr recruitment and signaling without concomitant G protein activation [56]. What is more, the authors went on to show in an *ex vivo* murine heart model that this stretch-activated $AT_{1A}R$ -elicited β arr signaling resulted in enhanced ERK1/2 and Akt kinase (protein kinase B, PKB) activation, as well as EGFR transactivation, effects believed to mediate enhanced cardiomyocyte survival and protection (inhibition of apoptosis) [56]. In mouse hearts lacking β arrs or $AT_{1A}Rs$, mechanical stretch failed, of course, to produce these responses and

led, instead, to enhanced myocyte apoptosis [56]. Thus, it appears that the heart is also capable of responding to acute increases in mechanical stress by activating cardiac βarr-mediated cell survival signals, which again argues in favor of a beneficial and therapeutically desirable physiological role for cardiac AT₁R βarr-dependent signaling (at least for the cardiac βarr2 isoform-dependent one).

5. Other Cardiac GPCRs and βarrs

Unfortunately, and although there is a plethora of other GPCRs expressed in cardiac myocyte membranes, whose signaling also plays important roles in regulation of cardiac function, e.g., glucagon receptors that stimulate contractility, vasopressin receptors (V_1Rs) and α_1ARs that promote hypertrophy and cell proliferation, adenosine receptors that regulate heart rhythm, *etc*. [4], the involvement of cardiac β arrs (if any) in their signaling and function specifically in the heart has not so far been investigated. As their importance for cardiac physiology and for heart disease treatments continues to unravel though, investigations of the effects of cardiac β arrs on signaling and function of these GPCRs, as well, are bound to come to light.

6. Unanswered Questions on Cardiac Barrs

Clearly, several interesting and important questions about the roles of the two βarrs in cardiac physiology and disease have arisen from the preceding sections, which currently await elucidation in future studies. With regards to β_1AR signaling and function in the heart, the emerging consensus is that βarr1 is generally detrimental, and βarr2 might actually be beneficial for β₁AR-regulated cardiac function. Barr1 appears to be cardiotoxic, since (a) it is this cardiac Barr isoform that is responsible for β₁AR desensitization (G protein uncoupling) and downregulation, processes significantly contributing to the decline of cardiac β-adrenergic and inotropic reserves that underlies the pathophysiology of HF [27]; and (b) the G protein-independent signaling from cardiac β_1AR it mediates is largely cardiotoxic (inhibition of EGFR transactivation and CaMKII induction leading to cardiac apoptosis, SERCA2a activity lowering leading to reduced contractility, etc.) [27] (Figure 1). Coupled with its negative actions in the adrenal glands (stimulation of catecholamine secretion and of aldosterone production and secretion) during HF [27,57-59], which substantially increase the neurohormonal burden of the failing heart, βarr1 seems to be a valid and potentially important therapeutic target in post-MI HF (and maybe also in other heart diseases). Conversely, βarr2 appears to be cardioprotective, since (a) it does not seem to participate in β_1AR desensitization/downregulation [27]; and (b) the G protein-independent signaling from cardiac β₁AR it mediates is largely beneficial (EGFR transactivation and ERK activation leading to survival/proliferation, SERCA2a activity enhancement, via SUMOylation, leading to increased contractility, etc.) [27] (Figure 1). Also, it does not seem to have significant neurohormonal effects in the adrenals [56–58]. However, whether it is also involved in cardiac β_1AR -induced CaMKII activation, as β arr1 is [31], is an open question that needs to be addressed in future investigations.

With regards to β arr involvement in β_2AR signaling and function in the heart, even less is known at present. Do both β arrs desensitize this receptor in the failing heart or one of the two selectively? Do both β arrs recruit PDE4D to the cardiac β_2AR thereby diminishing its pro-contractile signaling or one of the two selectively (Figure 1) [37,39]? Do both β arrs participate in its purportedly beneficial,

anti-apoptotic signaling (e.g., towards ERK activation and NF- κ B inhibition) [60,61] in the heart or, again, one of the two selectively? Finally, do the β -blockers that stimulate β arr signaling from the β_2 AR (such as carvedilol) and exert beneficial effects on the failing heart induce binding of both β arrs equally to the cardiac β_2 AR *in vivo* or preferentially induce binding of one β arr over the other [42,62]? Since cardiac β arr1-dependent signaling appears to be damaging and cardiac β arr2-dependent signaling beneficial, preferential induction of β arr2 over β arr1 at the cardiomyocyte β_2 AR by " β arr-biased" β -blockers, like carvedilol, might be one mechanism by which these drugs exert their beneficial effects in the heart. Nevertheless, all these are very important questions that absolutely need to be elucidated in order to fully assess the impact of β arrs on cardiac physiology and their potential as therapeutic targets for treatment of HF and of other heart diseases.

As far as cardiac AT_1R signaling and function are concerned, important, currently unanswered questions on the roles of β arrs include: (a) whether β arrs are actually involved in regulation of AT_1R -dependent contractility (and if cardiac AT_1R is capable at all of signaling to increased contractility to begin with) and whether they oppose each other in that regard, *i.e.*, $GRK2/\beta$ arr1 block and $GRK6/\beta$ arr2 promote AT_1R -dependent contractility (Figure 1), as suggested by *in vitro* studies in cultured cardiomyocytes [49]; (b) whether β arrs differ in their signaling to ERK and Akt activation and EGFR transactivation from the cardiac AT_1R ; (c) whether both contribute to the anti-apoptotic, pro-survival and pro-proliferative (e.g., ERK1/2) signaling of AT_1R in the heart or one cardiac β arr mediates that signaling more than the other [63]; (d) whether they differ in desensitization (uncoupling) of the classical $G_{q/11}$ protein signaling of the AT_1R , which is known to result in maladaptive hypertrophy and adverse remodeling in the heart (Figure 1) [50,51], and, finally; (e) whether mechanical stretch preferentially activates one cardiac β arr over the other at the cardiac AT_1R or both equally [56]. Given that β arr "biased" agonists for the AT_1R are currently in development for HF [64,65], answering these questions would tremendously aid work towards the full realization of the potential and the prospects of these kind of compounds for cardiovascular therapy.

In summary, the majority of the current literature on β arrs and cardiac β AR and AT₁R signaling and function seems to overwhelmingly support the notion that β arr2 has (mainly) positive roles in cardiac physiology and function, whereas β arr1 has an overall negative impact on cardiac homeostasis and function, while also opposing several of the beneficial effects of cardiac β arr2. Thus, selective β arr2 stimulation in the heart and/or selective β arr1 inhibition (which leads to β arr2 stimulation indirectly, since the two β arrs always compete with each other inside the cell for binding to any given agonist-activated receptor) might be a valid therapeutic strategy in HF.

7. Conclusions and Future Perspectives

The two β arrs were discovered over two decades ago as plain negative regulators of G protein signaling by GPCRs. The realization that they can also act as signal transducers for these receptors, which occurred around the last turn of the century, revolutionized the fields of β arr biology, physiology and pharmacology. Nowadays, they are known to play important roles in function and pathophysiology of almost every organ/system in mammals, including the heart, via their actions on signaling of cardiac GPCRs such as the β_1 AR, the β_2 AR, and the AT₁R. The accelerating pace at which their roles in the heart are being uncovered has already brought cardiac β arrs to the attention of

pharmaceutical scientists and medicinal chemists, and may soon bring them also into the clinic as valid targets for cardiovascular therapy. Before they can be considered for therapeutic targeting in the heart however, full delineation of their effects on cardiac adrenergic, angiotensin and other GPCRs is warranted. With regards to cardiac βARs , a consensus had already begun to emerge: $\beta arr2$ appears cardioprotective and $\beta arr1$ cardiotoxic. Thus, $\beta arr2$ stimulation and/or $\beta arr1$ inhibition at cardiac βARs , either directly with pharmacological tools (e.g., small molecules or peptides) or indirectly with biased receptor ligands that bind the cardiac βAR extracellular side and selectively activate $\beta arr2$ on its intracellular side, seems to be a good strategy for future cardiovascular drug development. However, there is still a long list of question marks regarding cardiac $\beta arrs$ and the ΔT_1R , and even more questions regarding cardiac $\beta arrs$ and other GPCRs, not to mention that the puzzle of cardiac $\beta arrs$ and βARs is still far from being complete, as well. What we already know with certainty is that the two βarr isoforms, once thought of as complementary to each other, are anything but equal or interchangeable when it comes to their actions in the heart. The task of the future investigations in the field of cardiovascular $\beta arrs$ is thus to fully delineate their differences in heart physiology and disease.

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Conflicts of Interest

The authors declare no conflict of interest.

References

- 1. Lymperopoulos, A.; Bathgate, A. Arrestins in the cardiovascular system. *Prog. Mol. Biol. Transl. Sci.* **2013**, *118*, 297–334.
- 2. Kenakin, T. Making receptors a reality: The 2012 Nobel Prize in Chemistry. *Trends Pharmacol. Sci.* **2013**, *34*, 2–5.
- 3. Lymperopoulos, A.; Rengo, G.; Koch, W.J. Adrenergic nervous system in heart failure: Pathophysiology and therapy. *Circ. Res.* **2013**, *113*, 739–753.
- 4. Siryk-Bathgate, A.; Dabul, S.; Lymperopoulos, A. Current and future G protein-coupled receptor signaling targets for heart failure therapy. *Drug Des. Dev. Ther.* **2013**, 7, 1209–1222.
- 5. Lymperopoulos, A. Physiology and pharmacology of the cardiovascular adrenergic system. *Front. Physiol.* **2013**, *4*, 240.
- 6. Lymperopoulos, A. β-arrestin biased agonism/antagonism at cardiovascular seven transmembrane-spanning receptors. *Curr. Pharm. Des.* **2012**, *18*, 192–198.
- 7. Lymperopoulos, A.; Rengo, G.; Koch, W.J. GRK2 inhibition in heart failure: Something old, something new. *Curr. Pharm. Des.* **2012**, *18*, 186–191.
- 8. Gurevich, V.V.; Gurevich, E.V. The structural basis of arrestin-mediated regulation of G-protein-coupled receptors. *Pharmacol. Ther.* **2006**, *110*, 465–502.

- 9. Shukla, A.K.; Violin, J.D.; Whalen, E.J.; Gesty-Palmer, D.; Shenoy, S.K.; Lefkowitz, R.J. Distinct conformational changes in β-arrestin report biased agonism at seven-transmembrane receptors. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 9988–9993.
- 10. Gurevich, V.V.; Pals-Rylaarsdam, R.; Benovic, J.L.; Hosey, M.M.; Onorato, J.J. Agonist-receptor-arrestin, an alternative ternary complex with high agonist affinity. *J. Biol. Chem.* **1997**, *272*, 28849–28852.
- 11. Luttrell, L.M.; Gesty-Palmer, D. Beyond desensitization: Physiological relevance of arrestin dependent signaling. *Pharmacol. Rev.* **2010**, *62*, 305–330.
- 12. Ferguson, S.S. Evolving concepts in G protein-coupled receptor endocytosis: The role in receptor desensitization and signaling. *Pharmacol. Rev.* **2001**, *53*, 1–24.
- 13. Oakley, R.H.; Laporte, S.A.; Holt, J.A.; Caron, M.G.; Barak, L.S. Differential affinities of visual arrestin, βarrestin1, and βarrestin2 for G protein-coupled receptors delineate two major classes of receptors. *J. Biol. Chem.* **2000**, *275*, 17201–17210.
- 14. Charest, P.G.; Terrillon, S.; Bouvier, M. Monitoring agonist-promoted conformational changes of β-arrestin in living cells by intramolecular BRET. *EMBO Rep.* **2005**, *6*, 334–340.
- 15. Daaka, Y.; Luttrell, L.M.; Lefkowitz, R.J. Switching of the coupling of the β2-adrenergic receptor to different G proteins by protein kinase A. *Nature* **1997**, *390*, 88–91.
- 16. Communal, C.; Singh, K.; Sawyer, D.B.; Colucci, W.S. Opposing effects of β_1 and β_2 -aadrenergic receptors on cardiac myocyte apoptosis: Role of a pertussis toxin-sensitive G protein. *Circulation* **1999**, *100*, 2210–2212.
- 17. Chesley, A.; Lundberg, M.S.; Asai, T.; Xiao, R.P.; Ohtani, S.; Lakatta, E.G.; Crow, M.T. The β₂-adrenergic receptor delivers an antiapoptotic signal to cardiac myocytes through G_i-dependent coupling to phosphatidylinositol 3-kinase. *Circ. Res.* **2000**, *87*, 1172–1179.
- 18. Zhu, W.Z.; Zheng, M.; Koch, W.J.; Lefkowitz, R.J.; Kobilka, B.K.; Xiao, R.P. Dual modulation of cell survival and cell death by β₂-adrenergic signalling in adult mouse cardiomyocytes. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 1607–1612.
- 19. Dorn, G.W.; Tepe, N.M.; Lorenz, J.N.; Koch, W.J.; Liggett, S.B. Low- and high-level transgenic expression of β₂-adrenergic receptors differentially affect cardiac hypertrophy and function in Gαq-overexpressing mice. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 6400–6405.
- 20. Conner, D.A.; Mathier,, M.A.; Mortensen, R.M.; Christe, M.; Vatner, S.F.; Seidman, C.E.; Seidman, J.G. β-Arrestin1 knockout mice appear normal but demonstrate altered cardiac responses to β-adrenergic stimulation. *Circ. Res.* **1997**, *81*, 1021–1026.
- 21. Xiang, Y.; Kobilka, B.K. Myocyte adrenoceptor signaling pathways. *Science* **2003**, *300*, 1530–1532.
- 22. Bristow, M.R. β-adrenergic receptor blockade in chronic heart failure. *Circulation* **2000**, *101*, 558–569.
- 23. Packer, M.; Bristow, M.R.; Cohn, J.N.; Colucci, W.S.; Fowler, M.B.; Gilbert, E.M.; Shusterman, N.H. The effect of carvedilol on morbidity and mortality in patients with chronic heart failure. *N. Engl. J. Med.* **1996**, *334*, 1349–1355.
- 24. Noma, T.; Lemaire, A.; Naga Prasad, S.V.; Barki-Harrington, L.; Tilley, D.G.; Chen, J.; le Corvoisier, P.; Violin, J.D.; Wei, H.; Lefkowitz, R.J.; *et al.* β-arrestin-mediated β1-adrenergic receptor transactivation of the EGFR confers cardioprotection. *J. Clin. Invest.* **2007**, *117*, 2445–2458.

- 25. Tilley, D.G. G protein-dependent and G protein-independent signaling pathways and their impact on cardiac function. *Circ. Res.* **2011**, *109*, 217–230.
- 26. Noor, N.; Patel, C.B.; Rockman, H.A. β-arrestin: A signaling molecule and potential therapeutic target for heart failure. *J. Mol. Cell. Cardiol.* 2011, *51*, 534–41.
- 27. Bathgate-Siryk, A.; Dabul, S.; Pandya, K.; Walklett, K.; Rengo, G.; Cannavo, A.; de Lucia, C.; Liccardo, D.; Gao, E.; Leosco, D.; *et al.* Negative impact of β-arrestin-1 on post-myocardial infarction heart failure via cardiac and adrenal-dependent neurohormonal mechanisms. *Hypertension* **2013**, doi:10.1161/HYPERTENSIONAHA.113.02043.
- 28. Bers, D.M. Calcium cycling and signaling in cardiac myocytes. *Annu. Rev. Physiol.* **2008**, 70, 23–49.
- 29. Kho, C.; Lee, A.; Jeong, D.; Oh, J.G.; Chaanine, A.H.; Kizana, E.; Park, W.J.; Hajjar, R.J. SUMO1-dependent modulation of SERCA2a in heart failure. *Nature* **2011**, *477*, 601–605.
- 30. Lymperopoulos, A. Nova Southeastern University, Unpublished Observation, 2013.
- 31. Mangmool, S.; Shukla, A.K.; Rockman, H.A. β-Arrestin-dependent activation of Ca²⁺/calmodulin kinase II after β₁-adrenergic receptor stimulation. *J. Cell Biol.* **2010**, *189*, 573–587.
- 32. Anderson, M.E. CaMKII and a failing strategy for growth in heart. *J. Clin. Invest.* **2009**, *119*, 1082–1085.
- 33. Xiao, R.P.; Zhu, W.; Zheng, M.; Chakir, K.; Bond, R.; Lakatta, E.G.; Cheng, H. Subtype-specific β-adrenoceptor signaling pathways in the heart and their potential clinical implications. *Trends Pharmacol. Sci.* **2004**, *25*, 358–365.
- 34. Luttrell, L.M.; Roudabush, F.L.; Choy, E.W.; Miller, W.E.; Field, M.E.; Pierce, K.L.; Lefkowitz, R.J. Activation and targeting of extracellular signal-regulated kinases by β-arrestin scaffolds. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 2449–2454.
- 35. Witherow, D.S.; Garrison, T.R.; Miller, W.E.; Lefkowitz, R.J. β-Arrestin inhibits NF-κB activity by means of its interaction with the NF-κB inhibitor IkappaBalpha. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 8603–8607.
- 36. Gao, H.; Sun, Y.; Wu, Y.; Luan, B.; Wang, Y.; Qu, B.; Pei, G. Identification of β-arrestin2 as a G protein-coupled receptor-stimulated regulator of NF-κB pathways. *Mol. Cell.* **2004**, *14*, 303–317.
- 37. Richter, W.; Day, P.; Agrawal, R.; Bruss, M.D.; Granier, S.; Wang, Y.L.; Rasmussen, S.G.; Horner, K.; Wang, P.; Lei, T.; *et al.* Signaling from β1- and β2-adrenergic receptors is defined by differential interactions with PDE4. *EMBO J.* **2008**, *27*, 384–393.
- 38. Xiang, Y.; Naro, F.; Zoudilova, M.; Jin, S.L.; Conti, M.; Kobilka, B. Phosphodiesterase 4D is required for β2-adrenoceptor subtype-specific signaling in cardiac myocytes. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 909–914.
- 39. Salazar, N.C.; Vallejos, X.; Siryk, A.; Rengo, G.; Cannavo, A.; Liccardo, D.; de Lucia, C.; Gao, E.; Leosco, D.; Koch, W.J.; *et al.* GRK2 blockade with βARKct is essential for cardiac β2-adrenergic receptor signaling towards increased contractility. *Cell Commun. Signal.* **2013**, 11, 64.
- 40. Houslay, M.D.; Baillie, G.S.; Maurice, D.H. cAMP-Specific phosphodiesterase-4 enzymes in the cardiovascular system: A molecular toolbox for generating compartmentalized cAMP signaling. *Circ. Res.* **2007**, *100*, 950–966.

- 41. Drake, M.T.; Violin, J.D.; Whalen, E.J.; Wisler, J.W.; Shenoy, S.K.; Lefkowitz, R.J. β-arrestin-biased agonism at the β₂-adrenergic receptor. *J. Biol. Chem.* **2008**, *283*, 5669–5676.
- 42. Wisler, J.W.; DeWire, S.M.; Whalen, E.J.; Violin, J.D.; Drake, M.T.; Ahn, S.; Shenoy, S.K.; Lefkowitz, R.J. A unique mechanism of β-blocker action: Carvedilol stimulates β-arrestin signaling. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 16657–16662.
- 43. Billet, S.; Aguilar, F.; Baudry, C.; Clauser, E. Role of angiotensin II AT₁ receptor activation in cardiovascular diseases. *Kidney Int.* **2008**, *74*, 1379–1384.
- 44. Balmforth, A.J.; Shepherd, F.H.; Warburton, P.; Ball, S.G. Evidence of an important and direct role for protein kinase C in agonist-induced phosphorylation leading to desensitization of the angiotensin AT_{1A} receptor. *Br. J. Pharmacol.* **1997**, *122*, 1469–1477.
- 45. Olivares-Reyes, J.A.; Smith, R.D.; Hunyady, L.; Shah, B.H.; Catt, K.J. Agonist-induced signaling, desensitization, and internalization of a phosphorylation-deficient AT_{1A} angiotensin receptor. *J. Biol. Chem.* **2001**, *276*, 37761–37768.
- 46. Thomas, W.G.; Thekkumkara, T.J.; Baker, K.M. Cardiac effects of AII. AT_{1A} receptor signaling, desensitization, and internalization. *Adv. Exp. Med. Biol.* **1996**, *396*, 59–69.
- 47. Zhai, P.; Yamamoto, M.; Galeotti, J.; Liu, J.; Masurekar, M.; Thaisz, J.; Irie, K.; Holle, E.; Yu, X.; Kupershmidt, S.; *et al.* Cardiac-specific overexpression of AT1 receptor mutant lacking G αq/G α i coupling causes hypertrophy and bradycardia in transgenic mice. *J. Clin. Invest.* **2005**, 115, 3045–3056.
- 48. Holloway, A.C.; Qian, H.; Pipolo, L.; Ziogas, J.; Miura, S.; Karnik, S.; Southwell, B.R.; Lew, M.J.; Thomas, W.G. Side-chain substitutions within angiotensin II reveal different requirements for signaling, internalization, and phosphorylation of type 1A angiotensin receptors. *Mol. Pharmacol.* **2002**, *61*, 768–777.
- 49. Rajagopal, K.; Whalen, E.J.; Violin, J.D.; Stiber, J.A.; Rosenberg, P.B.; Premont, R.T.; Coffman, T.M.; Rockman, H.A.; Lefkowitz, R.J. β-arrestin2-mediated inotropic effects of the angiotensin II type 1A receptor in isolated cardiac myocytes. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 16284–16289.
- 50. Mishra, S.; Ling, H.; Grimm, M.; Zhang, T.; Bers, D.M.; Brown, J.H. Cardiac hypertrophy and heart failure development through Gq and CaM kinase II signaling. *J. Cardiovasc. Pharmacol.* **2010**, *56*, 598–603.
- 51. Dorn, G.W., II; Force, T. Protein kinase cascades in the regulation of cardiac hypertrophy. *J. Clin. Invest.* **2005**, *115*, 527–537.
- 52. Kukkonen, J.P. Regulation of receptor-coupling to (multiple) G proteins. A challenge for basic research and drug discovery. *Recept. Channels* **2004**, *10*, 167–183.
- 53. Zidar, D.A.; Violin, J.D.; Whalen, E.J.; Lefkowitz, R.J. Selective engagement of G protein coupled receptor kinases (GRKs) encodes distinct functions of biased ligands. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 9649–9654.
- 54. Aplin, M.; Christensen, G.L.; Schneider, M.; Heydorn, A.; Gammeltoft, S.; Kjølbye, A.L.; Sheikh, S.P.; Hansen, J.L. Differential extracellular signal-regulated kinases 1 and 2 activation by the angiotensin type 1 receptor supports distinct phenotypes of cardiac myocytes. *Basic Clin. Pharmacol. Toxicol.* **2007**, *100*, 296–301.

- 55. Boerrigter, G.; Lark, M.W.; Whalen, E.J.; Soergel, D.G.; Violin, J.D.; Burnett, J.C., Jr. Cardiorenal actions of TRV120027, a novel β-arrestin-biased ligand at the angiotensin II type I receptor, in healthy and heart failure canines: A novel therapeutic strategy for acute heart failure. *Circ. Heart Fail.* **2011**, *4*, 770–778.
- 56. Rakesh, K.; Yoo, B.; Kim, I.M.; Salazar, N.; Kim, K.S.; Rockman, H.A. β-Arrestin-biased agonism of the angiotensin receptor induced by mechanical stress. *Sci. Signal.* **2010**, *3*, ra46.
- 57. Lymperopoulos, A.; Rengo, G.; Funakoshi, H.; Eckhart, A.D.; Koch, W.J. Adrenal GRK2 upregulation mediates sympathetic overdrive in heart failure. *Nat. Med.* **2007**, *13*, 315–323.
- 58. Lymperopoulos, A.; Rengo, G.; Zincarelli, C.; Kim, J.; Soltys, S.; Koch, W.J. An adrenal β-arrestin 1-mediated signaling pathway underlies angiotensin II-induced aldosterone production *in vitro* and *in vivo*. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 5825–5830.
- 59. Lymperopoulos, A.; Rengo, G.; Zincarelli, C.; Kim, J.; Koch, W.J. Adrenal β-arrestin 1 inhibition *in vivo* attenuates post-myocardial infarction progression to heart failure and adverse remodeling via reduction of circulating aldosterone levels. *J. Am. Coll. Cardiol.* **2011**, *57*, 356–365.
- 60. Hasseldine, A.R.; Harper, E.A.; Black, J.W. Cardiac-specific overexpression of human β2 adrenoceptors in mice exposes coupling to both G_s and G_i proteins. *Br. J. Pharmacol.* **2003**, *138*, 1358–1366.
- 61. DeWire, S.M.; Ahn, S.; Lefkowitz, R.J.; Shenoy, S.K. β-arrestins and cell signaling. *Annu. Rev. Physiol.* **2007**, *69*. 483–510.
- 62. Liu, J.J.; Horst, R.; Katritch, V.; Stevens, R.C.; Wüthrich, K. Biased signaling pathways in β2-adrenergic receptor characterized by ¹⁹F-NMR. *Science* **2012**, *335*, 1106–1110.
- 63. Kim, J.; Ahn, S.; Ren, X.R.; Whalen, E.J.; Reiter, E.; Wei, H.; Lefkowitz, R.J. Functional antagonism of different G protein-coupled receptor kinases for β-arrestin-mediated angiotensin II receptor signaling. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 1442–1447.
- 64. DeWire, S.M.; Violin, J.D. Biased ligands for better cardiovascular drugs: Dissecting G-protein-coupled receptor pharmacology. *Circ. Res.* **2011**, *109*, 205–216.
- 65. Violin, J.D.; Soergel, D.G.; Boerrigter, G.; Burnett, J.C., Jr.; Lark, M.W. GPCR biased ligands as novel heart failure therapeutics. *Trends Cardiovasc. Med.* **2013**, *23*, 242–249.
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