

Supplementary Information

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Abstract: Oligonucleotides carrying amino, thiol groups, as well as fluorescein, *c-myc* peptide sequence and nanogold at internal positions were prepared and used for the assembly of bidimensional DNA arrays.

Keywords: DNA tile; DNA bidimensional arrays; gold nanoparticles; self-assembly

Figure S1. 10% Denaturing (8M urea) polyacrylamide gel analysis of long oligonucleotides. Buffer conditions: 1 x Tris-borate-EDTA (TBE) buffer. The gel was run at 50 °C. Lane 1: 10-base pairs DNA marker. Lane 2: unmodified oligonucleotide B5, **10**. Lane 3: oligonucleotide B5 carrying *c-myc* peptide, **15**. Lane 4: unmodified oligonucleotide B1, **6**. Lane 5: oligonucleotide B5 carrying fluoresceine, **14**.

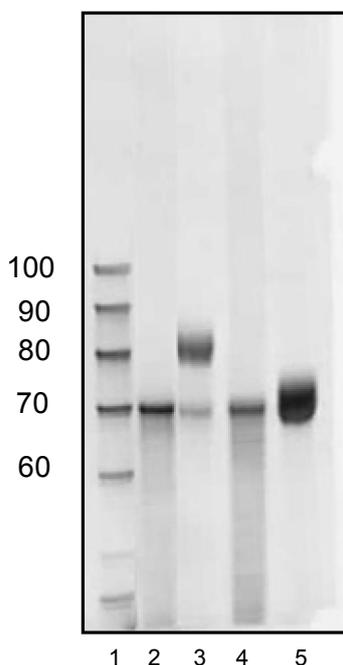


Figure S2. 8% Native polyacrylamide gel analysis of DNA-tile formation. Buffer conditions: TAE-Mg²⁺ buffer. The gel was run at 20 °C. Lane 1: 25-base pairs DNA marker. Lane 2: DNA-tile A (equimolar mixture of oligonucleotides A1-A5, **1-5**). Lane 3: DNA-tile B* (equimolar mixture of oligonucleotides B1-B5, **6-10**). Lane 4: DNA-tile B*-Fluoresceine (equimolar mixture of oligonucleotides B1-B4 and B5-fluoresceine, **6-9** and **14**). Lane 5: DNA-tile B*-*c-myc* peptide (equimolar mixture of oligonucleotides B1-B4 and B5- *c-myc* peptide, **6-9** and **15**).

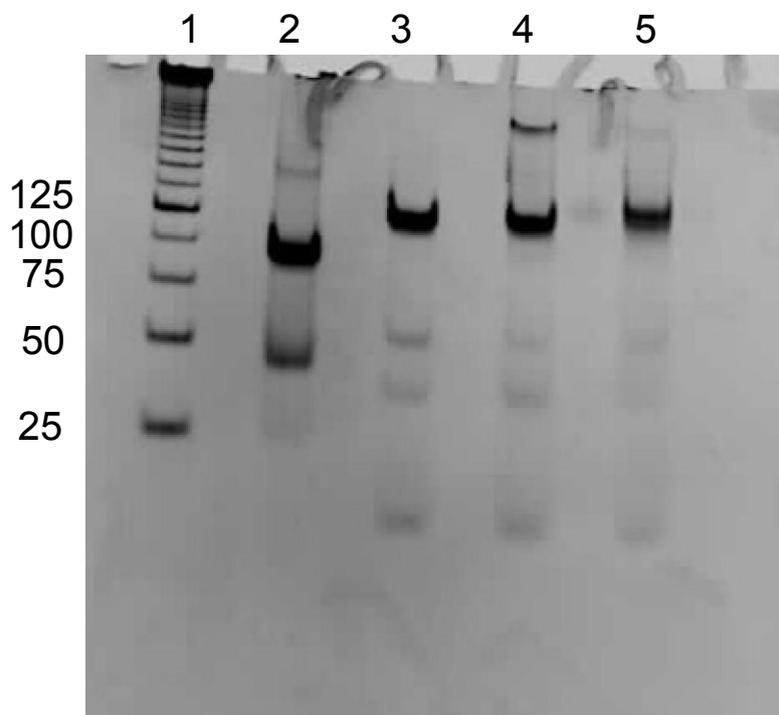


Figure S3. 3% Agarose gel of oligonucleotide-Nanogold conjugate, **16**. Buffer conditions: 0.5 x Tris-borate-EDTA (TBE) buffer. Lane 1: reaction of thiol-oligonucleotide **12** (B1-thiol) with maleimido-NANOGOLD after removal of the *t*-butylthio group. Lane 2: bromophenol blue and xylenecyanol dyes.

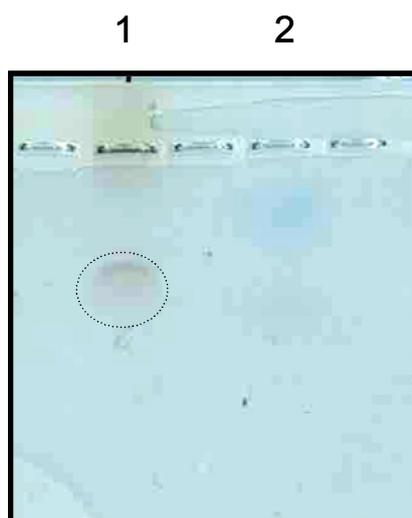
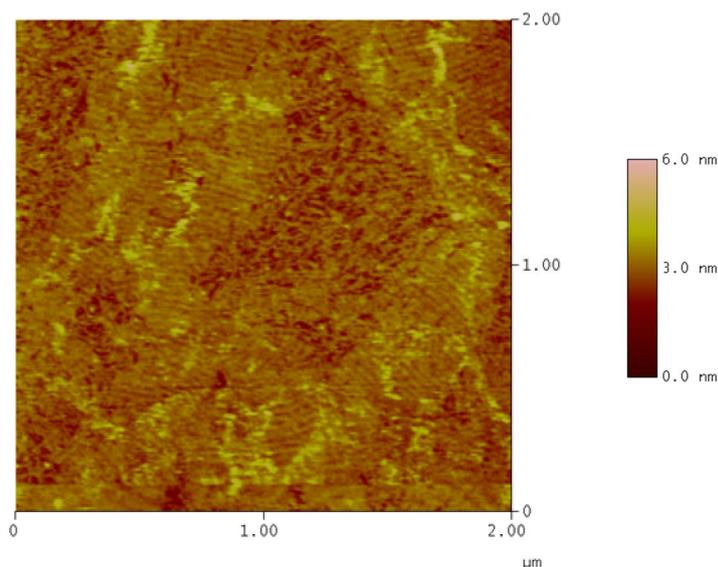
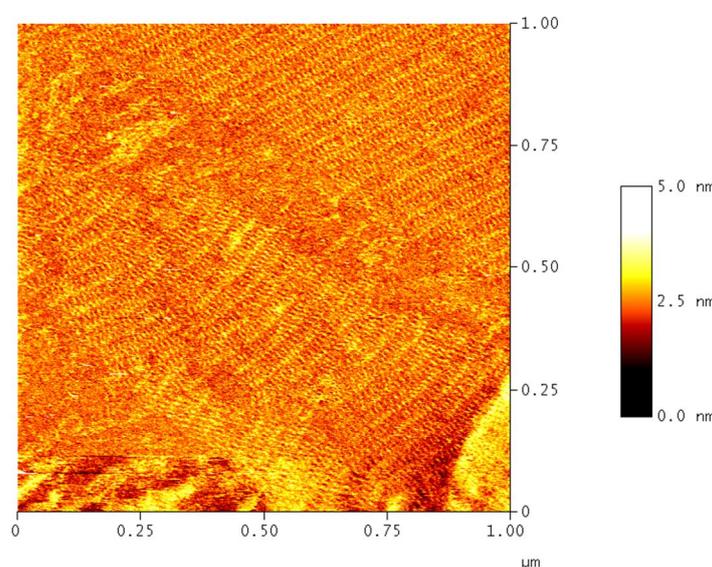


Figure S4. Influence of the purity of the oligonucleotides in the formation of the bidimensional DNA lattices.



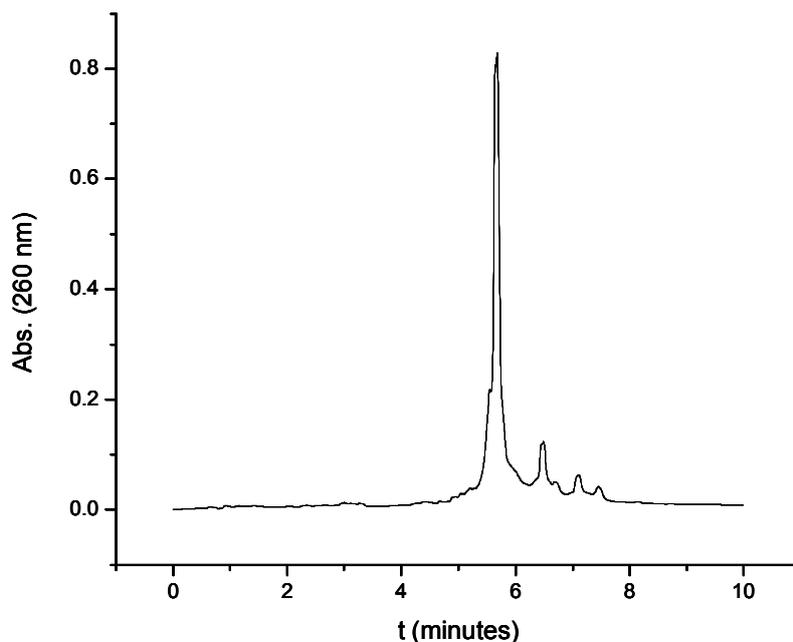
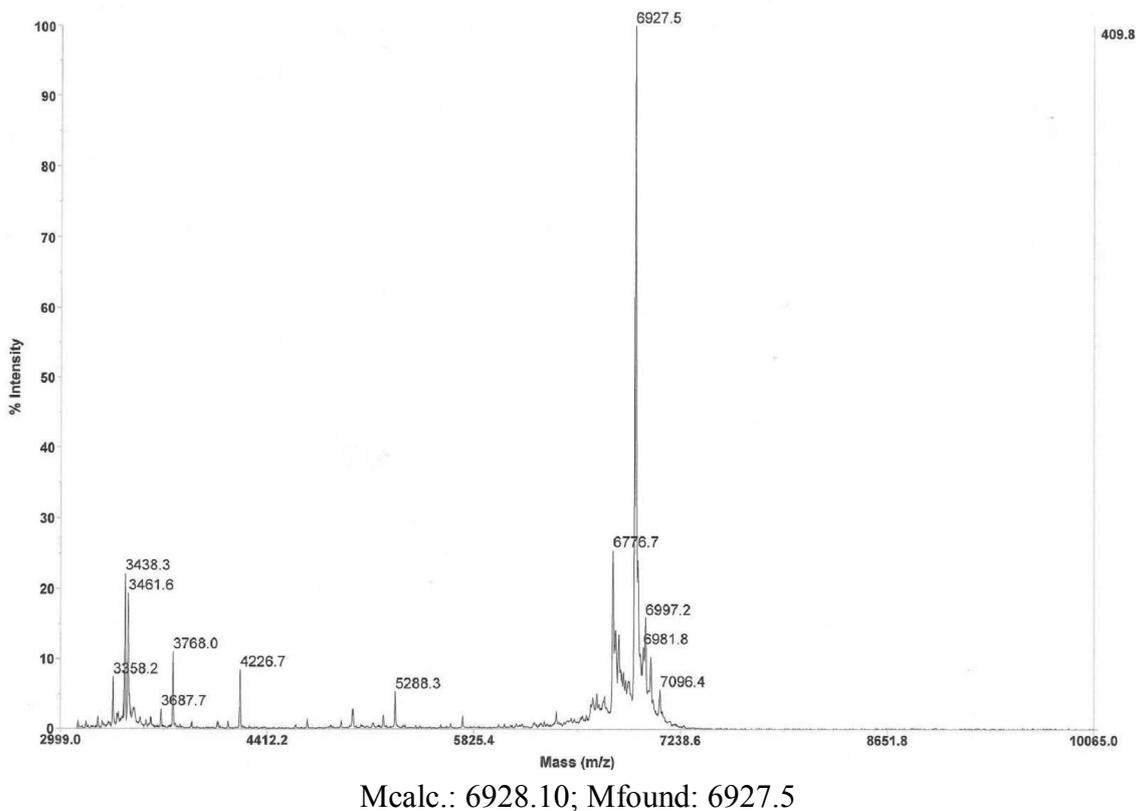
A) HPLC purified oligonucleotides (90–95% purity)



B) Gel-electrophoresis purified oligonucleotides (>99% purity)

Synthesis, Purification and Characterization a 20mer Oligonucleotide Carrying Amino-dT Residues

The assembly of the sequence (5'-CGAGXCAXXGAGXCAXCGAG-3'; X = amino-dT) was performed using standard 200 nmol scale synthesis cycle (LV200). Ammonia deprotection was performed at 50 °C overnight. The resulting crude was analyzed by HPLC (Column: X-BridgeTMOST C₁₈ (2.5 μm, 4.6 × 50 mm). Conditions: solvent A: 5% ACN in 100 mM triethylammonium acetate (pH = 7) and solvent B: 70% ACN in 100 mM triethylammonium acetate (pH = 7). Flow rate: 1 mL/min. 10 min linear gradient from 0–30% B. The main peak observed in the HPLC profile was also analyzed by mass spectrometry (MALDI-TOF). $t_R = 5.7$ min, MALDI-TOF m/z (negative mode [M+H]⁻) calc for C₂₃₆H₃₁₆N₈₇O₁₂₃P₁₉ 6928.10, found 6927.50.

Figure S5. HPLC profile of a 20mer oligonucleotide carrying amino-dT residues.**Figure S6.** Mass spectrometry of a 20mer oligonucleotide carrying amino-dT residues.

Synthesis, Purification and Characterization of a 13mer Carrying Fluoresceine and the c-myc Peptide

As oligonucleotides **14** and **15** are too long for mass spectrometry analysis we have prepared a 13mer DNA sequence carrying the *t*-butyldithio-ethyl-5-methyl-dC residue in an internal position of

the oligonucleotide as model compound to set up the conjugation reactions. The assembly of the sequence **17** (5'-TTCCAYATTACCG; Y = *t*-butyldithio-ethyl-5-methyl-dC) was performed as described in the experimental section. The resulting crude was analyzed by HPLC (Column: X-Bridge™OST C₁₈ (2.5 μm, 4.6 × 50 mm). Conditions: solvent A: 5% ACN in 100 mM triethylammonium acetate (pH = 7) and solvent B: 70% ACN in 100 mM triethylammonium acetate (pH = 7). Flow rate: 1 mL/min. 10 min linear gradient from 5–40% B. The main peak observed in the HPLC profile was analyzed by mass spectrometry (MALDI-TOF). *t*_R = 4.9 min, MALDI-TOF *m/z* (negative mode [M+H]⁻) calc 4031.90, found 4032.52.

To cleave the disulfide bond, compound **17** was treated with tris(2-carboxyethyl)phosphine hydrochloride (TCEP.HCl) solution and allow to react at 55 °C overnight. Under these conditions, the *tert*-butylthiol group was completely removed yielding a new peak at 3.0 min (Figure S7B).

Unprotected thiol oligonucleotide **17** was treated fluorescein diacetate 5-maleimide and maleimido-*N*-Ala-Glu-Gln-Lys-Leu-Ile-Ser-Glu-Glu-Asp-Leu-Asn-CONH₂ (*c-myc* peptide) as described in the experimental section yielding the desired conjugates **18** and **19** that were characterized by mass spectrometry (Figures S9-S12). Compound **18**: *t*_R = 3.6 min, MALDI-TOF *m/z* (negative mode [M+H]⁻) calc for C₁₅₂H₁₈₀N₄₄O₈₄P₁₂S 4372.08, found 4369.44. Compound **19**: *t*_R = 7.8 min, ESI-MS *m/z* (negative mode [M+H]⁻) calc for C₁₉₇H₂₇₀N₆₁O₁₀₂P₁₂S 5529.90, found 5531.93. Also found 5203.34, 4915.00, 4625.39, 4313.32, 4009.95, 3706.07, 3393.19 corresponding to different fragments of the oligonucleotide-peptide conjugate. Each fragment corresponded to the loss of one nucleobase, beginning at the 3'-end and the last fragment corresponded to the oligonucleotide-peptide conjugate having the modified nucleobase the 3'-end.

Table S1. Sequences of oligonucleotides prepared.

#	Sequence (5'-3')
17	TTCCAYATTACCG; Y = <i>t</i> -butyldithio-ethyl-5-methyl-dC
18	TTCCAZATTACCG; Z = <i>N</i> -(fluoresceine-maleimido-S-ethyl)-5-methyl-dC
19	TTCCAZATTACCG; Z = <i>N</i> (<i>c-myc</i> -peptide-maleimido-S-ethyl)-5-methyl-dC; <i>c-myc</i> peptide sequence: maleimido-Ala-Glu-Gln-Lys-Leu-Ile-Ser-Glu-Glu-Asp-Leu-Asn-CONH ₂

Figure S7. HPLC profiles of A) protected oligonucleotide **17** and B) deprotected oligonucleotide **17**.

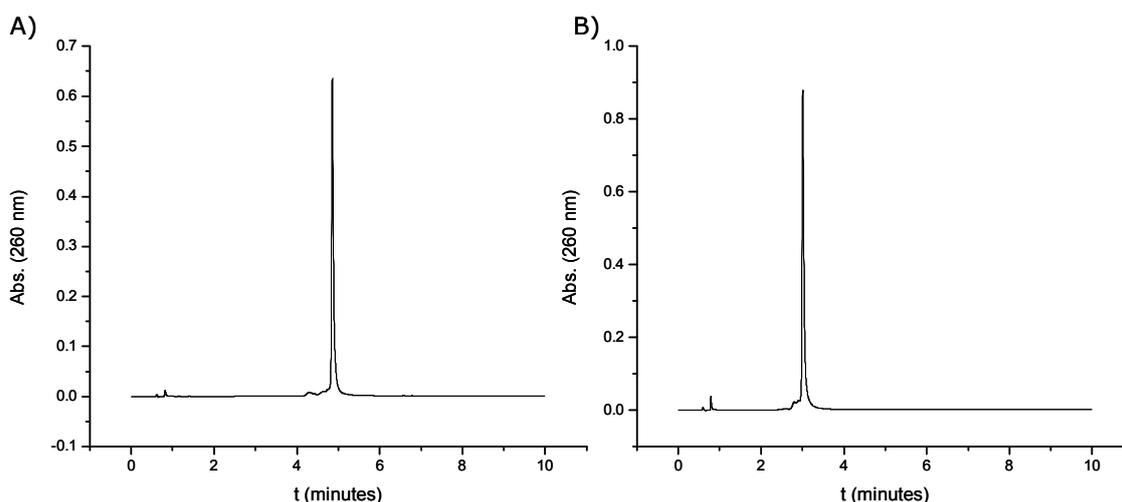
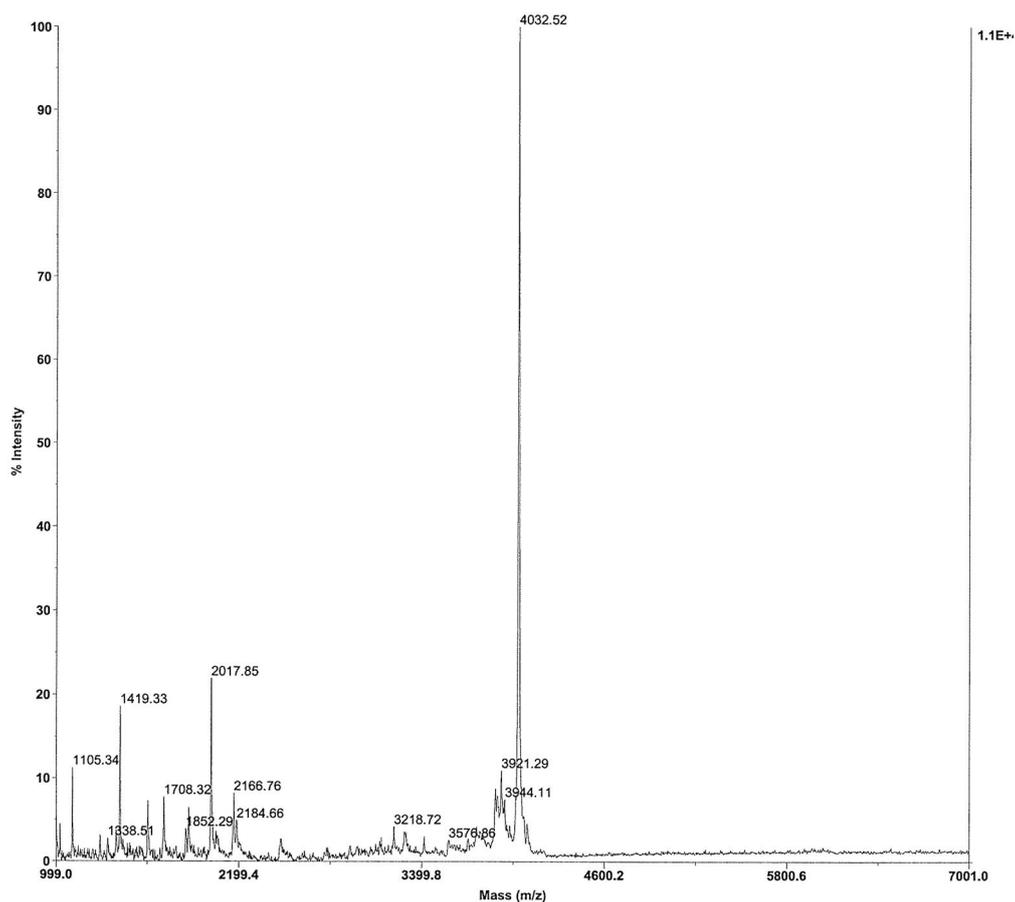


Figure S8. Mass spectrometry of compound 17.



Mcalc.: 4031.90; Mfound: 4032.52

Figure S9. HPLC profile of fluorescein conjugate 18.

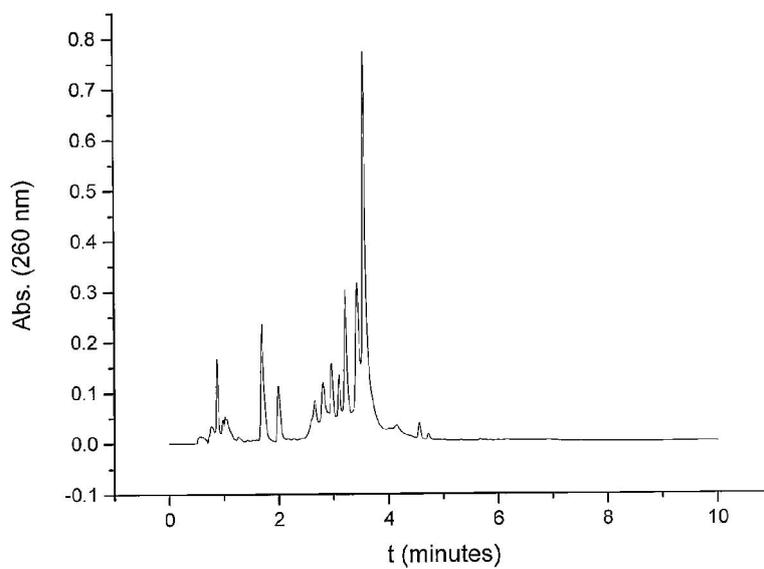
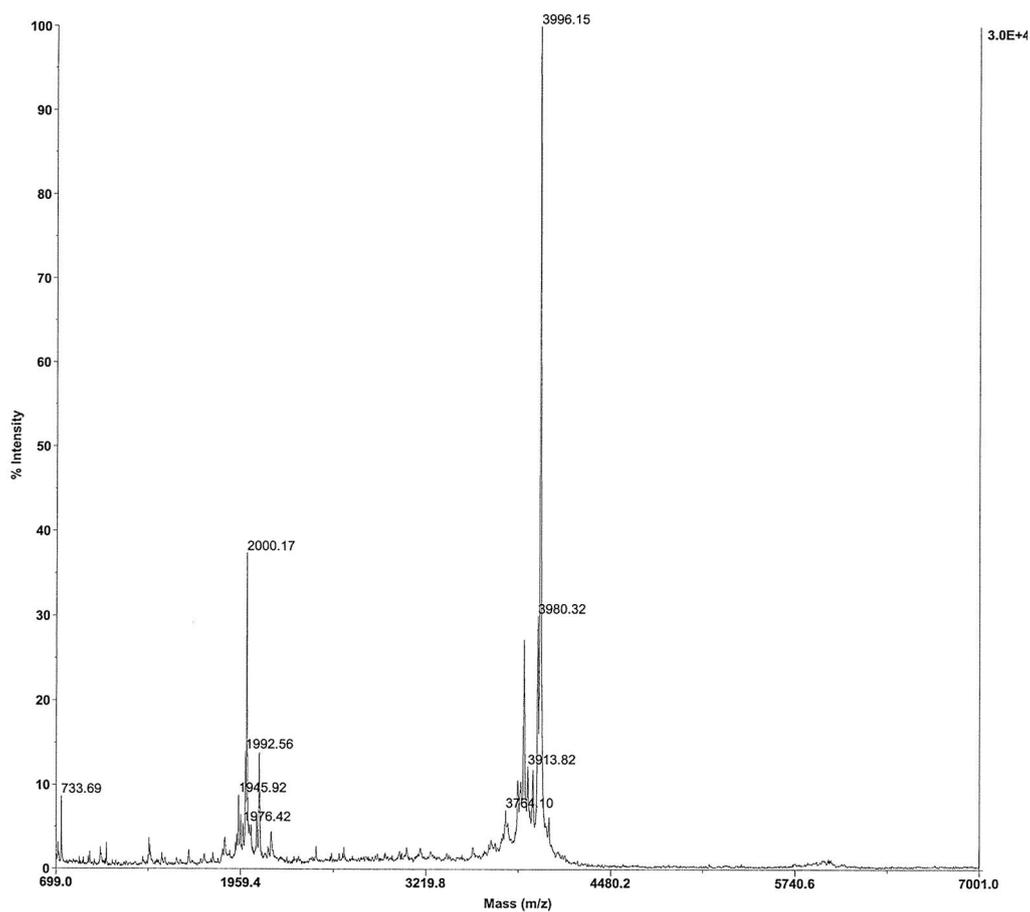


Figure S10. Mass spectrometry of compound 18.

Mcalc.: 4372.08; Mfound: 4369.44

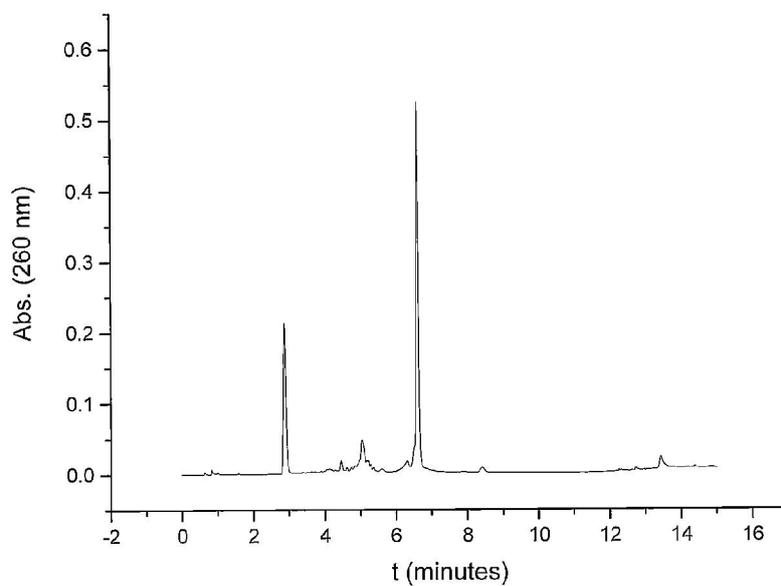
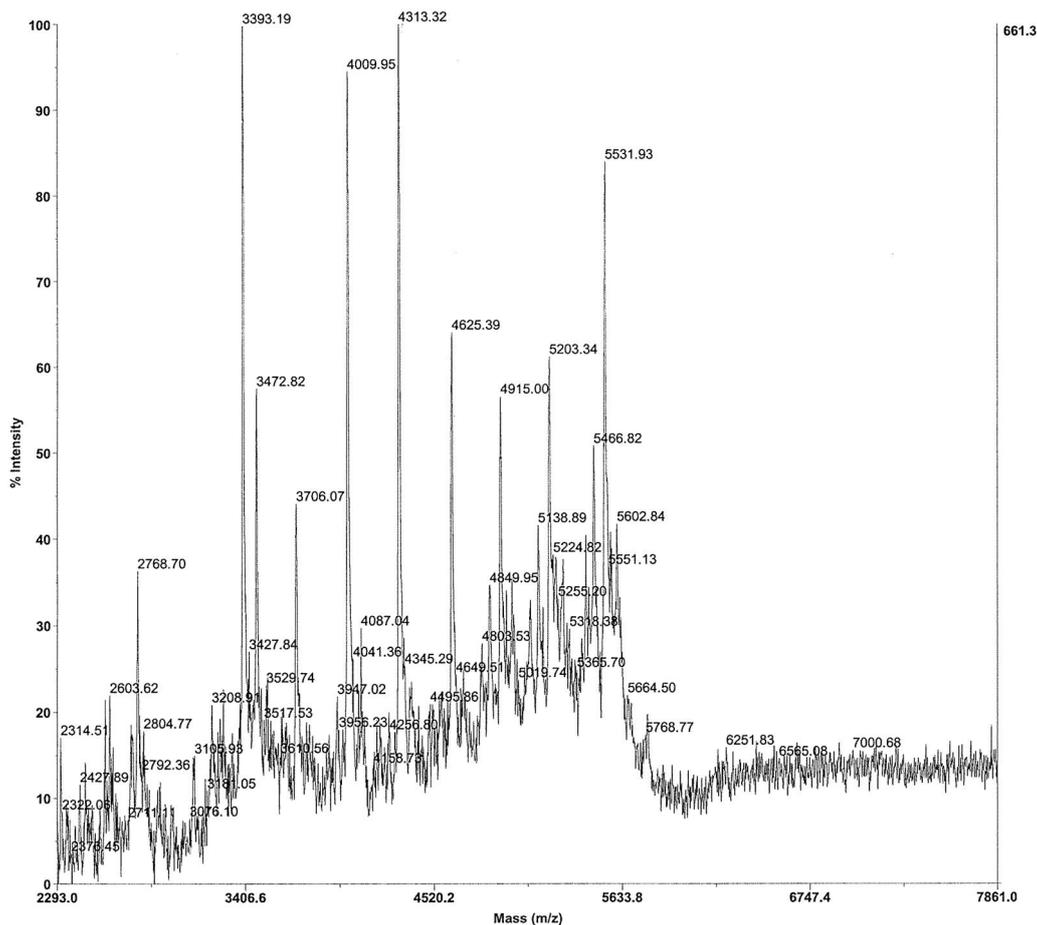
Figure S11. HPLC profile of *c-myc* peptide conjugate 19.

Figure S12. Mass spectrometry of *c-myc* peptide conjugate 19.



Mcalc.: 5529.90; Mfound: 5531.93

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