

Article

Ultra Low-Dose Radiation: Stress Responses and Impacts Using Rice as a Grass Model

Randeep Rakwal^{1,2,*}, Ganesh Kumar Agrawal², Junko Shibato¹, Tetsuji Imanaka³, Satoshi Fukutani³, Shigeru Tamogami⁴, Satoru Endo⁵, Sarata Kumar Sahoo⁶, Yoshinori Masuo¹ and Shinzo Kimura⁷

¹ Health Technology Research Center, National Institute of Advanced Industrial Science and Technology (AIST), West, Tsukuba, Ibaraki 305-8569, Japan; E-Mails: rjunko@nifty.com (J.S.); y-masuo@aist.go.jp (Y.M.)

² Research Laboratory for Biotechnology and Biochemistry (RLABB), GPO Box 8207, Kathmandu, Nepal; E-Mails: plantproteomics@gmail.com (R.R.); gkagrawal123@gmail.com (G.K.A.)

³ Research Reactor Institute, Kyoto University (KURRI), Osaka, Japan; E-Mails: imanaka@rri.kyoto-u.ac.jp (T.I.); fukutani@rri.kyoto-u.ac.jp (S.F.)

⁴ Laboratory of Growth Regulation Chemistry, Akita Prefectural University, Akita 010-0195, Japan; E-Mail: tamo_chem@akita-pu.ac.jp

⁵ Graduate School of Engineering, Hiroshima University (HU), Hiroshima, Japan; E-Mail: endos@hiroshima-u.ac.jp

⁶ Research Center for Radiation Protection, National Institute of Radiological Sciences (NIRS), Chiba 263-8555, Japan; E-Mail: sahoo@nirs.go.jp

⁷ Department of Research Planning and Coordination, Japan NIOSH, Kawasaki, Japan; E-Mail: kimura@h.jniosh.go.jp

* Author to whom correspondence should be addressed; E-Mail: rakwal-68@aist.go.jp; Tel. +81-29-861-8508; Fax: +81-29-861-8508

Received: 5 February 2009; in revised form: 11 March 2009 / Accepted: 13 March 2009 /

Published: 16 March 2009

Abstract: We report molecular changes in leaves of rice plants (*Oryza sativa* L. - reference crop plant and grass model) exposed to ultra low-dose ionizing radiation, first using contaminated soil from the exclusion zone around Chernobyl reactor site. Results revealed induction of stress-related marker genes (Northern blot) and secondary metabolites (LC-MS/MS) in irradiated leaf segments over appropriate control. Second,

employing the same *in vitro* model system, we replicated results of the first experiment using in-house fabricated sources of ultra low-dose gamma (γ) rays and selected marker genes by RT-PCR. Results suggest the usefulness of the rice model in studying ultra low-dose radiation response/s.

Keywords: Contaminated soil; γ -ray; Marker genes; *Oryza sativa*; Phytoalexins.

1. Introduction

It is now over 20 years since the Chernobyl nuclear accident, and the resulting environmental impact of radiation on living organisms that followed is still being studied [1]. The initial 2800 km² radius exclusion zone around the Chernobyl reactor site where radioactive fallout heavily contaminated the environment was later modified and extended to cover 4,300 km². The aftereffects are still being felt in Southern Belarus, where the levels of cesium-137 (¹³⁷Cs) and strontium-90 (⁹⁰Sr) – both fission products from the reactor core – are and will remain high for decades to come. Other than human life directly or indirectly affected by radiation the plant life is also continuously affected, being unable to move out of the hot zone. The soil carries most of the radio-contamination, although use of fertilization and liming of the fields has been used to expel or bind the radionuclides. Nevertheless, a low to high level of contamination still exists in and around areas surrounding the Chernobyl nuclear reactor site.

With this background, the radioactively contaminated soil should provide us with a good source material for studying the effects of ionizing radiation (IR) on living organisms. Indeed, a single study in two genetically identical wheat populations revealed a remarkably strong induction of germline mutation upon chronic exposure to low-dose IR (0.3 Gy over a period of 10 months) from the Chernobyl accident [2]. Various physiological factors for the observed increased frequency of heterozygous structural variants in the offspring derived from exposed plants were considered before reaching to a conclusion that heterozygous variation was probably due to a radiation-induced increases in microsatellite mutation in the exposed plants. The results itself surprised Kovalchuk and co-workers, and we quote them – theoretically, this low-level exposure should not cause such a large increase in the mutation rate, suggesting that chronic exposure to IR has effects as yet unknown [2]. This one report highlighted the effect of low doses of IR in causing plant stress response. Therefore, the use of plants to study low- or ultra low-dose IR becomes a reality, and which is communicated in this report.

The plant model used was rice (*Oryza sativa* L.) – a grass plant, reference monocot plant, and potential human model. Why rice? In previous extensive studies, our and other research groups have been examining the effects of a wide variety of abiotic and biotic stresses in rice (for a review see [3]) primarily because its genome is sequenced. Moreover, rice is also a critical food crop for almost half the human population. With its sequenced genome rice has been termed the Rosetta Stone (by Dr. Robin Buell, a lead investigator for The Institute for Genomic Research, TIGR) and a cornerstone for cereal crop functional genomics by our group [4]. During the past 3 years, Rakwal and coworkers (at AIST, RLABB, NIRS, KURRI, HU and Japan NIOSH) have extended the on-going examination of the

stress responses against a wide variety of environmental stimuli to the relatively unstudied IR [including gamma (γ)-, X-rays, and neutrons, and heavy ions] in rice. For this the established two-week-old rice seedling *in vitro* model system [3,5,6] is being followed. In this study, ongoing in stages since the year 2005, we examined the effects of ultra low-dose radiation in rice leaves by observing the changes in selected marker genes and metabolites related to the defense/stress response pathways against radioactively contaminated Chernobyl soil (CCS), and γ -rays. Our present study has showed that ultra low-dose radiation elicits a defense/stress response in rice cells, which is a novel finding. More importantly, it suggests the possibility of using rice plant as a simple and good model for investigating the responses to IR, including the relatively uninvestigated ultra low-doses.

2. Experimental Section

2.1. Soil sample and gamma- and beta- ray measurements

The radioactive CCS was collected during an on-site visit to Masany (Belarus; the site is 10 Km from the Chernobyl nuclear reactor) by one of the co-authors, Dr. Shinzo Kimura, in November of 2002. The γ -ray measurement was performed on 20 g of contaminated soil before the decomposition of samples, using a Ge-detector (CANBERRA GC1318, Connecticut, USA; 1.9 keV half bandwidth), coupled to a multichannel analyzer (Laboratory Equipment Co., MCA-48F, Ibaraki, Japan) using standard procedures. During measurement, one channel was set to 0.5 keV so that measurement up to 4,000 keV was possible. The available ^{90}Sr (β -ray emission spectra) in 1 g of contaminated soil was measured by a low background beta-ray spectrometer (LBC-471Q, ALOKA Co. LTD, Tokyo Japan) according to standard procedures. For the in-house radiation experiment, the γ -ray exposure rate emitted from the fabricated disc shaped radiation sources was measured using glass fluorescence dosimeters (GD-301, Dose Ace FGD-1000, ALOKA) under the same conditions as the experiment using the same glass Petri dishes (Supplementary Figure 1).

2.2. Test plants and *in vitro* rice seedling system

Rice (*O. sativa* L. cv. Nipponbare) seedlings were grown for two-weeks (after germination) under white fluorescent light (wavelength 390-500 nm, $150 \text{ mmol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$, 12 h photoperiod) at 25°C and 70% relative humidity [5,6]. For treatments, the middle portions of fully expanded leaves (approximately 2 cm long segments cut with clean scissors) were placed on 25 mL MQTM water (Millipore, Waters) in clean Petri dishes, which were subsequently placed over 20 g CCS contained in the cover of the same Petri dish, and incubated in the growth cabinet under continuous light ($150 \text{ mmol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$). There was no direct connection between the soil and the leaf segments, which were separated by the water in the Petri dish. Leaf segments floated on MQ water and placed over uncontaminated soil served as an appropriate control (CON). Sixteen individual leaf segments were sampled and pooled (from two independent experiments performed in the year 2005) at the times indicated and immediately frozen in liquid nitrogen and stored at -80°C. Leaf segments (at 96 h) were also photographed with a digital camera. In the second experiment using in-house fabricated radiation doses of ultra-low γ -ray, rice plants were grown exactly as described above and leaf segments were

also prepared for irradiation in exactly the same way. Following irradiation, the leaf segments (ca. 16 per Petri dish) were pooled and immediately immersed in liquid nitrogen. A dummy experiment using cut leaf segments without γ -ray (control, C: $5.0 \pm 0.4 \mu\text{Gy}/3$ days) in the same growth cabinet as the treatments, along with leaves from whole plants (WP) kept in the experimental room for 3 days (background: $4.5 \pm 0.2 \mu\text{Gy}/3$ days) was also sampled. The positive control (WP) at 3 days was taken by cutting the rice leaves in exactly the same manner as on day 1, followed by immediate freezing in liquid nitrogen. For time-course experiment, another set of rice plants were cultivated as above, and then rice leaves were cut into segments and treated with a dummy (no radiation source) or ultra low-dose γ -ray for 1 to 72 h. These experiments (γ -irradiation) were performed four times at different periods of the year 2008 and in the beginning of 2009.

2.3. Total RNA extraction and Northern analysis

Total RNA isolation and Northern blot analysis was performed as described previously [5,6]. Briefly, deep frozen leaf segments were ground to a very fine powder in liquid nitrogen followed by extraction of total RNA with QIAGEN RNeasy Plant Maxi Kit (QIAGEN, Maryland, USA). The quality and equal quantity of the RNA was determined spectrophotometrically (NanoDrop, Wilmington, Delaware, USA) and visually confirmed using formaldehyde-agarose gel electrophoresis. Gene specific cDNA probes for rice: PR genes, *OsPR1b* (basic, accession number U89895), *OsPR5* (thaumatin-like protein, accession number X68197) and *OsPR10a* (accession number D38170); OB-related genes, *OsCATc* (catalase, accession number AB020502), *OsPOX* (peroxidase, accession number X66125) and *OsAPX1/2* (ascorbate peroxidase, accession numbers AB053297/BAB20889); SM-related genes, *OsPAL* (phenylalanine lyase, accession number X87946) and *OsCHS* (chalcone synthase, accession number X89859); P-related genes, *OsRbsL* (ribulose-1,5-bisphosphate carboxylase oxygenase, RuBisCO large subunit; [7]), *OsRbsS* (RuBisCO small subunit; [7]) and *OsCab* (chlorophyll *a/b* binding protein; [7]). Membranes were washed with $2 \times \text{SSC}$ and 0.1% (w/v) SDS at 65°C for 1 h, and exposed to an X-ray film (Kodak, Tokyo, Japan) using two intensifying screens for 2 d at -80°C . Rice *actin* gene was used as a control. Equal loading (20 μg) of total RNA was also confirmed by staining the blotted membrane with methylene blue (rRNA).

2.4. Semi-quantitative RT-PCR analysis

Total RNA was extracted as described above for subsequent RT-PCR analysis with eight selected “marker” genes based on the first experiment using CCS (see Figure 1). First-strand cDNA was synthesized in a 50 μL reaction mixture with a StartaScriptTM RT-PCR Kit (Stratagene) using 10 μg total RNA isolated from leaves of irradiated and control plants. Specific primers were designed from the 3'-UTR regions (forward and reverse primer sequences are provided in Supplementary Table 1) of each of the genes used in this study by comparison and alignment with all available related genes in the databases, NCBI and KOME (Knowledge-based Oryza Molecular biological Encyclopedia, <http://cdna01.dna.affrc.go.jp/cDNA/>). The RT-PCR was carried out exactly as described [8].

2.5. Phytoalexin estimation

Sakuranetin and momilactone A in leaves were analyzed and estimated using standards by a liquid chromatography tandem mass spectrometry technique as previously established and described [9].

3. Results and Discussion

3.1. Radioactive Chernobyl soil triggers defense/stress responses in rice leaves

Measurements of radioactivity revealed that the CCS emits radiation, which is due to the fact that it consists of long-lived radioisotopes ^{137}Cs and ^{90}Sr as fission products (Figure 1A). Measurements on CCS showed radiation level to be 5.34 $\mu\text{Gy/day}$ (Table 1). This dose is ca. 10.90-fold over the (shielded) control level indicating an extremely low (ultra) level of emitted radiation.

Table 1. The γ -ray measurement was performed on 20 g contaminated soil before the decomposition of samples, using a Ge-detector, coupled to a multichannel analyzer. The available ^{90}Sr (β -ray emission spectra) in 1 g contaminated soil was measured by a low background β -ray spectrometer. CCS: Contaminated Chernobyl Soil -5.34 $\mu\text{Gy/day}$; CON: Control Soil -0.49 $\mu\text{Gy/day}$.

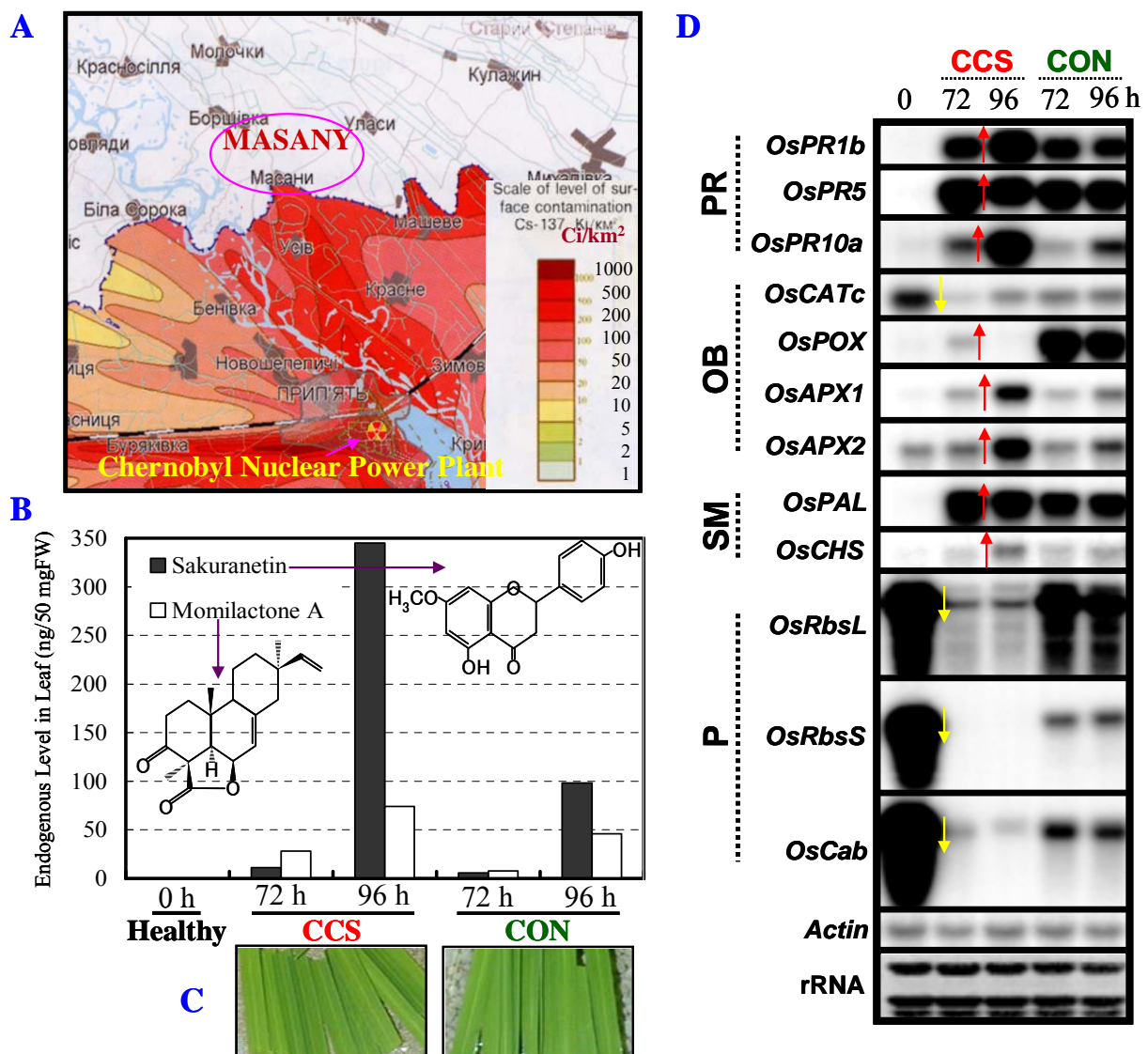
Radionuclide	Half-life	Energy (keV) [emission ratio]	Radioactivity [Bq/g-dry]
^{241}Am *1)	432.2 year	59.54 (35.9%)	0.67 ± 0.013
^{137}Cs *1)	30.07year	661.7 (85.1%)	67.9 ± 0.024
^{134}Cs *1)	2.065 year	604.7 (97.6%) 795.8 (85.5%)	0.16 ± 0.003 0.18 ± 0.002
^{90}Sr *2)	28.78 year	546 (100%)	3.18 ± 0.33

*1) Measurement by Gamma-ray spectrometer.

*2) Measurement by Beta-ray spectrometer.

Examination of rice stress-inducible molecular markers in the rice leaves exposed to CCS first revealed strong accumulation of two major rice phytoalexins (sakuranetin and momilactone A) at 72 and 96 h post-exposure (Figure 1B). The phytoalexins are prominent and established secondary metabolite (SM) markers commonly used to investigate stress responses in rice [reviewed in 3]. A slight increase in these phytoalexins in the control (CON) may be part of a stress response triggered by continuous light exposure. Nonetheless, a strong induction of phytoalexins is also seen upon pathogen attack and other abiotic stresses such as exposure to ozone [reviewed in 3], indicating a conservation in the induction of these secondary metabolites among numerous abiotic stresses, including to ultra low-dose IR. There also appeared to be a change in leaf color after exposure to CCS. At 96 h post-exposure leaf segments showed subtle change in color from dark green (CON) to light green (CCS), suggesting a possible damage to the photosynthetic apparatus (Figure 1C). In the case of ozone, such damage also correlated well with phytoalexins accumulation (reviewed in [3]).

Figure 1. The effect of Contaminated Chernobyl Soil (CCS) on rice leaf biology. (A) Soil was collected during an on-site visit to Masany. (B) Levels of sakuranetin and momilactone A increase in leaves at 72 and 96 h after exposure (5.34 μ Gy/day) to CCS over control (CON), compared to non-detection in healthy leaves (0 h) at the start of the experiment. (C) Photograph (Digital Camera, Sony Cyber-shot 5.0 Megapixels) of representative leaf segments 96 h after exposure to CCS. (D) Northern blot analysis reveals changes in mRNA level of marker genes at 72 and 96 h using gene specific cDNA probes for rice genes: *OsPR1b*, *OsPR5* and *OsPR10a*; *OsCATc*, *OsPOX* and *OsAPX1/2*; *OsPAL* and *OsCHS*;; *OsRbsL*, *OsRbsS* and *OsCab*. Leaf segments cut from the 3rd/4th leaves of two-week-old seedlings were used as the experimental material (see also Section 2.2 for details).



Secondly, northern blot analysis revealed distinct changes in the mRNA expression of specific marker genes in CCS over CON (Figure 1D). The induction (red arrows) or suppression (yellow arrows) of these genes is marked for clarity. Of note, these genes were selected based on our studies of the rice defense/stress responses (reviewed in [3]). These prominently included the pathogenesis-related (PR) genes *OsPR1b*, *OsPR5*, and *OsPR10a*. The oxidative burst (OB)- (*OsCATc*, *OsPOX*,

OsAPX1/2), SM- (*OsPAL* and *OsCHS*), and photosynthesis (P)- (*OsRbsL*, *OsRbsS*, and *OsCab*) related genes also showed changed expression profiles. A strong induction of the three PR gene transcripts in CCS over CON suggests radiation from CCS causes an activation of the defense/stress response pathway. On the other hand, a differential (induction or suppression) response of the antioxidant-related transcripts by CCS exposure was observed. The *OsCATc* transcript is strongly suppressed by CCS (at 72 h), whereas, the *OsAPX1* and 2 transcripts are strongly induced at 96 h over CON. On the other hand, the *OsPOX* transcript was weakly induced at 72 h in CCS over the potent induction of its transcript in CON at both times points. This result suggests that the genes responsible for antioxidant defense may be modulated by the cellular levels of reactive oxygen species (ROS) upon radiation. In future studies, the type of ROS and their levels will have to be measured and compared in irradiated and control samples to clarify this point. Moreover, as expected the P-related *OsRbsL*, *OsRbsS*, and *OsCab* transcripts, the best studied examples of plant nuclear encoded genes [10,11], were found to be strongly suppressed by exposure to CCS. This might relate to the observed loss of green color in irradiated leaves (Figure 1C). Damage to the photosynthesis may also lead to an unbalanced ROS level leading to the differential modulation of the OB-related gene expressions seen in Figure 1C. This Northern blot analysis data are also in good agreement with a preliminary DNA microarray analysis for rice leaf segments exposed to CCS, which showed large changes in gene expressions in defense/stress-related categories [12].

Differential expression of marker metabolites and genes in leaves upon CCS exposure suggest a subtle effect of ultra low-dose of radiation in rice cells. The study of Kovalchuk and co-workers shows the rate of mutation by long-term internal and external exposure from radiation in wheat [2]. However, we carried out a study limited to external short-term ultra low-dose irradiation exposure, in order to clarify the effect of radiation by observing changes of biological active metabolites and genes. Therefore, our rice *in vitro* model system is indicative of an active reaction triggered by ultra low-level radiation in the living cells.

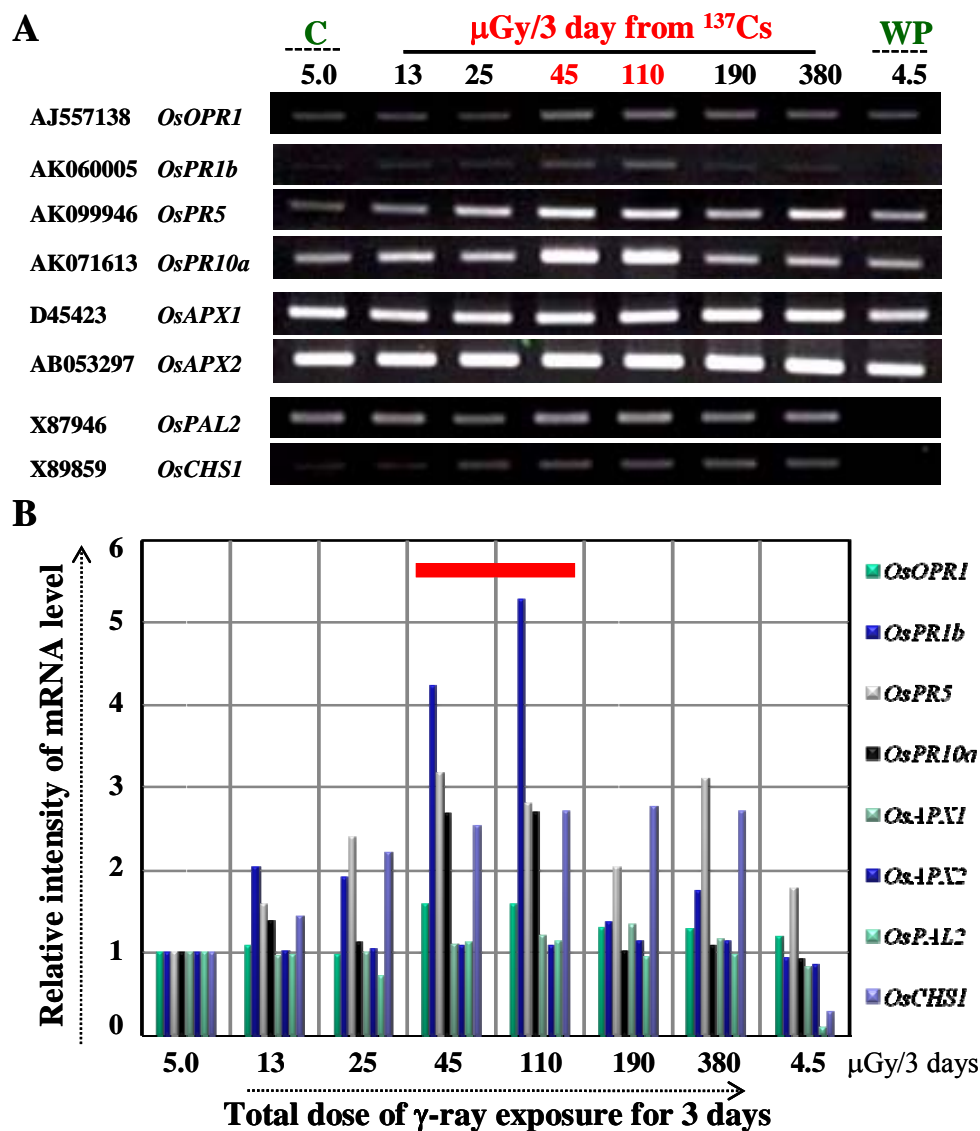
3.2. Ultra low-dose gamma (γ)-radiation also triggers the expression of defense/stress marker genes

As this striking observation detailed above (Figure 1) mainly due to γ -ray from CCS was never reported before, we further asked a simple question: whether ultra low-dose γ -radiation mimics the observed responses? To investigate this, we designed an in-house experiment where a disc-shaped γ -ray ^{137}Cs source was fabricated to irradiate leaf segments in a similar manner as done with CCS. The experiment was designed to perform six γ -radiation doses (13 ± 1 , 25 ± 2 , 45 ± 2 , 110 ± 10 , 190 ± 10 and 380 ± 20 $\mu\text{Gy}/3$ days) in parallel to appropriate control as illustrated and described in Supplementary Figure 1.

Using semi-quantitative RT-PCR, we found that ultra low-dose γ -radiation (2.6-times the background) activated the expression of classical marker genes at mRNA level for stress responses in plants associated with jasmonic acid biosynthesis (*OsOPR1*), and the defense/stress-related [PR (*OsPR1b*, *OsPR5*, *OsPR10a*), OB (*OsAPX1*, *OsAPX2*), and SM (*OsPAL2*, *OsCHS1*)] pathways (Figure 2A). The relative intensity of mRNA level is presented graphically for clarity (Figure 2B). Interestingly, we observed that the induction of these marker genes was dependent on the applied dose of ultra low-dose γ -ray, specifically and strikingly in the dose range of 45 to 110 $\mu\text{Gy}/3$ days (red bar

in Figure 2B). This result suggests a potential threshold for ultra low-dose in triggering a defense/stress response, which nevertheless remains to be further investigated. Nonetheless, these results indicate that ultra low-dose γ -radiation mimics the stress-related responses observed initially with CCS, where all these genes were dramatically changed at the mRNA level along with phytoalexins accumulation (see [12] and Figure 1).

Figure 2. Effects of ultra low-dose γ -radiation using in-house fabricated radiation source on mRNA expression level of marker genes in leaves of rice seedlings by RT-PCR. (A) Leaf segments were prepared as described in Figure 1, pooled together after treatment for total RNA isolation followed by quality check, cDNA synthesis, and RT-PCR analysis. The gene specific primers used are provided in Supplementary Table 1. Leaf segments were also sampled from dummy (C: control), and whole plants (WP: whole plant control) that served as a positive control for background radiation in the room where the experiment (in an incubator) was carried out. (B) The relative intensity of mRNA level of each transcript in (A) are presented. The intensities were calculated by the ATTO image analysis software (ATTO, Tokyo, Japan). Results are the representative of two independent experiments.



3.3. The *OsPR10a* is a potential candidate marker gene for radiation responses

From these two independent set of experimental results we found a candidate marker gene expression that showed the strongest dose-dependent response (*OsPR10a*, Figures 1D, 2, and 3, and [12]). Hence we propose that it could be utilized as a potential marker for ultra low-dose γ -radiation. For example, the *OsPR10a* promoter can be fused to fluorescent reporter genes to monitor the environmental radiation, either man-made or natural. Recently, it has been proposed that reference plants including wild grass (consisting of the well known cereal crops such as rice) should be considered as model test organisms in radiation studies by the International Commission on Radiological Protection [13]. Therefore, from the results presented here we can unravel the usefulness of a natural living organism (plant), in this case the grass model rice, for investigating ultra low-dose radiation responses. The only other report that comes close to this study is the effect of single whole-body X-radiation (acute dose) on clear induction of chromosomal inversion in the spleen and prostrate at ultra low-doses in a mammalian pKZ1 recombination mutagenesis mouse model [14].

4. Conclusions

The results arising from the present study using both natural CCS and in-house fabricated γ -ray sources provide evidence for ultra low-dose γ -radiation in causing perturbations in the multi-layered stress-associated biological processes in rice leaves. Further experiments are planned in the coming year to conduct high-throughput omic analysis post- γ -ray irradiation in both leaf segments (*in vitro*) and whole plants (*in planta/in vivo*) in short- and long-time exposures. This will help in a global profiling of radiation-induced molecular changes giving i) new candidate genes/proteins/metabolites for further study and ii) providing potential insight into the how ultra low-dose γ -ray triggers the defense/stress response. We wish to clarify one additional point in future studies. Do the molecular changes observed in leaves reflect a stress response leading to plant damage and/or death or are they indicative of an adaptation mechanism against the applied radiation? Pending these future studies, it is believed that the present study will lead to a new method to investigate radiation effects on gene: biological response to stimulation by ultra low-dose IR, which might also give some clue to the problem of by-stander effect of radiation. We also hope to rekindle the debate on understanding biological effects of ultra-, low-, and high-level radiation in living organisms, including our stationary life-sustaining “distant cousins” – the plants. It may be envisioned that rice will serve as a human model for investigating the responses to IR, due to conservation of genes/gene families in living organisms.

Acknowledgements

We appreciate the technical support during the first phase CCS experiment of the late Mr. Satoru Ebisawa (NIRS), who passed away due to cancer. The survey of Chernobyl was made possible by kind support from Dr. Kenzo Fujimoto (NIRS). The authors appreciate intensive discussions on the study from Drs. Hiroaki Koide (KURRI) and Kouya Ogino (Environmental Microwave Research Institute, Kyoto, Japan).

References

1. Stone, R. Return to the inferno: Chernobyl after 20 years. *Science* **2006**, *312*, 180-182.
2. Kovalchuk, O.; Dubrova, Y.E.; Arkhipov, A.; Hohn, B.; Kovalchuk, I. Wheat mutation rate after Chernobyl. *Nature* **2000**, *407*, 583-584.
3. Jwa, N.S.; Agrawal, G.K.; Tamogami, S.; Yonekura, M.; Han, O.; Iwahashi, H.; Rakwal, R. Role of defense/stress-related marker genes, proteins and secondary metabolites in defining rice self-defense mechanisms. *Plant Physiol. Biochem.* **2006**, *44*, 261-273.
4. Agrawal, G.K.; Rakwal, R. Rice proteomics: A cornerstone for cereal food crop proteomes. *Mass Spec. Rev.* **2006**, *25*, 1-53.
5. Agrawal, G.K.; Rakwal, R.; Jwa, N.S.; Agrawal, V.P. Signaling molecules and blast pathogen attack activates rice *OsPR1a* and *OsPR1b* genes: A model illustrating components participating during defense/stress response. *Plant Physiol. Biochem.* **2001**, *39*, 1095-1103.
6. Agrawal, G.K.; Jwa, N.S.; Iwahashi, H.; Rakwal, R. Importance of ascorbate peroxidases *OsAPX1* and *OsAPX2* in the rice pathogen response pathways and growth and reproduction revealed by their transcriptional profiling. *Gene* **2003**, *322*, 93-103.
7. Kusumi, K.; Komori, H.; Satoh, H.; Iba, K. Characterization of a zebra mutant of rice with increased susceptibility to light stress. *Plant Cell Phys.* **2000**, *41*, 158-164.
8. Cho, K.; Shibato, J.; Agrawal, G.K.; Jung, Y.H.; Kubo, A.; Jwa, N.S.; Tamogami, S.; Satoh, K.; Kikuchi, S.; Higashi, T.; Kimura, S.; Saji, H.; Tanaka, Y.; Iwahashi, H.; Masuo, Y.; Rakwal, R. Integrated transcriptomics, proteomics and metabolomics analyses to survey ozone responses in the leaves of rice seedling. *J. Proteome Res.* **2008**, *7*, 2980-2998.
9. Tamogami, S.; Rakwal, R.; Kodama, O. Phytoalexin production by amino acid conjugates of jasmonic acid through induction of naringenin-7-O-methyltransferase, a key enzyme on phytoalexin biosynthesis in rice (*Oryza sativa* L.). *FEBS Lett.* **1997**, *401*, 239-242.
10. Tobin, E.M.; Silverthorne, J. Light regulation of gene expression in higher plants. *Annu. Rev. Plant Physiol.* **1985**, *36*, 569-593.
11. Dean, C.; Pichersky, E.; Dunsmuir, P. Structure, evolution and regulation of RbcS genes in higher plants. *Annu. Rev. Plant Physiol.* **1989**, *40*, 415-439.
12. Kimura, S.; Shibato, J.; Agrawal, G.K.; Kim, Y.K.; Nahm, B.H.; Jwa, N.S.; Iwahashi, H.; Rakwal, R. Microarray analysis of rice leaf response to radioactivity from contaminated Chernobyl soil. *Rice Gen. Newsl.* **2008**, *24*, 52-54.
13. *Annals of the ICRP*; ICRP: Oxford, UK, 2007; No. 103, pp. 1-135.
14. Hooker, A.M.; Bhat, M.; Day, T.K.; Lane, J.M.; Swinburne, S.J.; Morley, A.A.; Sykes, P.J. The linear no-threshold model does not hold for low dose ionizing radiation. *Rad. Res.* **2004**, *162*, 447-452.