



Article Rapid Assembly of Pyrrole-Ligated 1,3,4-Oxadiazoles and Excellent Antibacterial Activity of Iodophenol Substituents

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Abstract: Pyrrole-ligated 1,3,4-oxadiazole is a very important pharmacophore which exhibits broad therapeutic effects such as anti-tuberculosis, anti-epileptic, anti-HIV, anti-cancer, anti-inflammatory, antioxidant, and antibacterial activities. A one-pot Maillard reaction between D-Ribose and an L-amino methyl ester in DMSO with oxalic acid at 2.5 atm and 80 °C expeditiously produced pyrrole-2-carbaldehyde platform chemicals in reasonable yields, which were utilized for the synthesis of pyrrole-ligated 1,3,4-oxadiazoles. Benzohydrazide reacted with the formyl group of the pyrrole platforms to provide the corresponding imine intermediates, which underwent I₂-mediated oxidative cyclization to the pyrrole-ligated 1,3,4-oxadiazole skeleton. The structure and activity relationship (SAR) of the target compounds with varying alkyl or aryl substituents of the amino acids and electron-withdrawing or electron-donating substituents on the phenyl ring of benzohydrazide were evaluated for antibacterial activity against *Escherichia coli*, *Staphylococcus aureus*, and *Acinetobacter baumannii* as representative Gram(–) and Gram(+) bacteria. Branched alkyl groups from the amino acid showed better antibacterial activities. Absolutely superior activities were observed for **5f-1** with an iodophenol substituent against *A. baumannii* (MIC < 2 μ g/mL), a bacterial pathogen that displays a high resistance to commonly used antibiotics.

Keywords: pyrrole; 1,3,4-oxadiazole; Maillard reaction; D-ribose; L-amino acid; pyrrole-2-carbaldehyde; iodine effect; antibacterial activity

1. Introduction

Oxadiazole is a five-membered heterocyclic aromatic compound composed of four structural isomers depending on the positions of two nitrogen atoms relative to an oxygen atom [1]. Among them, 1,3,4-oxadiazole has received intensive attention in the field of medicinal chemistry due to its broad metabolic profile [2–4] and in the field of material science for its excellent optoelectronic properties [5–8]. As an isostere of an amide and an ester, 1,3,4-oxadiazole serves as a promising pharmacophore for the discovery of new drugs exhibiting antimicrobial, anticonvulsant, anti-inflammatory, analgesic, antitumor, antiviral, antihypertensive, and enzyme inhibitory activities [9]. There have been extensive literature reviews on the specific synthetic methods and diverse biological activities of 1,3,4-oxadiazole derivatives [10–12].

Motivated by Raltegravir [13,14], an antiretroviral drug used to treat HIV/AIDS, and Zibotentan [15,16], an anti-cancer drug candidate, a number of poly heterocyclic



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). compounds containing a 1,3,4-oxadiazole pharmacophore were constructed, and their biological activities were investigated. Indole-ligated 1,3,4-oxadiazoles [A] were synthesized and evaluated to show antimicrobial, anti-inflammatory, and antiproliferative activities (Figure 1) [17,18]. Their biological importance was also identified by their antioxidants and acetylcholinesterase inhibition properties [19]. Likewise, various 2-benzofuranyl-1,3,4-oxadiazoles [B] were synthesized [20,21], demonstrating their biological activities in α -Glucosidase inhibition as well as the inhibition of glycogen synthase kinase 3 β for treating diabetes and Alzheimer's disease, respectively [22–24].



Figure 1. Heterocycle-ligated 1,3,4-oxadiazoles [A] and [B], and retrosynthesis of **1** from pyrrole **2** through the cyclization using key precursors [C] and [D].

Pyrrole is a very important structural motif in drug discovery projects because of the wide presence of natural, biologically active pyrrole alkaloid products [25,26]; thus, pyrrole-ligated 1,3,4-oxadiazole would be a perfect base structure for the development of potential lead compounds [27]. Considering the efficacy of the procedures of constructing a 1,3,4-oxadiazole ring, 2-pyrrolyl-5-phenyl-1,3,4-oxadizole **1** would be an ideal core structure, achieved either through dehydration from aroylhydrazide [C] or by cyclization from aroylhydrazone [D]. In the benzene ring, the substituent effects of the core structure **1** on antibacterial activity have been reported for the cases of 4,5-dibromopyrrole [28,29] and 4-nitropyrrole [30], respectively.

Pyrrole-2-carbaldehydes **2**, derived from the conversion of D-ribose with L-amino acids [31], were demonstrated to be useful, sustainable platform chemicals for the construction of highly functionalized poly heterocyclic compounds [32]. Since natural amino acids themselves demonstrate specific biological activities [33–35], it was envisioned that 1,3,4-oxadiazoles **1** from pyrrole-2-carbaldehydes **2** with the *N*-amino acid moiety would be very interesting core structures for the investigation of their biological activities. The effect of the amino acid moiety of **1** on antimicrobial activities was screened first, and the substituent effects on the benzene ring were then investigated for **5** and **6** with some selected amino acid moieties. We found a marginal size effect of the alkyl groups from amino acids (Val and Ile, etc.) and superior antibacterial activity of the iodophenol substituents of pyrrole-ligated 1,3,4-oxadiazoles **5** and **6** against *S. aureus* and *A. baumannii*. All the syntheses of pyrrole-ligated 1,3,4-oxadiazoles **1**, **5**, and **6** and their antibacterial activities are closely described herein.

2. Results and Discussion

Structure and activity relationships (SARs) for pyrrole-ligated 1,3,4-oxadiazoles **1** were generally studied by changing substituent groups in the aromatic rings [28–30]. We were interested in the SAR of **1** by *N*-alkyl substituents because pyrrole-2-carbaldehydes **2**, the starting materials for 1,3,4-oxadiazoles **1**, are easily prepared from L-amino acids, and each amino acid has its own biological activity. Ten L-amino acids with hydrogen, alkyl, aralkyl, ester, and sulfide substituents R were selected to assess the size effect (linear or branched) or potential electronic effect. Pyrrole-2-carbaldehydes **2** were efficiently prepared by a one-pot ribose conversion with an L-amino methyl ester in the presence of oxalic acid in DMSO,

following an improved procedure under 2.5 atm argon at 80 °C [32]. The corresponding pyrrole platform chemicals 2a-2j with the *N*-amino acid moiety were prepared in yields of 32~63% (Scheme 1 and Table 1).



Scheme 1. Preparations of pyrrole-2-carbaldehydes **2**, *N*-benzoylhydrazones **4**, and 1,3,4-oxadiazoles **1** from D-ribose conversion with L-amino acids.

Table 1. Yields of pyrrole-2-carbaldehydes 2, N-benzoylhydrazones 4, and 1,3,4-oxadiazoles 1 from



Entry	Amino Acid (R)	Yield 2 (%) ¹	Yield 4 (%) ²	Yield 1 (%) ²
а	Gly (H)	32	96	80
b	Ala (Me)	38	88	91
с	Val (<i>i</i> -Pr)	42	83	98
d	Leu (i-Bu)	63	77	87
e	Ile (s-Bu)	40	86	90
f	Phe (PhCH ₂)	54	83	86
g	Bn (PhCH ₂ CH ₂)	53	79	86
h	AsP (MeO ₂ CCH ₂)	47	70	88
i	Glu (MeO ₂ CCH ₂ CH ₂)	37	81	95
j	Met (MeSCH ₂ CH ₂)	46	82	50 ³

D-ribose conversion with L-amino acids in Scheme 1.

 $\overline{1}$ Yields from one-pot reaction [32]. $\overline{2}$ Isolated yields after SiO₂ flash column chromatography. $\overline{3}$ Cyclized product **1k** was also obtained in 49% yield.

Two representative procedures are generally utilized for the construction of the 1,3,4oxadizole core, as depicted in Figure 1 [12]. The cyclodehydration route from diacylhydrazine [C] is suitable for pyrrole-2-carboxylic acids [36], whereas the oxidative cyclization route from N-acylhydrazone [D] is widely used for pyrrole-2-carbaldehydes as starting materials [37]. There were various cyclization conditions for 1,3,4-oxadiazoles reported for each conversion [12]. We adopted the oxidative cyclization route of N-acylhydrazones 4, which can be obtained from pyrrole-2-carbaldehydes 2 by condensation with benzohydrazide **3a**. The corresponding *N*-benzoylhydrazones **4a–4j** were obtained in decent yields (70~96%) at the reflux temperature of toluene. Oxidative cyclization conditions were then screened using NBS, NIS, and I₂ under K₂CO₃, DBU, Et₃N, and NaOH as a base. The condition using NIS/NaOH in DMSO at 100 °C was optimal for providing pyrrole-ligated 1,3,4-oxadiazoles 1 in yields of 80~98%. It is noteworthy that the NIS-mediated further cyclization of the methylsulfide chain on the pyrrole ring occurred partly for 1j derived from methionine to produce 1k (at a yield of 49%), which explains the lower yield of 1j (50%). All eleven pyrrole-ligated oxadiazoles **1a–1j** were rapidly assembled from pyrrole platform chemicals 2 with different amino acid residues and ready for antibacterial assays against Escherichia coli and Staphylococcus aureus as two representative Gram(-) and Gram(+) bacteria (Table 2).

Entry	Compound	Amino Acid (R)	MIC/E. <i>coli</i> (µg/mL)	MIC/S. aureus (µg/mL)
1	1a	Gly (H)	>2048	2048
2	1b	Ala (Me)	512	512
3	1c	Val (<i>i</i> -Pr)	256	128
4	1d	Leu (i-Bu)	512	128
5	1e	Ile (s-Bu)	256	64
6	1f	Phe (PhCH ₂)	1024	512
7	1g	Bn (PhCH ₂ CH ₂)	1024	512
8	1h	Asp (MeO_2CCH_2)	2048	1024
9	1i	Glu (MeO ₂ CCH ₂ CH ₂)	2048	2048
10	1j	Met (MeSCH ₂ CH ₂)	>2048	2048
11	2k	c-Met (SCH ₂ CH ₂)	256	256

Table 2. Minimum inhibition activity (MIC) of 2-pyrrolyl-5-phenyl-1,3,4-oxadiazoles **1** against *E. coli* and *S. aureus*.

There was a definite size effect of the alkyl substituent R on antibacterial activity in 1,3,4-oxadiazoles **1**. The highest MIC value was required for **1a** from glycine (R = H), and it decreased as the size (branch) of the alkyl group increased from alanine (R = Me) to isoleucine (R = *s*-Bu) (entries 1–5, Table 2). A benzene ring seemed be unimportant, judging from the cases of the benzyl and homobenzyl substituents (entries 6–7). There was no functional group effect for the ester and sulfide, reflecting a lack of electronic interactions between the substituent R and the bacterial enzymes. A comparison of the MIC values for **1** and its cyclized derivative **1k** confirmed the importance of the size (or rigidity) effect of R on antibacterial activity (entries 10 and 11). An additional point to mention is that the MIC values for **1** were not much different between Gram(–) and Gram(+) bacteria, indicating that there would be no transport barriers through membranes for these small molecules.

The electronic effects of the substituents on the phenyl ring against antibacterial activities were then investigated for 2-pyrrolyl-5-phenyl-1,3,4-oxadiazoles **5** and **6** with the maximum size effect in the series, derived from valine (R = isopropyl) and isoleucine (R = *sec*-butyl), respectively. Commercial benzohydrazides **3** with a substituent X of a different electronic nature (e.g., F, Cl, OH, and OMe) were utilized in the synthesis of **5** and **6** (Scheme 2 and Table 3). The condensation reaction of pyrrole-2-carbaldehyde **2c** (R = *i*-Pr) and **2e** (R = *s*-Bu) with various benzohydrazides **3** produced the corresponding *N*-benzoylhydrazone intermediates in refluxing toluene, which underwent an oxidative cyclization reaction (without purification) to afford 2-pyrrolyl-5-phenyl-1,3,4-oxadiazoles **5** (R = *i*-Pr) and **6** (R = *s*-Bu) with various electronic substituents X on the phenyl ring.



Scheme 2. Two-step preparation of various 1,3,4-oxadiazoles **5** and **6** with various electronic substituents X, derived from valine and isoleucine.

A milder oxidative cyclization condition was required in these cases, for which I_2/K_2CO_3 in 1,4-dioxane at 85 °C was optimal for the production of **5** in yields of 40~85% and **6** in yields of 62~89% in two steps [38]. Serendipitously, we found extra iodination reactions under the oxidative cyclization conditions for 1,3,4-oxadiazoles **5e**, **5f**, **6e**, and **6f** with phenol substituents, obtaining the corresponding di-iodination or tetra-iodination products **5e-1**, **5f-1**, **6e-1**, and **6f-1**, respectively in yields of 32~45% (Figure 2). It was very fortunate to achieve further iodination on the phenol rings so that we were able to find the superior Iodophenol antibacterial effects for these oxadiazole derivatives (vide infra). The corresponding deiodination (originally intended) products were prepared by reduction

using Zn dust in AcOH to produce **5f** (X = p-OH) in a yield of 97%, **6e** (X = p-OH) in a yield of 50%, and **6f** (X = p-OH) in a yield of 42%.



Figure 2. Extra iodination products from 2-pyrrolyl-5-phenyl-1,3,4-oxadiazoles 5 and 6.

Table 3. Yields of various 1,3,4-oxadiazoles **5** and **6** with various electronic substituents X, derived from valine and isoleucine in Scheme 2.

Entry	X	Yield 5 (%)	Yield 6 (%)
а	<i>o</i> -F	40	89
b	<i>m</i> -F	70	88
с	p-F	54	62
d	p-Cl	62	79
e-1	<i>o</i> -OH (I)	33 ¹	45 ²
e	o-OH	-	50 ³
f-1	<i>p-</i> OH (I)	32 ⁴	43 ⁵
f	p-OH	97 ⁶	42 ⁷
g	o-OMe	85	80
g-1	o-OMe	61 ⁸	-
ĥ	<i>p</i> -OMe	70	79
h-1	<i>p</i> -OMe	69 ⁹	83 ¹⁰

The structures of ¹ **5e-1**, ² **6e-1**, ⁴ **5f-1**, ⁵ **6f-1**. ⁸ **5g-1**, ⁹ **5h-1**, and ¹⁰ **6h-1** are depicted in Figure 2, which were produced by extra iodination reactions. The compounds ³ **6e**, ⁶ **5f**, and ⁷ **6f** were prepared by deiodination using Zn in AcOH at 25 °C from **6e-1**, **5f-1**, and **6f-1**, respectively. The compounds ⁸ **5g-1**, ⁹ **5h-1**, and ¹⁰ **6h-1** were prepared by separate iodination reaction (I₂ in DMSO) at 85 °C for 6 h from **5g**, **5h**, and **6h**, respectively.

To assess the "iodine effect" on antibacterial activity, separate iodination reactions (I₂ in DMSO at 85 °C) were intentionally carried out for the 1,3,4-oxadiazoles **5g**, **5h**, and **6h** with the electron-rich anisole substituent in which 3-mono-iodination or 3,4-diiodination reactions proceeded on the pyrrole ring (not on the anisole ring) to provide the corresponding **5g-1** (X = o-OMe) in a yield of 61%, **5h-1** (X = p-OMe) in a yield of 69% as a 1.3:1 mixture of di- and 3'-mono-iodination products, and **6h-1** (X = p-OMe) in a yield of 83%, respectively. All twenty-two pyrrole-ligated oxadiazoles **5** and **6** were rapidly assembled from pyrrole-2-carbaldehydes **2c** (R = i-Pr) and **2e** (R = s-Bu) with diversely X-substituted benzohydrazide **3** and ready for antibacterial assays against *Escherichia coli, Staphylococcus aureus*, and *Acinetobacter baumannii* together with Vancomycin and Erythromycin as positive controls (Table 4).

SARs of the phenyl substituent X of 2-pyrrolyl-5-phenyl-1,3,4-oxadiazoles **5** and **6** can be deduced from the MIC (μ g/mL) in Table 4. *ortho*-F substitutions provided generally better antibacterial activities than the *meta-* and *para*-F counterparts (entries 1–3 and 12–14), and chloride was better than fluoride (entries 4 and 15 versus 3 and 14). Iodophenol substituents exhibited superior antibacterial activities against *A. baumannii* and *S. aureus* regardless of the position of the hydroxyl substituent (entries 5, 6, 16, and 18). The MIC values of <2 μ g/mL for **5f-1** and 8 μ g/mL for **6f-1** against *A. baumannii* were much lower than those of the positive controls (>1024 μ g/mL for vancomycin and 128 μ g/mL for erythromycin). The "iodophenol effect" on antibacterial activity is obvious when compared with the cases

of deiodination products **5f** and **6f**, the MIC values for which were significantly increased to 128 μ g/mL and 512 μ g/mL, respectively (entries 7 and 19). The mechanism of the "iodophenol effect" is not clear at present, but it is reasonable to explain that iodide or molecular I₂ may be liberated by the neighboring OH group [39]. No effects or only slight improvements in antibacterial activities were observed for the pyrrole iodination products **5g-1** and **5h-1** from **5g** and **5h** (entries 8–11), whereas the reverse effect was clear for the pyrrole iodination product **6h-1** from **6h** (entries 21 and 22).

Table 4. Minimum inhibition concentration (MIC) of 1,3,4-oxadiazoles **5** and **6** against *E. coli, S. aureus,* and *A. baumannii.*

Entry	Comp'd	x	E. coli (μg/mL)	S. aureus (μg/mL)	A. baumannii (μg/mL)
1	5a	<i>o</i> -F	512	256	128
2	5b	<i>m</i> -F	>1024	>1024	512
3	5c	<i>p</i> -F	>1024	>1024	256
4	5d	p-Cl	512	256	512
5	5e-1	<i>o</i> -OH (I) ¹	1024	4	64
6	5f-1	<i>p</i> -OH (I) ¹	1024	4	<2
7	5f	p-OH	>1024	128	128
8	5g	o-OMe	>1024	512	256
9	5g-1	<i>o</i> -OMe (I) ²	512	512	256
10	5h	<i>p</i> -OMe	>1024	512	256
11	5h-1	<i>p</i> -OMe (I) ²	512	256	128
12	6a	<i>o</i> -F	512	256	256
13	6b	<i>m</i> -F	1024	256	256
14	6c	<i>p-</i> F	>1024	512	512
15	6d	p-Cl	512	512	256
16	6e-1	<i>o</i> -OH (I) ¹	1024	<2	512
17	6e	o-OH	256	64	64
18	6f-1	<i>p</i> -OH (I) ¹	1024	<2	8
19	6f	<i>p</i> -OH	1024	8	512
20	6g	o-OMe	>1024	512	256
21	6h	<i>p</i> -OMe	>1024	256	256
22	6h-1	<i>p</i> -OMe (I) ²	1024	512	512
23	Vancomycin	-	>1024	4	>1024
24	Erythromycin	-	1024	4	128

 $\overline{1}$ Extra iodination product on the phenol ring. ² Extra iodination product on the pyrrole ring. The structures of extra iodination products are depicted in Figure 2.

3. Materials and Methods

3.1. Experimental

3.1.1. General Chemical Syntheses

¹H- and ¹³C-NMR spectra were recorded on 400 MHz and 100 MHz NMR spectrometers, respectively, in a deuterated solvent (notified in parenthesis) with tetramethylsilane (TMS) as an internal reference. The column chromatography was performed using the method of Still with silica gel 60 and a 70–230 mesh ASTM, using a gradient mixture of EtOAc/hexanes. Reactions were performed in a well-dried flask under an argon atmosphere unless mentioned otherwise.

3.1.2. General Procedure for the Preparation of 1

Formation of Hydrazone 4 from pyrrole-2-carbaldehyde (Step-1): The solution of pyrraline **2** (~1.00 g, 1 equiv.) and benzohydrazide **3a** (1 equiv.) in toluene (10 mL) was heated at 110 °C for 6 h. The reaction mixture was cooled to room temperature, diluted with EtOAc, washed with brine and H₂O, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was purified by SiO₂ flash column chromatography to obtain the corresponding benzohydrazone **4**.

2-pyrrolyl-5-phenyl-1,3,4-oxadiazole 1 from hydrazone 4 (Step-2): The mixture of benzohydrazone 4 (1 equiv.), NaOH (2 equiv.), and N-iodosuccinimide (1 equiv.) in DMSO (10 mL) was heated at 110 °C for 2 h. The reaction mixture was cooled to room temperature, diluted with EtOAc, washed with brine and H₂O, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The crude product was purified by SiO₂ flash column chromatography to obtain the corresponding 2-pyrrolyl-5-phenyl-1,3,4-oxadiazole 1.

Methyl 2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*acetate* (**1a**). Data for **4a**: white solid in a yield of 96% (1.64 g, 5.76 mmol); ¹H-NMR (DMSO-d₆) δ = 3.69 (s, 3H), 5.21 (s, 2H), 6.16 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.53 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.02 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.48–7.54 (m, 2H), 7.54–7.60 (m, 1H), 7.86–7.90 (m, 2H), 8.28 (s, 1H), 11.49 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 51.1, 53.1, 109.9, 117.2, 128.4, 128.7, 129.7, 129.9, 132.7, 134.9, 142.0, 163.7, 170.7 ppm; IR (CH₂Cl₂) ν = 3237, 3063, 3006, 2954, 2848, 1750, 1649, 1616, 1552, 1494, 1468, 1433, 1345, 1322, 1279, 1216, 1188, 1140, 1086, 1031, 1001, 914, 801, 754, 690 cm⁻¹.

Data for **1a**: light-yellow solid in a yield of 80% (0.60 g, 2.13 mmol); ¹H-NMR (acetoned₆) δ = 3.73 (s, 3H), 5.39 (s, 2H), 6.32 (dd, *J* = 4.0, 2.4 Hz, 1H), 7.06 (dd, *J* = 4.0, 1.6 Hz, 1H), 7.18 (dd, *J* = 2.4, 1.6 Hz, 1H), 7.57–7.65 (m, 3H), 8.08–8.14 (m, 2H) ppm; ¹³C-NMR (acetone-d₆) δ = 50.1, 51.6, 109.2, 114.3, 117.5, 124.0, 126.5, 129.1, 129.2, 131.6, 159.1, 162.4, 168.9 ppm; IR ν = 3119, 3069, 2957, 2927, 2857, 1752, 1713, 1614, 1556, 1501, 1490, 1455, 1424, 1401, 1376, 1328, 1304, 1297, 1264, 1213, 1106, 1081, 995, 956, 787, 773, 760, 725, 693, 583 cm⁻¹; HRMS (ESI) calcd for C₁₅H₁₃N₃O₃+Na, 306.0849: found 306.0851.

Methyl (*S*)-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*propanoate* (**1b**). Data for **4b**: white solid in a yield of 88% (1.45 g, 4.85 mmol); ¹H-NMR (DMSO-d₆) δ = 1.70 (d, *J* = 7.2 Hz, 3H), 3.67 (s, 3H), 6.02 (q, *J* = 7.2 Hz, 1H), 6.20 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.54 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.17 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.48–7.61 (m, 3H), 7.86–7.92 (m, 2H), 8.32 (s, 1H), 11.51 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 19.1, 53.4, 56.1, 110.2, 117.5, 126.6, 128.2, 128.7, 129.6, 132.7, 134.9, 142.4, 163.7, 172.8 ppm; IR ν = 3232, 3065, 2954, 1748, 1716, 1644, 1612, 1556, 1494, 1461, 1426, 1359, 1286, 1225, 1146, 1091, 1063, 1031, 962, 910, 887, 857, 802, 714, 695 cm⁻¹; HRMS (ESI) calcd for C₁₆H₁₇N₃O₃+Na, 322.1162: found 322.1164.

Data for **1b**: light-yellow solid in a yield of 91% (0.70 g, 2.35 mmol); ¹H-NMR (CD₃OD) δ = 1.82 (d, *J* = 7.2 Hz, 3H), 3.71 (s, 3H), 6.00 (q, *J* = 7.2 Hz, 1H), 6.33 (dd, *J* = 4.0, 2.8 Hz, 1H), 7.02 (dd, *J* = 4.0, 1.6 Hz, 1H), 7.23 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.50–7.59 (m, 3H), 8.00–8.04 (m, 2H) ppm; ¹³C-NMR (CD₃OD) δ = 19.5, 54.5, 58.5, 112.2, 117.6, 119.7, 126.0, 128.1, 129.1, 131.7, 134.4, 162.1, 165.5, 174.7 ppm; IR ν = 3129, 3003, 2955, 2848, 1751, 1663, 1608, 1553, 1505, 1450, 1384, 1336, 1289, 1224, 1203, 1110, 1091, 1062, 1015, 981, 962, 853, 813, 776, 729, 691, 610 cm⁻¹; HRMS (ESI) calcd for C₁₆H₁₅N₃O₃+Na, 320.1006: found 320.1007.

Methyl (*S*)-3-*methyl*-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*butanoate* (**1c**). Data for **4c**: white solid in a yield of 83% (1.30 g, 3.98 mmol); ¹H-NMR (DMSO-d₆) δ = 1.03 (d, *J* = 6.8 Hz, 3H), 1.35 (d, *J* = 6.8 Hz, 3H), 2.76 (m, 1H), 4.04 (s, 3H), 6.35 (d, *J* = 8.8 Hz, 1H), 6.57 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.87 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.51 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.82–7.97 (m, 3H), 8.21–8.30 (m, 2H), 8.73 (s, 1H), 11.90 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 17.6, 18.4, 31.3, 51.4, 62.8, 108.8, 114.6, 124.7, 126.6, 126.7, 127.6, 130.7, 132.8, 140.3, 161.7, 169.9 ppm; IR ν = 3241, 3071, 2975, 2878, 1750, 1644, 1615, 1557, 1495, 1459, 1433, 1392, 1356, 1286, 1211, 1160, 1133, 1083, 1057, 1023, 1005, 952, 915, 890, 835, 800, 755, 715, 695, 617 cm⁻¹.

Data for **1c**: light-yellow solid in a yield of 98% (0.15 g, 0.46 mmol); ¹H-NMR (CD₃OD) $\delta = 0.81$ (d, J = 7.2 Hz, 3H), 1.08 (d, J = 7.2 Hz, 3H), 2.53 (m, 1H), 3.76 (s, 3H), 5.99 (d, J = 10.0 Hz, 1H), 6.38 (dd, J = 4.0, 2.8 Hz, 1H), 7.03 (dd, J = 4.0, 1.6 Hz, 1H), 7.36 (dd, J = 2.8, 1.6 Hz, 1H), 7.54–7.64 (m, 3H), 8.06–8.12 (m, 2H) ppm; ¹³C-NMR (CD₃OD) $\delta = 20.4, 21.1$, 35.2, 54.1, 67.1, 112.3, 119.0, 128.5, 130.0, 130.5, 131.1, 134.4, 136.0, 145.0, 167.8, 174.2 ppm; IR $\nu = 2958, 2927, 2851, 1744, 1669, 1609, 1560, 1500, 1454, 1376, 1260, 1221, 1105, 1076, 1013, 775, 727, 691 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₉N₃O₃+Na, 348.1319: found 348.1319.$

Methyl (*S*)-4-*methyl*-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*pentanoate* (**1d**). Data for **4d**: white solid in a yield of 77% (1.18 g, 3.46 mmol); ¹H-NMR (DMSO-d₆) δ = 0.86 (d, *J* = 6.4 Hz, 3H), 0.92 (d, *J* = 6.4 Hz, 3H), 1.92 (dd of A of ABq, *J*_{AB} = 14.0, *J*_d = 9.2, 4.8 Hz, 1H), 2.12 (dd of B of ABq, *J*_{AB} = 14.0, *J*_d = 11.6, 4.4 Hz, 1H), 3.67 (s, 3H), 6.21 (dd, *J* = 3.6,

2.8 Hz, 1H), 6.30 (dd, J = 9.2, 4.4 Hz, 1H), 6.52 (dd, J = 3.6, 1.6 Hz, 1H), 7.19 (dd, J = 2.8, 1.6 Hz, 1H), 7.48–7.62 (m, 3H), 7.86–7.92 (m, 2H), 8.34 (s, 1H), 11.52 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 22.6, 24.1, 25.7, 42.1, 53.5, 58.5, 110.6, 117.6, 127.2, 128.4, 128.7, 129.6, 132.7, 134.8, 142.5, 163.6, 172.8 ppm; IR ν = 3234, 3065, 2956, 2874, 1743, 1646, 1614, 1556, 1495, 1459, 1427, 1348, 1278, 1240, 1203, 1174, 1086, 1034, 998, 953, 928, 903, 795, 754 cm⁻¹.

Data for **1d**: light-yellow solid in 87% yield (1.35g, 4.01 mmol); ¹H-NMR (acetone-d₆) $\delta = 0.94$ (d, J = 6.4 Hz, 3H), 0.95 (d, J = 6.4 Hz, 3H), 1.49 (m, 1H), 2.02–2.16 (m, 1H), 2.16–2.30 (m, 1H), 3.71 (s, 3H), 6.38 (dd, J = 3.6, 2.8 Hz, 1H), 7.06 (dd, J = 3.6, 1.6 Hz, 1H), 7.34 (dd, J = 2.8, 1.6 Hz, 1H), 7.56–7.65 (m, 3H), 8.08–8.16 (m, 2H) ppm; ¹³C-NMR (acetone-d₆) $\delta = 20.9$, 22.3, 24.7, 41.0, 51.9, 58.0, 109.8, 114.4, 117.6, 123.9, 126.1, 126.6, 129.2, 131.6, 159.3, 162.3, 171.2 ppm; IR $\nu = 3119$, 3069, 2957, 2867, 1750, 1704, 1664, 1609, 1557, 1504, 1454, 1412, 1369, 1333, 1273, 1240, 1197, 1176, 1133, 1106, 1080, 1021, 1000, 964, 925, 880, 836, 811, 730, 690, 613 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₁N₃O₃+Na, 362.1475: found 362.1474.

Methyl (2*S*,3*S*)-3-*methyl*-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*pentanoate* (**1e**). Data for **4e**: white solid in a yield of 86% (0.91 g, 2.67 mmol); ¹H-NMR (DMSO-d₆) δ = 0.79 (t, J = 7.2 Hz, 3H), 0.96 (d, J = 6.4 Hz, 3H), 1.01–1.20 (m 2H), 2.15–2.26 (m 2H), 3.69 (s, 3H), 6.03 (d, J = 6.8 Hz, 1H), 6.22 (dd, J = 3.6, 2.8 Hz, 1H), 6.52 (dd, J = 3.6, 1.6 Hz, 1H), 7.18 (dd, J = 2.8, 1.6 Hz, 1H), 7.49–7.62 (m, 3H), 7.88–7.95 (m, 2H), 8.39 (s, 1H), 11.55 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 10.7, 15.5, 24.3, 38.1, 52.2, 62.8, 109.7, 115.5, 125.4, 127.5, 127.6, 128.4, 131.6, 133.6, 141.1, 162.5, 170.9 ppm.

Data for **1e**: light-yellow solid in 90% yield (0.42 g, 1.23 mmol, a 2:1 mixture of stereoisomers); ¹H-NMR (major isomer, CDCl₃) δ = 0.84 (t, *J* = 7.2 Hz, 3H), 1.05 (d, *J* = 6.4 Hz, 3H), 1.06–1.28 (m, 2H), 2.27–2.40 (m, 1H), 3.75 (s, 3H), 6.18 (d, *J* = 10.4 Hz, 1H), 6.39 (dd, *J* = 4.0, 2.8 Hz, 1H), 7.04 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.40 (dd, *J* = 2.8, 2.0 Hz, 1H), 7.58–7.66 (m, 3H), 8.10–8.16 (m, 2H) ppm; ¹³C-NMR (major isomer, CDCl₃) δ = 10.1, 15.0, 24.7, 38.8, 51.7, 63.7, 110.1, 114.0, 114.1, 123.9, 126.0, 126.6, 129.2, 131.6, 159.3, 162.4, 170.7 ppm; IR v = 3144, 3124, 3066, 2970, 2932, 2879, 1747, 1704, 1606, 1552, 1501, 1492, 1449, 1414, 1387, 1334, 1284, 1236, 1196, 1178, 1100, 1077, 1020, 964, 926, 883, 727, 694, 618 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₁N₃O₃+Na, 362.1475: found 362.1475.

Methyl (*S*)-3-*phenyl*-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*propanoate* (**1f**). Data for **4f**: white solid in a yield of 83% (0.94 g, 2.51 mmol); ¹H-NMR (DMSO-d₆) δ = 3.42 (d of A of ABq, J_{AB} = 14.4, J_d = 10.0 Hz, 1H), 3.49 (d of B of ABq, J_{AB} = 14.4, J_d = 6.4 Hz, 1H), 3.67 (s, 3H), 6.11 (dd, J = 3.6, 2.8 Hz, 1H), 6.43 (dd, J = 3.6, 1.6 Hz, 1H), 6.49 (dd, J = 10.0, 6.4 Hz, 1H), 7.10–7.25 (m, 6H), 7.50–7.62 (m, 3H), 7.88–7.93 (m, 2H), 8.23 (s, 1H), 11.50 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 39.1, 53.6, 61.1, 110.4, 117.6, 127.5, 127.7, 128.2, 128.7, 129.3, 129.7, 130.4, 132.8, 134.9, 137.9, 142.2, 163.8, 171.7 ppm; IR ν = 3236, 3064, 3033, 2957, 2843, 1743, 1645, 1613, 1555, 1497, 1455, 1436, 1348, 1280, 1220, 1185, 1164, 1076, 1032, 1008, 904, 843, 802, 753, 699 cm⁻¹.

Data for **1**f: light-yellow solid in a yield of 86% (2.31 g, 6.19 mmol); ¹H-NMR (CD₃OD) δ = 3.38 (d of A of ABq, J_{AB} = 14.0, J_d = 10.0 Hz, 1H), 3.59 (d of B of ABq, J_{AB} = 14.0, J_d = 5.2 Hz, 1H), 3.75 (s, 3H), 6.29 (dd, J = 4.0, 2.8 Hz, 1H), 6.39 (dd, J = 10.0, 5.2 Hz, 1H), 6.90 (dd, J = 4.0, 1.6 Hz, 1H), 7.02–7.14 (m, 5H), 7.24 (dd, J = 2.8, 1.6 Hz, 1H), 7.54–7.62 (m, 3H), 8.01–8.05 (m, 2H) ppm; ¹³C-NMR (CD₃OD) δ = 41.4, 54.6, 63.9, 112.5, 117.3, 119.8, 126.0, 129.2, 129.2, 129.4, 130.7, 131.5, 131.8, 134.5, 139.0, 162.1, 165.5, 173.4 ppm; IR v = 3068, 3033, 3004, 2956, 2848, 1747, 1703, 1665, 1606, 1559, 1505, 1449, 1412, 1372, 1337, 1274, 1230, 1201, 1174, 1107, 1083, 1012, 982, 926, 884, 778, 728, 699, 615 cm⁻¹; HRMS (ESI) calcd for C₂₂H₁₉N₃O₃+Na 396.1319, found 396.1321.

Methyl (*S*)-4-phenyl-2-(2-(5-phenyl-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)butanoate (**1g**). Data for **4g**: white solid in a yield of 79% (1.31 g, 3.48 mmol); ¹H-NMR (DMSO-d₆) δ = 2.36–2.60 (m, 4H), 3.68 (s, 3H), 6.00–6.07 (m, 1H), 6.26 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.58 (dd, *J* = 3.6, 1.2 Hz, 1H), 7.11–7.28 (m, 6H), 7.50–7.61 (m, 3H), 7.87–7.92 (m, 2H), 8.34 (s, 1H), 11.52 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 32.8, 35.1, 53.5, 60.1, 110.6, 117.2, 127.2, 128.5, 128.7, 129.5, 129.6, 129.6, 132.7, 134.8, 141.8, 142.3, 163.7, 172.1 ppm; IR v = 3227, 3063, 3030, 2951, 2866,

1744, 1646, 1610, 1558, 1494, 1457, 1431, 1325, 1287, 1217, 1164, 1079, 1055, 1037, 1029, 1004, 952, 913, 755, 698 cm⁻¹.

Data for **1g**: light-yellow solid in 86% yield (0.42 g, 1.13 mmol); ¹H-NMR (CD₃OD) δ = 2.40–2.67 (m, 4H), 3.71 (s, 3H), 5.92 (dd, *J* = 10.4, 4.0 Hz, 1H), 6.43 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.99–7.05 (m, 3H), 7.06 (dd, *J* = 4.0, 1.6 Hz, 1H), 7.08–7.16 (m, 2H), 7.31 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.53–7.62 (m, 3H), 8.00–8.06 (m, 2H) ppm; ¹³C-NMR (CD₃OD) δ = 31.4, 33.3, 51.7, 58.9, 110.0, 114.5, 117.3, 123.2, 125.8, 126.0, 126.4, 128.0, 128.0, 129.0, 131.7, 139.8, 159.2, 162.7, 171.3 ppm; IR v = 3064, 3026, 2956, 2931, 2854, 1745, 1662, 1606, 1557, 1504, 1450, 1415, 1254, 1233, 1198, 1177, 1082, 1012, 979, 814, 772, 725, 698, 605 cm⁻¹; HRMS (ESI) calcd for C₂₃H₂₁N₃O₃+Na, 410.1475: found 410.1477.

Dimethyl (*S*)-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*succinate* (**1h**). Data for **4h**: white solid in a yield of 81% (1.01g, 2.84 mmol); ¹H-NMR (CD₃OD) δ = 3.27 (d of A of ABq, *J*_{AB} = 16.8, *J*_d = 8.0 Hz, 1H), 3.37 (d of B of ABq, *J*_{AB} = 16.8, *J*_d = 6.0 Hz, 1H), 3.63 (s, 3H), 3.73 (s, 3H), 6.20 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.32 (dd, *J* = 8.0, 6.0 Hz, 1H), 6.58 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.02 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.46–7.60 (m, 3H), 7.86–7.91 (m, 2H), 8.24 (s, 1H) ppm; ¹³C-NMR (CD₃OD) δ = 39.4, 53.8, 54.6, 59.5, 112.0, 120.3, 129.4, 129.9, 130.0, 131.1, 134.4, 136.0, 144.6, 167.8, 173.2, 173.8 ppm; IR ν = 3411, 2958, 1737, 1644, 1613, 1581, 1554, 1495, 1444, 1414, 1348, 1285, 1238, 1177, 1123, 1084, 1008, 979, 908, 803, 786, 712 cm⁻¹.

Data for **1h**: light-yellow solid in 88% yield (0.79 g, 2.22 mmol); ¹H-NMR (CD₃OD) δ = 3.25 (d of A of ABq, J_{AB} = 16.8, J_d = 8.0 Hz, 1H), 3.41 (d of B of ABq, J_{AB} = 16.8, J_d = 6.0 Hz, 1H), 3.63 (s, 3H), 3.72 (s, 3H), 6.32 (dd, J = 3.6, 2.8 Hz, 1H), 6.39 (dd, J = 8.0, 6.0 Hz, 1H), 7.03 (dd, J = 3.6, 1.6 Hz, 1H), 7.17 (dd, J = 2.8, 1.6 Hz, 1H), 7.50–7.60 (m, 3H), 8.00–8.07 (m, 2H) ppm; ¹³C-NMR (CD₃OD) δ = 36.4, 51.2, 52.0, 56.8, 109.8, 115.0, 117.0, 123.2, 126.4, 126.7, 129.0, 131.7, 159.1, 162.8, 169.9, 170.5 ppm; IR ν = 3123, 3006, 2958, 2854, 1741, 1662, 1605, 1553, 1505, 1485, 1439, 1417, 1371, 1271, 1224, 1173, 1083, 1010, 985, 860, 819, 781, 726, 692, 672, 611 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₇N₃O₅+Na, 378.1060: found 378.1062.

Dimethyl (*S*)-2-(2-(5-*phenyl*-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)*pentanedioate* (**1i**). Data for **4i**: white solid in a yield of 70% (1.52 g, 4.08 mmol), ¹H-NMR (DMSO-d₆) δ = 2.10–2.21 (m, 1H), 2.24–2.42 (m, 2H), 2.44–2.53 (m, 1H), 3.55 (s, 3H), 3.70 (s, 3H), 6.01–6.09 (m, 1H), 6.22 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.55 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.12 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.49–7.61 (m, 3H), 7.87–7.92 (m, 2H), 8.32 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 28.7, 31.3, 52.7, 53.6, 59.7, 110.7, 117.5, 127.6, 128.5, 128.7, 129.7, 132.8, 134.8, 142.2, 163.7, 171.7, 173.5 ppm; IR v = 3234, 3020, 2954, 1738, 1650, 1615, 1558, 1458, 1437, 1346, 1280, 1218, 1177, 1075, 1029, 913, 888, 801, 753 cm⁻¹.

Data for **1i**: light-yellow solid in a yield of 95% (1.04 g, 2.82 mmol); ¹H-NMR (acetoned6) δ = 2.24–2.39 (m, 2H), 2.44–2.54 (m, 1H), 2.63–2.73 (m, 1H), 3.57 (s, 3H), 3.73 (s, 3H), 6.24 (dd, *J* = 10.8, 5.2 Hz, 1H), 6.39 (dd, *J* = 3.6, 2.8 Hz, 1H), 7.06 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.30 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.58–7.65 (m, 3H), 8.10–8.16 (m, 2H) ppm; ¹³C-NMR (acetone-d6) δ = 29.4, 31.4, 52.6, 53.7, 60.7, 111.8, 116.2, 119.4, 125.6, 128.0, 128.3, 130.9, 133.3, 160.9, 164.1, 172.0, 173.7 ppm; IR ν = 3120, 3005, 2956, 2923, 2853, 1741, 1664, 1607, 1552, 1504, 1490, 1450, 1371, 1240, 1202, 1174, 1106, 1078, 1012, 989, 964, 930, 882, 847, 821, 727, 694, 612 cm⁻¹; HRMS (ESI) calcd for C₁₉H₁₉N₃O₅+Na, 392.1217: found 392.1220.

Methyl (S)-4-(methylthio)-2-(2-(5-phenyl-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)butanoate (**1j**) and methyl (S)-6-(5-phenyl-1,3,4-oxadiazol-2-yl)-3,4-dihydro-2H-pyrrolo[2,1-b][1,3]thiazine-4-carboxylate (**1k**). Data for **4j**: white solid in a yield of 82% (0.77 g, 2.14 mmol); ¹H-NMR (DMSO-d₆) δ = 2.02 (s, 3H), 2.24–2.46 (m, 4H), 3.69 (s, 3H), 5.97–6.05 (m, 1H), 6.22 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.55 (dd, *J* = 3.6, 1.2 Hz, 1H), 7.15 (dd, *J* = 2.8, 1.2 Hz, 1H), 7.48–7.61 (m, 3H), 7.86–8.02 (m, 2H), 8.33 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 15.8, 30.8, 32.9, 53.6, 59.7, 110.6, 117.3, 127.4, 128.5, 128.7, 129.7, 132.8, 134.8, 142.1, 163.7, 171.8 ppm; IR v = 3230, 3006, 2957, 12918, 2849, 1743, 1649, 1614, 1556, 1491, 1459, 1431, 1350, 1288, 1230, 1209, 1144, 1091, 1033, 1001, 955, 909, 888, 798, 756, 613 cm⁻¹.

Data for **1j**: light-yellow solid in a yield of 50% (0.31 g, 0.86 mmol); ¹H-NMR (CD₃OD) δ = 2.03 (s, 3H), 2.28–2.36 (m, 1H), 2.43–2.63 (m, 3H), 3.74 (s, 3H), 6.18 (dd, *J* = 10.0, 4.8 Hz, 1H), 6.39 (dd, *J* = 4.0, 2.8 Hz, 1H), 7.07 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.25 (dd, *J* = 2.8, 2.0 Hz, 1H),

7.55–7.63 (m, 3H), 8.06–8.10 (m, 2H) ppm; 13 C-NMR (CD₃OD) δ = 13.7, 29.6, 31.1, 51.8, 58.7, 109.8, 114.8, 117.2, 123.2, 126.4, 129.0, 131.7, 159.2, 162.8, 171.0, 179.6 ppm; IR ν = 2956, 2925, 2857, 1750, 1667, 1603, 1554, 1505, 1452, 1381, 1274, 1243, 1090, 1016, 961, 883, 817, 774, 732, 691 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₉N₃O₃S+Na, 380.1039: found 380.1040.

Data for **1k**: light-yellow solid in 49% yield (0.31g, 0.72 mmol); ¹H-NMR (CD₃OD) δ = 2.39–2.49 (m, 1H), 2.86–3.02 (m, 3H), 3.73 (s, 3H), 5.94 (dd, *J* = 5.2, 3.2 Hz, 1H), 6.06 (d, *J* = 4.0 Hz, 1H), 7.01 (d, *J* = 4.0 Hz, 1H), 7.49–7.57 (m, 3H), 7.97–8.01 (m, 2H) ppm; ¹³C-NMR (CD₃OD) δ = 22.0, 28.0, 53.3, 58.2, 108.5, 116.1, 119.0, 124.7, 127.7, 128.7, 130.3, 132.9, 160.2, 163.6, 172.5 ppm; IR ν = 3008, 2948, 2852, 1751, 1648, 1604, 1552, 1497, 1439, 1401, 1355, 1292, 1257, 1214, 1176, 1145, 1125, 1085, 1070, 1023, 974, 929, 898, 854, 758, 727, 696 cm⁻¹; HRMS (ESI) calcd for C₁₇H₁₅N₃O₃S+Na, 364.0726: found 364.0729.

3.1.3. General Procedure for the Preparation of 5 (R = i-Pr) and 6 (R = s-Bu)

Step-1: At 25 °C, under an argon atmosphere, benzohydrazide **3** (1.1 equiv.) was added to a stirred solution of pyrrole-2-carbaldehyde **2** (~1.0 g, 1 equiv.) in toluene/DMSO (v:v = 15:1). The mixture was heated at 110 °C for 12 h and cooled to room temperature. The solvent was removed under reduced pressure in a rotary evaporator, and the crude product was filtered through a short pad of SiO₂ (EtOAc eluent) and concentrated under reduced pressure.

Step-2: I₂ (1.2 equiv.) and K₂CO₃ (3 equiv.) were added to a stirred solution of the above imine in 1,4-dioxane (20 mL). The mixture was heated at 85 °C for 6h under an argon atmosphere and cooled to room temperature. The mixture was diluted with CH₂Cl₂, washed with a 10% Na₂S₂O₃ solution, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The product was purified by SiO₂ flash column chromatography to obtain pyrrole-fused 1,3,4-oxadiazole **5** or **6**.

Methyl (*S*)-2-(2-(5-(2-*fluorophenyl*)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylbutanoate (**5a**). Orange oil, 40% yield (663 mg, 1.93 mmol); $R_f = 0.45$ (4:1 hexane/EtOAc); Data for **5a**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (d, *J* = 6.4 Hz, 3H), 1.08 (d, *J* = 6.4 Hz, 3H), 2.48 (m, 1H), 3.75 (s, 3H), 6.15 (d, *J* = 10.0 Hz, 1H), 6.36 (dd, *J* = 4.0, 3.2 Hz, 1H), 6.98 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.23–7.28 (m, 1H), 7.28–7.34 (m, 1H), 7.31 (dd, *J* = 3.2, 2.0 Hz, 1H), 7.50–7.57 (m, 1H), 8.08–8.13 (m, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.2, 52.3, 64.6, 110.4, 114.5, 117.0 (d, *J* = 20.5 Hz), 117.8, 124.6 (d, *J* = 3.1 Hz), 126.0, 129.6 (d, *J* = 1.5 Hz), 133.3 (d, *J* = 8.4 Hz), 158.8, 159.4, (d, *J* = 5.4 Hz), 159.7 (d, *J* = 1.5 Hz), 161.3, 171.3 ppm; IR v = 2970, 2880, 1748, 1605, 1500, 1475, 1450, 1273, 1239, 1219, 1200, 1185, 1170, 1102, 1078, 1009, 827, 742, 669, 617 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₈FN₃O₃+Na 366.1224: found 366.1225.

Methyl (*S*)-2-(2-(5-(3-*fluorophenyl*)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylbutanoate (**5b**). Orange oil, 70% yield (759 mg, 2.21 mmol); $R_f = 0.59$ (4:1 hexane/EtOAc); Data for **5b**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (d, *J* = 6.4 Hz, 3H), 1.08 (d, *J* = 6.4 Hz, 3H), 2.49 (m, 1H), 3.75 (s, 3H), 6.14 (d, *J* = 10.0 Hz, 1H), 6.37 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.97 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.20–7.27 (m, 1H), 7.32 (dd, *J* = 2.8, 2.0 Hz, 1H), 7.47–7.54 (m, 1H), 7.76–7.81 (m, 1H), 7.87–7.91 (m, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.1, 52.3, 64.6, 110.4, 113.8 (d, *J* = 24.2 Hz), 114.4 117.7, 118.6 (d, *J* = 21.2 Hz), 122.5 (d, *J* = 3.0 Hz), 125.7 (d, *J* = 9.1 Hz), 126.1, 130.9 (d, *J* = 8.3 Hz), 159.7, 161.6, (d, *J* = 2.3 Hz), 164.0, 171.2 ppm; IR v = 2970, 2880, 1750, 1600, 1565, 1500, 1495, 1450, 1410, 1375, 1307, 1275, 1240, 1205, 1181, 1105, 1080, 1010, 995, 870, 800, 735, 680, 615 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₈FN₃O₃+Na 366.1224: found 366.1226.

Methyl (*S*)-2-(2-(5-(4-fluorophenyl)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylbutanoate (**5c**). Yellow oil, 54% yield (721 mg, 2.10 mmol); $R_f = 0.65$ (4:1 hexane/EtOAc); Data for **5c**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (d, *J* = 6.4 Hz, 3H), 1.08 (d, *J* = 6.4 Hz, 3H), 2.48 (m, 1H), 3.75 (s, 3H), 6.14 (d, *J* = 10.0 Hz, 1H), 6.36 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.94 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.18–7.25 (m, 2H), 7.31 (dd, *J* = 2.8, 2.0 Hz, 1H), 8.07–8.13 (m, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.1, 52.3, 64.6, 110.3, 114.1, 116.4 (d, *J* = 22.0 Hz), 120.2 (d, *J* = 3.0 Hz), 125.9, 129.0 (d, *J* = 9.1 Hz), 159.4, 161.8, 163.4, 165.9, 171.2 ppm; IR ν = 2970, 2880, 1750, 1605, 1500,

1470, 1455, 1435, 1415, 1270, 1235, 1215, 1200, 1160, 1100, 1075, 1015, 965, 845, 740, 625 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₈FN₃O₃+Na 366.1224, found 366.1228.

Methyl (*S*)-2-(2-(5-(4-*chlorophenyl*)-1,3,4-*oxadiazol*-2-*yl*)-1*H*-*pyrrol*-1-*yl*)-3-*methylbutanoate* (5d). Brown solid, 62% yield (533 mg, 1.48 mmol); $R_f = 0.61$ (4:1 hexane/EtOAc); Data for 5d: ¹H-NMR (CDCl₃) $\delta = 0.82$ (d, *J* = 6.4 Hz, 3H), 1.08 (d, *J* = 6.4 Hz, 3H), 2.48 (m, 1H), 3.75 (s, 3H), 6.14 (d, *J* = 10.0 Hz, 1H), 6.36 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.95 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.31 (dd, *J* = 2.8, 2.0 Hz, 1H), 7.48–7.53 (m, 2H), 8.01–8.06 (m, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.2, 52.3, 64.6, 110.3, 114.3, 117.8, 122.3, 126.0, 128.1, 129.5, 137.8, 159.6, 161.8, 171.2 ppm; IR $\nu = 2970$, 2880, 1750, 1605, 1500, 1485, 1455, 1410, 1270, 1237, 1200, 1180, 1100, 1080, 1015, 970, 840, 740 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₈ClN₃O₃+Na 382.0929, found 382.0933.

Methyl (*S*)-2-(2-(5-(2-*hydroxy*-3,5-*diiodophenyl*)-1,3,4-*oxadiazo*l-2-*y*])-1*H*-*pyrro*l-1-*y*])-3*methylbutanoate* (**5e-1**). White solid, 33% yield (748 mg, 1.26 mmol); $R_f = 0.54$ (4:1 hexane/EtOAc); Data for **5e-1**: ¹H-NMR (CDCl₃) $\delta = 0.82$ (d, *J* = 6.4 Hz, 3H), 1.09 (d, *J* = 6.4 Hz, 3H), 2.49 (m, 1H), 3.77 (s, 3H), 5.99 (d, *J* = 9.6 Hz, 1H), 6.40 (dd, *J* = 4.0, 2.8 Hz, 1H), 7.05 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.36 (dd, *J* = 2.8, 2.0 Hz, 1H), 8.06 (d, *J* = 2.0 Hz, 1H), 8.18 (d, *J* = 2.0 Hz, 1H) 11.02 (s, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.6$, 19.3, 33.2, 52.4, 64.9, 81.3, 86.7, 109.8, 110.8, 115.7, 116.9, 127.0, 134.7, 149.8, 156.2, 158.7, 160.1, 171.0 ppm; IR $\nu = 2970$, 2890, 1750, 1600, 1565, 1535, 1500, 1445, 1415, 1380, 1255, 1240, 1220, 1185, 1105, 1080, 1015, 1000, 915, 870, 740, 660, 615, 600 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₇I₂N₃O₄+Na 615.9201: found 615.9203.

Methyl (*S*)-2-(2-(5-(4-hydroxy-2,3,5,6-tetraiodophenyl)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1yl)-3-methylbutanoate (**5f-1**): Yellow solid, 32% yield (1.33 g, 1.57 mmol); $R_f = 0.55$ (4:1 hexane/EtOAc); Data for **5f-1**. ¹H-NMR (CDCl₃) $\delta = 0.81$ (d, *J* = 6.4 Hz, 3H), 1.07 (d, *J* = 6.4 Hz, 3H), 2.41–2.53 (m, 1H), 3.75 (s, 3H), 6.09 (d, *J* = 10.0 Hz, 1H), 6.36 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.96 (dd, *J* = 4.0, 1.6 Hz, 1H), 7.31 (dd, *J* = 2.8, 1.6 Hz, 1H), 8.40 (s, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.1, 52.3, 64.6, 82.4, 110.4, 114.5, 117.7, 119.9, 126.1, 137.6, 156.2, 159.5, 159.6, 171.2 ppm; IR $\nu = 3450$, 3145, 3125, 2970, 2875, 1745, 1610, 1595, 1500, 1450, 1395, 1300, 1270, 1235, 1220, 1200, 1180, 1160, 1135, 1105, 1075, 1010, 995, 965, 910, 890, 810, 735, 710, 685, 650, 615 cm⁻¹; HRMS (ESI) calcd for [C₁₈H₁₅I₄N₃O₄–I₂+H₂]+Na 615.9201: found 615.9213.

Methyl (*S*)-2-(2-(5-(2-*methoxyphenyl*)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylbutanoate (**5g**). Red oil, 85% yield (1.03 g, 2.89 mmol); $R_f = 0.66$ (3:2 hexane/EtOAc); Data for **5g**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (d, *J* = 6.4 Hz, 3H), 1.07 (d, *J* = 6.4 Hz, 3H), 2.48 (m, 1H), 3.74 (s, 3H), 4.00 (s, 3H), 6.19 (d, *J* = 10.0 Hz, 1H), 6.35 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.94 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.06–7.12 (m, 2H), 7.29 (dd, *J* = 2.8, 2.0 Hz, 1H), 7.48–7.53 (m, 1H), 7.99 (dd, *J* = 7.6, 2.0 Hz, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.2, 52.2, 56.0, 64.5, 110.1, 112.0, 112.9, 113.9, 118.2, 120.7, 125.5, 130.2, 132.8, 158.0, 159.0, 161.3, 171.4 ppm; IR v = 2970, 2875, 2840, 1750, 1600, 1545, 1500, 1470, 1455, 1440, 1270, 1260, 1235, 1215, 1185, 1170, 1130, 1100, 1075, 1025, 750, 735, 675, 616 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₁N₃O₄+Na 378.1424, found 378.1428.

Methyl (*S*)-2-(2-(5-(4-*methoxyphenyl*)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylbutanoate (**5h**). Ivory solid, 70% yield (1.12 g, 3.15 mmol); $R_f = 0.70$ (3:2 hexane/EtOAc); Data for **5h**: ¹H-NMR (CDCl₃) $\delta = 0.82$ (d, J = 6.4 Hz, 3H), 1.07 (d, J = 6.4 Hz, 3H), 2.48 (m, 1H), 3.74 (s, 3H), 3.89 (s, 3H), 6.15 (d, J = 10.0 Hz, 1H), 6.35 (dd, J = 4.0, 2.8 Hz, 1H), 6.92 (dd, J = 4.0, 2.0 Hz, 1H), 7.00–7.05 (m, 2H), 7.29 (dd, J = 2.8, 2.0 Hz, 1H), 8.01–8.05 (m, 2H), 7.99 (dd, J = 7.6, 2.0 Hz, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.1, 52.3, 55.5, 64.5, 110.1, 113.8, 114.5, 116.4, 118.2, 125.6, 128.6, 159.0, 162.2, 162.6, 171.3 ppm; IR $\nu = 2970$, 2875, 1745, 1610, 1600, 1505, 1450, 1395, 1300, 1270, 1240, 1220, 1200, 1180, 1160, 1135, 1100, 1080, 1015, 995, 910, 890, 735, 715, 685, 650, 615 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₁N₃O₄+Na 378.1424: found 378.1426.

Methyl (2*S*,3*S*)-2-(2-(5-(2-*fluorophenyl*)-1,3,4-oxadiazol-2-*y*])-1*H*-pyrrol-1-*y*])-3-methylpentanoate (**6a**). Orange-red oil, 89% yield (1.03 g, 2.89 mmol); $R_f = 0.47$ (4:1 hexane/EtOAc); Data for **6a**: ¹H-NMR (CDCl₃) $\delta = 0.84$ (t, *J* = 7.2 Hz, 3H), 1.05 (d, *J* = 6.4 Hz, 3H), 1.06–1.27 (m,

2H), 2.21–2.32 (m, 1H), 3.74 (s, 3H), 6.20 (d, J = 9.6 Hz, 1H), 6.36 (dd, J = 4.0, 2.8 Hz, 1H), 6.98 (dd, J = 4.0, 2.0 Hz, 1H), 7.23–7.29 (m, 1H), 7.30–7.34 (m, 1H), 7.31 (dd, J = 2.8, 2.0 Hz, 1H), 7.50–7.57 (m, 1H), 8.08–8.13 (m, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 11.0$, 15.6, 24.8, 39.2, 52.3, 63.8, 110.3, 114.6, 117.0 (d, J = 21.3 Hz), 117.9, 124.6 (d, J = 3.8 Hz), 126.0, 129.6 (d, J = 0.5 Hz), 133.3 (d, J = 8.4 Hz), 158.8, 159.4, (d, J = 5.4 Hz), 159.6 (d, J = 1.5 Hz), 161.3, 171.4 ppm; IR $\nu = 2970$, 2937, 2880, 1750, 1600, 1500, 1475, 1450, 1400, 1260, 1235, 1198, 1180, 1100, 1080, 1026, 1000, 910, 825, 767, 735, 698, 670, 615 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₀FN₃O₃+Na 380.1381:found 380.1386.

Methyl (2*S*,3*S*)-2-(2-(5-(3-*fluorophenyl*)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylpentanoate (**6b**). Orange-red oil, 88% yield (0.99 g, 2.77 mmol); $R_f = 0.47$ (4:1 hexane/EtOAc); Data for **6b**: ¹H-NMR (CDCl₃) $\delta = 0.84$ (t, J = 7.2 Hz, 3H), 1.05 (d, J = 6.4 Hz, 3H), 1.06–1.28 (m, 2H), 2.20–2.32 (m, 1H), 3.75 (s, 3H), 6.19 (d, J = 10.0 Hz, 1H), 6.36 (dd, J = 3.6, 2.8 Hz, 1H), 6.97 (dd, J = 3.6, 2.0 Hz, 1H), 7.20–7.27 (m, 1H), 7.33 (dd, J = 2.8, 2.0 Hz, 1H), 7.47–7.53 (m, 1H), 7.76–7.81 (m, 1H), 7.87–7.91 (m, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 10.9$, 15.5, 24.8, 39.1, 52.3, 63.8, 110.4, 113.8 (d, J = 24.2 Hz), 114.5, 117.7, 118.6 (d, J = 21.3 Hz), 122.5 (d, J = 3.0 Hz), 125.7 (d, J = 8.3 Hz), 126.1, 130.9 (d, J = 8.4 Hz), 159.6, 161.6 (d, J = 3.1 Hz), 164.0, 171.3 ppm; IR $\nu = 2970$, 2940, 2880, 1750, 1595, 1560, 1505, 1490, 1454, 1415, 1245, 1205, 1178, 1105, 1080, 995, 915, 870, 795, 735, 680, 615 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₀FN₃O₃+Na 380.1381, found 380.1384.

Methyl (2*S*,3*S*)-2-(2-(5-(4-fluorophenyl)-1,3,4-oxadiazol-2-yl)-1*H*-pyrrol-1-yl)-3-methylpentanoate (6c). Yellow oil, 62% yield (768 mg, 2.15 mmol); $R_f = 0.58$ (4:1 hexane/EtOAc); Data for 6c: ¹H-NMR (CDCl₃) $\delta = 0.83$ (t, *J* = 7.2 Hz, 3H), 1.04 (d, *J* = 6.4 Hz, 3H), 1.05–1.27 (m, 2H), 2.22–2.32 (m, 1H), 3.74 (s, 3H), 6.19 (d, *J* = 10.4 Hz, 1H), 6.36 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.94 (dd, *J* = 3.6, 2.0 Hz, 1H), 7.18–7.25 (m, 2H), 7.31 (dd, *J* = 2.8, 2.0 Hz, 1H), 8.07–8.13 (m, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 10.9$, 15.5, 24.8, 39.1, 52.3, 63.7, 110.3, 114.2, 116.4 (d, *J* = 22.8 Hz), 117.9, 126.0, 129.1 (d, *J* = 8.3 Hz), 159.4, 161.8, 163.4, 165.9, 171.4 ppm; IR v = 2970, 2940, 2880, 1750, 1670, 1605, 1500, 1455, 1415, 1240, 1200, 1180, 1160, 1080, 1015, 1000, 850, 740, 620 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₀FN₃O₃+Na 380.1381: found 380.1387.

Methyl (2*S*,3*S*)-2-(2-(5-(4-chlorophenyl)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylpentanoate (6d). Orange oil, 79% yield (254 mg, 0.68 mmol); $R_f = 0.65$ (4:1 hexane/EtOAc); Data for 6d: ¹H-NMR (CDCl₃) $\delta = 0.83$ (t, J = 7.2 Hz, 3H), 1.04 (d, J = 6.4 Hz, 3H), 1.06–1.27 (m, 2H), 2.20–2.32 (m, 1H), 3.74 (s, 3H), 6.19 (d, J = 10.4 Hz, 1H), 6.36 (dd, J = 4.0, 2.8 Hz, 1H), 6.95 (dd, J = 4.0, 1.6 Hz, 1H), 7.32 (dd, J = 2.8, 1.6 Hz, 1H), 7.48–7.52 (m, 2H), 8.01–8.05 (m, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 10.9$, 15.5, 24.8, 39.1, 52.3, 63.8, 110.3, 114.4, 117.8, 122.3, 126.1, 128.1, 129.4, 137.8, 159.5, 161.8, 171.3 ppm; IR $\nu = 2970$, 2935, 2880, 1750, 1605, 1505, 1485, 1455, 1410, 1255, 1235, 1200, 1178, 1095, 1080, 1015, 840, 735 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₀ClN₃O₃+Na 396.1085: found 396.1089.

Methyl (2*S*,3*S*)-2-(2-(5-(2-*hydroxy*-3,5-*diiodophenyl*)-1,3,4-*oxadiazo*l-2-*y*])-1*H*-*pyrro*l-1-*y*])-3-*methylpentanoate* (**6e-1**). Green solid, 45% yield (893 mg, 1.47 mmol); $R_f = 0.50$ (4:1 hexane/EtOAc); Data for **6e-1**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (t, J = 7.2 Hz, 3H), 1.05 (d, J = 6.4 Hz, 3H), 1.02–1.22 (m, 2H), 2.20–2.33 (m, 1H), 3.77 (s, 3H), 6.04 (d, J = 9.6 Hz, 1H), 6.39 (dd, J = 4.0, 2.8 Hz, 1H), 7.04 (dd, J = 4.0, 1.6 Hz, 1H), 7.37 (dd, J = 2.8, 1.6 Hz, 1H), 8.06 (d, J = 2.0 Hz, 1H), 8.17 (d, J = 2.0 Hz, 1H), 11.02 (s, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 10.9$, 15.5, 24.8, 38.1, 52.4, 64.0, 81.3, 86.7, 109.8, 110.8, 115.7, 116.9, 127.0, 134.7, 149.8, 156.2, 158.7, 160.1, 171.1 ppm; IR $\nu = 3415$, 2970, 2880, 1750, 1605, 1565, 1530, 1500, 1455, 1415, 1380, 1255, 1235, 1190, 1180, 1100, 1080, 990, 915, 875, 758, 740, 665, 598 cm⁻¹; HRMS (ESI) calcd for C₁₉H₁₉I₂N₃O₄+Na 629.9357: found 629.9358.

Methyl (2*S*,3*S*)-2-(2-(5-(4-hydroxy-3,5-diiodophenyl)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylpentanoate (**6f-1**). Yellow oil, 43% yield (935 mg, 1.54 mmol); $R_f = 0.45$ (4:1 hexane/EtOAc); Data for **6f-1**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (t, *J* = 7.2 Hz, 3H), 1.04 (d, *J* = 6.4 Hz, 3H), 1.05–1.24 (m, 2H), 2.20–2.32 (m, 1H), 3.75 (s, 3H), 6.14 (d, *J* = 10.0 Hz, 1H), 6.21 (br s, 1H), 6.36 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.96 (dd, *J* = 4.0, 2.0 Hz, 1H), 7.32 (dd, *J* = 2.8, 2.0 Hz, 1H), 8.40 (s, 2H), 8.17 (d, *J* = 2.0 Hz, 1H), 11.02 (s, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 10.9$, 15.5, 24.8, 39.1, 52.3, 63.8, 82.5, 110.4, 114.5, 117.7, 119.9, 126.1, 137.6, 156.2, 159.4, 159.6, 171.3 ppm; IR ν = 3450, 3145, 3125, 3070, 2970, 2880, 1745, 1608, 1595, 1500, 1450, 1395, 1300, 1235, 1195, 1178, 1155, 1105, 1075, 990, 890, 775, 736, 710, 685, 615 cm⁻¹; HRMS (ESI) calcd for C₁₉H₁₉I₂N₃O₄+Na 629.9357: found 629.9360.

Methyl (2*S*,3*S*)-2-(2-(5-(2-*methoxyphenyl*)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylpentanoate (**6g**). Orange-red oil, 80% yield (768 mg, 2.08 mmol); $R_f = 0.19$ (4:1 hexane/EtOAc); Data for **6g**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (t, *J* = 7.2 Hz, 3H), 1.04 (d, *J* = 6.4 Hz, 3H), 1.05–1.27 (m, 2H), 2.20–2.32 (m, 1H), 3.73 (s, 3H), 3.99 (s, 3H), 6.24 (d, *J* = 9.6 Hz, 1H), 6.34 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.94 (dd, *J* = 3.6, 1.6 Hz, 1H), 7.05–7.11 (m, 2H), 7.30 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.47–7.53 (m, 1H), 7.98 (dd, *J* = 7.6, 2.0 Hz, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 11.0$, 15.5, 24.8, 39.2, 52.2, 56.0, 63.7, 110.1, 112.0, 112.9, 114.0, 118.3, 120.7, 125.6, 130.2, 132.8, 157.9, 159.0, 161.3, 171.5 ppm; IR $\nu = 2970$, 2942, 2882, 2845, 1750, 1605, 1550, 1500, 1485, 1455, 1440, 1415, 1255, 1240, 1200, 1180, 1100, 1080, 1050, 1025, 915, 730, 678, 650, 620 cm⁻¹; HRMS (ESI) calcd for C₂₀H₂₃N₃O₄+Na 392.1581: found 392.1585.

Methyl (2*S*,3*S*)-2-(2-(5-(4-*methoxyphenyl*)-1,3,4-*oxadiazo*l-2-*y*))-1*H*-*pyrro*l-1-*y*))-3-*methylpentanoate* (**6h**). Orange oil; 79% yield (739 mg, 2.00 mmol); $R_f = 0.24$ (4:1 hexane/EtOAc); Data for **6h**: ¹H-NMR (CDCl₃) $\delta = 0.83$ (t, *J* = 7.2 Hz, 3H), 1.03 (d, *J* = 6.4 Hz, 3H), 1.05–1.23 (m, 2H), 2.20–2.30 (m, 1H), 3.74 (s, 3H), 3.88 (s, 3H), 6.20 (d, *J* = 9.6 Hz, 1H), 6.34 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.92 (dd, *J* = 3.6, 1.6 Hz, 1H), 6.99–7.03 (m, 2H), 7.29 (dd, *J* = 2.8, 1.6 Hz, 1H), 8.00–8.04 (m, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 11.0$, 15.5, 24.8, 39.1, 52.2, 55.4, 63.7, 110.1, 113.8, 114.5, 116.4, 118.2, 125.6, 128.6, 158.9, 162.2, 162.6, 171.4 ppm; IR $\nu = 2970$, 2940, 2880, 2840, 1750, 1610, 1500, 1460, 1445, 1310, 1255, 1180, 1100, 1080, 1030, 1000, 915, 840, 730, 700, 625, 610 cm⁻¹; HRMS (ESI) calcd for C₂₀H₂₃N₃O₄+Na 392.1581: found 392.1584.

3.2. General Procedure for Deiodination Reaction on the Phenol Ring

Zn dust (2~5 equiv.) and acetic acid (5 equiv.) were added to a stirred solution of 2pyrrolyl-5-(iodophenolic)-1,3,4-oxadiazole **5f-1** or **6e-1** (~0.5–1.0 g, 1 equiv.) in THF (30mL). The mixture was stirred at 25 °C for 1~6 h under an argon atmosphere. The mixture was quenched with saturated NaHCO₃ solution, extracted with Et₂O, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The product was purified by SiO₂ flash column chromatography to produce the deiodination product.

Methyl (*S*)-2-(2-(5-(4-hydroxyphenyl)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylbutanoate (**5**f). White solid, 97% yield (270 mg, 0.79 mmol); $R_f = 0.58$ (3:2 hexane/EtOAc); Data for **5**f: ¹H-NMR (CDCl₃) $\delta = 0.83$ (d, *J* = 6.4 Hz, 3H), 1.08 (d, *J* = 6.4 Hz, 3H), 2.43–2.54 (m, 1H), 3.75 (s, 3H), 6.13 (d, *J* = 9.6 Hz, 1H), 6.36 (dd, *J* = 4.0, 2.8 Hz, 1H), 6.93 (dd, *J* = 4.0, 1.6 Hz, 1H), 7.01 (d, *J* = 8.4 Hz, 2H), 7.30 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.99 (d, *J* = 8.4 Hz, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.5$, 19.3, 33.1, 52.3, 64.6, 110.2, 114.0, 116.0, 116.2, 118.0, 125.7, 128.8, 159.0, 159.1, 162.7, 171.3 ppm; IR $\nu = 3120$, 2970, 2940, 2880, 1750, 1610, 1595, 1500, 1440, 1394, 1375, 1335, 1285, 1270, 1240, 1200, 1105, 1090, 1075, 1010, 995, 915, 840, 775, 735, 719, 635 cm⁻¹; HRMS (ESI) calcd for C₁₈H₁₉N₃O₄+Na 364.1268: found 365.1271.

Methyl (25,35)-2-(2-(5-(2-*hydroxyphenyl*)-1,3,4-oxadiazol-2-*yl*)-1H-*pyrrol*-1-*yl*)-3-*methylpentanoate* (6e). White solid, 50% yield (210 mg, 0.59 mmol); R_f = 0.54 (3:2 hexane/EtOAc); Data for 6e: ¹H-NMR (DMSO-d₆) δ = 0.79 (t, *J* = 7.2 Hz, 3H), 0.96 (d, *J* = 6.4 Hz, 3H), 0.98–1.19 (m, 2H), 2.16–2.26 (m, 1H), 3.69 (s, 3H), 6.02 (d, *J* = 9.6 Hz, 1H), 6.24 (dd, *J* = 3.6, 2.8 Hz, 1H), 6.56 (dd, *J* = 3.6, 1.6 Hz, 1H), 6.92–6.99 (m, 2H), 7.20 (dd, *J* = 2.8, 1.6 Hz, 1H), 7.41–7.46 (m, 1H), 7.86–7.90 (m, 1H) 8.39 (s, 1H) ppm; ¹³C-NMR (DMSO-d₆) δ = 11.1, 16.0, 24.8, 38.5, 52.7, 63.3, 110.3, 115.9, 116.5, 117.8, 119.3, 126.2, 127.8, 128.6, 134.2, 142.5, 159.8, 164.9, 171.3 ppm; IR v = 3240, 3080, 2970, 2935, 2880, 1750, 1640, 1600, 1570, 1540, 1495, 1465, 1425, 1360, 1335, 1315, 1258, 1230, 1200, 1175, 1155, 1105, 1075, 1060, 1040, 995, 950, 895, 825, 757, 740, 670, 618 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₃N₃O₄+Na 380.1581: found 380.1586.

Methyl (2*S*,3*S*)-2-(2-(5-(4-hydroxyphenyl)-1,3,4-oxadiazol-2-yl)-1H-pyrrol-1-yl)-3-methylpentanoate (**6f**). White solid, 42% yield (98 mg, 0.28 mmol); $R_f = 0.67$ (3:2 hexane/EtOAc); Data for **6f**: ¹H-NMR (CDCl₃) $\delta = 0.77$ (t, J = 7.2 Hz, 3H), 0.98 (d, J = 6.8 Hz, 3H), 0.98–1.18 (m, 2H), 2.26–2.35 (m, 1H), 3.70 (s, 3H), 5.96 (d, J = 10.0 Hz, 1H), 6.38 (dd, J = 3.6, 2.8 Hz, 1H), 6.97 (d, J = 8.4 Hz, 2H), 6.98 (dd, J = 3.6, 1.6 Hz, 1H), 7.40 (dd, J = 2.8, 1.6 Hz, 1H), 7.90 (d,

J = 8.4 Hz, 2H), 10.34 (s, 1H) ppm; ¹³C-NMR (CDCl₃) δ = 10.9, 15.5, 24.8, 39.1, 52.3, 63.7, 110.3, 114.2, 115.5, 117.9, 125.9, 129.0, 136.9, 157.7, 159.2, 161.1, 171.4 ppm; IR ν = 3137, 2965, 2875, 2838, 1742, 1610, 1579, 1558, 1497, 1464, 1466, 1438, 1338, 1307, 1287, 1254, 1224, 1202, 1173, 1092, 1060, 1027, 961, 938, 911, 837, 810, 798, 731, 696 cm⁻¹; HRMS (ESI) calcd for C₁₉H₂₁N₃O₄+Na 378.1424: found 378.1427.

3.3. General Procedure for Iodination Reaction on the Pyrrole Ring

I₂ (4 equiv.). was added to a stirred solution of 2-pyrrolyl-5-(anisyl)-1,3,4-oxadiazoles **5g**, **5h**, or **6h** (~1.0 g, 1 equiv.) in DMSO (10 mL). The mixture was heated at 90 °C for $3\sim5$ h under an argon atmosphere. After cooling to room temperature, the mixture was diluted with Et₂O, washed with 10% Na₂S₂O₃ solution, dried over anhydrous Na₂SO₄, filtered, and concentrated under reduced pressure. The product was purified by SiO₂ flash column chromatography to produce iodination products **5g-1**, **5h-1**, or **6h-1** on the pyrrole ring.

Methyl (*S*)-2-(3,4-*diiodo*-2-(5-(2-*methoxyphenyl*)-1,3,4-*oxadiazo*l-2-*y*])-1*H*-*pyrro*l-1-*y*])-3*methylbutanoate* (**5g-1**). Orange oil, 61% yield (923 mg, 1.52 mmol); $R_f = 0.60$ (3:2 hexane/EtOAc); Data for **5g-1**: ¹H-NMR (CDCl₃) $\delta = 0.86$ (d, *J* = 6.4 Hz, 3H), 1.04 (d, *J* = 6.4 Hz, 3H), 2.42 (m, 1H), 3.73 (s, 3H), 4.03 (s, 3H), 6.12 (d, *J* = 10.0 Hz, 1H), 7.08–7.14 (m, 2H), 7.45 (s, 1H), 7.51–7.56 (m, 1H), 8.12 (dd, *J* = 7.6, 1.6 Hz, 1H) ppm; ¹³C-NMR (CDCl₃) $\delta = 18.6$, 19.2, 33.1, 52.5, 56.0, 65.6, 78.0, 80.4, 112.0, 112.3, 120.8, 121.4, 130.4, 130.5, 133.2, 156.7, 158.1, 162.1, 170.6 ppm; IR $\nu = 2970$, 2937, 2880, 2840, 1750, 1665, 1605, 1590, 1545, 1500, 1475, 1470, 1435, 1390, 1285, 1265, 1220, 1205, 1185, 1165, 1130, 1060, 1050, 1025, 943, 915, 768, 755, 735, 677, 650 cm⁻¹; HRMS (ESI) calcd for C₁₉H₁₉I₂N₃O₄+Na 629.9357: found 629.9363.

Methyl (*S*)-2-(3,4-*diiodo*-2-(5-(4-*methoxyphenyl*)-1,3,4-*oxadiazo*l-2-*y*])-1*H*-*pyrro*l-1-*y*])-3*methylbutanoate* (**5h-1**). Yellow oil (a 1.3:1 mixture of di- and mono-iodide products); diiodide, 39% yield (199 mg, 0.33 mmol, calcd), mono-iodide, 30% yield (121 mg, 0.25 mmol, calcd); R_f = 0.60 (3:2 hexane/EtOAc); Data for di-iodide at pyrrole: ¹H-NMR (CDCl₃) δ = 0.86 (d, *J* = 6.4 Hz, 3H), 1.05 (d, *J* = 6.4 Hz, 3H), 2.43 (m, 1H), 3.74 (s, 3H), 3.89 (s, 3H), 6.09 (d, *J* = 10.0 Hz, 1H), 7.05 (d, *J* = 8.8 Hz, 2H), 7.44 (s, 1H), 8.11 (d, *J* = 8.8 Hz, 2H) ppm; ¹³C-NMR (CDCl₃) δ = 18.6, 19.1, 33.0, 52.5, 55.5, 65.7, 78.0, 80.4, 114.6, 116.0, 120.3, 128.9, 130.6, 156.7, 162.5, 163.4, 170.5 ppm; IR ν = 2970, 1750, 1615, 1590, 1560, 1500, 1485, 1460, 1440, 1390, 1335, 1255, 1205, 1175, 1160, 1065, 1030, 945, 840, 755, 745, 625 cm⁻¹; HRMS (ESI) calcd for C₁₉H₁₉I₂N₃O₄+Na 629.9357, found 629.9358. Data for mono-iodide at pyrrole (C-3): ¹H-NMR (CDCl₃) δ = 0.61 (d, *J* = 6.4 Hz, 3H), 1.06 (d, *J* = 6.4 Hz, 3H), 2.43 (m, 1H), 3.76 (s, 3H), 3.89 (s, 3H), 6.13 (d, *J* = 9.6 Hz, 1H), 7.01 (d, *J* = 2.0 Hz, 1H), 7.02 (d, *J* = 8.4 Hz, 2H), 7.35 (d, *J* = 2.0 Hz, 1H), 8.02 (d, *J* = 8.4 Hz, 2H) ppm; ¹³C-NMR (CDCl₃) δ = 18.5, 19.2, 33.3, 52.4, 61.3, 64.9, 77.2, 114.5, 116.0, 120.1, 121.5, 128.6, 130.1, 157.7, 162.4, 162.8, 170.8 ppm; HRMS (ESI) calcd for C₁₉H₂₀IN₃O₄+Na 504.0396: found 504.0387.

Methyl (2*S*,3*S*)-2-(3-*iodo*-2-(5-(4-*methoxyphenyl*)-1,3,4-*oxadiazo*l-2-*y*l)-1*H*-*pyrro*l-1-*y*l)-3*methylpentanoate* (**6h-1**). Yellow oil, 83% yield (308 mg, 0.67 mmol); $R_f = 0.40$ (4:1 hexane/EtOAc); Data for **6h-1**: ¹H-NMR (CDCl₃) $\delta = 0.84$ (t, J = 7.2 Hz, 3H), 1.02 (d, J = 6.4 Hz, 3H), 1.06–1.26 (m, 2H), 2.16–2.28 (m, 1H), 3.75 (s, 3H), 3.89 (s, 3H), 6.18 (d, J = 10.0 Hz, 1H), 6.99–7.04 (m, 2H), 7.01 (d, J = 1.6 Hz, 1H), 7.35 (d, J = 1.6 Hz, 1H), 7.99–8.03 (m, 2H) ppm; ¹³C-NMR (CDCl₃) $\delta = 10.9$, 15.4, 24.8, 39.2, 52.4, 55.5, 64.1, 77.2, 114.6, 116.0, 120.1, 120.3, 128.7, 130.1, 157.6, 162.4, 162.9, 171.0 ppm; IR $\nu = 2970$, 2935, 2880, 2840, 1750, 1607, 1498, 1464, 1440, 1310, 1260, 1176, 1100, 1065, 1030, 1000, 915, 840, 815, 745, 640, 625, 607 cm⁻¹; HRMS (ESI) calcd for C₂₀H₂₂IN₃O₄+Na 518.0547: found 518.0548.

3.4. Biological Evaluation

The minimum inhibitory concentrations (MICs) were determined using the broth microdilution method in a 96-well plate [40,41]. The 96-well plates containing chemicals in two-fold serial dilutions (4 µg/mL to 2048 µg/mL for series 1; 2 µg/mL to 1024 µg/mL for series 5 and 6) were prepared in Luria–Bertani (LB) medium. *E. coli, S. aureus,* and *A. baumannii* cells were grown in LB broth to the exponential phase. A 10 µL volume of cells diluted with LB broth to a concentration of 10^8 cells/mL was inoculated on the plates. The

MIC was determined after incubation at 37 °C for 16 h under aerobic conditions. The optical density was measured in triple at 600 nm (OD_{600}) using a microplate reader (Bio-Rad, USA) at 20 h after treatment of the chemicals in concentrations of 2, 4, 8, 16, 32, 64, 128, 256, 512, and 1024 µg/mL. The average and standard deviation values of OD_{600} are reported in Table S1 and Table S2 of the Supporting Information. Vancomycin and Erythromycin were used as positive controls (see Table S2 for the average and standard deviation values of OD_{600}). As the chemicals were dissolved in 100% DMSO, 100% DMSO and triple-distilled water were used as negative controls. The minimum inhibitory concentration (MIC) in Tables 2 and 4 is defined as the lowest concentration of chemicals which provides an average OD_{600} value of less than 0.100.

4. Conclusions

We extended the synthetic utility of pyrrole platform chemicals **2**, which can be readily prepared from the sustainable ribose conversion with amino acids, to the pyrrole-ligated 1,3,4-oxadiazole core structure **1** through the reaction with benzohydrazide **3a**. The size effect of the R group from the amino acids clearly offered better antibacterial activities for 1,3,4-oxadiazoles **1c** and **1e**, which were derived from valine and isoleucine, respectively. Benzohydrazides **3** with various electronic X-substituents were utilized for the construction of 2-pyrrolyl-5-phenyl-1,3,4-oxadiazoles **5** and **6** with *N*-valine and *N*-isoleucine residues, respectively. Relationships of structure and antibacterial activity were deduced from MIC values for 1,3,4-oxadiazoles **5** and **6** against *E. coli, S. aureus* and *A. baumannii*. A positive *ortho* effect was marginally observed for fluoride substituents. Most importantly, a superior iodophenol effect was evident in the antibacterial activities of 1,3,4-oxadiazoles **5f-1** and **6f-1**, which provided much lower MIC values against *A. baumannii* than those of the vancomycin and erythromycin as positive controls. These findings provide a guiding principle for the design of superior future antimicrobial agents.

Supplementary Materials: The following supporting information can be downloaded at: https://www. mdpi.com/article/10.3390/molecules28083638/s1, (1) ¹H/¹³C-NMR spectra; (2) MIC data against *E. coli, S. aureus*, and *A. baumannii*; (3) High-Resolution Mass Spectra for the entire 1,3,4-oxadiazoles synthesized in this paper. Table S1. Determination of MIC for 1 for *E. coli* and *S. aureus* by OD600 (20 h). Each value was obtained as an average of at least triple measurements. Table S2. Determination of MIC for 5 and 6 for *E. coli, S. aureus*, and *A. baumannii* by OD600 (20 h). Each value was obtained as an average of at least triple measurements. Table S2. Determination of MIC for 5 and 6 for *E. coli, S. aureus*, and *A. baumannii* by OD600 (20 h). Each value was obtained as an average of at least triple measurements.

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