

Supplementary Materials

Novel Pyridothienopyrimidine Derivatives: Design, Synthesis and Biological Evaluation as Antimicrobial and Anticancer Agents

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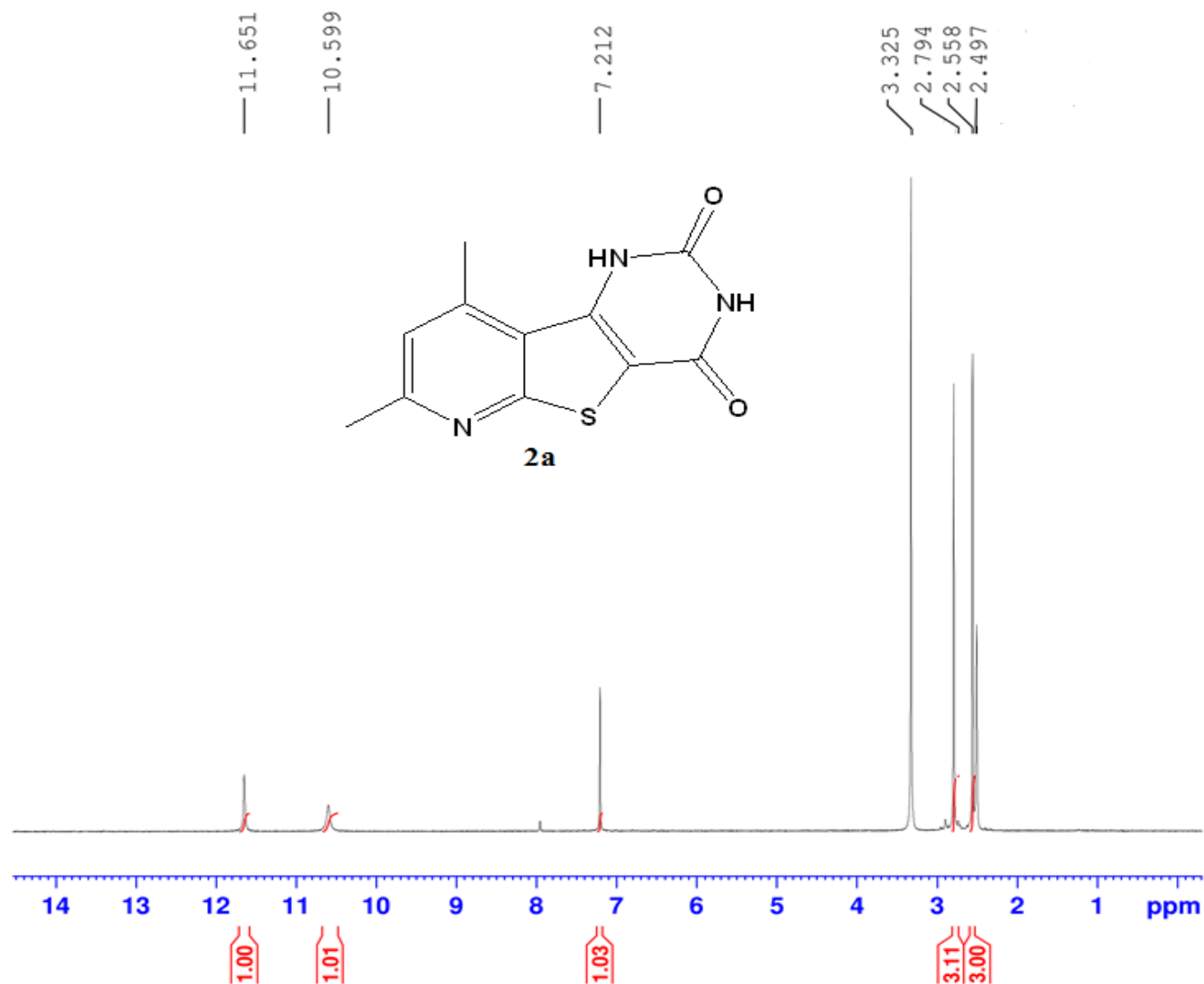
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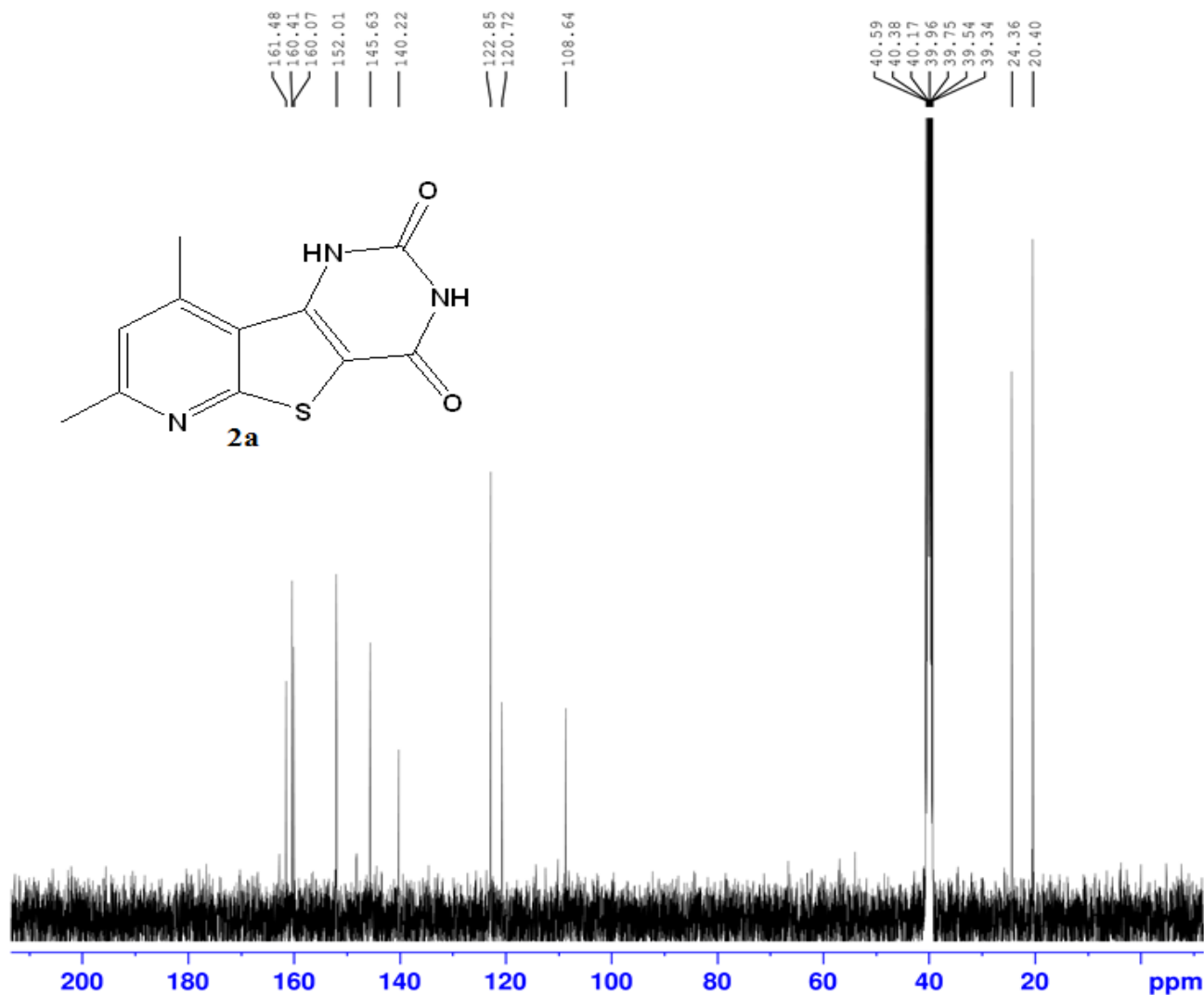


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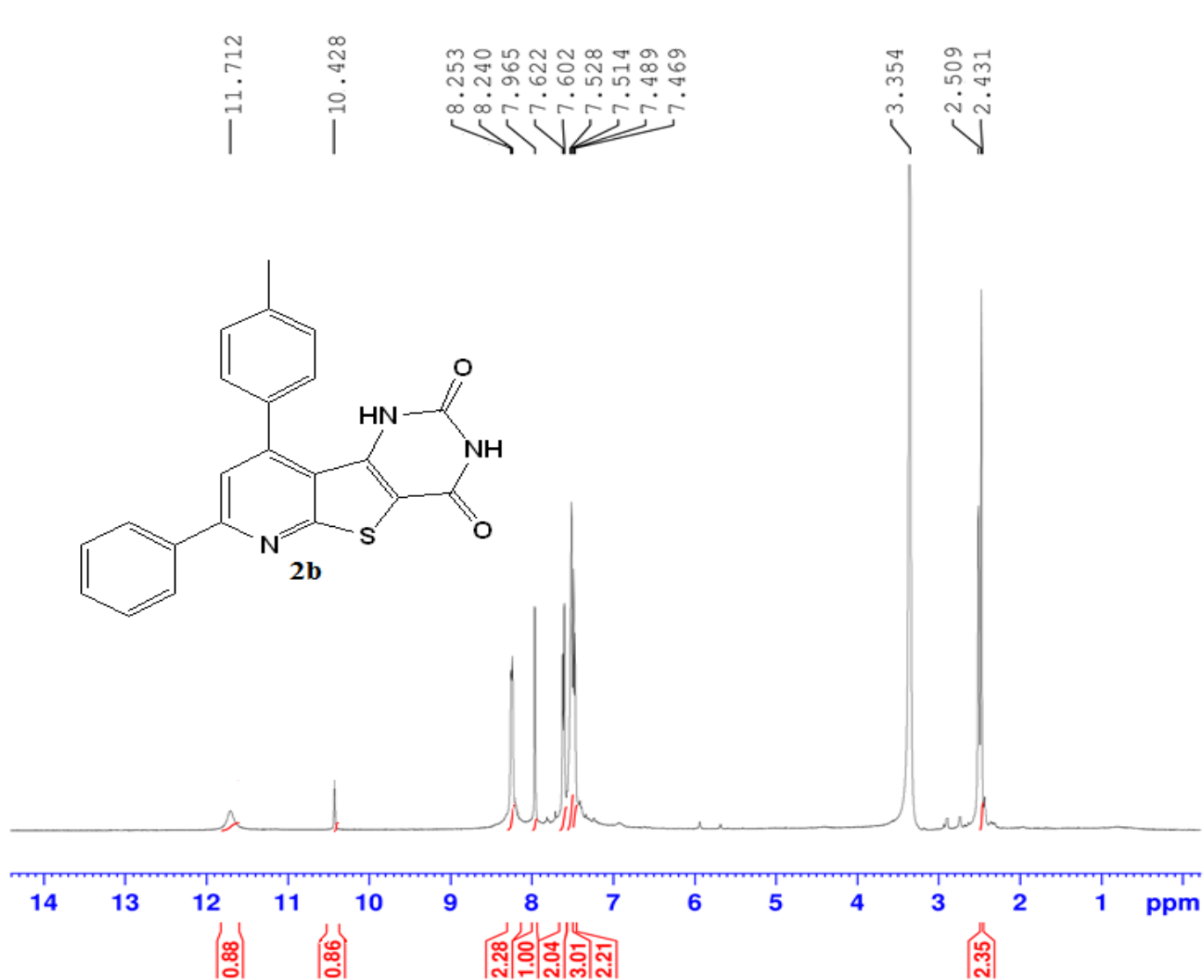
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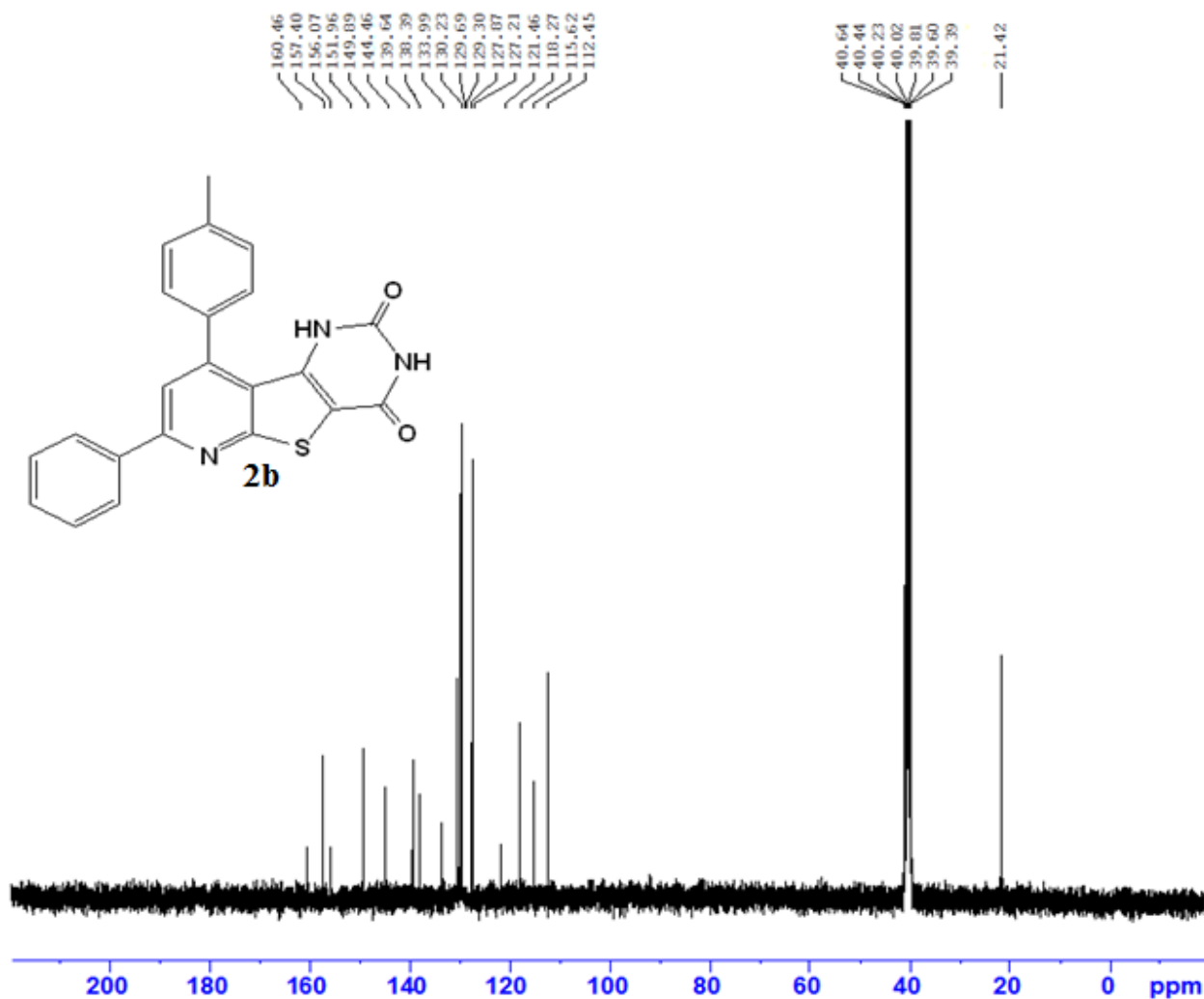


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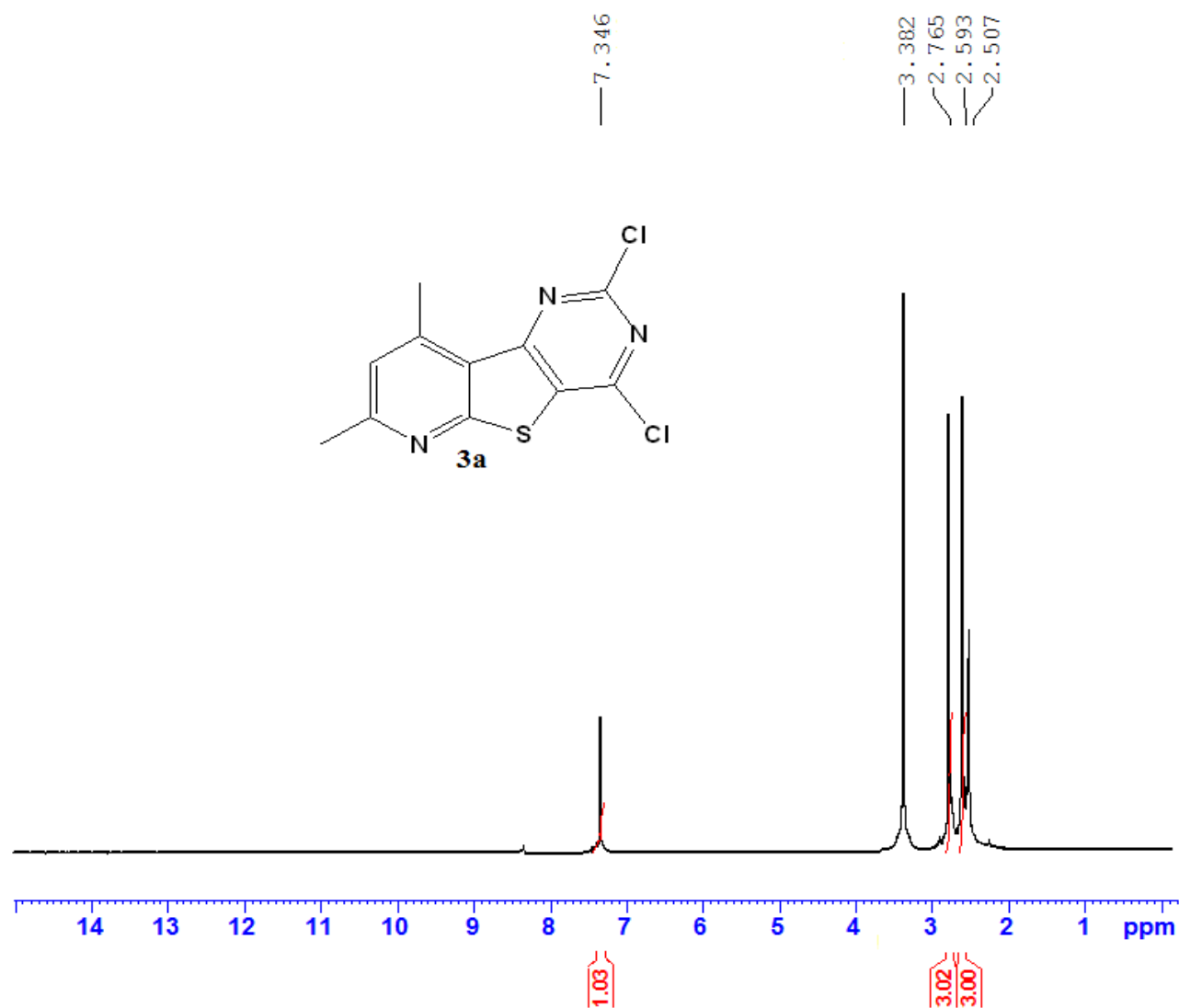
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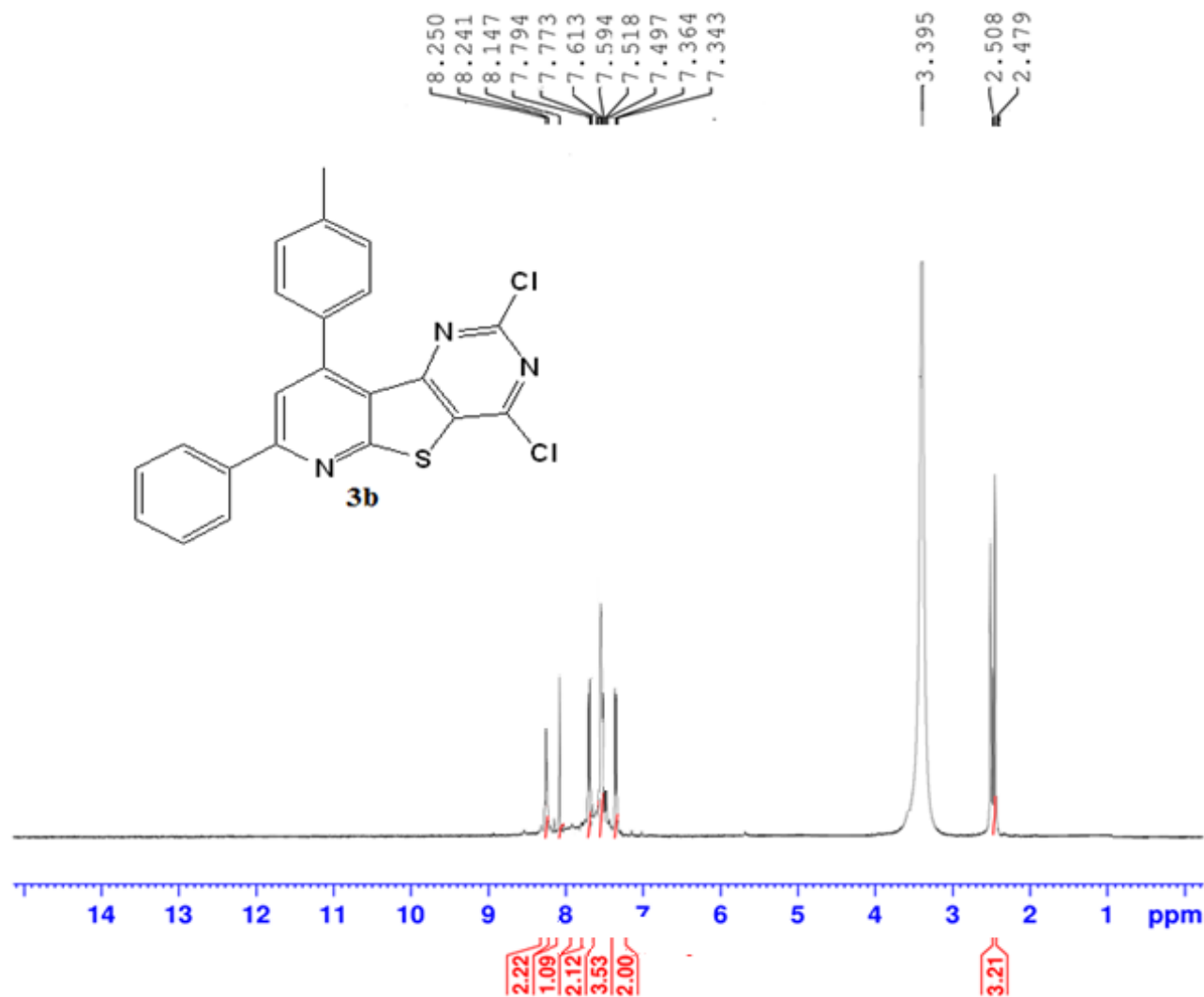


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 SOLVENT DMSO
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 FIDRES 0.122266 Hz
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 DE 6.50 usec
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 TD0 1

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 NUC1 1H
 P1 12.00 usec
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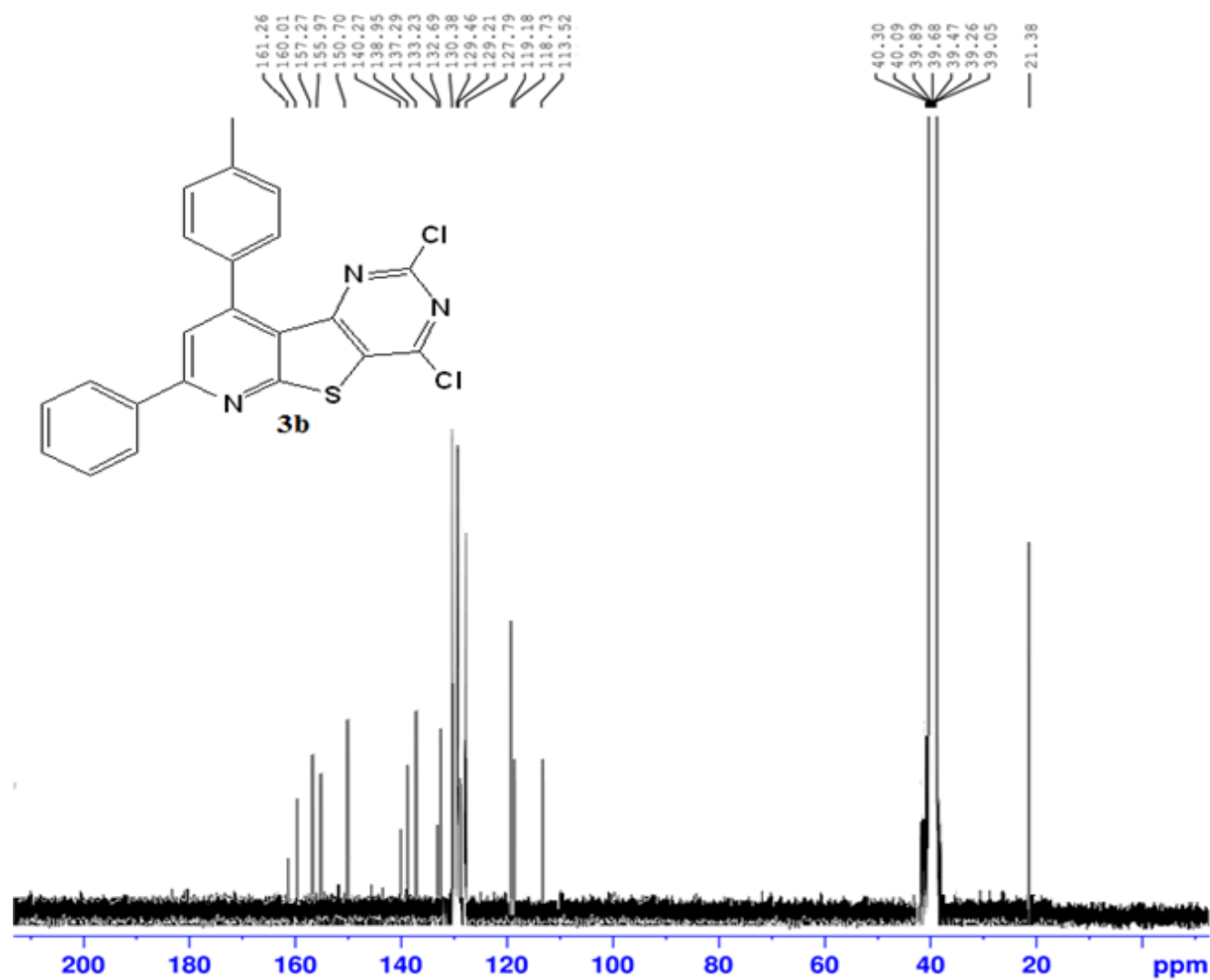


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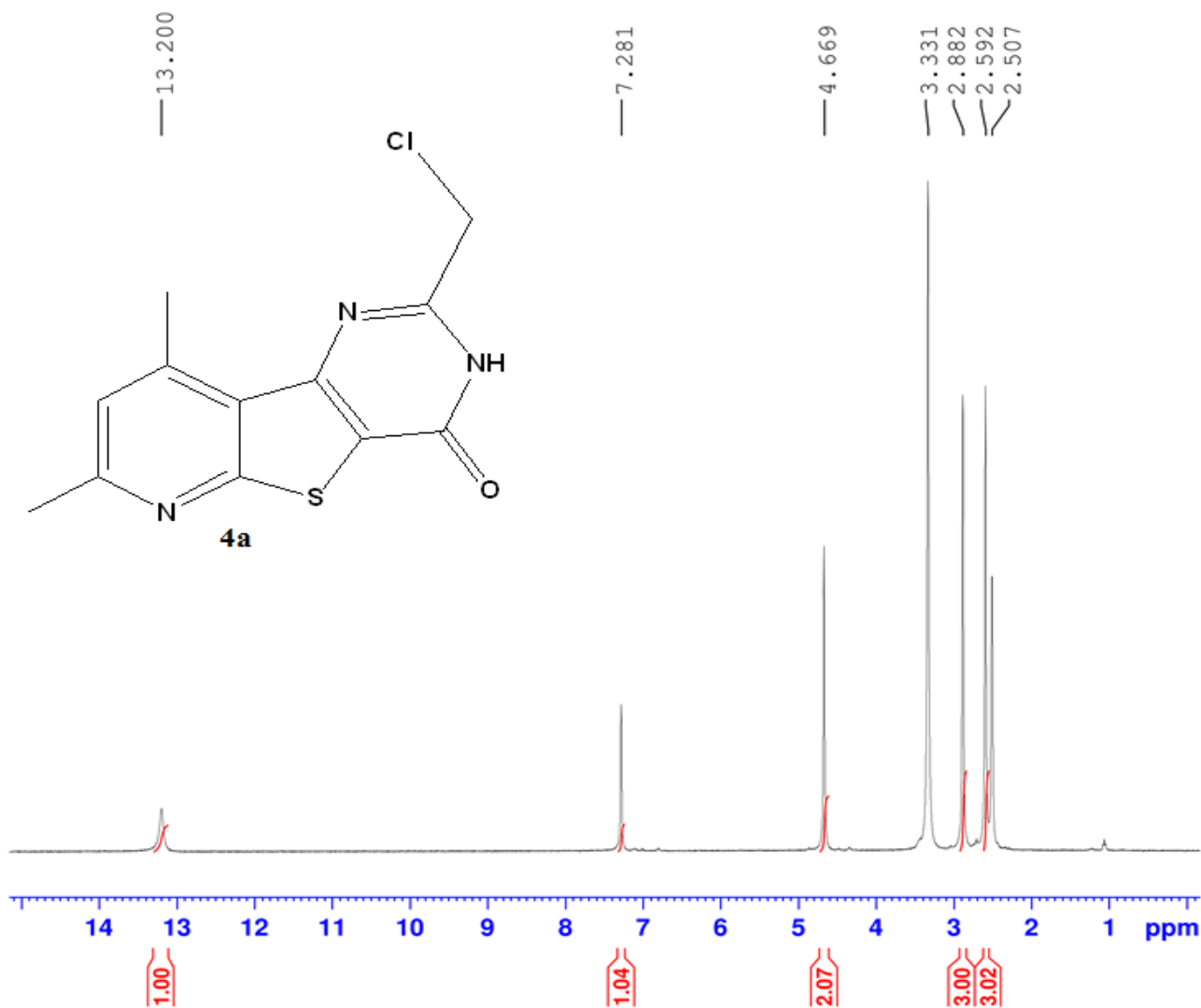
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NS 19000
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DW 20.800 usec
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TE 300.0 K
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d11 0.03000000 sec
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NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 18.00000000 W
PLW12 0.34722000 W
PLW13 0.28125000 W

F2 - Processing parameters
SI 32768
SF 100.6177975 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

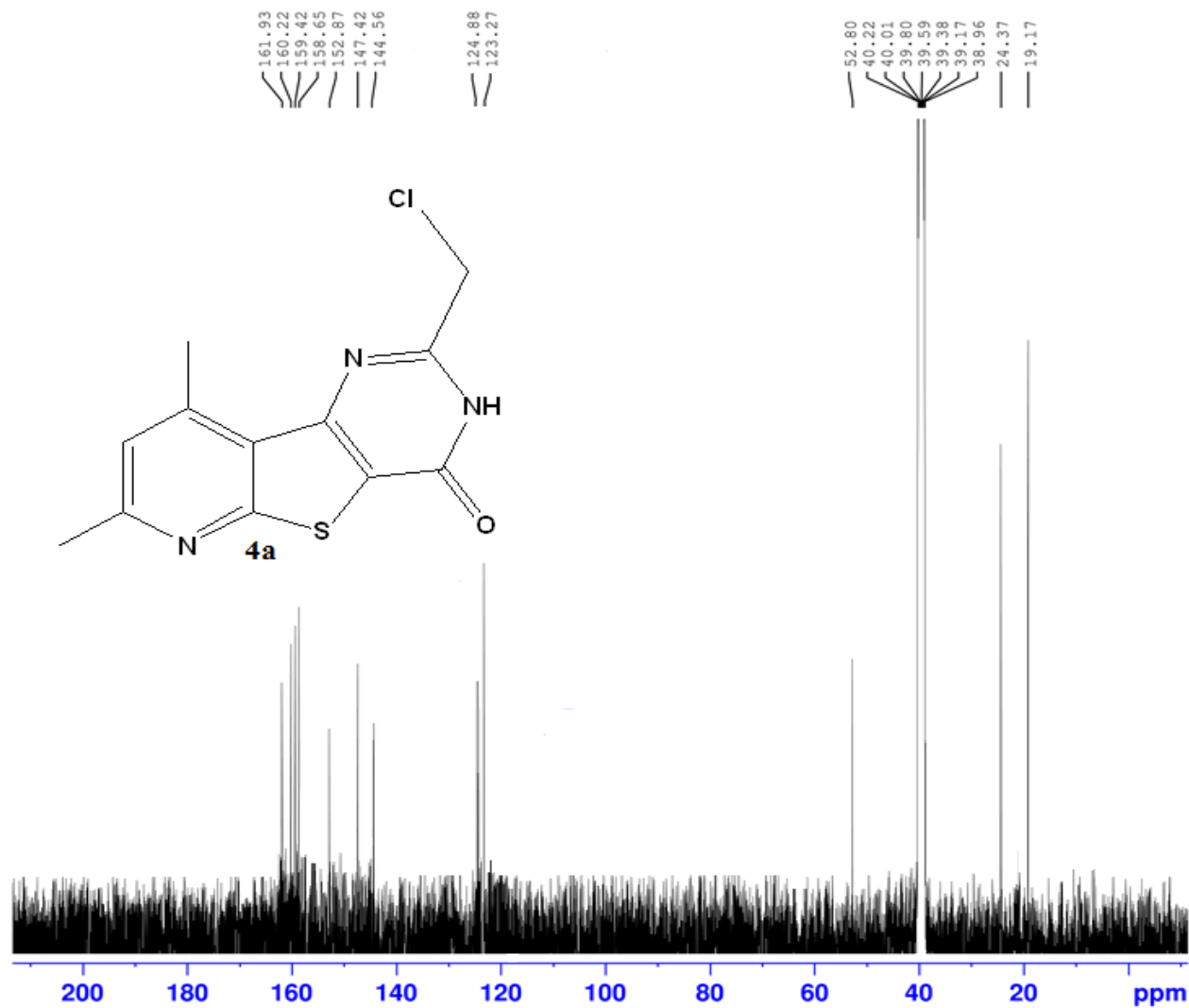


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 PULPROG zg30
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 SOLVENT DMSO
 NS 56
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

F2 - Processing parameters
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 SF 400.1500000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



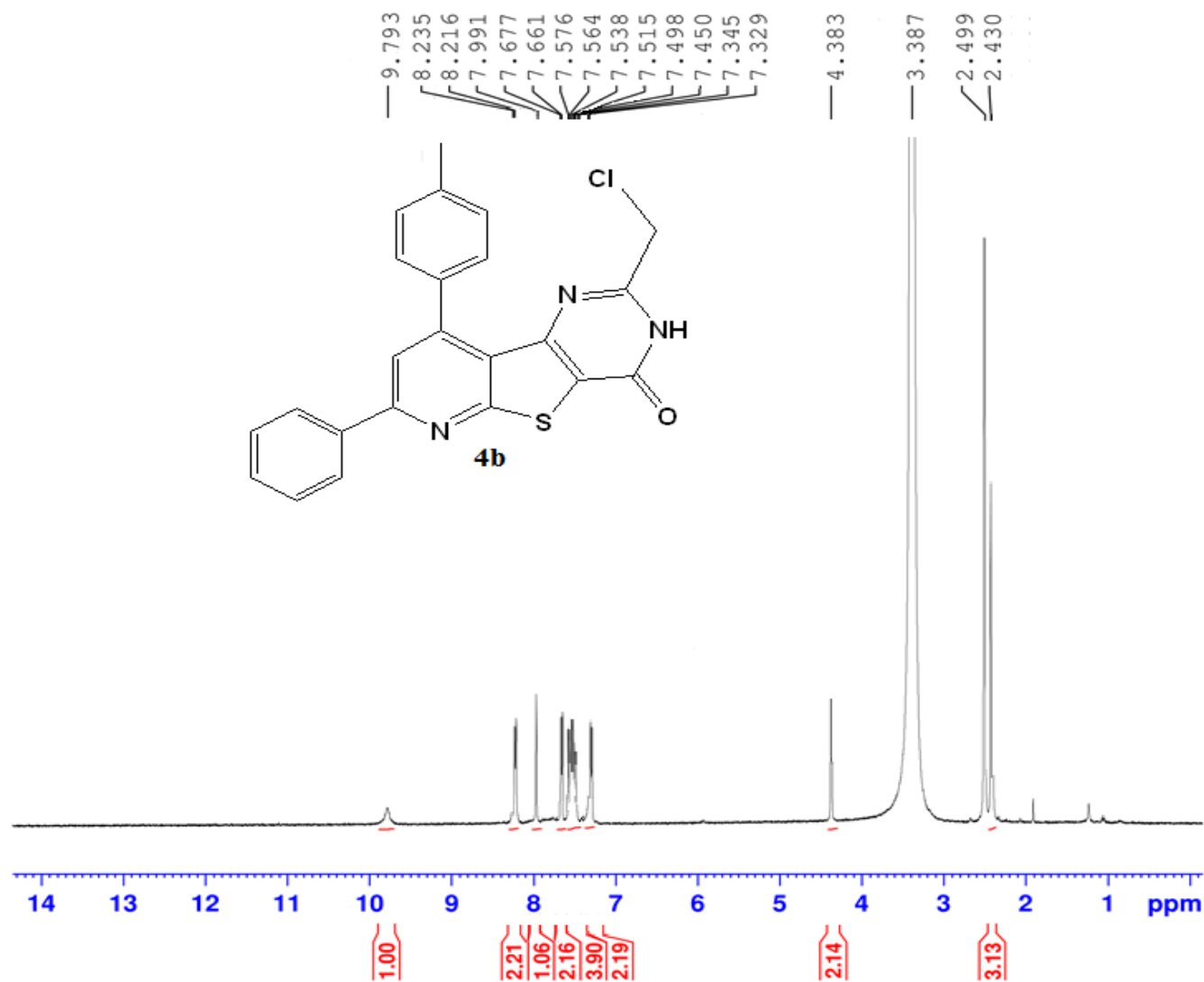
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 SOLVENT DMSO
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 FIDRES 0.366798 Hz
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 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
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 NUC1 13C
 P1 10.00 usec
 PLW1 47.00000000 W

===== CHANNEL f2 =====
 SFO2 400.1516006 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 90.00 usec
 PLW2 18.00000000 W
 PLW12 0.34722000 W
 PLW13 0.28125000 W

F2 - Processing parameters
 SI 32768
 SF 100.6177975 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

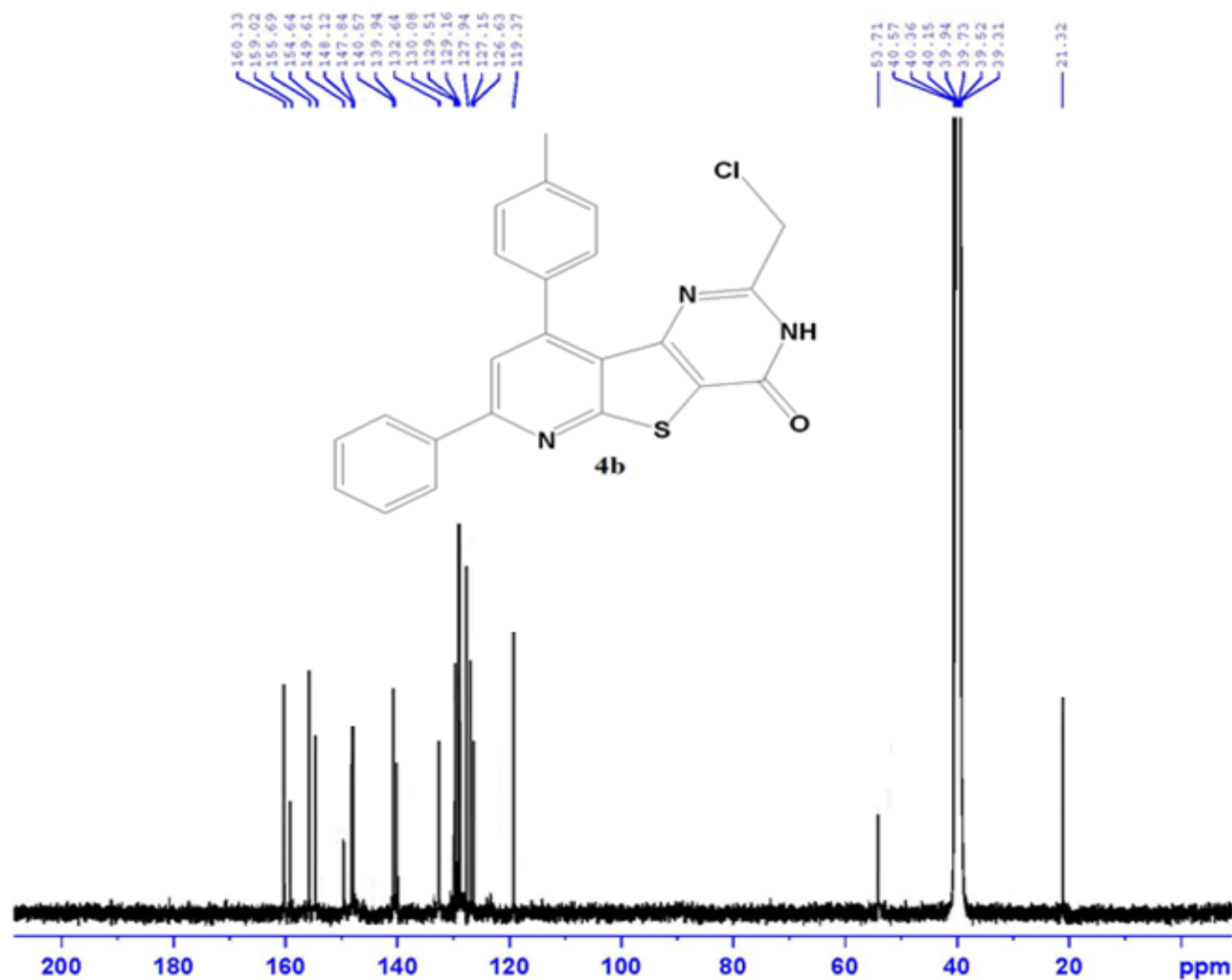


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 SOLVENT DMSO
 NS 91
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 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

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 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

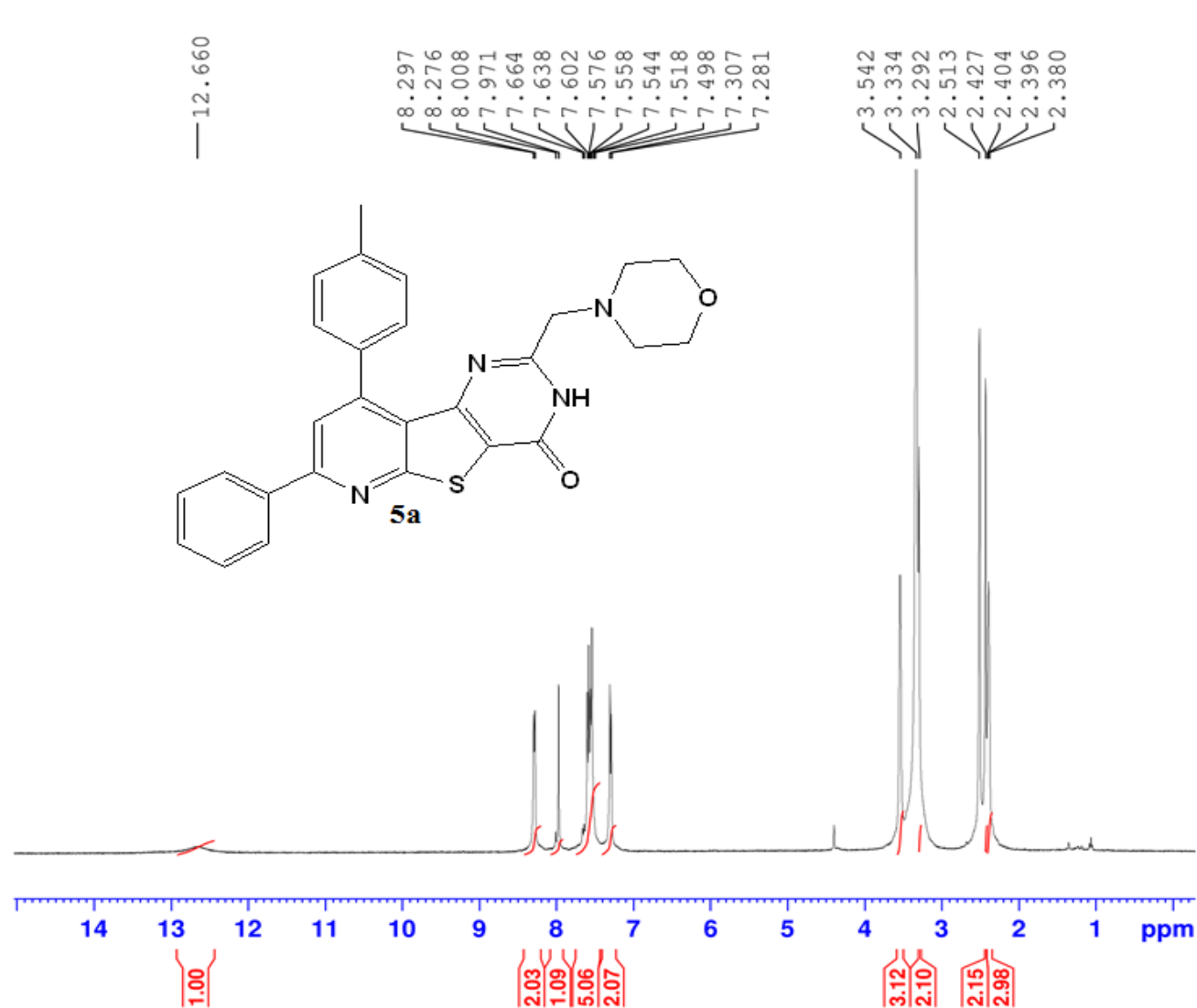
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EXPNO 10
PROCNO 1

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Time 20.12
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TD 65536
SOLVENT DMSO
NS 2130
DS 4
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FIDRES 0.713596 Hz
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RG 197.77
CW 20.800 usec
DE 6.50 usec
TE 293.2 K
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D11 0.02000000 sec
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NUC1 13C
P1 10.00 usec
PLM1 47.00000000 N
SFO2 400.2016038 MHz
NUC2 1H
CPDPRG2 waltz16
PCPD2 90.00 usec
PLM2 13.00000000 N
PLM12 0.29249999 N
PLM13 0.14713000 N

F2 - Processing parameters
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GB 0
PC 1.40

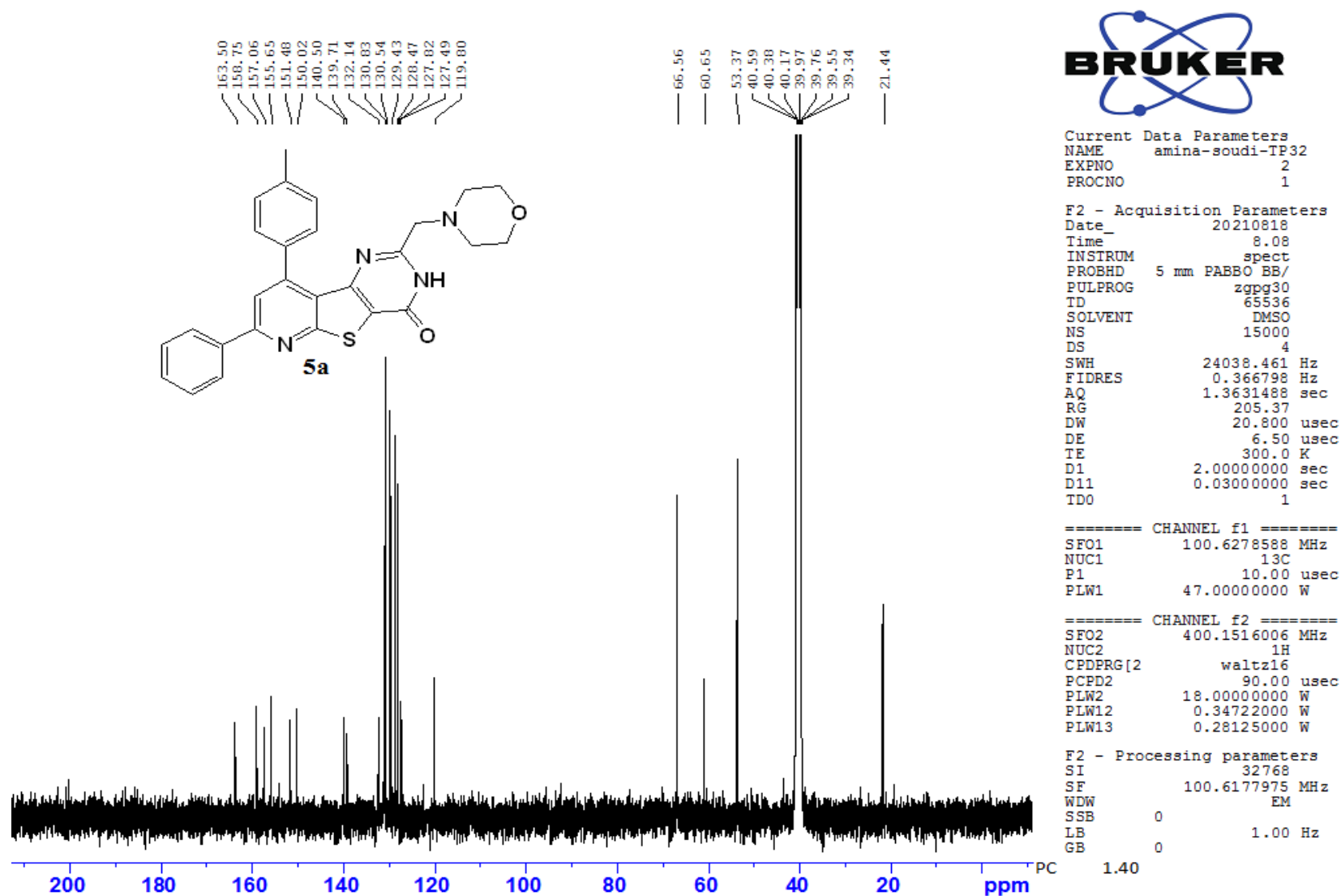


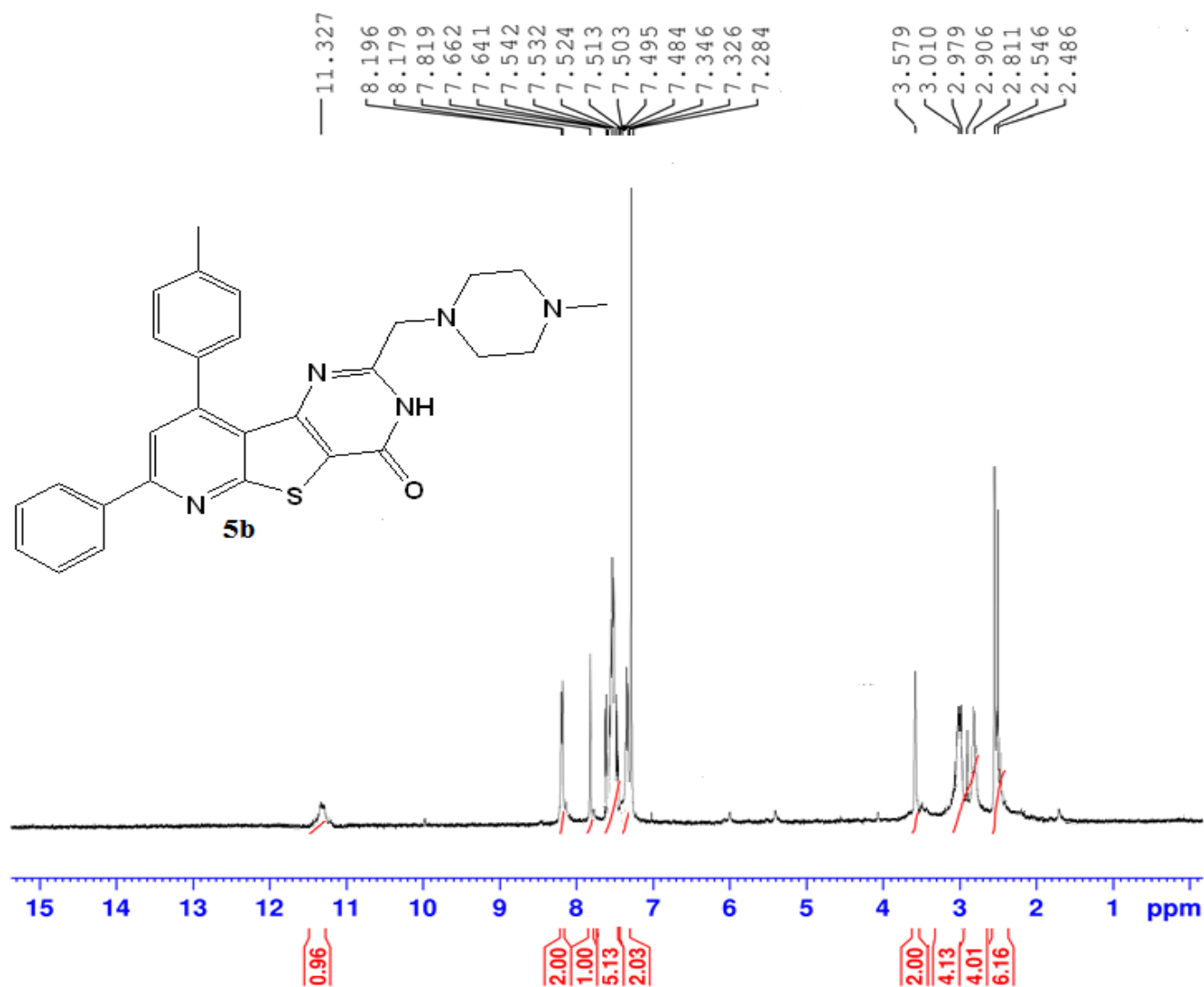
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PULPROG zg30
TD 65536
SOLVENT DMSO
NS 128
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 205.37
DW 62.400 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 1H
P1 12.00 usec
PLW1 18.00000000 W

F2 - Processing parameters
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SF 400.1500000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



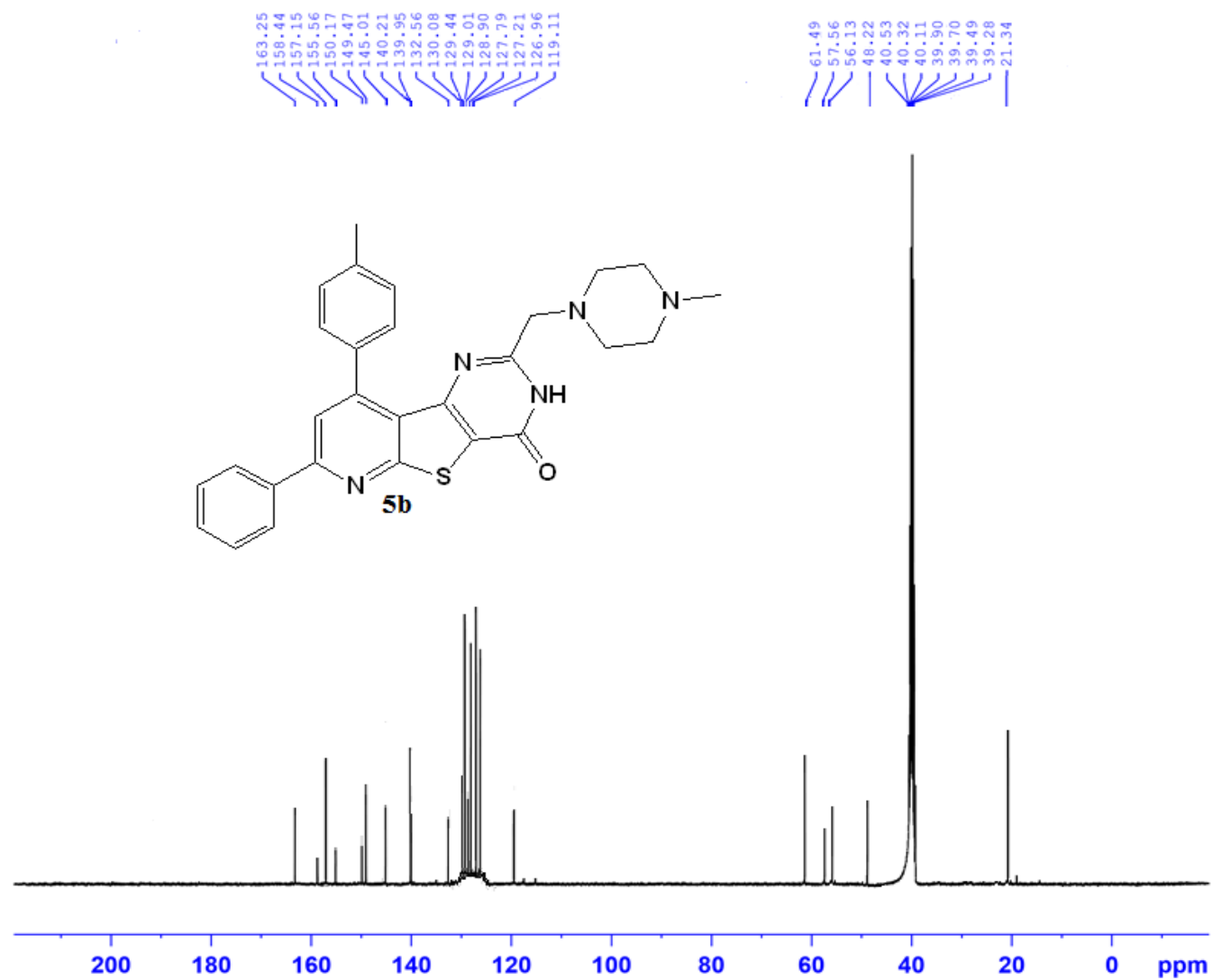


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 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 93
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

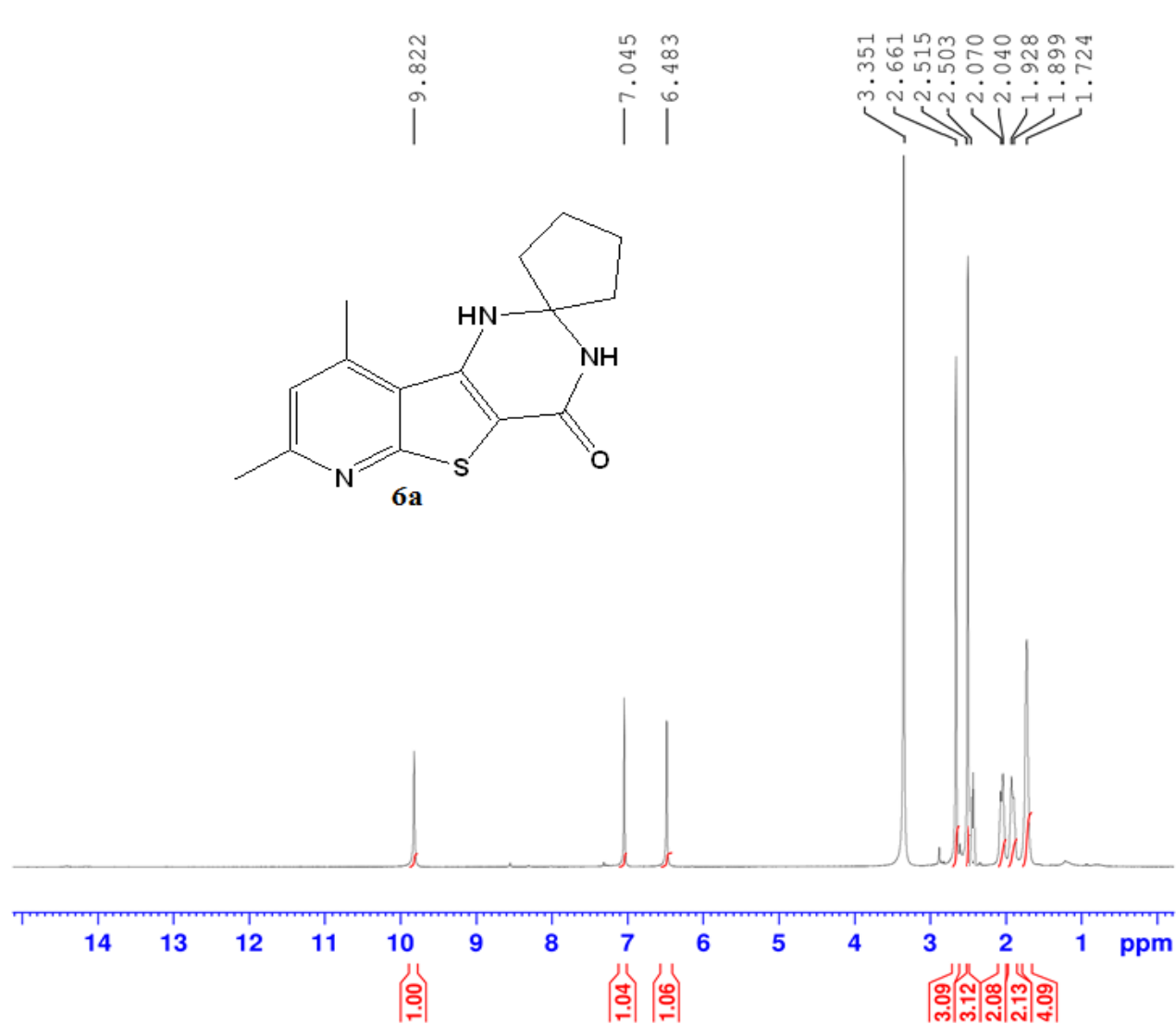
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Current Data Parameters
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EXPNO 10
PROCNO 1

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SOLVENT DMSO
NS 1024
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FIDRES 0.733596 Hz
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RG 197.77
DW 20.800 use
DE 6.50 use
TE 295.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6404331 MHz
NUC1 13C
P1 10.00 use
PLW1 47.00000000 W
SFO2 400.2016008 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 use
PLW2 13.00000000 W
PLW12 0.29249999 W
PLW13 0.14713000 W

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GB 0
PC 1.40

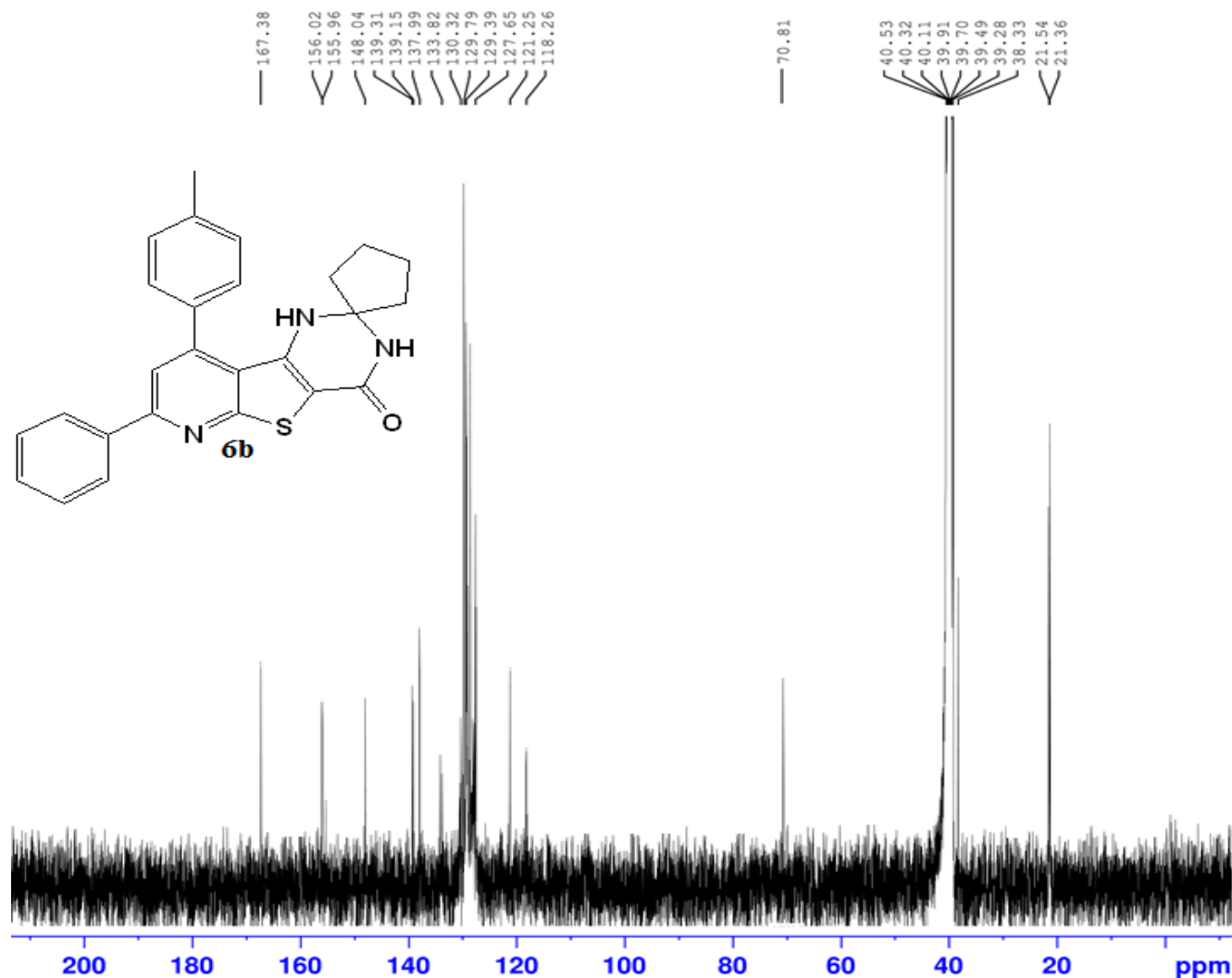


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 SOLVENT DMSO
 NS 54
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 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1500000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



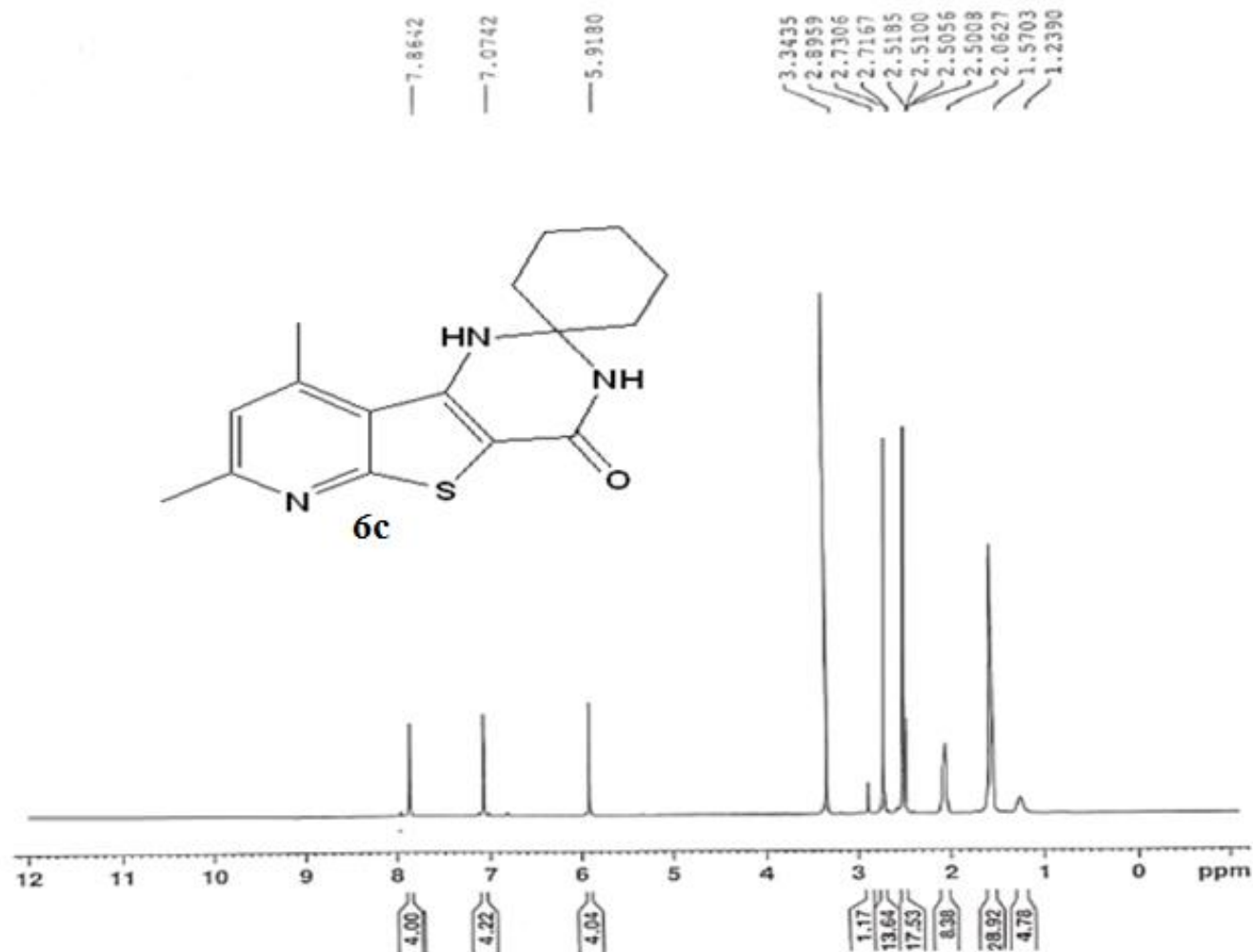
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RG 205.37
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 18.00000000 W
PLW12 0.34722000 W
PLW13 0.28125000 W

F2 - Processing parameters
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SF 100.6177975 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

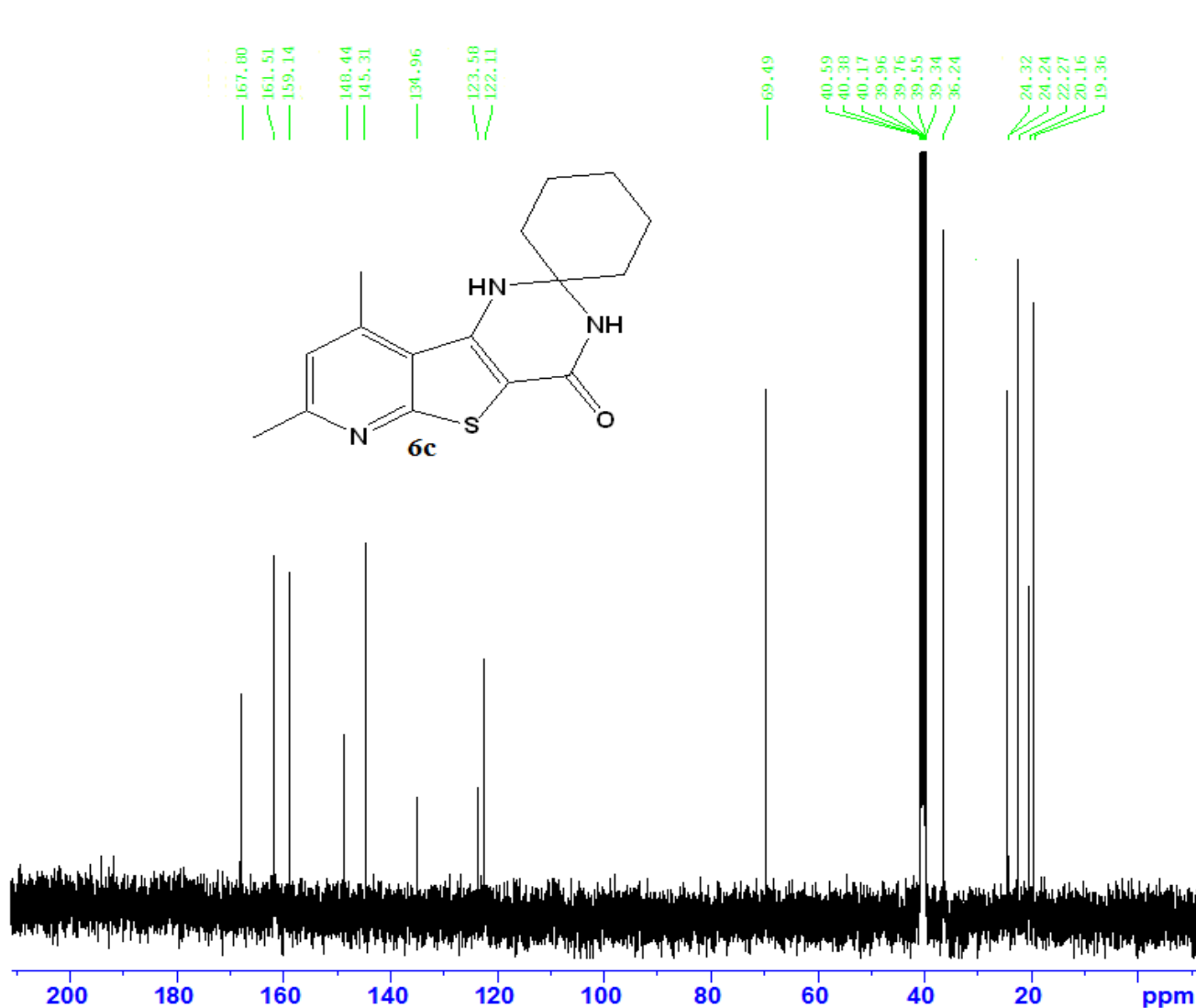


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 PULPROG zg30
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 SOLVENT DMSO
 NS 121
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 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

F2 - Processing parameters
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 SF 400.1500000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz



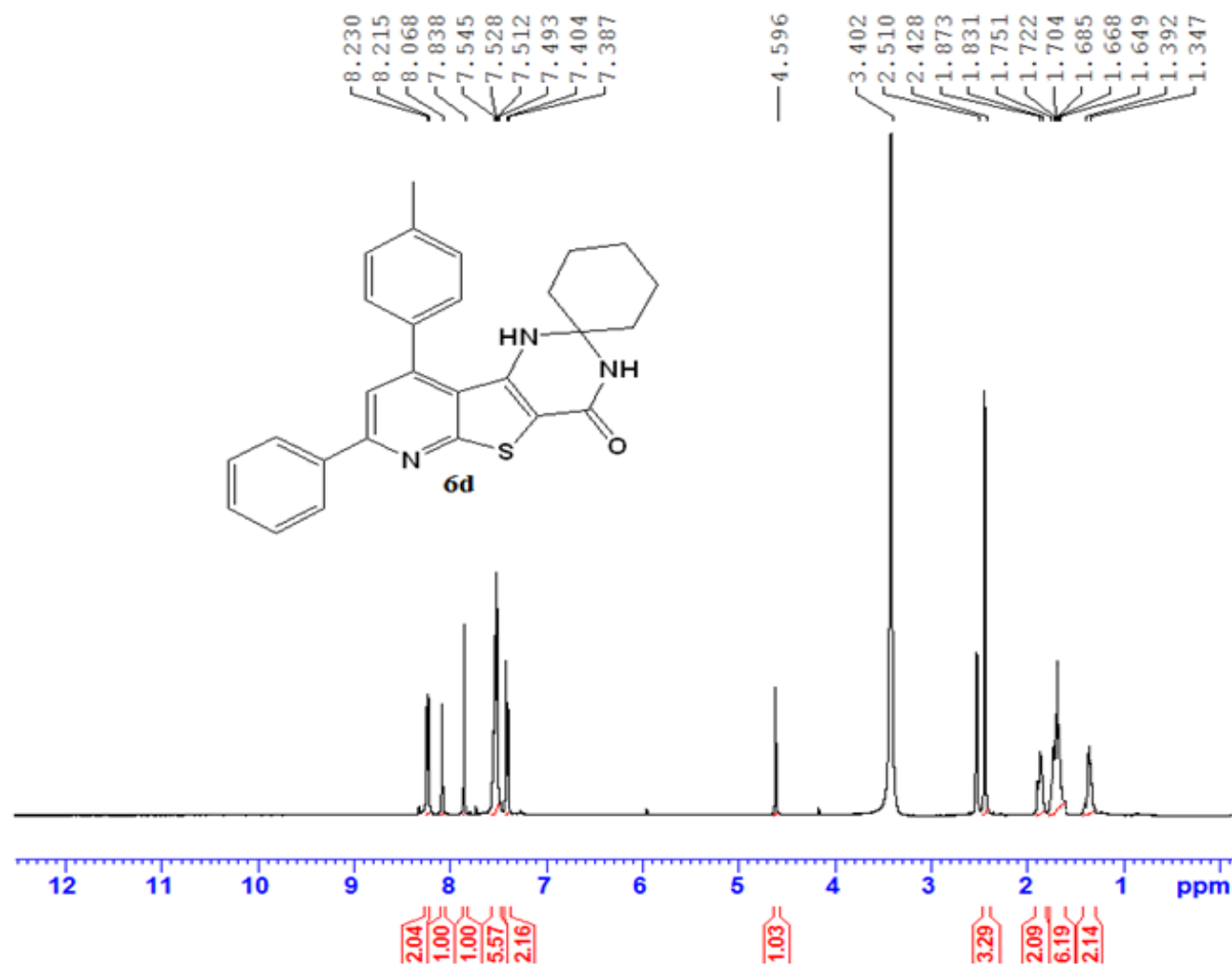
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PROCNO 1

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PULPROG zgpg30
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SOLVENT DMSO
NS 603
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SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 205.37
DW 20.800 usec
DE 6.50 usec
TE 299.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 18.00000000 W
PLW12 0.34722000 W
PLW13 0.28125000 W

F2 - Processing parameters
SI 32768
SF 100.6177975 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

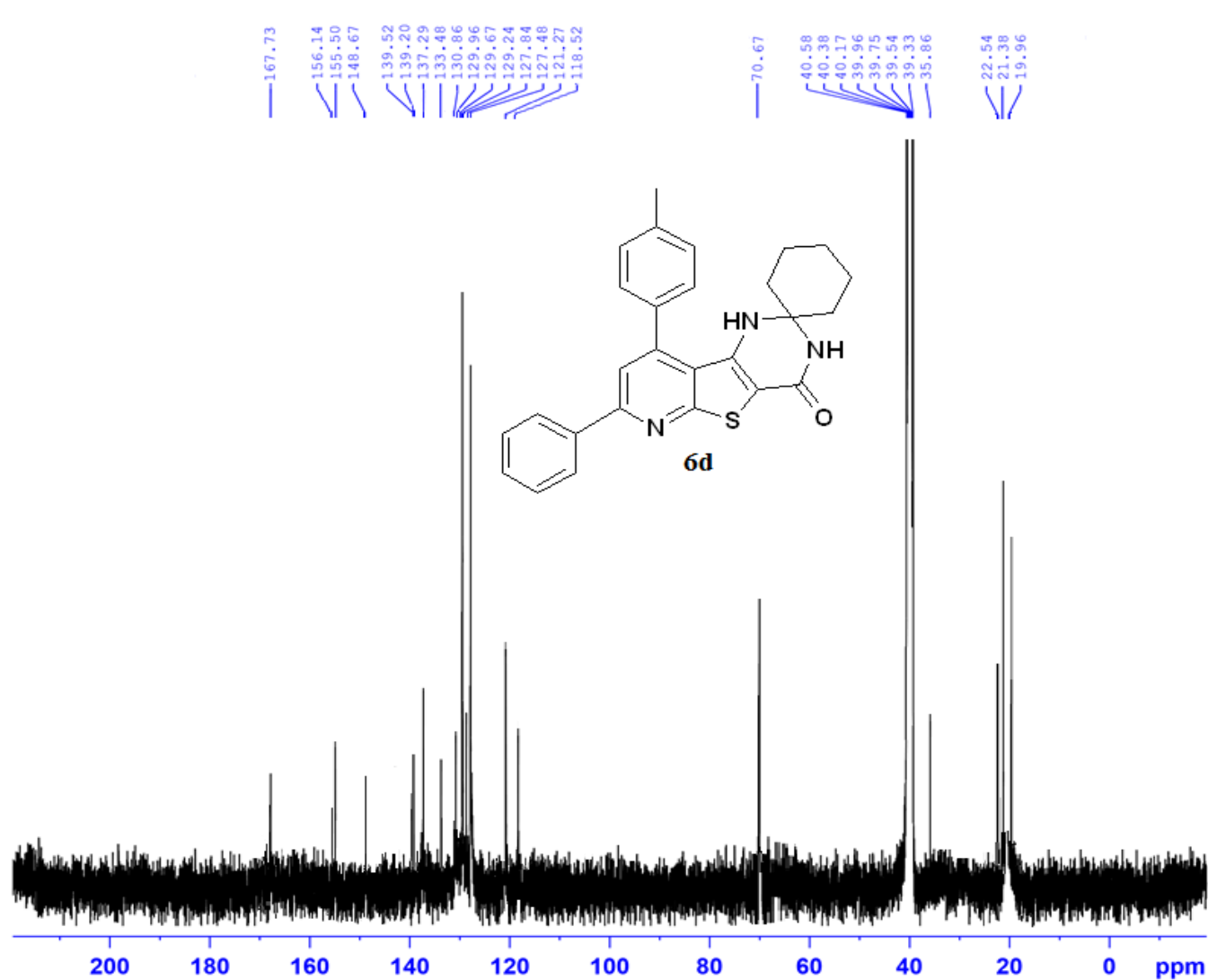


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 PROCNO 1

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 Time 11.17
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 PULPROG zg30
 ID 65536
 SOLVENT DMSO
 NS 64
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 298.0 K
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 TD0 1

===== CHANNEL f1 =====
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 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

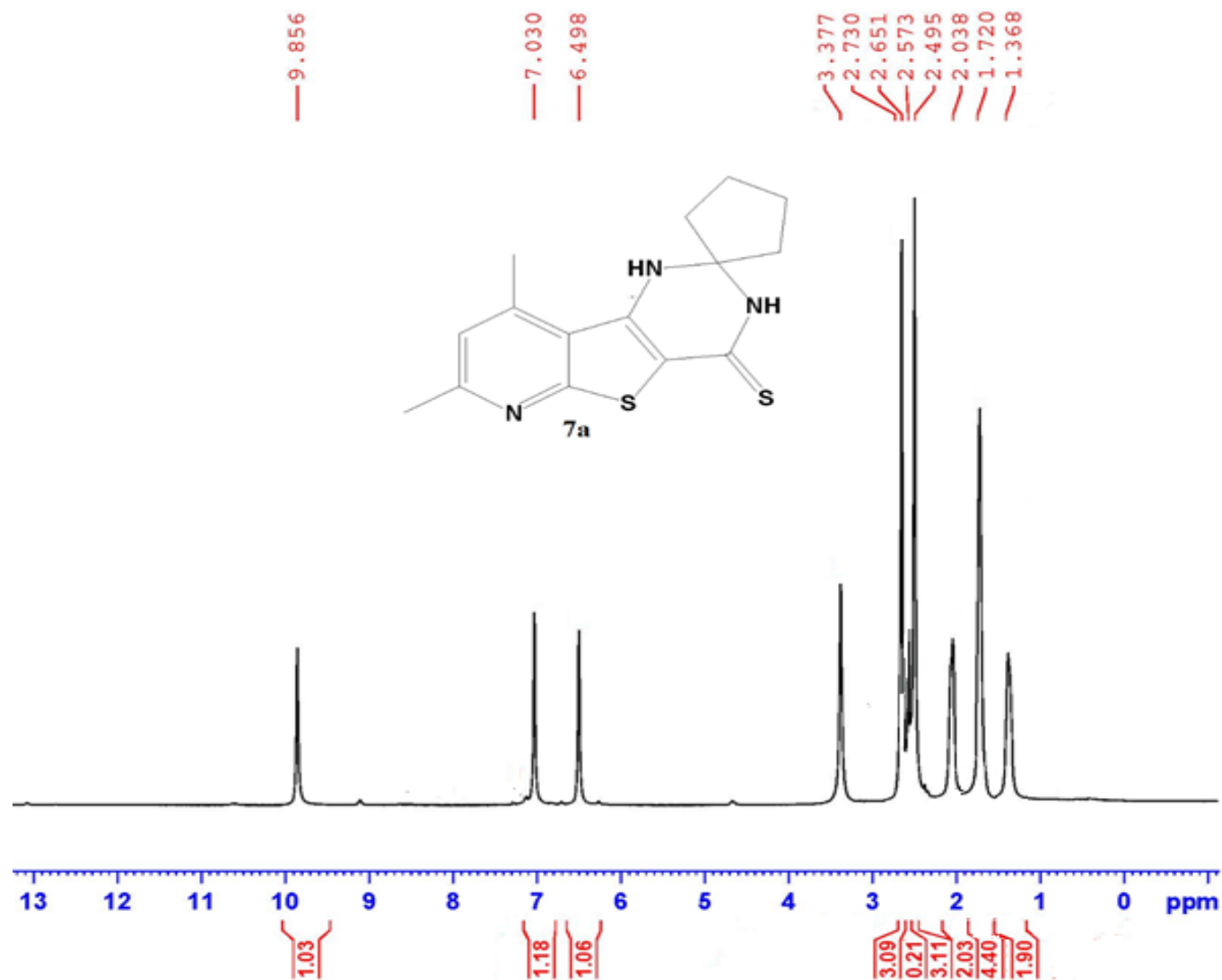
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 GB 0
 PC 1.00



Current Data Parameters
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EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20211209
Time 16.54 h
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PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 3000
DS 4
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 1.3631488 sec
RG 197.77
DW 20.800 usec
DE 6.50 usec
TE 294.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6404331 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W
SFO2 400.2016008 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 13.00000000 W
PLW12 0.29249999 W
PLW13 0.14713000 W

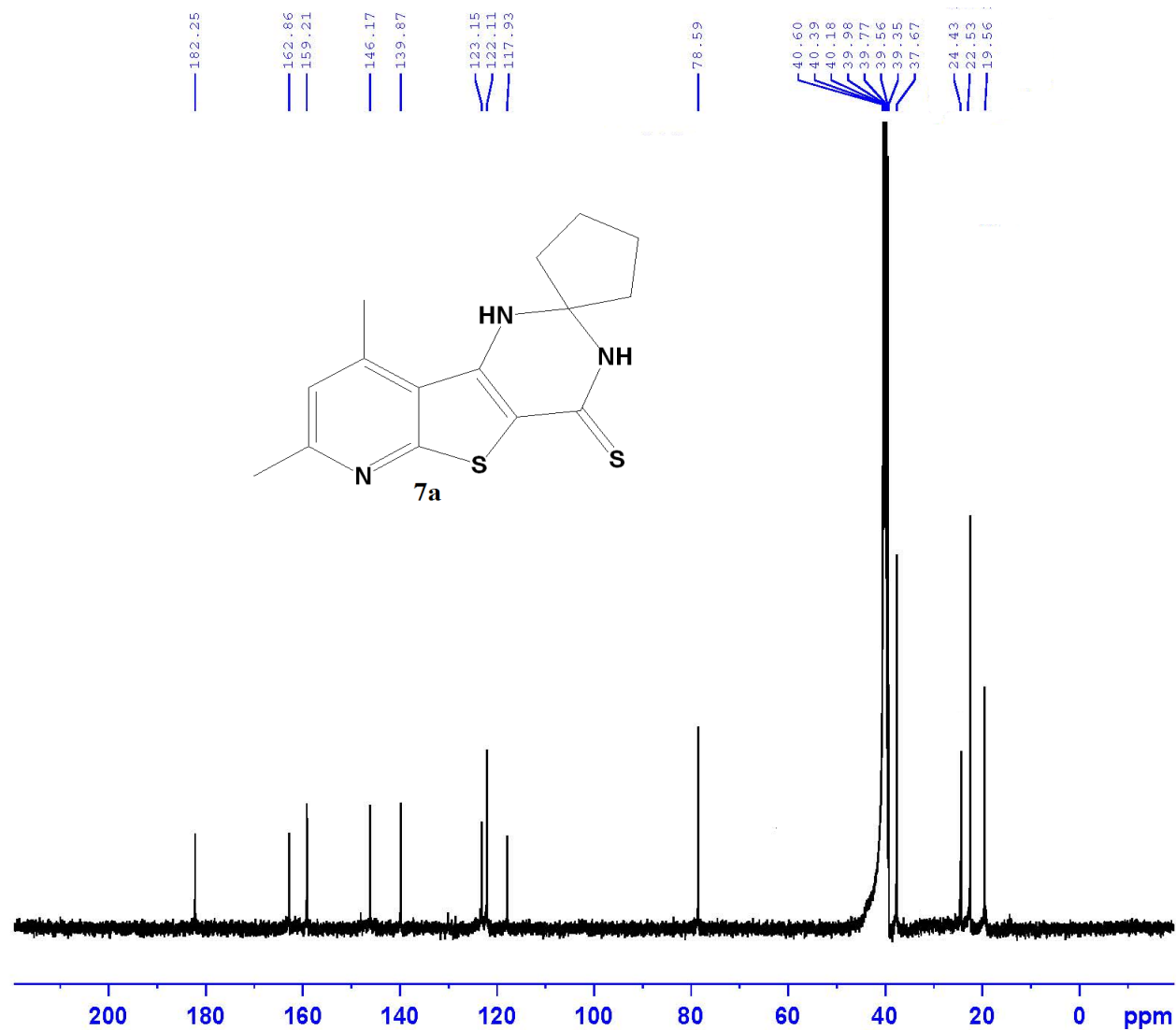
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SF 100.6303700 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



Current Data Parameters
NAME Omina Alsoudi-MP11-SA-
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190721
Time 11.02 h
INSTRUM spect
PROBHD Z108618_0945 (
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.244532 Hz
AQ 4.0894465 sec
RG 68.17
DQ 62.400 usec
DE 6.50 usec
TE 292.9 K
D1 1.00000000 sec
TD0 1
SFO1 400.2024712 MHz
NUC1 1H
P1 13.50 usec
PLW1 13.00000000 W

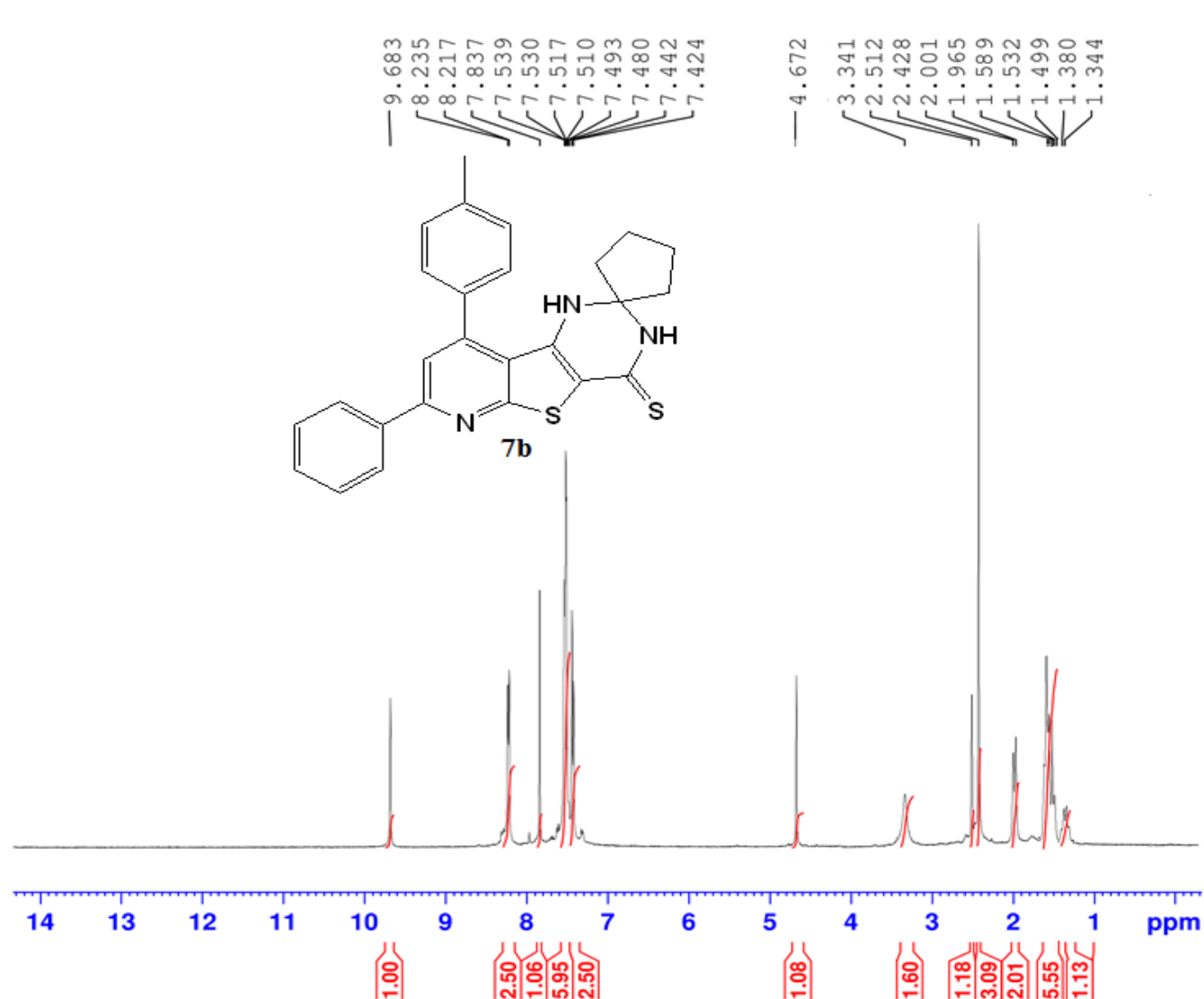
F2 - Processing parameters
SI 65536
SF 400.2000000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
 NAME Amina elsaudy-MP11-carbon-NM
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191111
 Time_ 0.34 h
 INSTRUM spect
 PROBHD Z108618 0945 (
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 2100
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.733596 Hz
 AQ 1.3631488 sec
 RG 197.77
 DW 20.800 usec
 DE 6.50 usec
 TE 297.8 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 100.6404331 MHz
 NUC1 13C
 P1 10.00 usec
 PLM1 47.00000000 W
 SFO2 400.2016009 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 90.00 usec
 PLW2 13.00000000 W
 PLW12 0.29249999 W
 PLW13 0.14713000 W

F2 - Processing parameters
 SI 32768
 SF 100.6303700 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

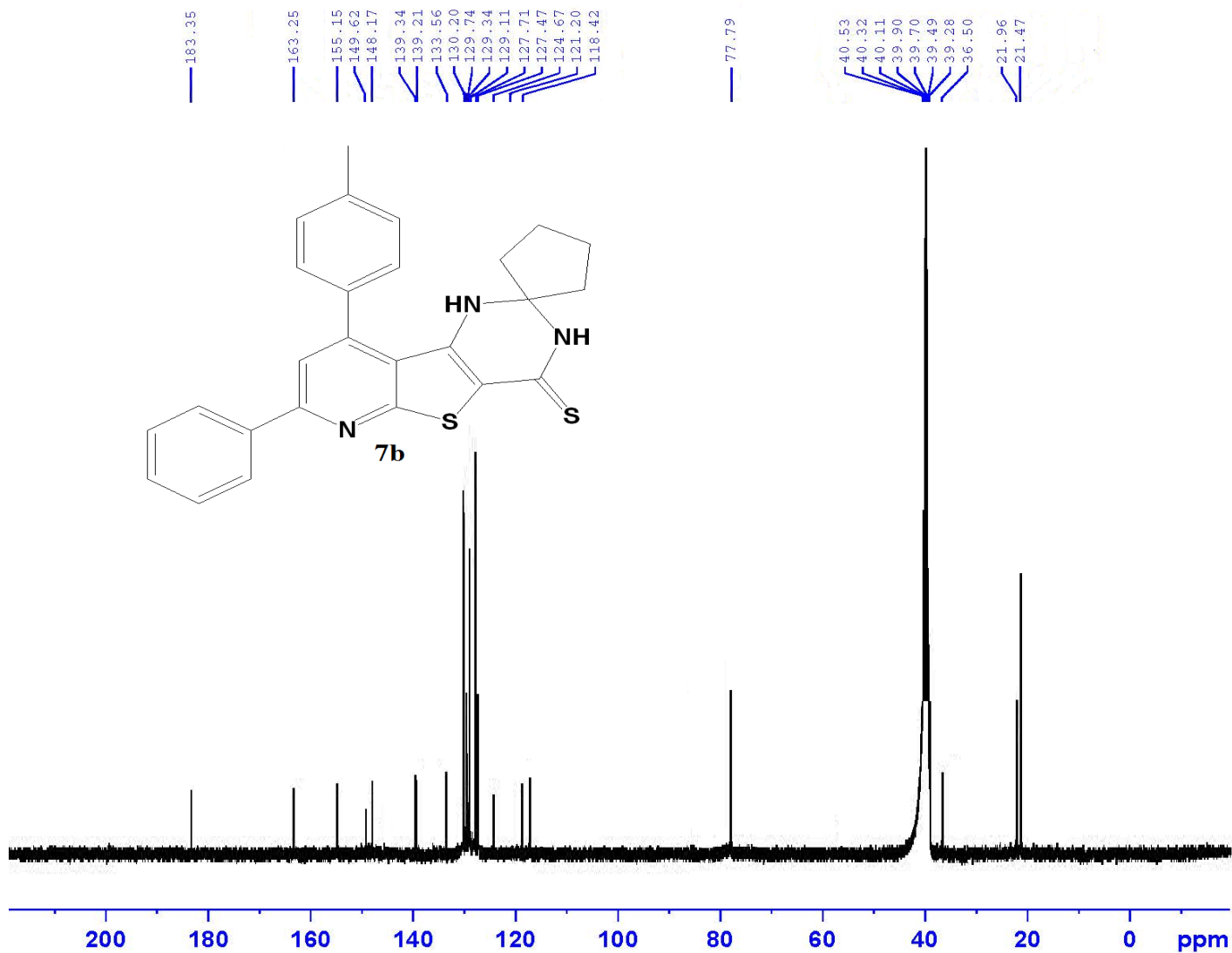


Current Data Parameters
 NAME amina-nasr-T.11
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210218
 Time 12.00
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 33
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

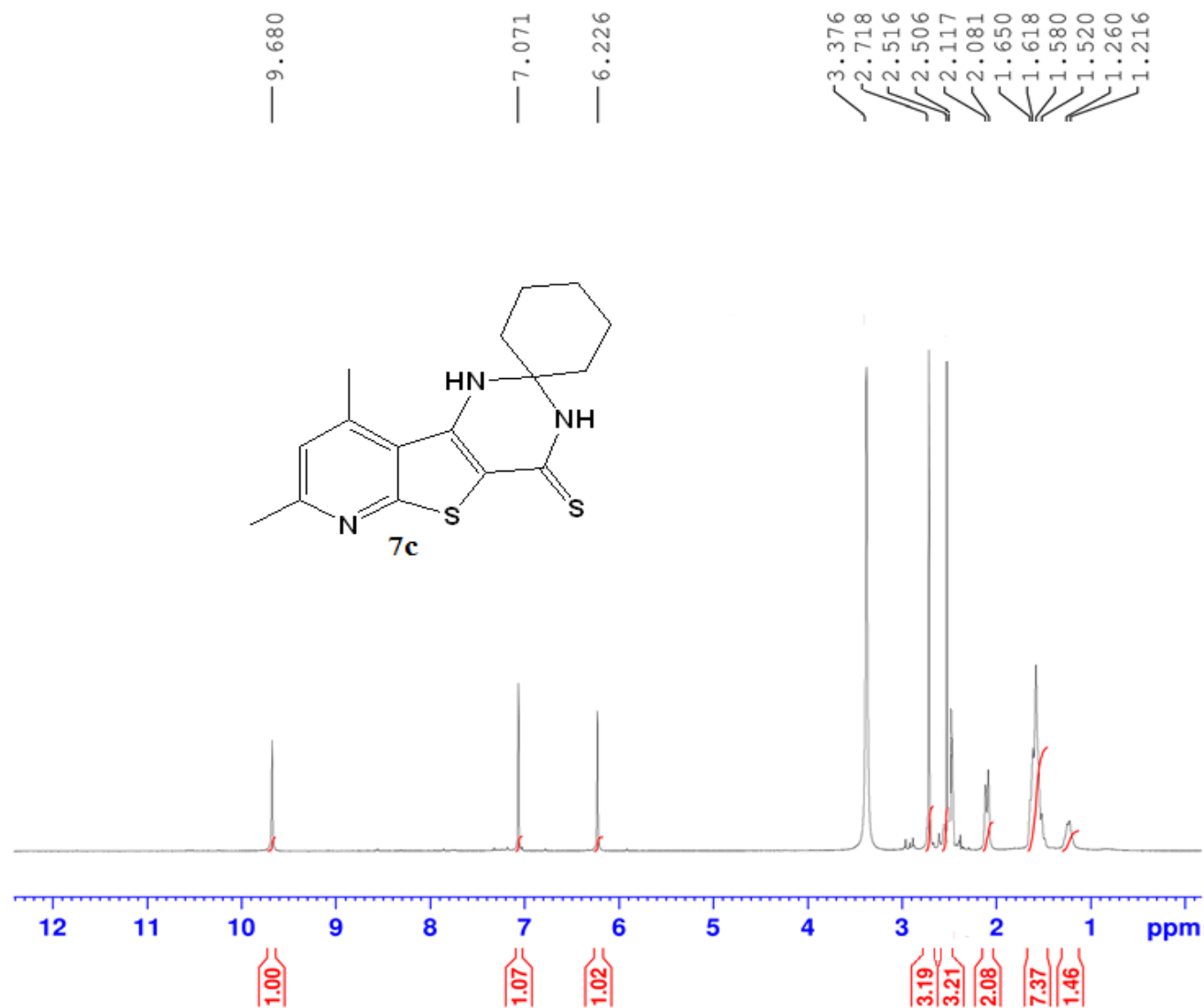
F2 - Processing parameters
 SI 65536
 SF 400.1500000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
NAME Eman-TPP11 -c13-R
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20211208
Time_ 12.14 h
INSTRUM spect
PROBHD Z108618_0945 (
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 1.3631488 sec
RG 197.77
DW 20.800 usec
DE 6.50 usec
TE 295.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6404331 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W
SFO2 400.2016008 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 13.00000000 W
PLW12 0.29249999 W
PLW13 0.14713000 W

F2 - Processing parameters
SI 32768
SF 100.6303700 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

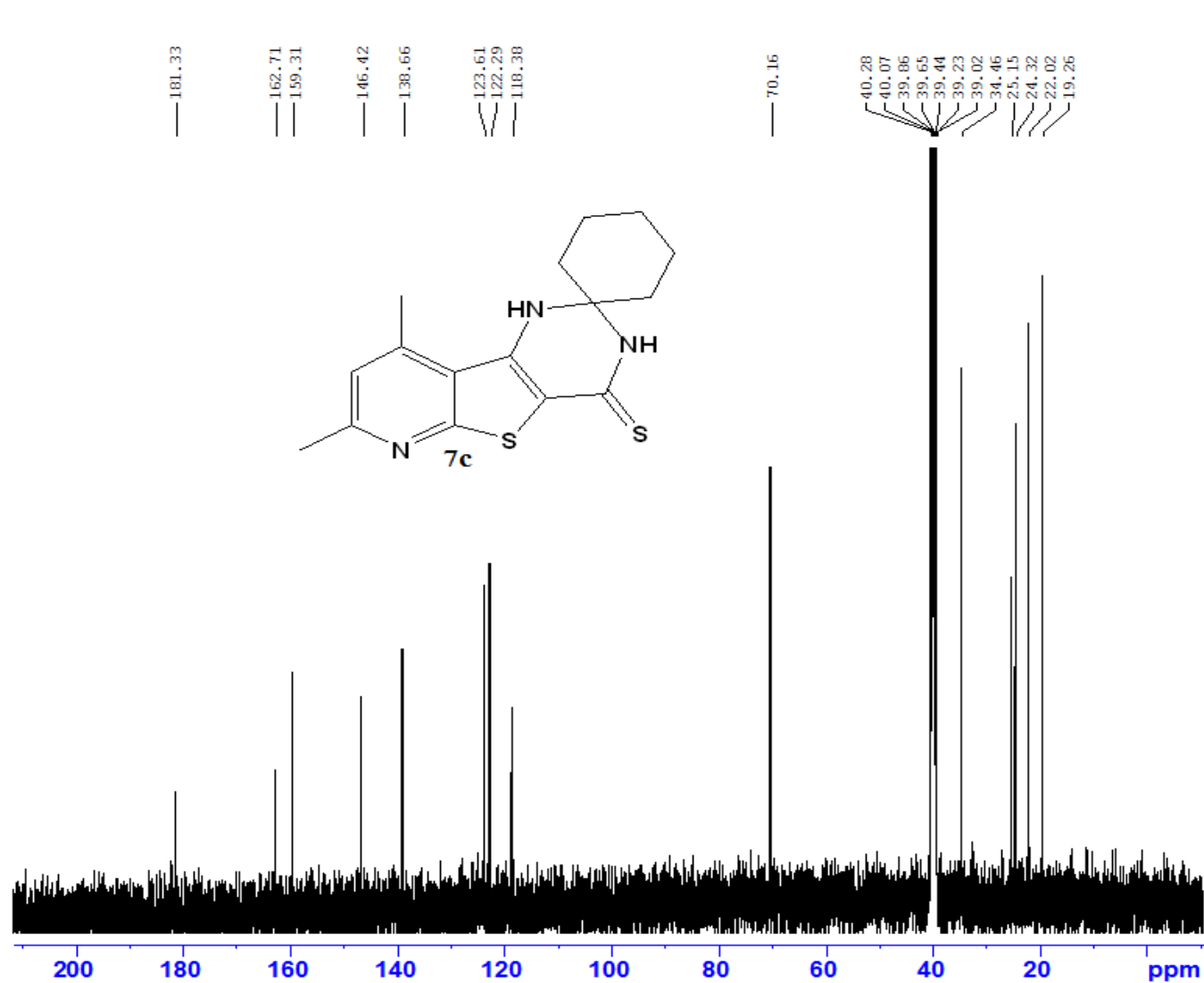


Current Data Parameters
 NAME amina-nasr-M4
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190103
 Time 10.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT DMSO
 NS 41
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894465 sec
 RG 205.37
 DW 62.400 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1500000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



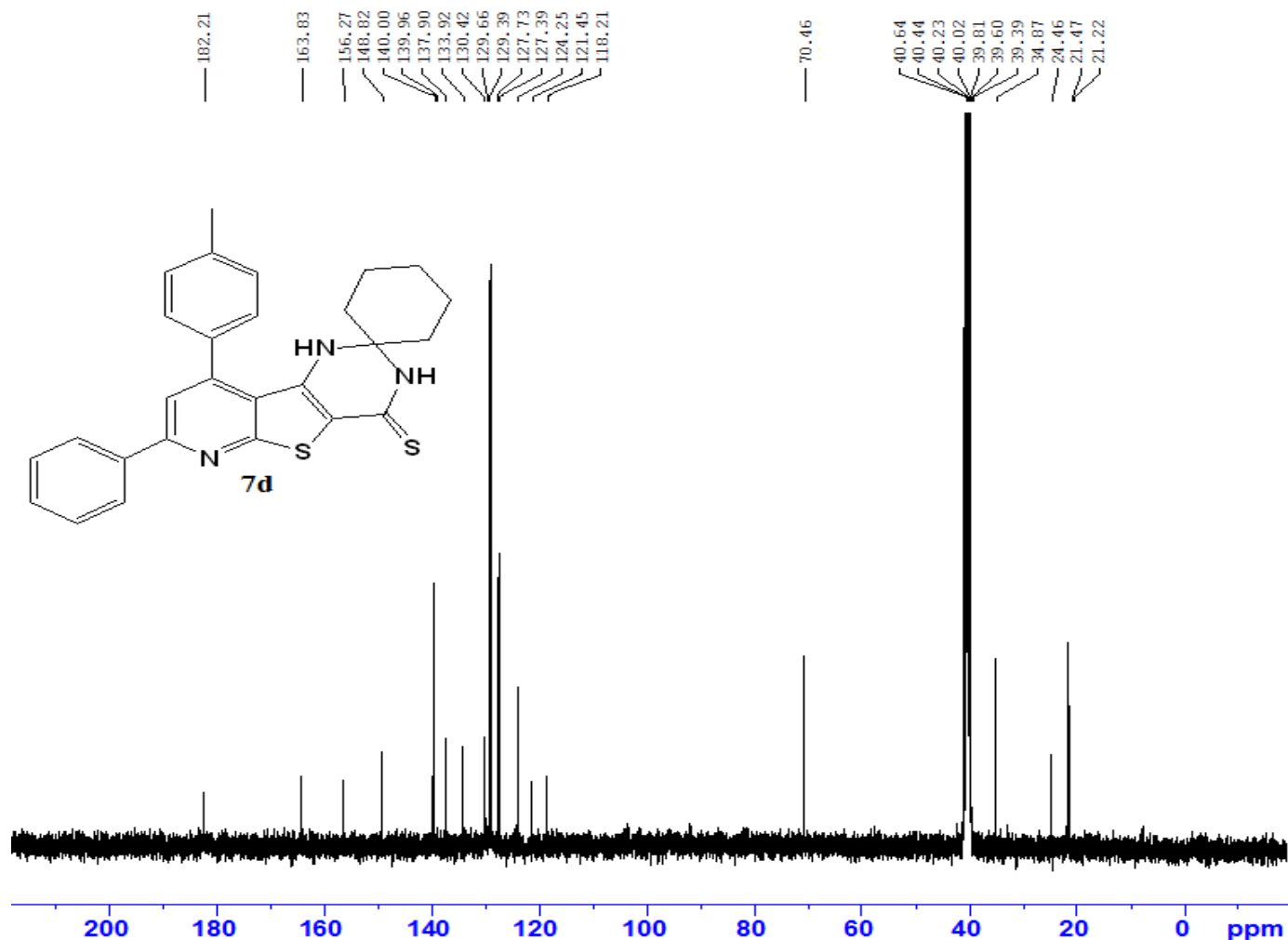
Current Data Parameters
NAME amina-nasr-M4
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190113
Time_ 12.29
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1092
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 205.37
DW 20.800 usec
DE 6.50 usec
TE 298.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 18.00000000 W
PLW12 0.34722000 W
PLW13 0.28125000 W

F2 - Processing parameters
SI 32768
SF 100.6177975 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



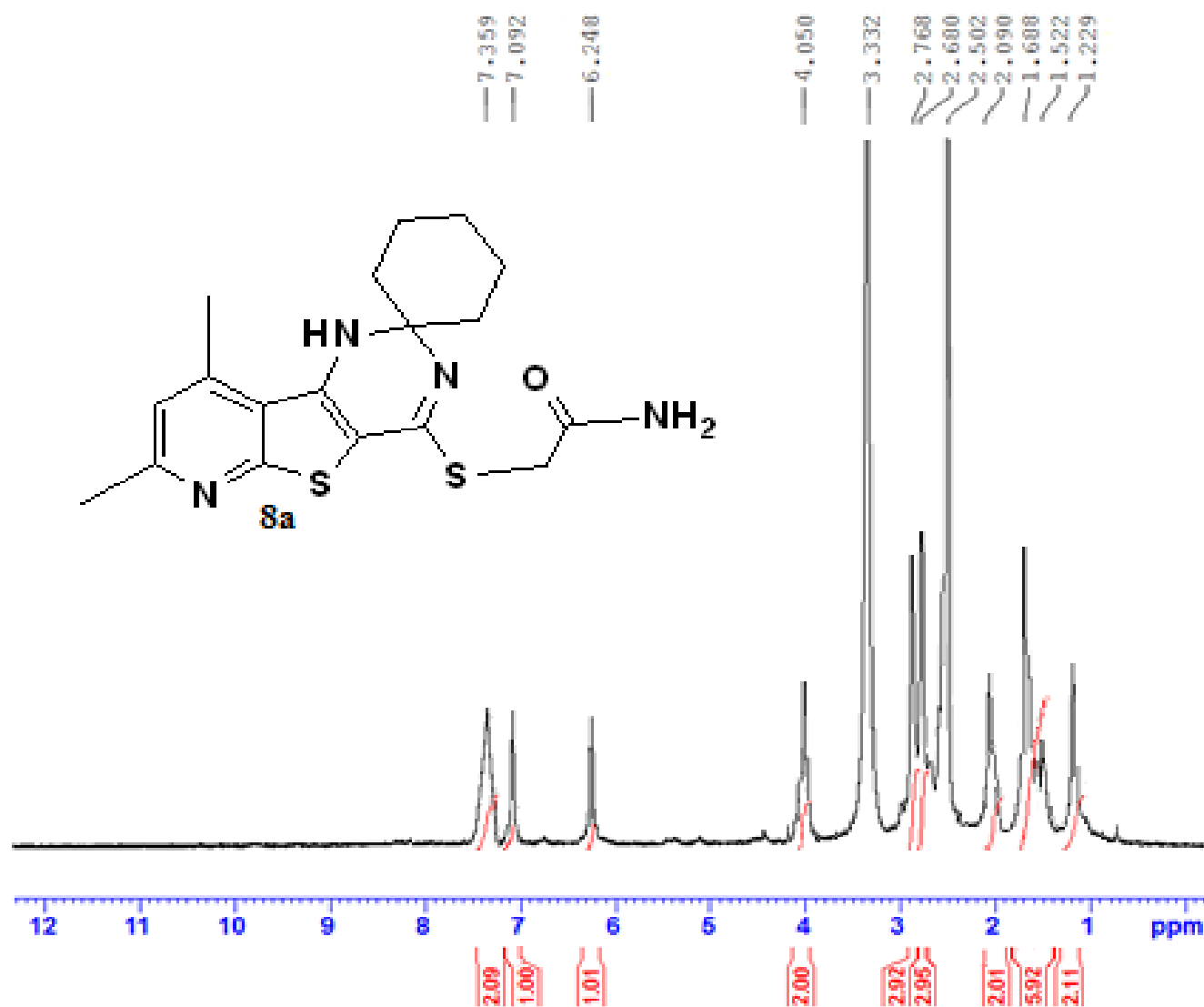
Current Data Parameters
 NAME amina-nasr-TP11
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210218
 Time 13.05
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 1098
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 205.37
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 SF01 100.6278588 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 47.00000000 W

===== CHANNEL f2 =====
 SF02 400.1516006 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 90.00 usec
 PLW2 18.00000000 W
 PLW12 0.34722000 W
 PLW13 0.28125000 W

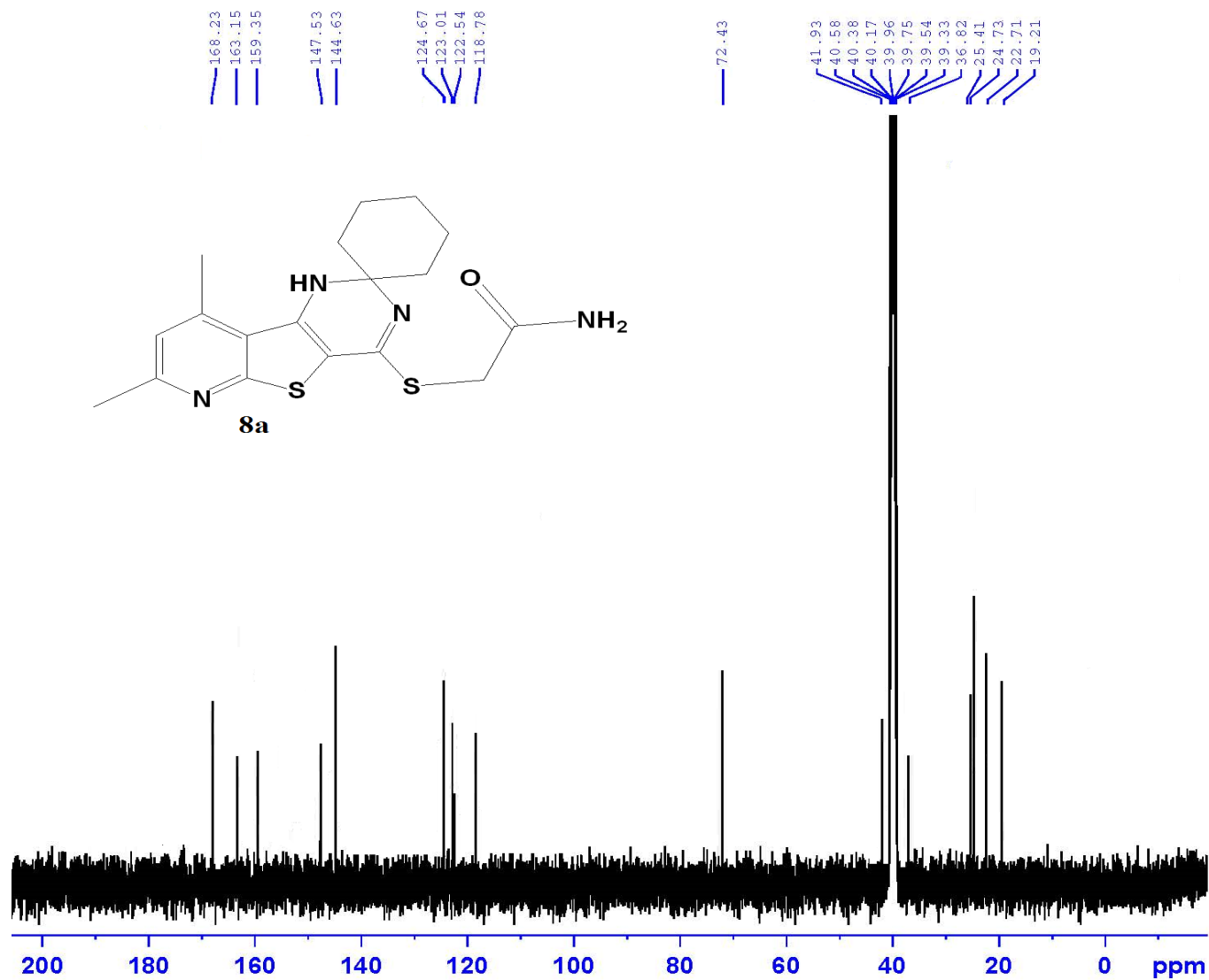
F2 - Processing parameters
 SI 32768
 SF 100.6177975 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



Current Data Parameters
NAME EYAN-M24
EXPNO 1
PROCNO 1
F2 - Acquisition Parameters
Date_ 20211212
Time 12.18
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 128
DS 2
SWE 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894465 sec
RG 209.37
DM 62.400 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1524711 MHz
NUC1 1H
P1 12.00 usec
PLW1 18.00000000 W

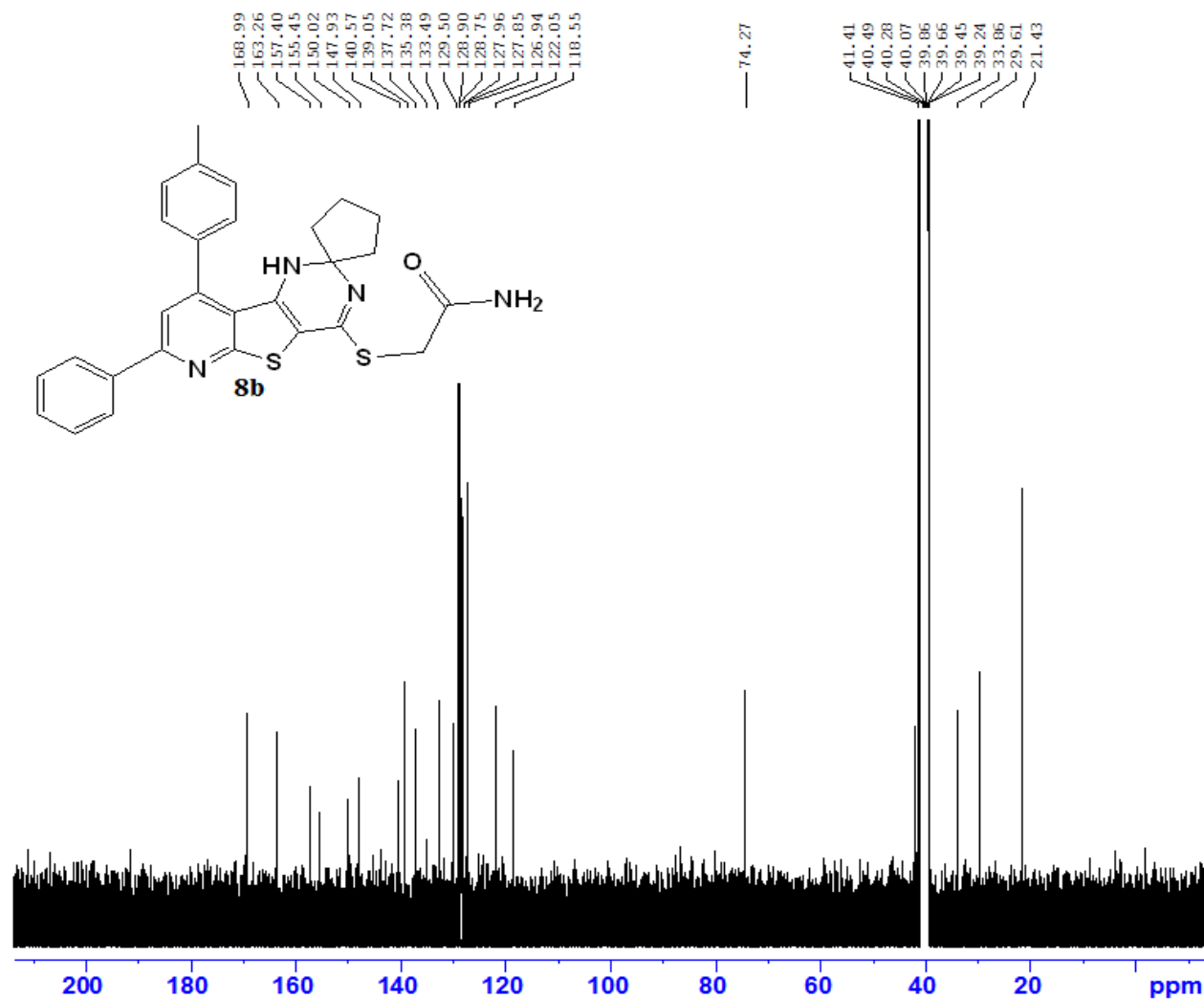
F2 - Processing parameters
SI 65536
SF 400.1500000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME Eman-M 2g
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20211209
Time_ 6.20 h
INSTRUM spect
PROBHD Z108618_0945 (
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.733596 Hz
AQ 1.3631488 sec
RG 197.77
DW 20.800 usec
DE 6.50 usec
TE 293.9 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6404331 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W
SFO2 400.2016008 MHz
NUC2 1H
PCPD2 waltz16
PLW2 13.00000000 W
PLW12 0.29249999 W
PLW13 0.14713000 W

F2 - Processing parameters
SI 32768
SF 100.6303700 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



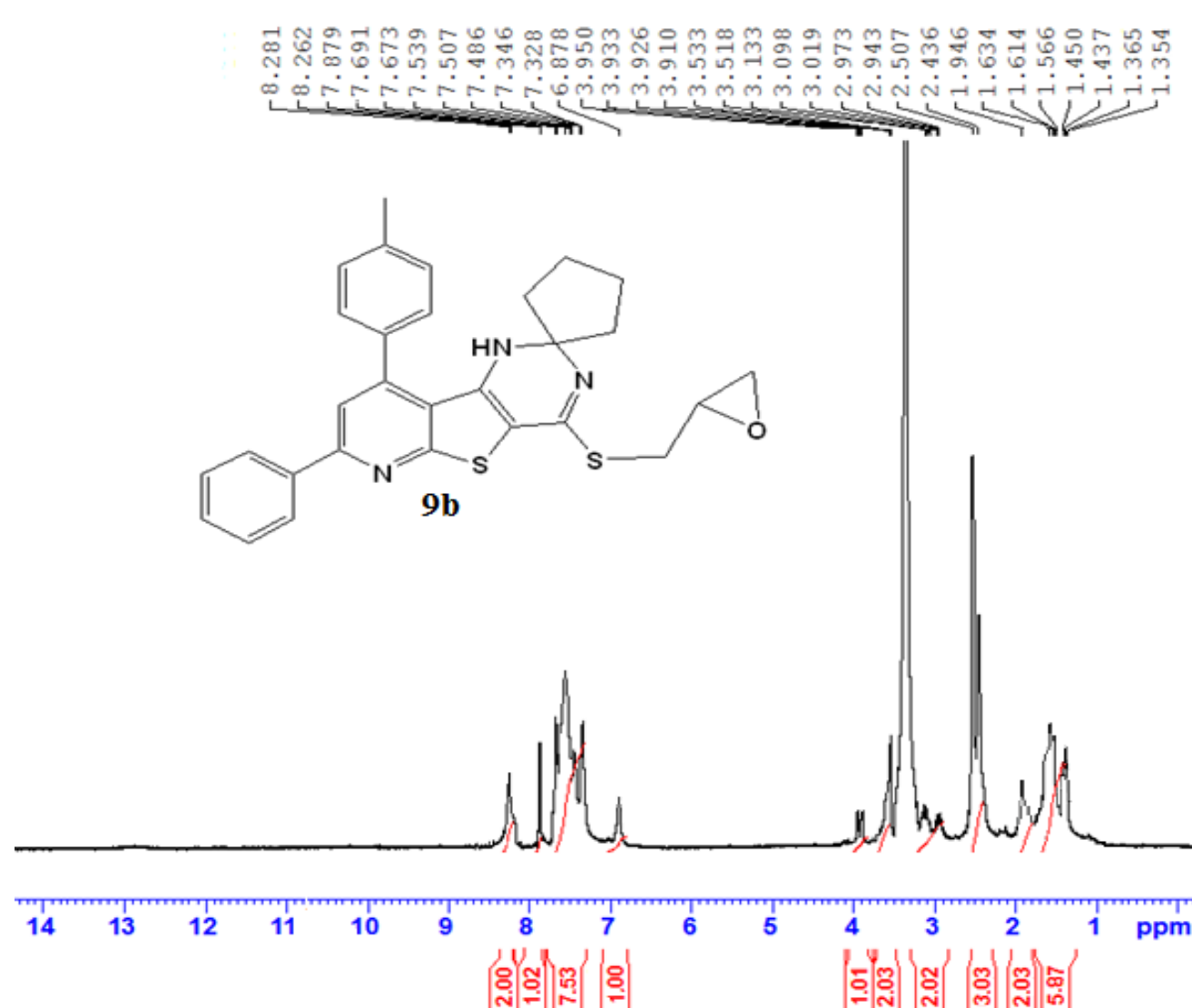
Current Data Parameters
 NAME amina-nasr-TPP23
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211018
 Time 8.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 15295
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 205.37
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 100.6278588 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 47.00000000 W

===== CHANNEL f2 =====
 SFO2 400.1516006 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 90.00 usec
 PLW2 18.00000000 W
 PLW12 0.34722000 W
 PLW13 0.28125000 W

F2 - Processing parameters
 SI 32768
 SF 100.6177975 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

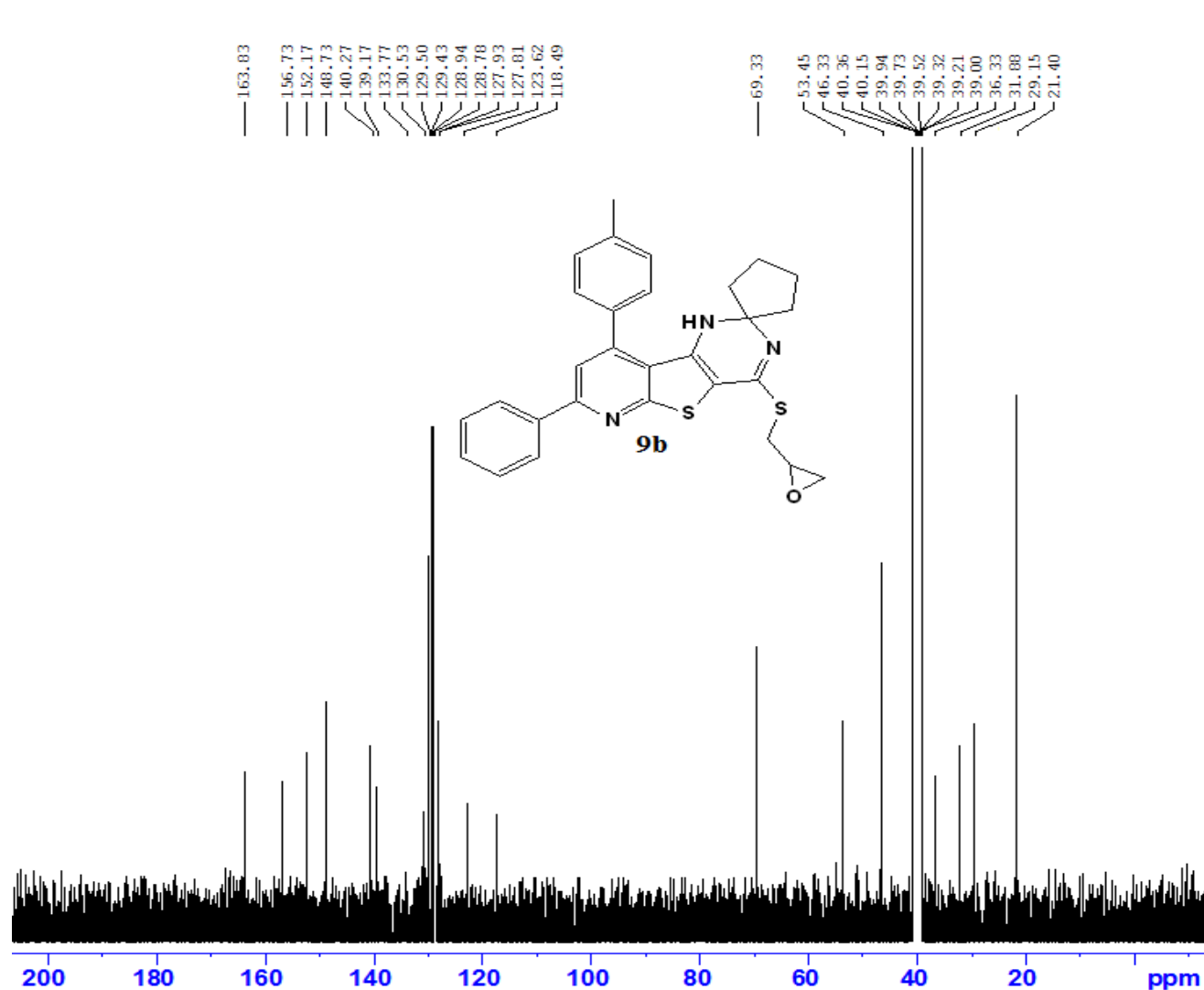


Current Data Parameters
 NAME Eman-TPP25
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211212
 Time 12.22
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 ID 65536
 SOLVENT DMSO
 NS 3000
 DS 4
 SWH 24012.820 Hz
 FIDRES 0.153066 Hz
 AQ 4.2054465 sec
 RG 205.37
 DW 30.400 usec
 DE 6.50 usec
 TE 300.0 K
 D1 2.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 SFO1 400.1524711 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 18.00000000 W

F2 - Processing parameters
 SI 65536
 SF 400.1500000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
NAME EMAN - TPP25
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20211212
Time 11.25
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 335
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 205.37
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 100.6278588 MHz
NUC1 13C
P1 10.00 usec
PLW1 47.00000000 W

===== CHANNEL f2 =====
SFO2 400.1516006 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 90.00 usec
PLW2 18.00000000 W
PLW12 0.34722000 W
PLW13 0.28125000 W

F2 - Processing parameters
SI 32768
SF 100.6177975 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Minimum Inhibitory Concentration (MIC) Measurement

The minimum Inhibitory Concentration activity of the compounds was then evaluated using broth dilution method. Whereas, two-fold serial dilution at the concentrations (0.25, 0.5, 1, 2, 4, 8, 16, 32, 64, 128 µg/ml) was used to investigate the Minimum Inhibitory Concentration (MIC) values (expressed in µg/mL) for the target compounds and the reference drugs. The tubes were then inoculated with the test organisms, grown in their suitable broth for 24 h at 37 °C for bacteria, yeast and for 48 h at 30°C for fungi activity (1×10^8 CFU/mL for bacteria and 1×10^6 CFU/mL of yeast and fungi), each 2 mL received 0.1 mL of the above inoculums. Positive controls were prepared separately for either bacteria, yeast or fungi with respective organisms in the same culture media without the target compounds. After incubation, the tube with lowest concentration of extract that shows no growth was taken as the MIC value for the respective organism.

In vitro anticancer screening

The cell lines were purchased from the American Type Culture collection as follows: breast carcinoma cell line (MCF-7) and the liver carcinoma cell line (HepG2). Cytotoxic activity screening was performed using MTT assay at Regional Center for Mycology and Biotechnology, Al- Azhar University. Exponentially, cells were placed in 10^4 cells/ well for 24 h, and then add fresh medium which containing different concentration of the tested sample. Serial two-fold dilutions of the tested sample were added using a multichannel pipette. Moreover, all cells were cultivated at 37 °C, 5% CO₂ and 95% humidity. Also, incubation of control cells occurred at 37 °C. However, after incubation for 24 h different concentrations of sample (50, 25, 12.5, 6.25, 3.125, 1.56 and 0 µg L⁻¹) were added and continued the incubation for 48 h, then, add the crystal violet solution 1% to each well for 0.5 h to examine viable cells. Rinse the wells using water until no stain. After that, add 30% glacial acetic acid to all wells with shaking plates on Microplate reader (TECAN, Inc.) to measure the absorbance, using a test wavelength of 490 nm. Besides, compare the treated samples with the control cell. The cytotoxicity was estimated by IC₅₀ (the concentration that inhibits 50% of growth of cancer cell) in µM for the tested compounds and the reference drugs doxorubicin and cisplatin.

In vitro EGFR kinase assay

EGFR kinase inhibitory assay were performed for the target compounds **3b**, **4a**, **5a**, **6b**, **8b** and **9b** with erlotinib as a reference inhibitor, by using the EGFR kinase assay kit (Cat. # 40321). The assay Kit is designed to measure EGFR Kinase activity for screening applications using Kinase-Glo® MAX as a detection reagent using Kinase-Glo® MAX as a detection reagent. Thaw 5x Kinase Buffer 1: ATP and PTK substrate Poly (Glu:Tyr 4:1) (10 mg/ml) kinase was provided. Then the master mixture was prepared: N wells x (6 µl 5x Kinase Buffer 1 + 1 µl ATP (500 µM) + 1 µl PTK substrate Poly (Glu:Tyr 4:1) (10 mg/ml)+ 17 µl water) and 25 µl of the mixture was added to every well. The tested compounds were dissolved in DMSO, then 5 µl of their serial two-fold dilutions at concentrations (150, 100, 50, 25, 12.5, 6.25, 3.125 and 1.56 nM) was added in each well labeled as "Test Inhibitor". For the well labeled as "Positive Control", 5 µl of the same solution of the reference drug erlotinib and "Blank", 5 µl of the same solution without inhibitor (Inhibitor buffer). (3 ml of 1x Kinase Buffer 1) was prepared by mixing 600 µl of 5x Kinase Buffer 1 with 2400 µl water. To the wells designated as "Blank", add 20 µl of 1x Kinase Buffer 1. The reaction initiated by adding 20 µl of diluted EGFR enzyme to the wells designated "Positive Control" and "Test Inhibitor Control". Incubate at 30°C for 40 minutes. Thaw Kinase-Glo Max reagent. After the 40 minute reaction, 50 µl of Kinase-Glo Max reagent was added to each well. The plate was covered with aluminum foil and incubate at room temperature for 15 minutes. Then the luminescence was measured using the microplate reader (Infinite M200 microplate reader, Tecan, Männedorf, Switzerland). All assays were performed in triplicate. The relative inhibition (%) of inhibitors were then calculated compared to the control with no inhibitor. Then the IC₅₀ values and their standard deviation (SD) for the tested compounds and the reference drug were determined in (nM).

Molecular docking study

The molecular modeling studies were carried out using Molecular Operating Environment (MOE, 2019.0102) software. All minimizations were performed with MOE until an RMSD gradient of $0.1 \text{ kcal}\cdot\text{mol}^{-1}\text{\AA}^{-1}$ with MMFF94x force field and the partial charges were automatically calculated. The X-ray crystallographic structure of Epidermal Growth Factor Receptor (**EGFR**) kinase domain complexed with a quinazoline inhibitor erlotinib (**ERL**) (**PDB ID: 1M17**) was downloaded from the protein data bank(<https://www.rcsb.org/structure/1M17>). For each co-crystallized enzyme; water molecules and ligands which are not involved in the binding were removed, the protein was prepared for the docking study using *Protonate 3D* protocol in MOE with default options. The co-crystalized ligand (**ERL**) was used to define the binding site for docking. Triangle Matcher placement method and London dG scoring function were used for docking

The Dock workflow begins with a prepared receptor and ligand loaded in MOE. The general algorithm is divided into stages,

1. Get the protein structure from protein data bank (PDB file), it's prepared through protonation and adding the required charges, water molecules not involved in interaction are removed.
2. Ligands' structures in suitable format were collected to form a structure database as MDB file and energy minimized to get the suitable conformers for docking procedure.
3. Use triangle matching placement method to generate a collection of poses from the pool of ligand conformations. Each of the generated poses is assigned a score.
4. Initial Scoring, poses generated by the placement methodology can be rescored using London dG method. Typically, scoring functions emphasize favorable hydrophobic, ionic and hydrogen bond contacts. For the Dock framework to work properly, all new scoring methods must assign low scores to good poses.
5. Refinement, poses resulting from the placement stage can be refined using either the explicit molecular mechanics forcefield method or the grid-based energetics method.
6. Final Scoring. The final poses can be rescored using one of several scoring schemes, we use London dG scoring function.
7. Generation of docking results database.