



# Article Development of Hydroxamate Derivatives Containing a Pyrazoline Moiety as APN Inhibitors to Overcome Angiogenesis

Yangyang Liu <sup>(D)</sup>, Dongsheng Zhao <sup>(D)</sup>, Chenghua Zhang, Hui Fang, Qingsitong Shen, Zhixian Wang and Jiangying Cao \*

School of Pharmacy, Shandong University of Traditional Chinese Medicine, Jinan 250355, China \* Correspondence: 60030103@sdutcm.edu.cn or caojiangyingmed@163.com; Tel.: +86-531-89628234

Abstract: Aminopeptidase N (APN) was closely associated with cancer invasion, metastasis, and angiogenesis. Therefore, APN inhibitors have attracted more and more attention of scientists as antitumor agents. In the current study, we designed, synthesized, and evaluated one new series of pyrazoline-based hydroxamate derivatives as APN inhibitors. Moreover, the structure–activity relationships of those were discussed in detail. 2,6-Dichloro substituted compound **140** with  $R_1 = CH_3$ , showed the best capacity for inhibiting APN with an IC<sub>50</sub> value of 0.0062 ± 0.0004 µM, which was three orders of magnitude better than that of the positive control bestatin. Compound **140** possessed both potent anti-proliferative activities against tumor cells and potent anti-angiogenic activity. At the same concentration of 50 µM, compound **140** exhibited much better capacity for inhibiting the micro-vessel growth relative to bestatin in the rat thoracic aorta ring model. Additionally, the putative interactions of **140** with the active site of APN are also discussed. The hydroxamate moiety chelated the zinc ion and formed four hydrogen bonds with His<sup>297</sup>, Glu<sup>298</sup> and His<sup>301</sup>. Meanwhile, the terminal phenyl group and another phenyl group of **140** interacted with S<sub>2</sub>' and S<sub>1</sub> pockets via hydrophobic effects, respectively.

Keywords: aminopeptidase N; CD13; inhibitors; antitumor agents; anti-angiogenesis

# 1. Introduction

Aminopeptidase N (APN, CD13, EC 3.4.11.2), a type II transmembrane glycoprotein, is a zinc-dependent exopeptidase belonging to the M1 superfamily of the MA clan of peptidases [1]. APN is widely expressed on the surfaces of diverse cells, such as liver, renal, intestinal, fibroblast, endothelial, tumor cells and so on. Due to its multiple functions in the hydrolysis of peptides, signal transduction and animal coronavirus infection, APN is also called the "moonlighting enzyme" [2]. As an exopeptidase, APN preferentially cleaves the hydrophobic and basic amino acid residues from the N-terminus of biologically active peptides with broad substrate specificity [3]. For example, in the central nervous system, APN and neutral endopeptidase (NEP) cooperatively hydrolyzes enkephalin and endorphin to regulate pain [4]. Moreover, APN catalyzes the conversion of angiotensin III to angiotensin IV to regulate the renin–angiotensin system [5].

Ongoing interest is focused on APN due to its significant role in the metastasis and angiogenesis of tumors. Metastasis is a complex biological process involving cell migration, cell invasion and angiogenesis, which leads to more than 90% of cancer related deaths [6,7]. APN can metabolize the extracellular matrix (ECM) to promote the process of invasion and metastasis of tumor cells [8,9]. Moreover, expression of APN had been observed on the surfaces of angiogenic blood vessels rather than normal ones [10]. Impaired neovas-cularization was found in APN knockout mice [11]. Furthermore, it was reported that APN on the surfaces of both tumor cells and nonmalignant stromal cells cooperatively induced angiogenesis [12]. In addition, a strong correlation between up-regulated APN



Citation: Liu, Y.; Zhao, D.; Zhang, C.; Fang, H.; Shen, Q.; Wang, Z.; Cao, J. Development of Hydroxamate Derivatives Containing a Pyrazoline Moiety as APN Inhibitors to Overcome Angiogenesis. *Molecules* 2022, *27*, 8339. https://doi.org/ 10.3390/molecules27238339

Academic Editor: Pascale Moreau

Received: 30 October 2022 Accepted: 24 November 2022 Published: 29 November 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). expression and poor prognosis of tumors has been certified through much research [13]. A recent study showed that APN was over-expressed in semi-quiescent liver cancer cells and considered as one of surface markers of liver cancer stem cells (CSCs), which mainly led to tumor recurrence [14]. All of this evidence supports the rationale that APN is an effective target for the chemotherapy of tumors.

Structurally, APN is composed of 967 amino acids, contains a short intracellular tail, a transmembrane anchor, a small Ser-/Thr-rich extracellular stalk and a large ectodomain [15]. Domain II, one component of the ectodomain, contains one catalytic zinc ion and two characteristic motifs (HEXXHX18E zinc-binding motif and GXMEN substrate-recognition motif) from the M1 aminopeptidase family [16]. Around the zinc ion, there were three hydrophobic pockets  $S_1$ ,  $S_1'$  and  $S_2'$ , which could bind the  $P_1$ ,  $P_1'$  and  $P_2'$  sites of the substrates, respectively. So far, many natural products and synthetic small molecules are reported to be APN inhibitors (APNIs). The natural APNIs are mostly peptides or peptidomimetics, such as bestatin 1, amastatin 2, probestin 3, AHPA-Val 4, and so on (Figure 1) [17,18]. The synthetic APNIs are structurally diverse, containing 1,2,3-triazol ureido peptidomimetics 5 [19], an aminophosphinic derivative 6 (PL253) [20], a flavone derivative 7 [21], a dicarbonyl derivative 8 [22], a chloramphenicol amine derivative 9 [23], an amino benzosuberone derivative 10 [24], a L-isoserine derivative 11 [25], an amide derivative 12 and so on (Figure 1) [26]. Among them, the dipeptide derivative bestatin is marketed as an immunomodulating agent for the adjuvant treatment of acute myeloid leukemia. Many studies have subsequently revealed that bestatin could prevent the invasion, metastasis and angiogenesis associated with tumors [27,28].

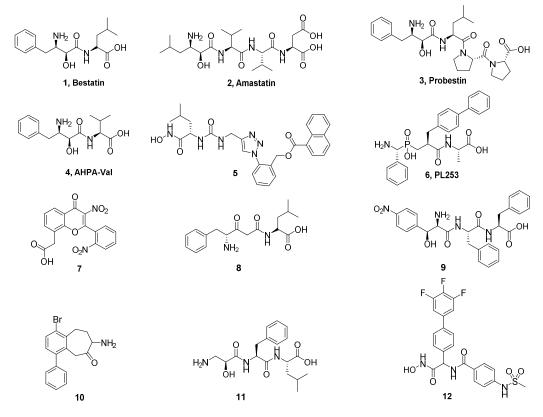


Figure 1. The structures of representative aminopeptidase N inhibitors (APNIs).

The structure–activity relationships (SARs) of APNIs revealed that the potent AP-NIs possessed zinc binding groups (ZBGs) and hydrophobic groups interacting with at least one of three hydrophobic pockets ( $S_1$ ,  $S_1'$  and  $S_2'$ ) of APN. In our previous work, pyrazoline-based hydroxamate derivatives **13a** and **13b** showed the capacity of inhibiting APN (Figure 2) [29]. Subsequently, optimization focused on the terminal phenyl group of the diphenyl pyrazoline core gave compound **13c** with improved APN inhibitory activity relative to compounds **13a** and **13b** [30]. In this work, in order to extend the SARs of these series of compounds and find more potent APNIs, further optimization was focused on two phenyl groups of compounds **13a** and **13b**, leading to compound **14**. The introduction of hydrophobic substituents on the phenyl groups of compound **14** or the replacement of the terminal phenyl group with a naphthyl moiety may strengthen the hydrophobic interactions with APN. Interestingly, among all the newly synthesized compounds, compound **14o** presented the best APN inhibitory activity, being two orders of magnitude better than those of lead compounds **13a** and **13b**.

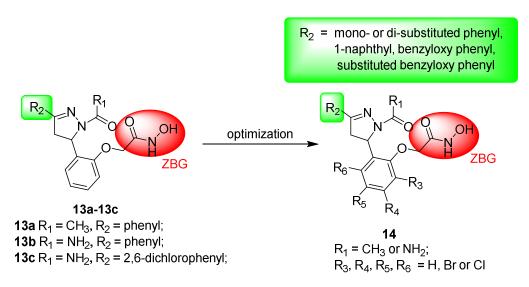
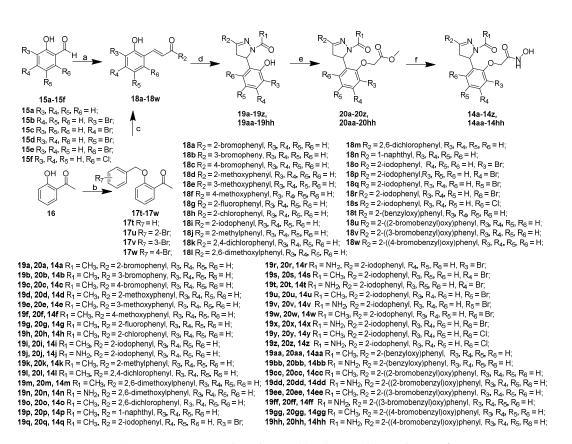


Figure 2. Design strategy of APNIs containing pyrazoline moiety.

# 2. Results and Discussion

# 2.1. Chemistry

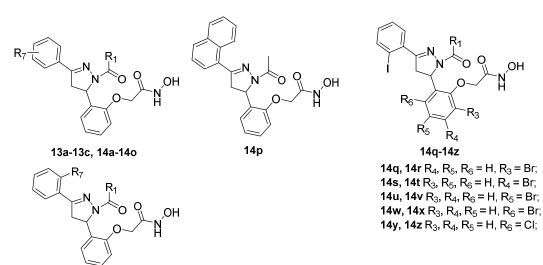
The target compounds were synthesized following the procedures shown in Scheme 1. Firstly, benzaldehyde derivatives **15a–15f** reacted with the corresponding ketone to form the key chalcone intermediates **18a–18s** via the Claisen–Schmidt reaction in the presence of potassium hydroxide. Additionally, 2-hydroxyacetophenone **16** reacted with benzyl bromide or substituted benzyl bromide to give compounds **17t–17w**, which further reacted with 2-hydroxybenzaldehyde **15a** to give chalcone intermediates **18a–18w**, respectively, following the same procedure of compounds **18a–18s**. Subsequently, compounds **18a–18w** were cyclized with excess hydrazine hydrate or semicarbazide hydrochloride to give the pyrazoline derivatives **19a–19z** and **19aa–19hh**, which further underwent nucleophilic substitution reaction with methyl bromoacetate to yield compounds **20a–20z** and **20aa–20hh**, respectively. Ultimately, the freshly prepared methanolic solution of NH<sub>2</sub>OK transformed the methyl ester groups of **20a–20z** and **20aa–20hh** to hydroxamate groups to form the target compounds **14a–14z** and **14aa–14hh**, respectively.



Scheme 1. Reagents and conditions: (a) substituted acetophenone for 18a–18m, 18o–18s, 1-(naphthalen-1-yl)ethan-1-one for 18n, KOH, 80% EtOH, 25 °C, 48 h; (b) benzyl bromide or substituted benzyl bromide, NaH, DMF, 25 °C, 12 h; (c) for 18t–18w, 15a, KOH, 80% EtOH, 25 °C, 48 h; (d)  $R_1 = CH_3$ , acetic acid, 80% hydrazine hydrate, 130 °C, 5 h or  $R_1 = NH_2$ , semicarbazide hydrochloride, NaOH, EtOH, 78 °C, 5 h; (e) methyl bromoacetate, NaH, DMF, 25 °C, 12 h; (f) NH<sub>2</sub>OK, MeOH, 25 °C, 0.5 h.

#### 2.2. Inhibitory Activities of Target Compounds against APN

All the synthesized pyrazoline derivatives were first evaluated for their APN inhibitory activities using bestatin and compounds 13a–13c as the positive controls. The results are listed in Table 1. Similar to SARs previously reported, the substituents on the terminal phenyl group of the diphenyl pyrazoline moiety could significantly impact the inhibitory activities against APN [30]. Comparing mono-substituted compounds 14a-14f with a methyl group as  $R_1$ , it was easily found that compounds (14a, 14d) with orthosubstituents showed better inhibitory potencies against APN than their counterparts with *meta*-substituents (14b, 14e) or *para*-substituents (14c, 14f), respectively. Moreover, among ortho mono-substituted compounds 14a, 14d, 14g-14i and 14k, compound 14i containing an *ortho*-iodine substitution exhibited the best capacity of inhibiting APN, with an  $IC_{50}$  value of 0.015  $\pm$  0.001  $\mu$ M. The APN inhibitory order of *ortho* mono-substituted compounds with  $R_1 = CH_3$ , was 14i (2-I) > 14h (2-Cl) > 14a (2-Br) > 14k (2-CH\_3) > 14g (2-F) > 14d (2-OCH\_3). Among dual-substituted compounds (14l, 14m, 14o) with  $R_1 = CH_3$ , compound 14o with 2,6-dichlorine substitution was two orders of magnitude better against APN than the analogues with 2,4-dichlorine (14l) or 2,6-dimethoxy (14m) substitution, suggesting that the substituents and substituted position on the terminal phenyl group significantly affected the inhibitory activities against APN. Moreover, ortho dual-substituted compounds showed improved APN inhibitory activities relative to the corresponding ortho mono-substituted analogs (14m vs. 14d; 14o vs. 14h). In addition, the APN inhibitory activities of compounds 14j and 14n, with  $NH_2$  as  $R_1$ , were inferior to those of counterparts 14i and 14m, with  $CH_3$ as R<sub>1</sub>, respectively. Replacement of the terminal phenyl group of compound 13a with a naphthyl group led to compound 14p, with comparable APN inhibitory activity to 13a.



**Table 1.** The structures and  $IC_{50}$  values of target compounds against porcine kidney aminopeptidase N (APN).

14aa-14hh

Compd	<b>R</b> <sub>1</sub>	<b>R</b> <sub>7</sub>	APN IC <sub>50</sub> (μM) <sup>a</sup>	Compd	<b>R</b> <sub>1</sub>	R <sub>7</sub>	APN IC <sub>50</sub> (µM) <sup>a</sup>
13a	CH <sub>3</sub>	Н	$0.32\pm0.01$	14q	CH <sub>3</sub>	_	$3.5\pm0.2$
13b	$NH_2$	Н	$0.16\pm0.01$	14r	NH <sub>2</sub>	_	$3.4\pm0.3$
13c	$NH_2$	2,6-diCl	$0.059\pm0.002$	14s	CH <sub>3</sub>	_	$2.1\pm0.2$
14a	CH <sub>3</sub>	2-Br	$0.046\pm0.003$	14t	NH <sub>2</sub>	-	$1.6\pm0.1$
14b	CH <sub>3</sub>	3-Br	$0.32\pm0.03$	14u	CH <sub>3</sub>	-	$0.37\pm0.02$
14c	CH <sub>3</sub>	4-Br	$4.4\pm0.3$	14v	NH <sub>2</sub>	-	$0.54\pm0.04$
14d	CH <sub>3</sub>	2-OCH <sub>3</sub>	$0.32\pm0.01$	14w	CH <sub>3</sub>	-	$0.024\pm0.001$
14e	CH <sub>3</sub>	3-OCH <sub>3</sub>	$0.37\pm003$	14x	NH <sub>2</sub>	-	$0.090\pm0.005$
14f	CH <sub>3</sub>	$4-OCH_3$	$3.8\pm0.2$	14y	CH <sub>3</sub>	-	$0.023\pm0.001$
14g	CH <sub>3</sub>	2-F	$0.25\pm0.01$	14z	NH <sub>2</sub>	-	$0.070\pm0.003$
14h	CH <sub>3</sub>	2-Cl	$0.029\pm0.002$	14aa	CH <sub>3</sub>	<sup>34</sup> O	$0.017\pm0.001$
14i	CH <sub>3</sub>	2-I	$0.015\pm0.001$	14bb	NH <sub>2</sub>	je <sup>s</sup> o	$0.015\pm0.001$
14j	NH <sub>2</sub>	2-I	$0.044\pm0.003$	14cc	CH <sub>3</sub>	<sup>z<sup>2</sup>CO</sup>	$0.068\pm0.004$
14k	CH <sub>3</sub>	2-CH <sub>3</sub>	$0.21\pm0.01$	14dd	NH <sub>2</sub>	<sup>25</sup> O Br	$0.23\pm0.01$
141	CH <sub>3</sub>	2,4-diCl	$0.58\pm0.04$	14ee	CH <sub>3</sub>	<sup>3<sup>4</sup>OBr</sup>	$0.031\pm0.002$
14m	CH <sub>3</sub>	2,6-diOCH <sub>3</sub>	$0.20\pm0.01$	14ff	NH <sub>2</sub>	<sup>3</sup> <sup>2</sup> O Br	$0.058\pm0.003$
14n	NH <sub>2</sub>	2,6-diOCH <sub>3</sub>	$0.44\pm0.03$	14gg	CH <sub>3</sub>	<sup>2<sup>2</sup></sup> O Br	$0.56\pm0.04$
<b>14o</b>	CH <sub>3</sub>	2,6-diCl	$0.0062\pm0.0004$	14hh	NH <sub>2</sub>	<sup>2<sup>f</sup></sup> O Br	$0.55\pm0.04$
14p	-	_	$0.31\pm0.02$	Bestatin	-	—	$8.8\pm0.4$

<sup>a</sup> Assays were performed in triplicate; data are shown as mean  $\pm$  SD.

Moreover, in order to explore the APN inhibitory effects of substituents on the other phenyl group, compounds **14q–14z** were designed using **14i** and **14j** as lead compounds. Compound **14w**, with  $R_1 = CH_3$  and  $R_6 = Br$ , exhibited better APN inhibitory activity than the counterparts **14q** ( $R_3 = Br$ ), **14s** ( $R_4 = Br$ ) and **14u** ( $R_5 = Br$ ), respectively. The same inhibitory tendency was also found among compounds **14r**, **14t**, **14v** and **14x**, with  $R_1 = NH_2$ .

Furthermore, compounds 14y (IC<sub>50</sub> =  $0.023 \pm 0.001 \mu$ M) and 14z (IC<sub>50</sub> =  $0.070 \pm 0.003 \mu$ M), with R<sub>6</sub> = Cl, displayed comparable or improved APN inhibition relative to corresponding bromo-substituted compounds 14w and 14x, respectively. However, the APN inhibitory activities of compounds 14y and 14z were inferior to those of corresponding lead compounds 14i and 14j, respectively, indicating that the introduction of substituents on the other phenyl group was detrimental.

Furthermore, introduction of one large hydrophobic *ortho*-benzyloxyl group on the terminal phenyl group of compounds **13a** and **13b** led to compounds **14aa** and **14bb**, respectively, with one order of magnitude improved APN inhibitory activities. However, bromo-substitution of the benzyloxyl group of **14aa** and **14bb** led to decreased APN inhibition (**14cc**, **14ee**, **14gg** vs. **14aa**; **14dd**, **14ff**, **14hh** vs. **14bb**). Moreover, *meta* bromosubstituted compounds presented better APN inhibitory activities than the counterparts with *ortho* or *para* bromo-substitution (**14ee** vs. **14cc**, **14gg**; **14ff** vs. **14dd**, **14hh**).

Generally, compounds with  $R_1 = CH_3$ , exhibited superior or inferior APN inhibitory activities to their counterparts with  $R_1 = NH_2$ , which depended on the substituents of the two phenyl groups of the diphenyl pyrazoline moiety. Interestingly, the APN inhibitory activities of compounds **14a**, **14h-14j**, **14o**, **14w**, **14y**, **14aa**, **14bb** and **14ee** were better than those of lead compounds **13a-13c**, supporting our design strategy. Notably, the most potent one, **14o** (IC<sub>50</sub> = 0.0062 ± 0.0004  $\mu$ M) was over 9-fold more potent than lead compound **13c** (IC<sub>50</sub> = 0.059 ± 0.002  $\mu$ M) and over 1419-fold more potent than positive control bestatin (IC<sub>50</sub> = 8.8 ± 0.4  $\mu$ M).

# 2.3. Anti-Proliferative Activities of Selected Compounds against Tumor Cells

Compound **140**, the most potent against APN, was selected to further evaluate the anti-proliferative activities against six tumor cell lines (U937, K562, PLC/PRF/5, PC-3, ES-2, HepG2). As results listed in Table 2, compound **140** exhibited much better anti-proliferative potencies than bestatin against all the tested tumor cell lines. Among them, K562 cells were most sensitive to compound **140**.

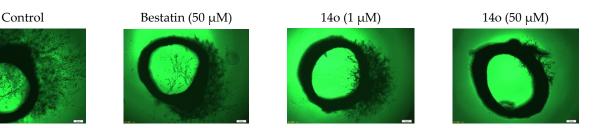
Compound -	IC <sub>50</sub> (μM) <sup>a</sup>								
Compound	U937	K562	PLC/PRF/5	PC-3	ES-2	HepG2			
<b>140</b> Bestatin	38.3 ± 3.6 >500	$10.4 \pm 1.2 > 500$	88.5 ± 7.4 >500	$45.8 \pm 4.1 > 500$	114.6 ± 9.7 >500	84.2 ± 7.6 >500			

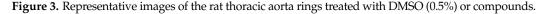
Table 2. In vitro anti-proliferative activities of selected compounds against tumor cells.

<sup>a</sup> Assays were performed in triplicate; data are shown as mean  $\pm$  SD.

# 2.4. Ex Vivo Anti-Angiogenesis Assay

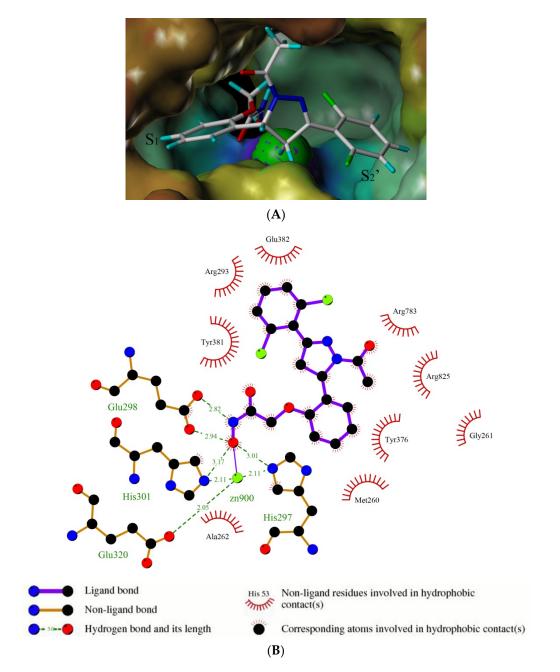
The anti-angiogenic potency of selected compound **140** was preliminarily measured using the rat thoracic aorta ring model. As the results in Figure 3 show, compound **140** could prevent the growth of micro-vessels derived from the thoracic aorta rings in a dose-dependent manner. Compared with the group treated with 1  $\mu$ M of **140**, obviously reduced micro-vessels were observed in the group treated with 50  $\mu$ M of **140**. Furthermore, at the same concentration of 50  $\mu$ M, the anti-angiogenic activity of **140** was much better than that of bestatin, which was consistent with the enzymatic inhibitory activities against APN.





# 2.5. Docking Assay

In order to investigate its binding mode in APN, compound **140** was docked into the active site of APN (PDB code: 2DQM) using Sybyl 2.1. As depicted in Figure 4A, the terminal phenyl group with 2,6-diclorine substitution and another phenyl group occupied the  $S_2'$  and  $S_1$  hydrophobic pockets, respectively. Meanwhile, the hydroxamate moiety chelated the zinc ion. Moreover, one more detailed binding mode of compound **140** with APN was illustrated in Figure 4B, generated with LIGPLOT. The terminal phenyl group could form hydrophobic interactions with the  $Arg^{293}$ ,  $Tyr^{381}$  and  $Glu^{382}$ . Meanwhile, another phenyl group could form hydrophobic interactions with  $Met^{260}$  and  $Tyr^{376}$ . In addition, the  $CH_3CO$  group hydrophobically interacted with  $Arg^{783}$  and  $Arg^{825}$ . Except for chelating the zinc ion, the hydroxamate moiety could form four hydrogen bonds with  $His^{297}$ ,  $Glu^{298}$  and  $His^{301}$  to enhance the potency of the interaction with APN.



**Figure 4.** (A) The FlexX docking result of **14o** in the active site of APN (PDB code: 2DQM). The carbon atoms are marked in gray, oxygen atoms in red, hydrogen atoms in light blue, nitrogen atoms

in blue and chlorine atoms in green. The green sphere represents zinc ion.  $S_1$  and  $S_2'$  represent two hydrophobic pockets. (**B**) The docking result shown by LIGPLOT. The carbon atoms, oxygen atoms, nitrogen atoms and chlorine atoms are marked in black, red, blue and green, respectively. The backbone of compound **140** is labeled violet and the zinc ion is shown as green sphere. The hydrogen bonds are shown as dashed lines (distance in Å).

#### 3. Materials and Methods

# 3.1. Chemistry: General Procedures

All the commercially available materials were used without further purification otherwise noted. Thin-layer chromatography (TLC) was used for the monitoring of all the reactions on 0.25 mm silica gel plates (60 GF-254) and the product spots were visualized by ferric chloride, iodine vapor and UV light. The purification of the product was carried out by column chromatography. Melting points were determined on an electrothermal melting point apparatus without correction. <sup>1</sup>H-NMR spectra were conducted on a Brucker DRX spectrometer with TMS as an internal standard. The values of chemical shifts were described as  $\delta$  in parts per million and *J* in Hertz. HRMS were conducted by the Shandong Analysis and Test Center.

3.1.1. General Procedure for Synthesis of Compounds **17t–17w** 1-(2-(Benzyloxy)phenyl) ethan-1-one (**17t**)

To a solution of 2-hydroxyacetophenone **16** (10.88 g, 80.00 mmol) in DMF was added the 60% NaH (3.84 g, 96.00 mmol), gradually. After stirring at 0 °C for 5 min, benzyl bromide (20.52 g, 120.00 mmol) was added into the mixture. The mixture was stirred at 25 °C for 12 h, then poured into water, followed by extraction with EtOAc. The organic phase was washed with brine and dried over MgSO<sub>4</sub>. After filtration and concentration under vacuum, the residue was purified by column chromatography (PE/EtOAc = 10:1, v/v) to give 16.63 g of compound **17t** as oil. Yield: 92%; ESI-MS m/z 227.1 [M+H]<sup>+</sup>.

Compounds 17u–17w were prepared following the procedure described for compound 17t.

#### 3.1.2. General Procedure for Synthesis of Compounds 18i, 18l, 18o-18w

(E)-3-(5-Bromo-2-hydroxyphenyl)-1-(2-iodophenyl)prop-2-en-1-one (**18q**)

5-Bromo-2-hydroxybenzaldehyde **15d** (10.00 g, 50.00 mmol) and 2-iodoacetophen- one (14.76 g, 60.00 mmol) were dissolved in ethanol (200 mL), followed by the addition of potassium hydroxide aqueous solution (25.20 g, 450.00 mmol, in 50 mL water) at 0 °C. After stirring at 25 °C for 48 h, water was added into the mixture and pH was adjusted to 7 by 1N HCl. The mixture stood for 6 h. After filtration the residue was further purified by column chromatograph (DCM/MeOH = 100:5, *v/v*) to give 11.77 g of compound **18q** as a yellow solid. Yield: 55%, mp: 160–162 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.65 (s, 1H), 7.97 (d, *J* = 7.9 Hz, 1H), 7.94–7.92 (m, 1H), 7.56–7.54 (m, 1H), 7.52–7.45 (m, 2H), 7.42 (dd, *J* = 8.8 Hz, *J* = 2.5 Hz, 1H), 7.32–7.21 (m, 2H), 6.87 (d, *J* = 8.8 Hz, 1H). ESI-MS *m*/*z* 428.5 [M+H] <sup>+</sup>.

Compounds 18a–18h, 18j, 18k, 18m and 18n were reported before [30]. Compounds 18i, 18l, 18o, 18p and 18r–18w were prepared following the procedure described for the compound 18q.

3.1.3. General Procedure for Synthesis of Compounds **19a–19i**, **19k–19m**, **19o–19q**, **19s**, **19u**, **19w**, **19y**, **19aa**, **19cc**, **19ee** and **19gg** 

1-(5-(2-Hydroxyphenyl)-3-(2-methoxyphenyl)-4,5-dihydro-1H-pyrazol-1-yl)ethan-1one (**19d**)

To a solution of **18d** (5.04 g, 20.00 mmol) in hot acetic acid (25 mL) was added 80% hydrazine hydrate (5.00 g, 80.00 mmol). After stirring at 120  $^{\circ}$ C for 5 h, the mixture was poured into water and the pH was adjusted to 7 by sodium bicarbonate. The formed precipitate was filtered off, dried at 50  $^{\circ}$ C and washed with EtOAc to give 4.03 g of

compound **19d** as a white solid. Yield: 65%, mp: 206–208 °C. <sup>1</sup>H NMR (400 MHz, DMSO*d*<sub>6</sub>):  $\delta$  9.65 (s, 1H), 7.83 (dd, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.43 (td, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.11–6.99 (m, 3H), 6.81 (td, *J* = 7.8 Hz, *J* = 1.4 Hz, 2H), 6.73–6.69 (m, 1H), 5.59 (dd, *J* = 11.7 Hz, *J* = 4.2 Hz, 1H), 3.84 (dd, *J* = 18.4 Hz, *J* = 11.7 Hz, 1H), 3.78 (s, 3H), 3.02 (dd, *J* = 18.4 Hz, *J* = 4.2 Hz, 1H), 2.31 (s, 3H). ESI-MS *m*/*z* 311.2 [M+H] <sup>+</sup>.

Compounds **19a–19c**, **19e–19i**, **19k–19m**, **19o–19q**, **19s**, **19u**, **19w**, **19y**, **19aa**, **19cc**, **19ee** and **19gg** were prepared following the procedure described for the compound **19d**.

3.1.4. General Procedure for Synthesis of Compounds **19***j*, **19***n*, **19***r*, **19***t*, **19***v*, **19***x*, **19***z*, **19b***b*, **19dd**, **19ff** and **19h** 

5-(2-Hydroxyphenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (19j)

Semicarbazide hydrochloride (2.68 g, 24.00 mmol) and sodium hydroxide (3.60 g, 90.00 mmol) were added to the solution of **18i** (7.00 g, 20.00 mmol) in ethanol (100 mL). After stirring at 78 °C for 5 h, the mixture was poured into water and the excess base was neutralized by 10% HCl. The formed precipitate was filtered off, dried at 50 °C and washed with EtOAc to give 5.54 g of compound **19j** as a white solid. Yield: 68%, mp: 176–178 °C. ESI-MS m/z 408.3 [M+H]<sup>+</sup>.

Compounds **19n**, **19r**, **19t**, **19v**, **19x**, **19z**, **19bb**, **19dd**, **19ff** and **19hh** were prepared following the procedure described for the compound **19j**.

3.1.5. General Procedure for Synthesis of Compounds 20a-20z and 20aa-20hh

Methyl 2-(2-(1-acetyl-3-(2-methoxyphenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy) acetate (20d)

To a solution of compound **19d** (1.50 g, 4.84 mmol) in DMF (20 mL) was added 60% NaH (0.23 g, 5.81 mmol) gradually. After stirring at 25 °C for 5 min, methyl bromoacetate (1.11 g, 7.26 mmol) was added. Then the mixture was stirred at 25 °C for 12 h, followed by the addition of water (100 mL). The formed precipitate was filtered off and further purified by column chromatography (DCM/MeOH = 50:1, *v/v*) to give 1.04 g of compound **20d** as a white solid. Yield: 56%, mp: 126–128 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  7.81 (dd, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.43 (td, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.22–7.18 (m, 1H), 7.10 (d, *J* = 8.3 Hz, 1H), 7.01 (t, *J* = 7.5 Hz, 1H), 6.95 (d, *J* = 7.8 Hz, 1H), 6.91–6.90 (m, 2H), 5.67 (dd, *J* = 11.7 Hz, *J* = 4.2 Hz, 1H), 4.95–4.86 (m, 2H), 3.85 (dd, *J* = 18.5 Hz, *J* = 11.7 Hz, 1H), 3.78 (s, 3H), 3.69 (s, 3H), 3.15 (dd, *J* = 18.5 Hz, *J* = 4.2 Hz, 1H), 2.32 (s, 3H). ESI-MS *m/z* 383.5 [M+H] <sup>+</sup>.

Compounds **20a–20c**, **20e–20z** and **20aa–20hh** were prepared following the procedure described for the compound **20d**.

#### 3.1.6. General Procedure for Synthesis of Compounds 14a–14z and 14aa–14hh

2-(2-(1-Acetyl-3-(2-bromophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14a)

A solution of potassium hydroxide (15.00 g, 267.86 mmol) in anhydrous methanol (50 mL) was dropwise added into a solution of hydroxylamine hydrochloride (12.54 g, 180.43 mmol) in anhydrous methanol (50 mL) at 0 °C. After stirring at 0 °C for 40 min, the mixture was filtered and the filtrate was used for the transformation of the methyl ester group to a hydroxamate group. Compound **20a** (1.00 g, 2.32 mmol) was dissolved in the above solution (10 mL) and the mixture was stirred at 25 °C for 0.5 h. Then the mixture was poured into water (100 mL) and 10% H<sub>3</sub>PO<sub>4</sub> solution was added to neutralize the excess base. The formed precipitate was filtered off and dried under vacuum to give 0.54 g of compound **14a** as a white solid. Yield: 54%, mp: 162–164 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.76 (s, 1H), 9.04 (s, 1H), 7.81 (dd, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.45–7.41 (m, 1H), 7.24–7.20 (m, 1H), 7.10 (d, *J* = 8.4 Hz, 1H), 7.03–6.96 (m, 2H), 6.94–6.86 (m, 2H), 5.81 (dd, *J* = 11.7 Hz, *J* = 4.3 Hz, 1H), 4.62–4.54 (m, 2H), 3.88 (dd, *J* = 18.6 Hz, *J* = 11.7 Hz, 1H), 3.79 (s, 3H), 3.14 (dd, *J* = 18.6 Hz, *J* = 4.3 Hz, 1H), 2.32 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  168.25, 164.76, 155.12, 154.77, 134.51, 132.62, 131.73, 131.47, 129.88, 128.89,

128.37, 126.02, 121.65, 121.14, 112.69, 66.53, 55.37, 44.23, 22.24; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>19</sub>BrN<sub>3</sub>O<sub>4</sub>: 432.0559, found: 432.0548.

Compounds **14b–14z** and **14aa–14hh** were prepared following the procedure described for the compound **14a**.

2-(2-(1-Acetyl-3-(3-bromophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14b)

White solid, yield: 55%, mp: 152–154 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.74 (s, 1H), 9.04 (s, 1H), 7.91 (t, *J* = 1.8 Hz, 1H), 7.76 (d, *J* = 7.8 Hz, 1H), 7.66 (dd, *J* = 7.8 Hz, *J* = 2.0 Hz, 1H), 7.42 (t, *J* = 7.9 Hz, 1H), 7.26–7.20 (m, 1H), 6.98 (d, *J* = 8.3 Hz, 1H), 6.91 (d, *J* = 4.5 Hz, 2H), 5.85 (dd, *J* = 11.8 Hz, *J* = 4.6 Hz, 1H), 4.62 (d, *J* = 14.2 Hz, 1H), 4.55 (d, *J* = 14.2 Hz, 1H), 3.80 (dd, *J* = 18.2 Hz, *J* = 11.8 Hz, 1H), 3.21 (dd, *J* = 18.2 Hz, *J* = 4.6 Hz, 1H), 2.36 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  168.07, 164.78, 154.73, 154.28, 134.01, 133.32, 131.40, 129.95, 129.34, 128.88, 126.05, 125.84, 122.57, 121.63, 112.72, 66.67, 55.62, 41.37, 22.21; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>19</sub>BrN<sub>3</sub>O<sub>4</sub>: 432.0559, found: 432.0549.

2-(2-(1-Acetyl-3-(4-bromophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14c)

White solid, yield: 55%, mp: 202–204 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.75 (s, 1H), 9.04 (s, 1H), 7.71–7.65 (m, 4H), 7.26–7.20 (m, 1H), 6.98 (d, *J* = 8.3 Hz, 1H), 6.91 (d, *J* = 4.5 Hz, 2H), 5.85 (dd, *J* = 11.8 Hz, *J* = 4.6 Hz, 1H), 4.62 (d, *J* = 14.2 Hz, 1H), 4.54 (d, *J* = 14.2 Hz, 1H), 3.81 (dd, *J* = 18.1 Hz, *J* = 11.8 Hz, 1H), 3.19 (dd, *J* = 18.1 Hz, *J* = 4.6 Hz, 1H), 2.35 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  167.96, 164.75, 154.73, 154.66, 132.23, 130.89, 129.99, 128.97, 128.86, 125.87, 124.09, 121.63, 112.71, 66.61, 55.59, 41.41, 22.19; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>19</sub>BrN<sub>3</sub>O<sub>4</sub>: 432.0559, found: 432.0563.

2-(2-(1-Acetyl-3-(2-methoxyphenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14d)

White solid, yield: 52%, mp: 170–172 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.76 (s, 1H), 9.04 (s, 1H), 7.81 (dd, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.45–7.41 (m, 1H), 7.24–7.20 (m, 1H), 7.10 (d, *J* = 8.4 Hz, 1H), 7.03–6.96 (m, 2H), 6.94–6.86 (m, 2H), 5.81 (dd, *J* = 11.7 Hz, *J* = 4.3 Hz, 1H), 4.62–4.54 (m, 2H), 3.88 (dd, *J* = 18.6 Hz, *J* = 11.7 Hz, 1H), 3.79 (s, 3H), 3.14 (dd, *J* = 18.6 Hz, *J* = 4.3 Hz, 1H), 2.32 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  167.82, 164.77, 158.45, 154.94, 154.64, 132.17, 130.42, 129.09, 128.72, 125.62, 121.68, 121.08, 120.57, 112.85, 112.62, 66.52, 56.18, 54.87, 44.94, 22.20; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>22</sub>N<sub>3</sub>O<sub>5</sub>: 384.1559, found: 384.1552.

2-(2-(1-Acetyl-3-(3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14e)

White solid, yield: 57%, mp: 170–172 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.76 (s, 1H), 9.06 (s, 1H), 7.40–7.32 (m, 2H), 7.28–7.20 (m, 2H), 7.06–6.98 (m, 2H), 6.94–6.89 (m, 2H), 5.85 (dd, *J* = 11.7 Hz, *J* = 4.4 Hz, 1H), 4.63 (d, *J* = 14.3 Hz, 1H), 4.56 (d, *J* = 14.3 Hz, 1H), 3.84–3.77 (m, 4H), 3.20 (dd, *J* = 18.1 Hz, *J* = 4.4 Hz, 1H), 2.36 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  167.90, 164.78, 159.83, 155.47, 154.70, 132.98, 130.37, 130.12, 128.82, 125.74, 121.65, 119.51, 116.56, 112.69, 112.03, 66.65, 55.68, 55.30, 41.65, 22.17; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>22</sub>N<sub>3</sub>O<sub>5</sub>: 384.1559, found: 384.1552.

2-(2-(1-Acetyl-3-(4-methoxyphenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14f)

White solid, yield: 55%, mp: 120–122 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.76 (s, 1H), 9.06 (s, 1H), 7.70 (d, *J* = 8.6 Hz, 2H), 7.25–7.19 (m, 1H), 7.02–6.97 (m, 3H), 6.93–6.85 (m, 2H), 5.83 (dd, *J* = 11.6 Hz, *J* = 4.2 Hz, 1H), 4.63 (d, *J* = 14.3 Hz, 1H), 4.56 (d, *J* = 14.3 Hz, 1H), 3.79 (s, 3H), 3.78 (dd, *J* = 18.2 Hz, *J* = 11.6 Hz, 1H), 3.15 (dd, *J* = 18.2 Hz, *J* = 4.2 Hz, 1H), 2.34 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  167.60, 164.79, 161.36, 155.35, 154.69, 130.23, 128.77, 128.71, 125.67, 124.12, 121.64, 114.65, 112.64, 66.62, 55.80, 55.04, 41.71, 22.17; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>22</sub>N<sub>3</sub>O<sub>5</sub>: 384.1559, found: 384.1564.

2-(2-(1-Acetyl-3-(2-fluorophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14g)

White solid, yield: 50%, mp: 176–178 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.75 (s, 1H), 9.03 (s, 1H), 7.90–7.85 (m, 1H), 7.54–7.48 (m, 1H), 7.33–7.29 (m, 2H), 7.27–7.20 (m, 1H), 6.99–6.91 (m, 3H), 5.84 (dd, *J* = 11.8 Hz, *J* = 4.5 Hz, 1H), 4.62–4.53 (m, 2H), 3.93–3.85 (m, 1H), 3.20–3.13 (m, 1H), 2.34 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  168.12, 164.79, 159.46, 154.73, 151.82, 132.70, 130.02, 129.53, 128.88, 125.83, 125.28, 121.66, 119.69, 117.18, 112.67, 66.51, 55.08, 43.74, 22.15; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>19</sub>FN<sub>3</sub>O<sub>4</sub>: 372.1360, found: 372.1365.

2-(2-(1-Acetyl-3-(2-chlorophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14h)

White solid, yield: 54%, mp: 166–168 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.74 (s, 1H), 9.02 (s, 1H), 7.72 (dd, *J* = 7.5 Hz, *J* = 2.1 Hz, 1H), 7.55 (dd, *J* = 7.7 Hz, *J* = 1.6 Hz, 1H), 7.48–7.40 (m, 2H), 7.26–7.22 (m, 1H), 6.99–6.91 (m, 3H), 5.84 (dd, *J* = 11.8 Hz, *J* = 4.5 Hz, 1H), 4.62–4.53 (m, 2H), 3.95 (dd, *J* = 18.1 Hz, *J* = 11.8 Hz, 1H), 3.21 (dd, *J* = 18.1 Hz, *J* = 4.5 Hz, 1H), 2.32 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  168.25, 164.80, 154.78, 154.25, 132.12, 131.64, 131.27, 131.17, 130.66, 129.90, 128.92, 127.90, 125.98, 121.68, 112.70, 66.53, 55.37, 44.25, 22.20; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>19</sub>ClN<sub>3</sub>O<sub>4</sub>: 388.1064, found: 388.1072.

2-(2-(1-Acetyl-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14i)

White solid, yield: 58%, mp: 174–176 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.92 (s, 1H), 9.02 (s, 1H), 8.04 (d, *J* = 7.8 Hz, 1H), 7.58–7.47 (m, 3H), 7.21–7.07 (m, 3H), 5.89 (dd, *J* = 12.0 Hz, *J* = 5.4 Hz, 1H), 4.64 (d, *J* = 13.2 Hz, 1H), 4.43 (d, *J* = 13.2 Hz, 1H), 3.94 (dd, *J* = 18.3 Hz, *J* = 12.0 Hz, 1H), 3.16 (dd, *J* = 18.3 Hz, *J* = 5.4 Hz, 1H), 2.31 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  168.05, 164.83, 156.19, 154.82, 137.70, 132.03, 130.45, 130.15, 129.88, 129.79, 128.82, 126.53, 125.79, 121.67, 112.70, 66.60, 54.37, 43.76, 23.37, 22.24; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>19</sub>IN<sub>3</sub>O<sub>4</sub>: 480.0420, found: 480.0418.

5-(2-(2-(Hydroxyamino)-2-oxoethoxy)phenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (14j)

White solid, yield: 51%, mp: 164–166 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.93 (s, 1H), 9.01 (s, 1H), 8.00 (d, *J* = 7.9 Hz, 1H), 7.57–7.52 (m, 2H), 7.50–7.46 (m, 1H), 7.23–7.11 (m, 3H), 6.48 (s, 2H), 5.77 (dd, *J* = 12.2 Hz, *J* = 6.2 Hz, 1H), 4.64 (d, *J* = 13.1 Hz, 1H), 4.41 (d, *J* = 13.1 Hz, 1H), 3.92 (dd, *J* = 18.1 Hz, *J* = 12.2 Hz, 1H), 3.12 (dd, *J* = 18.1 Hz, *J* = 6.2 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.84, 155.54, 154.59, 153.22, 141.08, 136.17, 131.32, 131.27, 130.80, 128.77, 128.72, 126.20, 121.66, 112.44, 95.70, 66.56, 55.10, 44.46; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>18</sub>IN<sub>4</sub>O<sub>4</sub>: 481.0373, found: 481.0371.

2-(2-(1-Acetyl-3-(o-tolyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxyacetamide (14k)

White solid, yield: 50%, mp: 170–172 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.75 (s, 1H), 9.03 (s, 1H), 7.47 (d, J = 7.6 Hz, 1H), 7.33 (d, J = 4.3 Hz, 2H), 7.29–7.21 (m, 2H), 6.99–6.92 (m, 3H), 5.78 (dd, J = 11.7 Hz, J = 4.4 Hz, 1H), 4.64–4.54 (m, 2H), 3.88 (dd, J = 17.8 Hz, J = 11.7 Hz, 1H), 3.22 (dd, J = 17.8 Hz, J = 4.4 Hz, 1H), 2.61 (s, 3H), 2.33 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  168.05, 164.83, 156.19, 154.82, 137.70, 132.03, 130.45, 130.15, 129.88, 129.79, 128.82, 126.53, 125.79, 121.67, 112.70, 66.60, 54.37, 43.76, 23.37, 22.24; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>22</sub>N<sub>3</sub>O<sub>4</sub>: 368.1610, found: 368.1607.

2-(2-(1-Acetyl-3-(2,4-dichlorophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxyacetamide (14l)

White solid, yield: 53%, mp: 128–130 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.67 (s, 1H), 8.95 (s, 1H), 7.68–7.64 (m, 2H), 7.43 (dd, J = 8.6 Hz, J = 2.1 Hz, 1H), 7.17–7.13 (m, 1H), 6.90–6.82 (m, 3H), 5.74 (dd, J = 11.7 Hz, J = 4.6 Hz, 1H), 4.52 (d, J = 14.2 Hz, 1H), 4.45 (d, J = 14.2 Hz, 1H), 3.86 (dd, J = 18.1 Hz, J = 11.7 Hz, 1H), 3.14 (dd, J = 18.1 Hz, J = 4.6 Hz, 1H), 2.24 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  168.28, 164.73, 154.78, 153.21, 135.30, 133.10, 132.40, 130.77, 129.76, 129.59, 128.93, 128.11, 126.10, 121.63, 112.70, 66.48, 55.57, 43.99, 22.20; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>Cl<sub>2</sub>N<sub>3</sub>O<sub>4</sub>: 422.0674, found: 422.0675.

2-(2-(1-Acetyl-3-(2,6-dimethoxyphenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxyacetamide (14m)

White solid, yield: 55%, mp: 178–180 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.70 (s, 1H), 8.98 (s, 1H), 7.36 (t, *J* = 8.4 Hz, 1H), 7.25–7.21 (m, 1H), 7.07 (d, *J* = 7.6 Hz, 1H), 7.01–6.95 (m, 2H), 6.71 (d, *J* = 8.4 Hz, 2H), 5.84 (dd, *J* = 11.7 Hz, *J* = 4.3 Hz, 1H), 4.62–4.53 (m, 2H), 3.73 (s, 3H), 3.66 (dd, *J* = 18.1 Hz, *J* = 11.7 Hz, 1H), 3.33 (s, 2H), 2.75 (dd, *J* = 18.1 Hz, *J* = 4.3 Hz, 1H), 2.21 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  167.69, 164.84, 158.58, 154.43, 153.17, 131.56, 130.84, 128.78, 125.93, 121.87, 112.46, 110.02, 104.78, 66.70, 56.45, 53.69, 45.71, 22.22; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>21</sub>H<sub>24</sub>N<sub>3</sub>O<sub>6</sub>: 414.1665, found: 414.1661.

3-(2,6-Dimethoxyphenyl)-5-(2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (**14n**)

White solid, yield: 52%, mp: 212–214 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.74 (s, 1H), 8.94 (s, 1H), 7.35 (t, *J* = 8.4 Hz, 1H), 7.24–7.17 (m, 2H), 7.00 (t, *J* = 7.5 Hz, 1H), 6.94 (d, *J* = 8.3 Hz, 1H), 6.70 (d, *J* = 8.5 Hz, 2H), 6.33 (s, 2H), 5.73 (dd, *J* = 11.9 Hz, *J* = 4.8 Hz, 1H), 4.61 (d, *J* = 14.4 Hz, 1H), 4.54 (d, *J* = 14.4 Hz, 1H), 3.73 (s, 6H), 3.63 (dd, *J* = 17.9 Hz, *J* = 11.9 Hz, 1H), 2.72 (dd, *J* = 17.9 Hz, *J* = 4.8 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.88, 158.65, 155.48, 154.25, 149.53, 132.23, 131.35, 128.59, 126.36, 121.80, 112.09, 110.34, 104.65, 66.56, 56.37, 53.38, 45.88; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>23</sub>N<sub>4</sub>O<sub>6</sub>: 415.1618, found: 415.1622.

2-(2-(1-Acetyl-3-(2,6-dichlorophenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxyacetamide (140)

White solid, yield: 54%, mp: 174–176 °C. <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.73 (s, 1H), 9.02 (s, 1H), 7.61–7.57 (m, 1H), 7.52 (t, *J* = 8.1 Hz, 1H), 7.29–7.20 (m, 1H), 7.04 (d, *J* = 7.7 Hz, 1H), 7.02–6.95 (m, 2H), 5.98 (dd, *J* = 11.9 Hz, *J* = 4.7 Hz, 1H), 4.60 (d, *J* = 14.2 Hz, 1H), 4.55 (d, *J* = 14.2 Hz, 1H), 3.82 (dd, *J* = 18.6 Hz, *J* = 11.9 Hz, 1H), 2.91 (dd, *J* = 18.6 Hz, *J* = 4.7 Hz, 1H), 2.28 (s, 3H); <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  168.21, 164.73, 154.49, 153.23, 134.25, 132.58, 130.55, 129.99, 129.04, 128.91, 125.53, 121.70, 112.68, 66.72, 54.86, 44.88, 22.17; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>Cl<sub>2</sub>N<sub>3</sub>O<sub>4</sub>: 422.0674, found: 422.0668.

2-(2-(1-Acetyl-3-(naphthalen-1-yl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxy acetamide (14p)

White solid, yield: 55%, mp: 198–200 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.81 (s, 1H), 9.25 (d, J = 8.7 Hz, 1H), 9.06 (s, 1H), 8.04–8.01 (m, 2H), 7.75 (d, J = 7.2 Hz, 1H), 7.72–7.68 (m, 1H), 7.63–7.60 (m, 1H), 7.55 (t, J = 7.7 Hz, 1H), 7.25 (td, J = 7.7 Hz, J = 1.8 Hz, 1H), 7.03–6.99 (m, 2H), 6.94 (t, J = 7.4 Hz, 1H), 5.86 (dd, J = 11.7 Hz, J = 4.3 Hz, 1H), 4.65 (d, J = 14.3 Hz, 1H), 4.58 (d, J = 14.3 Hz, 1H), 4.07 (dd, J = 17.8 Hz, J = 11.7 Hz, 1H), 3.42–3.38 (m, 1H), 2.45 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  168.05, 164.82, 155.98, 154.88, 134.09, 131.38, 130.32, 130.09, 129.41, 129.27, 128.84, 128.17, 127.90, 126.77, 125.89, 125.66, 112.66, 112.70, 66.58, 54.27, 44.11, 22.43; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>23</sub>H<sub>22</sub>N<sub>3</sub>O<sub>4</sub>: 404.1610, found: 404.1605.

2-(2-(1-Acetyl-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazol-5-yl)-6-bromophenoxy)-N-hydroxyacetamide (**14q**)

White solid, yield: 50%, mp: 96–98 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.92 (s, 1H), 9.02 (s, 1H), 8.04 (d, *J* = 7.8 Hz, 1H), 7.58–7.47 (m, 3H), 7.21–7.07 (m, 3H), 5.89 (dd, *J* = 12.0 Hz, *J* = 5.4 Hz, 1H), 4.64 (d, *J* = 13.2 Hz, 1H), 4.43 (d, *J* = 13.2 Hz, 1H), 3.94 (dd, *J* = 18.3 Hz, *J* = 12.0 Hz, 1H), 3.16 (dd, *J* = 18.3 Hz, *J* = 5.4 Hz, 1H), 2.31 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  168.41, 164.17, 155.77, 151.90, 141.24, 138.41, 135.59, 132.92, 131.68, 130.96, 128.86, 127.46, 126.00, 116.95, 95.70, 70.77, 54.74, 44.92, 22.40; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>BrIN<sub>3</sub>O<sub>4</sub>: 557.9525, found: 557.9528.

5-(3-Bromo-2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (**14r**)

White solid, yield: 52%, mp: 140–142 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.93 (s, 1H), 9.01 (s, 1H), 8.00 (d, *J* = 7.9 Hz, 1H), 7.57–7.52 (m, 2H), 7.50–7.46 (m, 1H), 7.23–7.11 (m, 3H), 6.48 (s, 2H), 5.77 (dd, *J* = 12.2 Hz, *J* = 6.2 Hz, 1H), 4.64 (d, *J* = 13.1 Hz, 1H),

4.41 (d, J = 13.1 Hz, 1H), 3.92 (dd, J = 18.1 Hz, J = 12.2 Hz, 1H), 3.12 (dd, J = 18.1 Hz, J = 6.2 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.28, 155.45, 152.79, 151.72, 140.93, 139.58, 136.11, 132.66, 131.39, 130.94, 128.79, 127.34, 126.41, 116.93, 95.96, 70.81, 55.10, 45.27; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>17</sub>BrIN<sub>4</sub>O<sub>4</sub>: 558.9478, found: 558.9482.

2-(2-(1-Acetyl-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazol-5-yl)-5-bromophenoxy)-N-hydroxyacetamide (14s)

White solid, yield: 54%, mp: 168–170 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.74 (s, 1H), 9.08 (s, 1H), 8.02 (d, *J* = 7.9 Hz, 1H), 7.54–7.43 (m, 2H), 7.19–7.10 (m, 3H), 6.96 (d, *J* = 8.2 Hz, 1H), 5.76 (dd, *J* = 11.8 Hz, *J* = 4.8 Hz, 1H), 4.68–4.58 (m, 2H), 3.88 (dd, *J* = 18.1 Hz, *J* = 11.8 Hz, 1H), 3.20 (dd, *J* = 18.1 Hz, *J* = 4.8 Hz, 1H), 2.34 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  168.42, 164.43, 156.35, 155.72, 141.31, 135.66, 131.60, 130.81, 129.41, 128.83, 128.08, 124.45, 121.20, 116.03, 95.54, 66.71, 55.28, 43.69, 22.40; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>BrIN<sub>3</sub>O<sub>4</sub>: 557.9525, found: 557.9520.

5-(4-Bromo-2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (14t)

White solid, yield: 57%, mp: 196–198 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.76 (s, 1H), 9.04 (s, 1H), 7.99 (d, *J* = 7.9 Hz, 1H), 7.50–7.44 (m, 2H), 7.18–7.13 (m, 3H), 7.01 (d, *J* = 8.5 Hz, 1H), 6.49 (s, 2H), 5.68 (dd, *J* = 12.0 Hz, *J* = 5.4 Hz, 1H), 4.69–4.58 (m, 2H), 3.87 (dd, *J* = 18.0 Hz, *J* = 12.0 Hz, 1H), 3.13 (dd, *J* = 18.0 Hz, *J* = 5.4 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.48, 155.51, 155.48, 153.43, 141.00, 136.14, 131.32, 130.82, 130.79, 128.77, 128.04, 124.47, 120.96, 115.78, 95.80, 66.73, 55.06, 44.12; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>17</sub>BrIN<sub>4</sub>O<sub>4</sub>: 558.9478, found: 558.9482.

2-(2-(1-Acetyl-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazol-5-yl)-4-bromophenoxy)-N-hydroxyacetamide (14u)

White solid, yield: 51%, mp: 106–108 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.76 (s, 1H), 9.04 (s, 1H), 8.03 (d, *J* = 7.9 Hz, 1H), 7.53–7.42 (m, 3H), 7.18 (t, *J* = 7.5 Hz, 1H), 7.11 (d, *J* = 2.5 Hz, 1H), 6.97 (d, *J* = 8.8 Hz, 1H), 5.79 (dd, *J* = 11.9 Hz, *J* = 4.9 Hz, 1H), 4.64–4.53 (m, 2H), 3.89 (dd, *J* = 18.2 Hz, *J* = 11.9 Hz, 1H), 3.23 (dd, *J* = 18.2 Hz, *J* = 4.9 Hz, 1H), 2.35 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  168.58, 164.46, 156.34, 154.25, 141.36, 135.52, 132.41, 131.65, 131.49, 130.85, 128.87, 115.21, 113.23, 95.47, 66.72, 55.30, 43.74, 22.41; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>BrIN<sub>3</sub>O<sub>4</sub>: 557.9525, found: 557.9522.

5-(5-Bromo-2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (**14v**)

White solid, yield: 54%, mp: 202–204 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.76 (s, 1H), 9.02 (s, 1H), 8.00 (d, *J* = 7.9 Hz, 1H), 7.51–7.41 (m, 3H), 7.18–7.14 (m, 2H), 6.96 (d, *J* = 8.8 Hz, 1H), 6.53 (s, 2H), 5.71 (dd, *J* = 12.0 Hz, *J* = 5.4 Hz, 1H), 4.64–4.54 (m, 2H), 3.89 (dd, *J* = 18.0 Hz, *J* = 12.0 Hz, 1H), 3.15 (dd, *J* = 18.0 Hz, *J* = 5.4 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.48, 155.53, 154.04, 153.53, 141.04, 136.04, 133.76, 131.38, 131.23, 130.82, 128.81, 128.76, 115.01, 113.24, 95.72, 66.77, 55.16, 44.18; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>17</sub>BrIN<sub>4</sub>O<sub>4</sub>: 558.9478, found: 558.9472.

2-(2-(1-Acetyl-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazol-5-yl)-3-bromophenoxy)-N-hydroxyacetamide (**14w**)

White solid, yield: 55%, mp: 152–154 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.65 (s, 1H), 8.98 (s, 1H), 8.08–8.02 (m, 1H), 7.61–7.59 (m, 1H), 7.53–7.45 (m, 1H), 7.24–7.06 (m, 3H), 6.93–6.82 (m, 1H), 6.28–5.86 (m, 1H), 4.66–4.45 (m, 2H), 3.93–3.73 (m, 1H), 3.61–3.23 (m, 1H), 2.26–2.20 (m, 3H); HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>BrIN<sub>3</sub>O<sub>4</sub>: 557.9525, found: 557.9520.

5-(2-Bromo-6-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (**14***x*)

White solid, yield: 59%, mp: 132–134 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.62 (m, 1H), 8.99 (s, 1H), 8.04–8.00 (m, 1H), 7.61–7.57 (m, 1H), 7.51–7.44 (m, 1H), 7.28–6.93 (m, 4H), 6.21–5.79 (m, 3H), 4.72–4.43 (m, 2H), 3.88–3.70 (m, 1H), 3.64–3.28 (m, 1H); HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>17</sub>BrIN<sub>4</sub>O<sub>4</sub>: 558.9478, found: 558.9484.

2-(2-(1-Acetyl-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazol-5-yl)-3-chlorophenoxy)-N-hydroxyacetamide (**14y**)

White solid, yield: 56%, mp: 168–170 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.68–10.63 (m, 1H), 9.01 (s, 1H), 8.08–8.03 (m, 1H), 7.61–7.46 (m, 2H), 7.30–7.15 (m, 2H), 7.08–6.88 (m, 2H), 6.26–5.89 (m, 1H), 4.68–4.45 (m, 2H), 3.95–3.73 (m, 1H), 3.64–3.24 (m, 1H), 2.25–2.21 (m, 3H); HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>19</sub>H<sub>18</sub>ClIN<sub>3</sub>O<sub>4</sub>: 514.0031, found: 514.0035.

5-(2-Chloro-6-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-3-(2-iodophenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (**14z**)

White solid, yield: 57%, mp: 190–192 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.62 (s, 1H), 8.97 (s, 1H), 8.04–8.00 (m, 1H), 7.59–7.56 (m, 1H), 7.51–7.44 (m, 1H), 7.28–7.00 (m, 4H), 6.19–5.83 (m, 3H), 4.73–4.44 (m, 2H), 3.90–3.70 (m, 1H), 3.67–3.28 (m, 1H); HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>17</sub>ClIN<sub>4</sub>O<sub>4</sub>: 514.9983, found: 514.9987.

2-(2-(1-Acetyl-3-(2-(benzyloxy)phenyl)-4,5-dihydro-1H-pyrazol-5-yl)phenoxy)-N-hydroxyacetamide (14aa)

White solid, yield: 55%, mp: 162–164 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.74 (s, 1H), 9.02 (s, 1H), 7.77 (d, *J* = 7.8 Hz, 1H), 7.44–7.39 (m, 3H), 7.36–7.28 (m, 3H), 7.24–7.20 (m, 2H), 7.02 (t, *J* = 7.5 Hz, 1H), 6.95 (d, *J* = 8.3 Hz, 1H), 6.92–6.86 (m, 2H), 5.79 (dd, *J* = 11.6 Hz, *J* = 4.2 Hz, 1H), 5.15 (s, 2H), 4.57 (d, *J* = 14.2 Hz, 1H), 4.47 (d, *J* = 14.2 Hz, 1H), 3.82 (dd, *J* = 18.5 Hz, *J* = 11.6 Hz, 1H), 3.18 (dd, *J* = 18.5 Hz, *J* = 4.2 Hz, 1H), 2.29 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  167.88, 164.76, 157.33, 154.92, 154.61, 136.97, 132.05, 130.28, 129.54, 128.90, 128.76, 128.34, 128.14, 125.58, 121.66, 121.32, 121.01, 114.02, 112.57, 70.37, 66.53, 54.82, 44.91, 22.19; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>26</sub>H<sub>26</sub>N<sub>3</sub>O<sub>5</sub>: 460.1872, found: 460.1868.

3-(2-(Benzyloxy)phenyl)-5-(2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-4,5-dihydro-1H-pyrazole-1-carboxamide (**14bb**)

White solid, yield: 52%, mp: 176–178 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.75 (s, 1H), 8.99 (s, 1H), 7.88 (d, J = 7.7 Hz, 1H), 7.40–7.28 (m, 6H), 7.22–7.17 (m, 2H), 7.02–6.88 (m, 4H), 6.47 (s, 2H), 5.70 (dd, J = 11.9 Hz, J = 4.8 Hz, 1H), 5.17–5.10 (m, 2H), 4.58 (d, J = 14.4 Hz, 1H), 4.47 (d, J = 14.4 Hz, 1H), 3.80 (dd, J = 18.2 Hz, J = 11.9 Hz, 1H), 3.15 (dd, J = 18.2 Hz, J = 4.8 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.79, 157.09, 155.48, 154.40, 151.32, 137.04, 131.75, 131.55, 129.58, 128.92, 128.55, 128.35, 128.15, 125.74, 121.64, 121.34, 121.23, 113.93, 112.29, 70.34, 66.48, 54.54, 45.22; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>25</sub>H<sub>25</sub>N<sub>4</sub>O<sub>5</sub>: 461.1825, found: 461.1830.

2-(2-(1-Acetyl-3-(2-((2-bromobenzyl)oxy)phenyl)-4,5-dihydro-1H-pyrazol-5-yl) phenoxy)-N-hydroxyacetamide (**14cc**)

White solid, yield: 53%, mp: 82–84 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.68 (s, 1H), 9.04 (s, 1H), 7.79 (d, *J* = 7.7 Hz, 1H), 7.64 (d, *J* = 8.0 Hz, 1H), 7.58 (d, *J* = 7.6 Hz, 1H), 7.47–7.36 (m, 2H), 7.30 (t, *J* = 7.6 Hz, 1H), 7.23–7.19 (m, 2H), 7.06 (t, *J* = 7.5 Hz, 1H), 6.94 (d, *J* = 8.3 Hz, 1H), 6.90–6.84 (m, 2H), 5.79 (dd, *J* = 11.7 Hz, *J* = 4.2 Hz, 1H), 5.20–5.12 (m, 2H), 4.56 (d, *J* = 14.3 Hz, 1H), 4.45 (d, *J* = 14.3 Hz, 1H), 3.78 (dd, *J* = 18.5 Hz, *J* = 11.7 Hz, 1H), 3.16 (dd, *J* = 18.5 Hz, *J* = 4.2 Hz, 1H), 2.29 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  167.82, 164.65, 157.14, 154.73, 154.54, 135.84, 133.12, 132.10, 130.92, 130.70, 130.18, 129.63, 128.70, 128.34, 125.51, 123.33, 121.59, 121.00, 113.91, 112.49, 70.31, 66.51, 54.73, 44.75, 22.15; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>26</sub>H<sub>25</sub>BrN<sub>3</sub>O<sub>5</sub>: 538.0978, found: 538.0982.

3-(2-((2-Bromobenzyl)oxy)phenyl)-5-(2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-4,5dihydro-1H-pyrazole-1-carboxamide (**14dd**)

White solid, yield: 56%, mp: 92–94 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.67 (s, 1H), 9.03 (s, 1H), 7.90 (d, *J* = 7.8 Hz, 1H), 7.64 (d, *J* = 7.8 Hz, 1H), 7.55 (d, *J* = 7.5 Hz, 1H), 7.43–7.27 (m, 3H), 7.22–7.17 (m, 2H), 7.04 (t, *J* = 7.5 Hz, 1H), 6.94–6.87 (m, 3H), 6.49 (s, 2H), 5.70 (dd, *J* = 11.8 Hz, *J* = 4.8 Hz, 1H), 5.19–5.11 (m, 2H), 4.57 (d, *J* = 14.4 Hz, 1H), 4.45 (d, *J* = 14.4 Hz, 1H), 3.75 (dd, *J* = 18.4 Hz, *J* = 11.8 Hz, 1H), 3.13 (dd, *J* = 18.4 Hz, *J* = 4.8 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  164.68, 156.92, 155.41, 154.33, 151.15, 135.87, 133.14, 131.64, 131.60, 130.94, 130.72, 129.63, 128.51, 128.36, 125.69, 123.38, 121.57, 121.48, 121.34, 113.79, 112.19,

70.28, 66.44, 54.46, 45.04; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>25</sub>H<sub>24</sub>BrN<sub>4</sub>O<sub>5</sub>: 539.0930, found: 539.0934.

2-(2-(1-Acetyl-3-(2-((3-bromobenzyl)oxy)phenyl)-4,5-dihydro-1H-pyrazol-5-yl) phenoxy)-N-hydroxyacetamide (**14ee**)

White solid, yield: 57%, mp: 164–166 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.73 (s, 1H), 9.01 (s, 1H), 7.74 (dd, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.66 (d, *J* = 2.2 Hz, 1H), 7.51 (d, *J* = 8.6 Hz, 1H), 7.43 (t, *J* = 7.4 Hz, 2H), 7.31 (t, *J* = 7.8 Hz, 1H), 7.23–7.18 (m, 2H), 7.03 (t, *J* = 7.5 Hz, 1H), 6.95 (d, *J* = 8.0 Hz, 1H), 6.89 (d, *J* = 7.4 Hz, 1H), 5.79 (dd, *J* = 11.6 Hz, *J* = 4.2 Hz, 1H), 5.17 (s, 2H), 4.58 (d, *J* = 14.4 Hz, 1H), 4.50 (d, *J* = 14.4 Hz, 1H), 3.85 (dd, *J* = 18.4 Hz, *J* = 11.6 Hz, 1H), 3.17 (dd, *J* = 18.4 Hz, *J* = 4.2 Hz, 1H), 2.29 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  167.87, 164.78, 157.01, 154.69, 154.63, 139.89, 132.01, 131.18, 131.14, 130.75, 130.24, 129.69, 128.77, 127.07, 125.61, 122.11, 121.65, 121.49, 121.06, 114.03, 112.57, 69.40, 66.51, 54.74, 44.81, 22.20; HRMS (AP-ESI) *m*/*z* [M+H]<sup>+</sup> calcd for C<sub>26</sub>H<sub>25</sub>BrN<sub>3</sub>O<sub>5</sub>: 538.0978, found: 538.0986.

3-(2-((3-Bromobenzyl)oxy)phenyl)-5-(2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-4,5dihydro-1H-pyrazole-1-carboxamide (**14ff**)

White solid, yield: 59%, mp: 120–122 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.74 (s, 1H), 8.98 (s, 1H), 7.87 (dd, J = 7.8 Hz, J = 1.8 Hz, 1H), 7.62 (t, J = 1.8 Hz, 1H), 7.51–7.49 (m, 1H), 7.41–7.36 (m, 2H), 7.30 (t, J = 7.8 Hz, 1H), 7.22–7.14 (m, 2H), 7.03–6.88 (m, 4H), 6.46 (s, 2H), 5.70 (dd, J = 11.9 Hz, J = 4.8 Hz, 1H), 5.16 (s, 2H), 4.59 (d, J = 14.4 Hz, 1H), 4.50 (d, J = 14.4 Hz, 1H), 3.81 (dd, J = 18.3 Hz, J = 11.9 Hz, 1H), 3.14 (dd, J = 18.3 Hz, J = 4.8 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  164.79, 156.81, 155.46, 154.40, 151.18, 139.92, 131.72, 131.54, 131.18, 130.80, 129.64, 128.56, 127.10, 125.77, 122.10, 121.64, 121.41, 113.97, 112.28, 69.36, 66.45, 54.53, 45.19; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>25</sub>H<sub>24</sub>BrN<sub>4</sub>O<sub>5</sub>: 539.0930, found: 539.0936.

2-(2-(1-Acetyl-3-(2-((4-bromobenzyl)oxy)phenyl)-4,5-dihydro-1H-pyrazol-5-yl) phenoxy)-N-hydroxyacetamide (**14gg**)

White solid, yield: 53%, mp: 96–98 °C. <sup>1</sup>H NMR (400 MHz, DMSO- $d_6$ ):  $\delta$  10.75 (s, 1H), 9.02 (s, 1H), 7.74 (dd, J = 7.7 Hz, J = 1.8 Hz, 1H), 7.55–7.52 (m, 2H), 7.44–7.36 (m, 3H), 7.24–7.17 (m, 2H), 7.03 (t, J = 7.5 Hz, 1H), 6.95 (d, J = 8.3 Hz, 1H), 6.91–6.85 (m, 2H), 5.78 (dd, J = 11.6 Hz, J = 4.2 Hz, 1H), 5.13 (d, J = 1.8 Hz, 2H), 4.58 (d, J = 14.2 Hz, 1H), 4.50 (d, J = 14.2 Hz, 1H), 3.82 (dd, J = 18.4 Hz, J = 11.6 Hz, 1H), 3.18 (dd, J = 18.4 Hz, J = 4.2 Hz, 1H), 2.29 (s, 3H); <sup>13</sup>C NMR (100 MHz, DMSO- $d_6$ ):  $\delta$  167.86, 164.78, 157.11, 154.95, 154.64, 136.45, 132.03, 131.84, 130.37, 130.23, 129.62, 128.74, 125.63, 121.64, 121.54, 121.44, 121.10, 114.01, 112.54, 69.57, 66.48, 54.85, 44.81, 22.19; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>26</sub>H<sub>25</sub>BrN<sub>3</sub>O<sub>5</sub>: 538.0978, found: 538.0980.

3-(2-((4-Bromobenzyl)oxy)phenyl)-5-(2-(2-(hydroxyamino)-2-oxoethoxy)phenyl)-4,5dihydro-1H-pyrazole-1-carboxamide (**14hh**)

White solid, yield: 55%, mp: 96–98 °C. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  10.76 (s, 1H), 8.99 (s, 1H), 7.85 (dd, *J* = 7.8 Hz, *J* = 1.8 Hz, 1H), 7.53 (d, *J* = 8.3 Hz, 2H), 7.40–7.35 (m, 3H), 7.23–7.14 (m, 2H), 7.02–6.88 (m, 4H), 6.47 (s, 2H), 5.69 (dd, *J* = 11.9 Hz, *J* = 4.7 Hz, 1H), 5.16–5.09 (m, 2H), 4.59 (d, *J* = 14.4 Hz, 1H), 4.50 (d, *J* = 14.4 Hz, 1H), 3.80 (dd, *J* = 18.3 Hz, *J* = 11.9 Hz, 1H), 3.14 (dd, *J* = 18.3 Hz, *J* = 4.7 Hz, 1H); <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  164.80, 156.86, 155.44, 154.42, 151.32, 136.53, 131.85, 131.68, 131.52, 130.36, 129.63, 128.54, 125.76, 121.61, 121.53, 121.44, 121.34, 113.95, 112.25, 69.51, 66.43, 54.58, 45.12; HRMS (AP-ESI) m/z [M+H]<sup>+</sup> calcd for C<sub>25</sub>H<sub>24</sub>BrN<sub>4</sub>O<sub>5</sub>: 539.0930, found: 539.0938.

#### 3.2. Biological Evaluation

#### 3.2.1. Enzymatic Inhibition Assay against APN In Vitro

The enzymatic inhibitory activities of target compounds were evaluated using *L*-Leu*p*-nitroanilide (Sigma-Aldrich, St. Louis, MO, USA) as substrate and soluble APN from porcine kidney microsomal (Enzo Life Sciences, Farmingdale, NY, USA) as enzyme. Firstly, the substrate was dissolved in DMSO to a concentration of 16 mmol/L and the enzyme was dissolved in buffer solution of 50 mM PBS (pH = 7.2) for a concentration of 0.15 IU/L.

16 of 18

Subsequently, inhibitors (40  $\mu$ L) and PBS (145  $\mu$ L) were added into the 96-well plates, followed by the addition of substrate (5  $\mu$ L) and enzyme (10  $\mu$ L). After incubation at 37 °C for 30 min, the absorbance values were measured at 405 nm with a plate reader (Varioskan, Thermo Fisher Scientific, Waltham, MA, USA).

# 3.2.2. Anti-Proliferative Assay

The MTT method was used for the determination of anti-proliferative activities of selected compounds against tumor cells. Human histiocytic lymphoma cells U937, human chronic myeloid leukemia cells K562, liver carcinoma alexander cells PLC/PRF/5, human prostate cancer cells PC-3, human ovarian clear cell carcinoma cells ES-2, human hepatocellular carcinoma cells HepG2 were selected as tested tumor cell lines. Firstly, the cells were cultured in RPMI 1640 medium with 10% FBS at 37 °C in 5% CO<sub>2</sub> humidified incubator. Cells (100  $\mu$ L) in logarithmic phase were inoculated in 96-well plates and allowed to grow for 4 h. Subsequently, inhibitors (100  $\mu$ L) at the tested concentration were added, followed by incubation for 48 h. Then, to the mixture was added MTT solution (20  $\mu$ L, 5 mg/mL) and further incubated for 4 h. The plates were centrifuged at 800 rpm for 3 min. The supernatant was poured off and the formed formazan was dissolved in DMSO (200  $\mu$ L). Finally, the mixture was shaken for 15 min and the absorbance values of the formazan solution were measured at 570 nm with a plate reader (Varioskan, Thermo, Waltham, MA, USA). All the tumor cells were owned by our laboratory.

# 3.2.3. Rat Aortic Ring Assay

The thoracic aortas were separated from 8- to 10-week-old male Sprague Dawley rats and cut into rings with a 1-mm-long cross-section after removal of the connective tissues around the aortas. The fresh aortic ring was added into the 96-well plates pre-coated with matrigel (50  $\mu$ L, BD bioscience), and covered with another 50  $\mu$ L of matrigel. After incubation at 37 °C in 5% CO<sub>2</sub> for 0.5 h, the tested compounds (100  $\mu$ L) in M199 culture medium with 10% FBS was added to the mixture and incubated at 37 °C in 5% CO<sub>2</sub> for 9 d. For the treatment group, the M199 culture medium with 10% FBS and inhibitors was changed every three days. For the control group, the M199 culture medium with 10% FBS and 0.5% DMSO was changed every three days. The formed micro-vessels were photographed by inverted microscope at 100× magnification. Experiments were repeated three times. All experiments involving laboratory animals were performed with the approval of the Laboratory Animal Welfare Review Committee of Shandong University of Traditional Chinese Medicine.

#### 3.3. Molecular Docking Studies

In the APN-bestatin co-crystal structure (PDB code: 2DQM), the residues in a 10.0 Å radius circle around bestatin was selected as the active site. Compound **140** was docked into it using Sybyl 2.1. Before the docking studies, the protein structure was prepared by deleting water molecules, adding hydrogen atoms, modifying atom types, and assigning with AMBER7 FF99 charges, followed by a 100-step minimization process. Using the Sybyl/Sketch module, the small molecular structures were prepared and further optimized using Powell's method with the Tripos force field with convergence criterion set at 0.05 kcal/Å mol and assigned using the Gasteiger–Hückel method. Other docking parameters implied in the program were default values. Molecular docking was carried out via the Sybyl/Surflex-Dock module. The top-scoring pose was selected for discussions.

# 4. Conclusions

Summarily, one series of pyrazoline-based hydroxamate derivatives was designed, synthesized and evaluated as APNIs. Interestingly, in the enzymatic assay against APN, some compounds exhibited improved APN inhibitory activities relative to the lead compounds **13a–13c** and positive control bestatin. Among them, the 2,6-dichloro substituted compound **14o** (IC<sub>50</sub> =  $0.0062 \pm 0.0004 \mu$ M) was the best one, with three orders of magni-

tude better APN inhibitory activity than bestatin. The SARs revealed that the substituents on the two phenyl groups of the diphenyl pyrazoline scaffold significantly impacted the capacities of inhibiting APN. Compounds with an *ortho* mono-substituent on the terminal phenyl group presented better APN inhibitory activities than their counterparts with a *meta* or *para* mono-substituent. Compounds with an *ortho*-substituent, such as 2-F (**14g**), 2-Cl (**14h**), 2-I (**14i**), 2-CH<sub>3</sub> (**14k**), showed improved capacities of APN inhibition relative to the unsubstituted lead compound **13a**. However, introduction of substituents on another phenyl group resulted in decreased APN inhibitory activities. The putative interactions of **14o** with the active site of APN was predicted using Surflex-Dock module of Sybyl 2.1., which further supported our design strategy. Moreover, compared with bestatin, compound **14o** demonstrated superior anti-proliferative activities against tumor cells and anti-angiogenic activity, which confirmed the potency of **14o** as an anti-tumor lead.

**Supplementary Materials:** The following supporting information can be downloaded at: https://www. mdpi.com/article/10.3390/molecules27238339/s1, the spectra data of some of intermediates **18**, **19** and **20**; the <sup>1</sup>HNMR and <sup>13</sup>C NMR spectra of target compounds **14h–14j**, **14o**, **14aa–14cc**, **14ee** and **14ff**.

**Author Contributions:** Conceptualization, J.C.; methodology, Y.L. and D.Z.; validation, C.Z.; formal analysis, H.F.; investigation, Y.L.; writing—original draft preparation, Y.L. and J.C.; writing—review and editing, Q.S. and Z.W.; supervision, J.C.; funding acquisition, J.C. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Natural Science Foundation of Shandong Province (Grant No. ZR2022QH186), Medical Science and Technology Development Project of Shandong Province (Grant No. 202013051070) and Strengthening Foundation Plan for Young Teachers of School of Pharmacy (Grant No. 2021-0013, No. 2021-009). The APC was funded by the Natural Science Foundation of Shandong Province (Grant No. ZR2022QH186).

**Institutional Review Board Statement:** The animal study protocol was approved by the Laboratory Animal Welfare Review Committee of Shandong University of Traditional Chinese Medicine (protocol code: SDUTCM20220303013). The date of approval was 3 March 2022.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds 14i and 14o are available from the authors.

# References

- 1. Antczak, C.; De Meester, I.; Bauvois, B. Ectopeptidases in pathophysiology. *Bioessays* 2001, 23, 251–260. [CrossRef] [PubMed]
- Mina-Osorio, P. The moonlighting enzyme CD13: Old and new functions to target. *Trends. Mol. Med.* 2008, 14, 361–371. [CrossRef] [PubMed]
- 3. Hooper, N.M. Families of zinc metalloproteases. *FEBS. Lett.* **1994**, *354*, 1–6. [CrossRef] [PubMed]
- 4. Roques, B.P.; Noble, F.; Daugé, V.; Fournie-Zaluski, M.C.; Beaumont, A. Neutral endopeptidase 24.11: Structure, inhibition, and experimental and clinical pharmacology. *Pharmacol. Rev.* **1993**, *45*, 87–146. [PubMed]
- Zini, S.; Fournie-Zaluski, M.C.; Chauvel, E.; Roques, B.P.; Corvol, P.; Llorens-Cortes, C. Identification of metabolic pathways of brain angiotensin II and III using specific aminopeptidase inhibitors: Predominant role of angiotensin III in the control of vasopressin release. *Proc. Natl. Acad. Sci. USA* 1996, 93, 11968–11973. [CrossRef] [PubMed]
- Wirtz, D.; Konstantopoulos, K.; Searson, P.C. The physics of cancer: The role of physical interactions and mechanical forces in metastasis. *Nat. Rev. Cancer* 2011, 11, 512–522. [CrossRef]
- 7. Saiki, I.; Fujii, H.; Yoneda, J.; Abe, F.; Nakajima, M.; Tsuruo, T.; Azuma, I. Role of aminopeptidase N (CD13) in tumor-cell invasion and extracellular matrix degradation. *Int. J. Cancer* **1993**, *54*, 137–143. [CrossRef]
- Fujii, H.; Nakajima, M.; Saiki, I.; Yoneda, J.; Azuma, I.; Tsuruo, T. Human melanoma invasion and metastasis enhancement by high expression of aminopeptidase N/CD13. *Clin. Exp. Metastasis* 1995, 13, 337–344. [CrossRef]
- Yoneda, J.; Saiki, I.; Fujii, H.; Abe, F.; Kojima, Y.; Azuma, I. Inhibition of tumor invasion and extracellular matrix degradation by ubenimex (bestatin). *Clin. Exp. Metastasis* 1992, 10, 49–59. [CrossRef]
- 10. Bhagwat, S.V.; Lahdenranta, J.; Giordano, R.; Arap, W.; Pasqualini, R.; Shapiro, L.H. CD13/APN is activated by angiogenic signals and is essential for capillary tube formation. *Blood* 2001, *97*, 652–659. [CrossRef]

- Rangel, R.; Sun, Y.; Guzman-Rojas, L.; Ozawa, M.G.; Sun, J.; Giordano, R.J.; Van Pelt, C.S.; Tinkey, P.T.; Behringer, R.B.; Sidman, R.L.; et al. Impaired angiogenesis in aminopeptidase N-null mice. *Proc. Natl. Acad. Sci. USA* 2007, 104, 4588–4593. [CrossRef] [PubMed]
- Guzman-Rojas, L.; Rangel, R.; Salameh, A.; Edwards, J.K.; Dondossola, E.; Kim, Y.G.; Saghatelian, A.; Giordano, R.J.; Kolonin, M.G.; Staquicini, F.I.; et al. Cooperative effects of aminopeptidase N (CD13) expressed by nonmalignant and cancer cells within the tumor microenvironment. *Proc. Natl. Acad. Sci. USA* 2012, 109, 1637–1642. [CrossRef] [PubMed]
- Wickström, M.; Larsson, R.; Nygren, P.; Gullbo, J. Aminopeptidase N (CD13) as a target for cancer chemotherapy. *Cancer Sci.* 2011, 102, 501–508. [CrossRef]
- 14. Haraguchi, N.; Ishii, H.; Mimori, K.; Tanaka, F.; Ohkuma, M.; Kim, H.M.; Akita, H.; Takiuchi, D.; Hatano, H.; Nagano, H.; et al. CD13 is a therapeutic target in human liver cancer stem cells. *J. Clin. Investig.* **2010**, *120*, 3326–3339. [CrossRef] [PubMed]
- Olsen, J.; Cowell, G.M.; Kønigshøfer, E.; Danielsen, M.; Møller, J.; Laustsen, L.; Hansen, O.C.; Welinder, K.G.; Engberg, J.; Hunziker, W.; et al. Complete amino acid sequence of human intestinal aminopeptidase N as deduced from cloned cDNA. *FEBS*. *Lett.* 1988, 238, 307–314. [CrossRef] [PubMed]
- 16. Wong, A.H.; Zhou, D.; Rini, J.M. The X-ray crystal structure of human aminopeptidase N reveals a novel dimer and the basis for peptide processing. *J. Biol. Chem.* **2012**, *287*, 36804–36813. [CrossRef] [PubMed]
- Bauvois, B.; Dauzonne, D. Aminopeptidase-N/CD13 (EC 3.4.11.2) inhibitors: Chemistry, biological evaluations, and therapeutic prospects. *Med. Res. Rev.* 2006, 26, 88–130. [CrossRef]
- 18. Mucha, A.; Drag, M.; Dalton, J.P.; Kafarski, P. Metallo-aminopeptidase inhibitors. Biochimie 2010, 92, 1509–1529. [CrossRef]
- Cao, J.Y.; Zang, J.; Kong, X.J.; Zhao, C.L.; Chen, T.; Ran, Y.Y.; Dong, H.; Xu, W.F.; Zhang, Y.J. Leucine ureido derivatives as aminopeptidase N inhibitors using click chemistry. Part II. *Bioorg. Med. Chem.* 2019, 27, 978–990. [CrossRef]
- Roques, B.P.; Fournié-Zaluski, M.C.; Wurm, M. Inhibiting the breakdown of endogenous opioids and cannabinoids to alleviate pain. Nat. Rev. Drug. Discov. 2012, 11, 292–310. [CrossRef]
- Bauvois, B.; Puiffe, M.L.; Bongui, J.B.; Paillat, S.; Monneret, C.; Dauzonne, D. Synthesis and biological evaluation of novel flavone-8-acetic acid derivatives as reversible inhibitors of aminopeptidase N/CD13. J. Med. Chem. 2003, 46, 3900–3913. [CrossRef]
- Ma, C.H.; Li, X.G.; Liang, X.W.; Jin, K.; Cao, J.Y.; Xu, W.F. Novel β-dicarbonyl derivatives as inhibitors of aminopeptidase N (APN). *Bioorg. Med. Chem. Lett.* 2013, 23, 4948–4952. [CrossRef]
- 23. Yang, K.H.; Wang, Q.; Su, L.; Fang, H.; Wang, X.J.; Gong, J.Z.; Wang, B.H.; Xu, W.F. Design and synthesis of novel chloramphenicol amine derivatives as potent aminopeptidase N (APN/CD13) inhibitors. *Bioorg. Med. Chem.* 2009, *17*, 3810–3817. [CrossRef]
- 24. Maiereanu, C.; Schmitt, C.; Schifano-Faux, N.; Le Nouën, D.; Defoin, A.; Tarnus, C. A novel amino-benzosuberone derivative is a picomolar inhibitor of mammalian aminopeptidase N/CD13. *Bioorg. Med. Chem.* **2011**, *19*, 5716–5733. [CrossRef] [PubMed]
- Pan, H.L.; Yang, K.H.; Zhang, J.; Xu, Y.Y.; Jiang, Y.Q.; Yuan, Y.M.; Zhang, X.P.; Xu, W.F. Design, synthesis and biological evaluation of novel l-isoserine tripeptide derivatives as aminopeptidase N inhibitors. *J. Enzyme. Inhib. Med. Chem.* 2013, 28, 717–726. [CrossRef] [PubMed]
- Lee, J.; Vinh, N.B.; Drinkwater, N.; Yang, W.; Sivaraman, K.; Schembri, L.S.; Gazdik, M.; Grin, P.M.; Butler, G.S.; Overall, C.M.; et al. Novel human aminopeptidase N inhibitors: Discovery and optimization of subsite binding interactions. J. Med. Chem. 2019, 62, 7185–7209. [CrossRef] [PubMed]
- 27. Mishima, Y.; Terui, Y.; Sugimura, N.; Matsumoto-Mishima, Y.; Rokudai, A.; Kuniypshi, R.; Hatake, K. Continuous treatment of bestatin induces anti-angiogenic property in endothelial cells. *Cancer. Sci.* 2007, *98*, 364–372. [CrossRef]
- 28. Saitoh, Y.; Koizumi, K.; Minami, T.; Sekine, K.; Sakurai, H.; Saiki, I. A derivative of aminopeptidase inhibitor (BE15) has a dual inhibitory effect of invasion and motility on tumor and endothelial cells. *Biol. Pharm. Bull.* **2006**, *29*, 709–712. [CrossRef]
- Cao, J.Y.; Zang, J.; Ma, C.H.; Li, X.G.; Hou, J.N.; Li, J.; Huang, Y.X.; Xu, W.F.; Wang, B.H.; Zhang, Y.J. Design, synthesis, and biological evaluation of pyrazoline-based hydroxamic acid derivatives as aminopeptidase N (APN) inhibitors. *ChemMedChem* 2018, 13, 431–436. [CrossRef]
- 30. Cao, J.Y.; Zhao, C.L.; Dong, H.; Xu, Q.F.; Zhang, Y.J. Development of pyrazoline-based derivatives as aminopeptidase N inhibitors to overcome cancer invasion and metastasis. *RSC Adv.* **2021**, *11*, 21426–21432. [CrossRef]