

## Article

# Phytochemical Composition and In Vitro Antioxidant, Anti-Inflammatory, Anticancer, and Enzyme-Inhibitory Activities of *Artemisia nilagirica* (C.B. Clarke) Pamp

Jawaher J. Albaqami <sup>1,†</sup>, Tancia P. Benny <sup>2,†</sup>, Hamida Hamdi <sup>1,3</sup> , Ammar B. Altemimi <sup>4,5</sup> ,  
Aswathi Moothakoottil Kuttithodi <sup>2</sup> , Joice Tom Job <sup>2,\*</sup> , Anju Sasidharan <sup>2</sup>  
and Arunaksharan Narayanankutty <sup>2,\*</sup> 

<sup>1</sup> Department of Biology, College of Science, Taif University, P.O. Box 11099, Taif 21944, Saudi Arabia

<sup>2</sup> Division of Cell and Molecular Biology, PG and Research Department of Zoology, St. Joseph's College (Autonomous), Devagiri, Calicut 673 008, Kerala, India

<sup>3</sup> Zoology Department, Faculty of Science, Cairo University, Giza 12613, Egypt

<sup>4</sup> Department of Food Science, College of Agriculture, University of Basrah, Basrah 61004, Iraq

<sup>5</sup> College of Medicine, University of Warith Al-Anbiyaa, Karbala 56001, Iraq

\* Correspondence: joicetomj@devagiricollege.org (J.T.J.); arunaksharann@devagiricollege.org (A.N.)

† These authors contributed equally to this work.



**Citation:** Albaqami, J.J.; Benny, T.P.; Hamdi, H.; Altemimi, A.B.; Kuttithodi, A.M.; Job, J.T.; Sasidharan, A.; Narayanankutty, A. Phytochemical Composition and In Vitro Antioxidant, Anti-Inflammatory, Anticancer, and Enzyme-Inhibitory Activities of *Artemisia nilagirica* (C.B. Clarke) Pamp. *Molecules* **2022**, *27*, 7119. <https://doi.org/10.3390/molecules27207119>

Academic Editor: Lucia Panzella

Received: 27 September 2022

Accepted: 18 October 2022

Published: 21 October 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

**Abstract:** Plants have been employed in therapeutic applications against various infectious and chronic diseases from ancient times. Various traditional medicines and folk systems have utilized numerous plants and plant products, which act as sources of drug candidates for modern medicine. *Artemisia* is a genus of the *Asteraceae* family with more than 500 species; however, many of these species are less explored for their biological efficacy, and several others are lacking scientific explanations for their uses. *Artemisia nilagirica* is a plant that is widely found in the Western Ghats, Kerala, India and is a prominent member of the genus. In the current study, the phytochemical composition and the antioxidant, enzyme-inhibitory, anti-inflammatory, and anticancer activities were examined. The results indicated that the ethanol extract of *A. nilagirica* indicated in vitro DPPH scavenging ( $23.12 \pm 1.28 \mu\text{g/mL}$ ), ABTS scavenging ( $27.44 \pm 1.88 \mu\text{g/mL}$ ),  $\text{H}_2\text{O}_2$  scavenging ( $12.92 \pm 1.05 \mu\text{g/mL}$ ), and FRAP ( $5.42 \pm 0.19 \mu\text{g/mL}$ ). The anti-inflammatory effect was also noticed in the Raw 264.7 macrophages, where pretreatment with the extract reduced the LPS-stimulated production of cytokines ( $p < 0.05$ ). *A. nilagirica* was also efficient in inhibiting the activities of  $\alpha$ -amylase ( $38.42 \pm 2.71 \mu\text{g/mL}$ ),  $\alpha$ -glucosidase ( $55.31 \pm 2.16 \mu\text{g/mL}$ ), aldose reductase ( $17.42 \pm 0.87 \mu\text{g/mL}$ ), and sorbitol dehydrogenase ( $29.57 \pm 1.46 \mu\text{g/mL}$ ). It also induced significant inhibition of proliferation in breast (MCF7  $\text{IC}_{50} = 41.79 \pm 1.07$ , MDAMB231  $\text{IC}_{50} = 55.37 \pm 2.11 \mu\text{g/mL}$ ) and colon ( $49.57 \pm 1.46 \mu\text{g/mL}$ ) cancer cells. The results of the phytochemical screening indicated a higher level of polyphenols and flavonoids in the extract and the LCMS analysis revealed the presence of various bioactive constituents including artemisinin.

**Keywords:** phytochemistry; *Artemisia nilagirica*; *Asteraceae*; antioxidant; anti-inflammatory activity; anticancer activity

## 1. Introduction

Medicinal plants are important sources of various biologically and pharmacologically active compounds [1,2]. Several traditional medicinal plants have been shown to have strong pharmacological properties, such as radical neutralizing, inflammation-preventing, antiproliferative, hypolipidemic, hepatoprotective, neuroprotective, antithrombotic, and immunomodulatory activities [3,4]. Among the various plant families, *Asteraceae* is one of the most widely utilized ones, and it is also equipped with numerous biological and pharmacological activities. Among the various genera, the *Artemisia* genus is well-known [5–7].

The *Artemisia* genus and the member species are well-studied for their various biological activities [8–13]. *Artemisia annua* L. has demonstrated significant medicinal benefits because of the presence of artemisinin [14]. *Artemisia mongolica* is another important member of the genus, which is rich in lactone derivatives of Sesquiterpene and a wide range of pharmacological activities [15]. The different species of the genus were found to have strong antibacterial and antifungal properties against pathogenic organisms in humans, livestock, and plants [16–20]. Antiproliferative and apoptotic effects are attributed to the bioactive compounds and extracts of various species of *Artemisia* [21–24].

*Artemisia nilagirica* is distributed throughout the Western Ghats, India; it has been traditionally applied by various tribal healers in the area for the treatment of infectious diseases and toxicity prevention. The plant has been shown to have significant biological and pharmacological activities based on various in vitro and in vivo studies. The initial studies by Ahameethunisa and Hopper [25] identified the antibacterial potential of the methanol extract of *A. nilagirica* against 15 bacterial strains. Further, the extract was found to be effective against *Mycobacterium smegmatis* and *M. bovis* [26]. The extract was also found to be effective against the malarial parasite *Plasmodium falciparum* [27].

The anticancer activities of the methanol and ethyl acetate extracts were also elucidated against the human monocytic leukemia cell (THP-1) [28]. Later, studies by Sahu, Meena, Shukla, Chaturvedi, Kumar, Datta, and Arya [24] also supported these results in colorectal cancer cell models. Studies by Raju et al. [29] indicated that the anticancer activity was mediated through the inhibition of TGF-beta signaling. The plant extract was also found to inhibit inflammatory insults in human red blood cell models [30]. The fruit of *A. nilagirica* was found to have significant antiradical activity via scavenging DPPH and nitric oxide radicals [31]. The essential oil extracted from *A. nilagirica* was a rich source of monoterpenoid compounds such as thujone, and by virtue of these compounds, the essential oil inhibited the growth of various fungal pathogens [32]. The essential oil was also effective against the phytopathogenic fungal groups of table grapes [33]. Additionally, the essential oil was also effective against various bacterial populations and capable of repelling mosquitoes [34].

Although several studies have reported the preliminary pharmacological activity of the plant, there is no clear-cut information on its quantitative chemical profile and nutritional value. Additionally, the anti-inflammatory properties are yet to be discovered in cell line models, and its mechanism of action is also not specified. Therefore, the present study aimed to analyze the chemical composition of the ethanol extract of *Artemisia nilagirica* leaves in terms of the bioactive compounds and proximate composition, as well as their antioxidant potential. Further, this study for the first time attempted to analyze the enzyme-inhibitory and anti-inflammatory activities of the extract in Raw 264.7 cells stimulated by lipopolysaccharides.

## 2. Results and Discussion

### 2.1. Determination of Proximate Composition of *A. nilagirica*

The *Artemisia* species, which includes 200–400 identified plants, are extensively spread in tropical and temperate areas [6]. The importance of the artemisia species in traditional medicine is well established [5]. The plant's antiviral, antifungal, antibiotic, insecticidal, hepatoprotective, and neuroprotective qualities make it useful in both Chinese and Ayurvedic medical systems [35]. The current study examined a specific member *Artemisia nilagirica*, its phytochemical makeup, and its pharmacological effects.

The physicochemical parameters of the *A. nilagirica* leaf powder are shown in Table 1. The predominant compounds were carbohydrate, protein, fat, and ash contents. The moisture content was estimated to be  $87.4 \pm 2.12\%$ .

### 2.2. Quantitative and Qualitative Estimation of Phytochemicals in *A. nilagirica*

The qualitative phytochemical screening identified the presence of compounds such as alkaloids, flavonoids, glycosides, sterols, and triterpenes (Table 2). The LCMS analysis

of the *A. nilagirica* ethanol extract indicated the presence of various phytochemicals, including artemisinin, quercetin, apigenin, B-caryophyllene, luteolin, and simple phenolic acids (Figure 1 and Table 3). Previous reports have also confirmed that numerous kinds of bioactive substances are found in *A. vulgaris*, *A. annua*, and other species, including flavonoids, sesquiterpenoids, essential oils, tannins, phenols, and saponins [15,36]. The total polyphenol content of *A. nilagirica* was estimated to be  $89.51 \pm 2.5$  mg gallic acid equivalent/g of extract. The total flavonoid content was  $14.35 \pm 0.9$  mg quercetin equivalent/g of extract (Table 4). Further, the HPLC quantification indicated higher levels of quercetin ( $240.39 \pm 4.87$   $\mu\text{g/g}$  extract), luteolin ( $146.87 \pm 5.29$   $\mu\text{g/g}$  extract), and apigenin ( $103.41 \pm 3.35$   $\mu\text{g/g}$  extract) in the *A. nilagirica* extract (Table 5). These compounds are known to possess strong anti-inflammatory, antiproliferative and antidiabetic activities [37–40].

**Table 1.** Physicochemical parameters of *A. nilagirica* leaf powder.

Physicochemical Parameters	Result
Moisture content (%)	$87.4 \pm 2.12$
Carbohydrate (%)	$55.80 \pm 4.1$
Protein (%)	$3.90 \pm 0.16$
Crude fat (%)	$2.12 \pm 0.18$
Ash content (%)	$0.74 \pm 0.04$

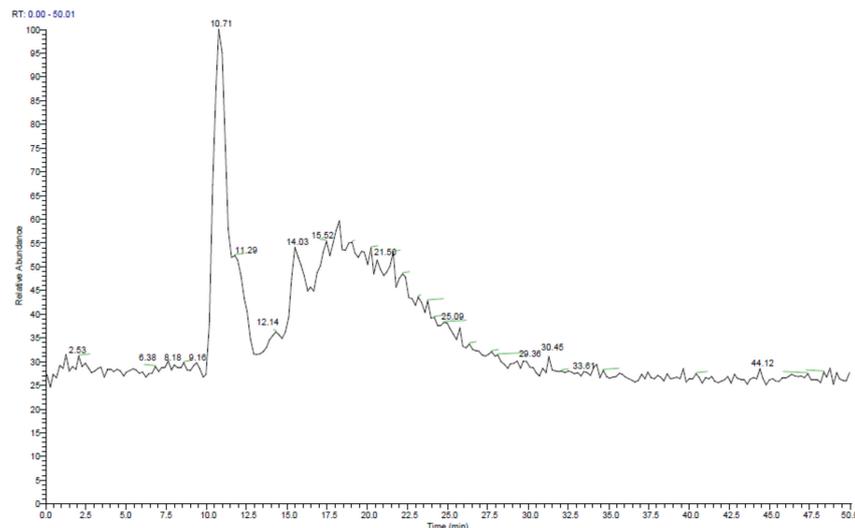
**Table 2.** Phytochemical constituents in the ethanol extract of *A. nilagirica*.

Test	Reaction
<b>Alkaloids</b>	
Marqui's test	++
Wagner's test	++
Mayer's test	+++
Hager's test	+
Froehde's test	++
Dragendorff test	++
<b>Glycosides</b>	
Legal's test	+
Keller-Kiliani test	+
<b>Flavonoids</b>	
Alkaline reagent test	++
Lead acetate test	++
Shinoda's test	+++
<b>Tannins</b>	
Ferric Chloride test	++
Gelatin test	++
<b>Phytosterols</b>	
Salkowski's test	++
Liebermann-Burchard test	+++
<b>Saponins</b>	
Froth test	+
Foam test	+
<b>Carbohydrates</b>	
Fehling test	++
Molish test	++
Benedict's test	++
<b>Phenols</b>	
Folin-Ciocalteu test	+++
<b>Resin</b>	
Acetone-water test	+

Table 2. Cont.

Test	Reaction
<b>Alkaloids</b>	
<b>Fixed oils and fats</b>	
Stain test	-
<b>Triterpenes</b>	
Liebermann-Burchardt's test	+++

Note: +++ high level, ++ moderate level, and + low-level presence of the compound.

Figure 1. The LC-MS total ion chromatogram of the *A. nilagirica* extract.Table 3. LCMS profiling of *A. nilagirica* with the retention time (RT), molecular mass, and chemical formula.

Sl. No.	RT (mins)	Compound Name	Formula	Mass
1	2.53	Ferulic acid	C <sub>10</sub> H <sub>10</sub> O <sub>4</sub>	194.00
2	6.38	Eugenol	C <sub>10</sub> H <sub>12</sub> O <sub>2</sub>	164.08
3	8.18	B-caryophyllene	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>	448.40
4	9.06	Luteolin	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	286.00
5	10.71	caffeic acid	C <sub>9</sub> H <sub>8</sub> O <sub>4</sub>	180.16
6	11.29	Quercetin	C <sub>15</sub> H <sub>10</sub> O <sub>7</sub>	302.00
7	12.14	Myricetin	C <sub>15</sub> H <sub>10</sub> O <sub>8</sub>	318.00
8	12.89	Apigenin	C <sub>15</sub> H <sub>10</sub> O <sub>5</sub>	270.05
9	14.03	Luteolin 5-0-beta-d-glucopyranoside	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>	448.13
10	15.52	Kaempferol	C <sub>15</sub> H <sub>10</sub> O <sub>6</sub>	286.23
11	21.56	Carnosic acid	C <sub>20</sub> H <sub>28</sub> O <sub>4</sub>	332.19
12	25.09	Artemisinin	C <sub>20</sub> H <sub>20</sub> O <sub>8</sub>	388.11
13	29.36	2alpha, 3 beta-Dihydroxyolean-12en-28-oic acid	C <sub>30</sub> H <sub>48</sub> O <sub>4</sub>	472.35
14	30.45	Menthyl acetate	C <sub>12</sub> H <sub>22</sub> O <sub>2</sub>	198.16
15	33.61	Oleanolic acid	C <sub>30</sub> H <sub>48</sub> O <sub>3</sub>	456.36
16	44.12	Basilimoside	C <sub>36</sub> H <sub>60</sub> O <sub>6</sub>	588.47

Table 4. The total polyphenol and flavonoid contents of *A. nilagirica* ethanol extract.

Assay	mg Equivalent/g
Total phenolic content	89.51 ± 2.5
Total flavonoid content	14.35 ± 0.9

**Table 5.** The quantification of selected compounds in the extract via HPLC.

RT (mins)	Compound Name	Quantity ( $\mu\text{g/g}$ Extract)
2.50	Ferulic acid	$18.51 \pm 1.82$
9.05	Luteolin	$146.87 \pm 5.29$
10.70	caffeic acid	$88.62 \pm 1.30$
11.30	Quercetin	$240.39 \pm 4.87$
12.87	Apigenin	$103.41 \pm 3.35$

### 2.3. In Vitro Antioxidant Activities of *A. nilagirica* Extract

The *Artemisia* genus members frequently display antioxidant activity [41]; our study also confirmed the antioxidant activity of *A. nilagirica* for the first time in terms of the radical generation inhibition and reducing potentials. The  $\text{IC}_{50}$  value of the *A. nilagirica* extract in the anti-DPPH radical assay was estimated to be  $23.12 \pm 1.28 \mu\text{g/mL}$ . Likewise, Table 6 shows the other antioxidant activities in terms of the ABTS radical scavenging activity, hydrogen peroxide scavenging potential, and ferric-reducing antioxidant power; the respective  $\text{IC}_{50}$  values were found to be  $27.44 \pm 1.88$ ,  $12.92 \pm 1.05$ , and  $5.42 \pm 0.19 \mu\text{g/mL}$ . On the contrary, the level of inhibition of nitric oxide radical generation ( $\text{IC}_{50}$ ) was determined to be  $367.09 \pm 12.05 \mu\text{g/mL}$  for the extract. However, in comparison with the standard antioxidant ascorbic acid (Table 6), the activity was much lower in the *A. nilagirica* extract; further purification of the extract may yield more active antioxidant compounds. The antioxidant properties are attributed to the bioactive compounds identified in the plant via LC-MS. Oxidative stress is the central independent factor that drives many chronic diseases, including cancers [42,43]; hence, the antioxidant properties of the plant may be useful in the management of diseases associated with oxidative stress.

**Table 6.** In vitro antioxidant activities of *A. nilagirica* extract (AN) expressed as  $\text{IC}_{50}$  values ( $\mu\text{g/mL}$ ).

Antioxidant Activity	$\text{IC}_{50}$ Value ( $\mu\text{g/mL}$ )	
	AN	Ascorbic Acid
DPPH scavenging	$23.12 \pm 1.28$	$9.64 \pm 0.89$
ABTS scavenging	$27.44 \pm 1.88$	$35.19 \pm 1.47$
$\text{H}_2\text{O}_2$ scavenging	$12.92 \pm 1.05$	$19.08 \pm 1.65$
FRAP value ( $\text{EC}_{50}$ )	$5.42 \pm 0.19$	$3.22 \pm 0.15$
Nitric oxide scavenging	$367.09 \pm 12.05$	$68.10 \pm 2.11$

### 2.4. Enzyme-Inhibitory Activities of *A. nilagirica* Ethanol Extract

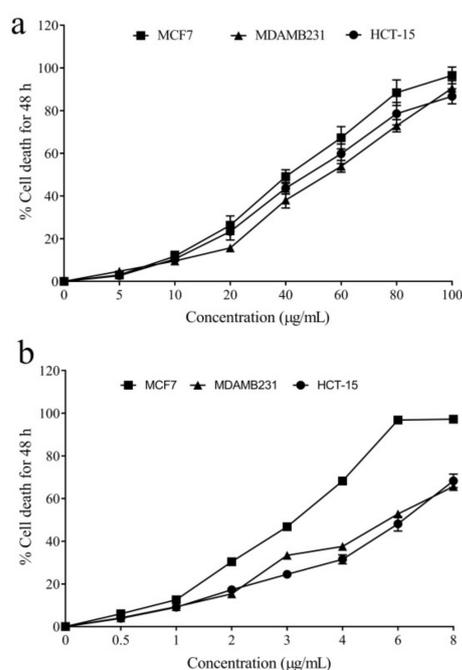
The enzyme-inhibitory properties of the extract were analyzed against four enzymes involved in type 2 diabetes mellitus, including  $\alpha$ -amylase,  $\alpha$ -glucosidase, aldose reductase, and sorbitol dehydrogenase (Table 7). The  $\text{IC}_{50}$  values for these enzymes were  $38.42 \pm 2.71$ ,  $55.31 \pm 2.16$ ,  $17.42 \pm 0.87$ , and  $29.57 \pm 1.46 \mu\text{g/mL}$ , respectively. Furthermore,  $\alpha$ -amylase and  $\alpha$ -glucosidase are enzymes involved in carbohydrate metabolism and are common targets of antidiabetic drugs [44]. Similarly, the polyol pathway enzymes, including aldose reductase and sorbitol dehydrogenase, are involved in diabetic complications [45,46]. Hence, the inhibition of these enzymes could result in strong antidiabetic activity for the *A. nilagirica* extract.

**Table 7.** In vitro enzyme-inhibitory properties of *A. nilagirica* expressed as  $\text{IC}_{50}$  values ( $\mu\text{g/mL}$ ).

Enzyme	$\text{IC}_{50}$ Value ( $\mu\text{g/mL}$ )
$\alpha$ -Amylase	$38.42 \pm 2.71$
$\alpha$ -Glucosidase	$55.31 \pm 2.16$
Aldose reductase	$17.42 \pm 0.87$
Sorbitol dehydrogenase	$29.57 \pm 1.46$

### 2.5. Antiproliferative Activity of the *A. nilagirica*

Additionally, the results showed the anticancer properties of *A. nilagirica* in human breast and colon cancer cells. The anticancer activity was analyzed in three cancer cell lines, including MCF-7, MDA-MB-231, and HCT-15. We observed dose-dependent cytotoxicity in these three cell lines (Figure 2). The IC<sub>50</sub> values against the three cells were estimated to be  $41.79 \pm 1.07$ ,  $55.37 \pm 2.11$ , and  $49.57 \pm 1.46$   $\mu\text{g}/\text{mL}$ , respectively. In comparison, the standard cyclophosphamide was more toxic to these cells, with respective IC<sub>50</sub> values of  $3.12 \pm 0.13$ ,  $5.74 \pm 0.20$ , and  $6.04 \pm 0.21$   $\mu\text{g}/\text{mL}$ . Previous studies have also shown different species of *Artemisia* in various cancer cells [47–50]. In addition, the green synthesized nanoparticles from different *Artemisia* species are also reported to exert antiproliferative effects on cancer cells mediated through apoptotic cell death [23,51,52]. A study by Sahu, Meena, Shukla, Chaturvedi, Kumar, Datta, and Arya [24] indicated that ethyl acetate and hexane fractions of *A. nilagirica* induced cell death in colon, lung, and breast cancer cells. In addition, the bioactive compounds, including quercetin, apigenin, and eugenol, have also been shown to have significant antiproliferative effects by modulating different signaling pathways [53,54].



**Figure 2.** The anticancer potentials of the leaf extract of *A. nilagirica* (a) and cyclophosphamide (b).

### 2.6. Anti-Inflammatory Activity of *A. nilagirica*

The *Artemisia nilagirica* extract was shown to inhibit the production of nitric oxide radicals in vitro. Further, the pretreatment of the extract also inhibited cytokine production and inflammatory insults in lipopolysaccharide-stimulated macrophages. The LPS is a microbial component that is known to stimulate inflammatory insults [55,56]. The *Artemisia nilagirica* leaf ethanol extract (AN) was found to inhibit the lipopolysaccharide-induced activation of macrophages and the subsequent cytokine release. The level of IL-1 $\beta$  was found to be significantly increased after LPS stimulation in macrophages; however, the pretreatment with AN at different doses significantly brought down the IL-1 $\beta$  levels in the macrophages (Table 8). Likewise, the levels of IL-6 and TNF- $\alpha$  also showed a similar increase during LPS exposure, which were successfully brought down by the treatment with different concentrations of *A. nilagirica*. The level of nitric oxide was determined biochemically and was also significantly elevated in LPS control cells. Pretreatment with 2.5, 5.0, and 7.5  $\mu\text{g}/\text{mL}$  of AN successfully brought down the levels to  $40.7 \pm 1.6$  ( $p < 0.05$ ),  $32.2 \pm 2.4$  ( $p < 0.05$ ), and  $25.7 \pm 2.1$  ( $p < 0.01$ ). In addition, it is noted that the high dose

of the extract resulted in stronger anti-inflammatory molecules compared to quercetin, which is a well-known anti-inflammatory molecule [57,58]. The LPS is known to stimulate cytokine production in macrophages by upregulating the NF- $\kappa$ B translocation to the nuclear compartment [59,60]. It is, therefore, possible that the *A. nilagirica* extract may also influence the LPS-induced activation of intracellular NF-KB signaling.

**Table 8.** Effect of the *Artemisia nilagirica* leaf ethanol extract (AN) against lipopolysaccharide-induced macrophage (Raw 264.7) activation, cytokine release (in pg/mg protein), and nitric oxide production ( $\mu$ M/mg protein).

Nature	Tumor Necrosis Factor $\alpha$	Interleukin 6	Interleukin 1 $\beta$	NO	
Untreated	97.6 $\pm$ 2.8	76.4 $\pm$ 3.1	67.8 $\pm$ 2.8	7.4 $\pm$ 0.57	
Negative Control (LPS alone)	420.8 $\pm$ 10.6	795.2 $\pm$ 11.7	628.9 $\pm$ 14.2	52.1 $\pm$ 2.0	
Quercetin (4.5 $\mu$ g/mL)	279.1 $\pm$ 11.3 **	414.2 $\pm$ 10.7 ***	334.8 $\pm$ 11.7 **	30.7 $\pm$ 1.2 *	
<i>Artemisia nilagirica</i> extract	2.5 $\mu$ g/mL	314.1 $\pm$ 14.5 *	698.0 $\pm$ 17.3 **	477.6 $\pm$ 11.8 **	40.7 $\pm$ 1.6 *
	5.0 $\mu$ g/mL	265.7 $\pm$ 10.7 **	524.3 $\pm$ 15.6 **	389.5 $\pm$ 14.6 **	32.2 $\pm$ 2.4 *
	7.5 $\mu$ g/mL	190.9 $\pm$ 14.8 ***	388.2 $\pm$ 15.8 ***	298.7 $\pm$ 15.2 ***	25.7 $\pm$ 2.1 **

*Artemisia nilagirica* leaf ethanol extract (AN), lipopolysaccharide (LPS), nitric oxide (NO). The significance is indicated as \* ( $p < 0.05$ ), \*\* ( $p < 0.01$ ), \*\*\* ( $p < 0.001$ ).

Thus, the study concludes that the *Artemisia nilagirica* ethanol extract exhibits antioxidant and anti-inflammatory properties in vitro and cultured cells. Further, the extract is also capable of inhibiting the proliferation of various cancer cells. The inhibition of enzymes associated with type 2 diabetes mellitus is also indicative of its anti-diabetic properties. The biological properties of the plant are expected to be due to the bioactive compounds identified in the *A. nilagirica* extract.

### 3. Materials and Methods

#### 3.1. *Artemisia Nilagirica* (C.B.Clarke) Pamp. Collection and Extraction Using 100% Ethanol

The *Artemisia nilagirica* plant samples were collected from the Wayanad District, Kerala (11.7917° N, 76.1716° E). The mature leaves were carefully cleaned of all kinds of dust via washing. These leaves were dried under shade for 2 weeks and powdered using a mixer grinder; the powder was extracted with 100% ethanol using the Soxhlet method. Briefly, 100 g of the powder was extracted with ethanol at 80 °C for 8 h and the extract was collected, filtered, and concentrated before storage.

#### 3.2. Phytochemical Analysis of *Artemisia nilagirica*

The leaf powder of *A. nilagirica* was analyzed for the proximate composition according to the methods used by Shukla et al. [61]. The qualitative phytochemical screening was carried out for the detection of alkaloids, flavonoids, glycosides, sterols, and triterpenes by referring to standard protocols [62,63]. The LC-MS analysis (Shimadzu LC- 8045, Kyoto, Japan) was used for phytochemical screening [64]; briefly, the C18 column measuring 4.6  $\times$  150 mm and 5  $\mu$ m in size was used for the study, with methanol (A) and water with 0.1% formic acid (B) as the mobile phase (gradient elution mode). The gradient was set as 95% solution A (0–5 min), 70% solution A (5 to 10 min), 65% solution A (10 to 20 min), 50% solution A (20 to 30 min), and 90% of solution B (until 50 min), with a flow rate of 1.0 mL/min.

The quantitative profiling was estimated in terms of the total polyphenols [65] and total flavonoids [66], and the concentrations of ferulic acid, luteolin, caffeic acid, quercetin, and apigenin were determined using an HPLC analysis according to the same LC-MS conditions mentioned above.

### 3.3. Analysis of the Antioxidant Activity of *A. nilagirica* Ethanol Extract

The antioxidant activities were determined as the scavenging potentials of different radicals, including diphenyl picryl hydrazyl (DPPH), ABTS [67], and hydrogen peroxide [68]; the reducing potential on ferric ions was also estimated using the procedures described in [69]. The nitric oxide radical removal rate was used as an indicator of the inflammatory process inhibition model [70]. The DPPH was dissolved in methanol (0.1 mM) and varying concentrations of the extract were mixed with it. The solution was incubated for 20 min in the dark at 30 °C and the change in absorbance was used to estimate the percentage inhibition. Likewise, the ABTS radical generated was mixed with different doses of the *A. nilagirica* extract and the % inhibition was calculated spectrophotometrically. The nitric oxide scavenging was determined using sodium nitroprusside (8 mM) as the radical source; the Griess reagent was used to estimate the nitrite remaining in the treated samples using spectrophotometry at 596 nm.

Ascorbic acid was used as a positive control and standard for the antioxidant assays. The percentage inhibition was determined using the formula

$$\text{Percentage inhibition} = \frac{\text{Absorbance of Control} - \text{Absorbance of Sample}}{\text{Absorbance of Control}} \times 100$$

### 3.4. Efficacy of *A. nilagirica* Ethanol Extract on Activities of Enzymes

The enzyme-inhibitory properties were analyzed against the selected enzymes involved in diabetes and secondary diabetic complications. The inhibitory effect on  $\alpha$ -amylase [71],  $\alpha$ -glucosidase [72], aldose reductase [73], and sorbitol dehydrogenase [46] was assessed according to the standard methods.

### 3.5. Effect of *A. nilagirica* Ethanol Extract on Cancer Cell Proliferation

The human breast cancer cell lines MCF7 and MDA-MB-231 and a colon cancer cell line (HCT-15) were collected from NCCS, Pune, India. These cells were maintained in complete MEM, Leibovitz's L-15, and RPMI-1640 media. The cells were selected as they are widely used in the anticancer screening of phytochemicals.

The inhibitory potential of the extract on human cancer cell proliferation (MCF7, MDA-MB-231, and HCT-15) was assessed using the MTT assay [74]. The IC<sub>50</sub> value was determined using probit analysis.

### 3.6. Effect of *A. nilagirica* Extract on Lipopolysaccharide-Induced Cytokine Production in Macrophages

The murine Raw 264.7 cells were allowed to attach ( $1 \times 10^7$  cells/mL) in a 24-well plate in complete growth media. The RPMI-1640 media was used to dilute the different concentrations of *A. nilagirica* (AN) (2.5, 5.0, and 7.5  $\mu\text{g/mL}$ ). Next, the cells were exposed to 1  $\mu\text{g/mL}$  lipopolysaccharide for another 24 h. The protein expression of cytokines such as interleukin-1 $\beta$  and interleukin-6 and the tumor necrosis factor- $\alpha$  release were determined using PeproTech ELISA kits (Rocky Hill, CT, USA), as per the commercially prescribed methods. The nitric oxide release was quantified using the Griess reaction method [64]. Quercetin was used as a standard anti-inflammatory compound in the study.

### 3.7. Presentation of the Data, Software Used, and Statistical Analysis

The accuracy of the results obtained was ensured by conducting three independent assignments, with each having four replicates. Microsoft Excel 2010 was used for data consolidation and verification. The processed data are presented as means  $\pm$  standard deviations; the IC<sub>50</sub> values were estimated using probit analysis (GraphPad Prism 7.0, San Diego, CA, USA).

## 4. Conclusions

*Artemisia nilagirica* is an ethnomedicinal plant in India. In our study, the ethanol extract of *A. nilagirica* leaves showed significant antiradical and reducing potentials, which are

indicative of its antioxidant potential. The IC<sub>50</sub> values were lower but comparable with those of the standard ascorbic acid. The extract also inhibited enzymes associated with diabetes mellitus, including alpha-amylase and  $\alpha$ -glucosidase. Additionally, the extract treatment significantly reduced the proliferative potential of breast and colon cancer cells. In Raw 264.7 macrophages, the pretreatment with the extract inhibited the LPS-stimulated production of cytokines and proved itself to be anti-inflammatory. Most importantly, the higher dose of the extract caused significantly higher activity than the standard quercetin used. Hence, we conclude that the ethanol extract of *A. nilagirica* leaves has antioxidant, anti-inflammatory, and anticancer properties; further studies on animal models and with bioassay-guided purification are necessary to identify the bioactive components.

**Author Contributions:** A.N.: Study design, methodology, analysis, manuscript editing. J.J.A. and H.H.: Study design, methodology, experimentation, analysis, funding acquisition, manuscript preparation, manuscript editing. A.M.K., A.S., T.P.B. and J.T.J.: Experimentation, analysis; manuscript draft preparation. A.B.A.: Analysis, manuscript preparation, manuscript editing. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study received no funding support.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Data is contained within the article.

**Acknowledgments:** ARN and JTJ acknowledge the DBT star scheme for the infrastructural developments in the college. ARN and JTJ is thankful to St. Joseph's College (Autonomous), Devagiri for the research promotion (seed grant) scheme RPSG-2022-23.

**Conflicts of Interest:** The authors declare no conflict of interest.

**Sample Availability:** Samples of the compounds are available from the corresponding author.

## References

1. Behera, N.K.; Mahalakshmi, G.S. A cloud based knowledge discovery framework, for medicinal plants from PubMed literature. *Inform. Med. Unlocked* **2019**, *16*, 100105. [[CrossRef](#)]
2. Karpavičienė, B. Traditional Uses of Medicinal Plants in South-Western Part of Lithuania. *Plants* **2022**, *11*, 2093. [[CrossRef](#)] [[PubMed](#)]
3. Noor, F.; Tahir ul Qamar, M.; Ashfaq, U.A.; Albutti, A.; Alwashmi, A.S.S.; Aljasir, M.A. Network Pharmacology Approach for Medicinal Plants: Review and Assessment. *Pharmaceuticals* **2022**, *15*, 572. [[CrossRef](#)] [[PubMed](#)]
4. Adeleke, B.S.; Babalola, O.O. Pharmacological Potential of Fungal Endophytes Associated with Medicinal Plants: A Review. *J. Fungi* **2021**, *7*, 147. [[CrossRef](#)] [[PubMed](#)]
5. Septembre-Malaterre, A.; Lalarizo Rakoto, M.; Marodon, C.; Bedoui, Y.; Nakab, J.; Simon, E.; Hoarau, L.; Savriama, S.; Strasberg, D.; Guiraud, P.; et al. *Artemisia annua*, a Traditional Plant Brought to Light. *Int. J. Mol. Sci.* **2020**, *21*, 4986. [[CrossRef](#)] [[PubMed](#)]
6. Bora, K.S.; Sharma, A. The genus *Artemisia*: A comprehensive review. *Pharm Biol.* **2011**, *49*, 101–109. [[CrossRef](#)] [[PubMed](#)]
7. Bisht, D.; Kumar, D.; Kumar, D.; Dua, K.; Chellappan, D.K. Phytochemistry and pharmacological activity of the genus *artemisia*. *Arch. Pharmacol Res.* **2021**, *44*, 439–474. [[CrossRef](#)]
8. Salaroli, R.; Andreani, G.; Bernardini, C.; Zannoni, A.; La Mantia, D.; Protti, M.; Forni, M.; Mercolini, L.; Isani, G. Anticancer activity of an *Artemisia annua* L. hydroalcoholic extract on canine osteosarcoma cell lines. *Res. Vet. Sci.* **2022**, *152*, 476–484. [[CrossRef](#)]
9. Baies, M.H.; Gherman, C.; Boros, Z.; Olah, D.; Vlase, A.M.; Cozma-Petrut, A.; Gyorke, A.; Miere, D.; Vlase, L.; Crisan, G.; et al. The Effects of *Allium sativum* L., *Artemisia absinthium* L., *Cucurbita pepo* L., *Coriandrum sativum* L., *Satureja hortensis* L. and *Calendula officinalis* L. on the Embryogenesis of *Ascaris suum* Eggs during an In Vitro Experimental Study. *Pathogens* **2022**, *11*, 1065. [[CrossRef](#)]
10. Suroowan, S.; Llorent-Martinez, E.J.; Zengin, G.; Dall'Acqua, S.; Sut, S.; Buskaran, K.; Fakurazi, S.; Mahomoodally, M.F. Phytochemical Characterization, Anti-Oxidant, Anti-Enzymatic and Cytotoxic Effects of *Artemisia verlotiorum* Lamotte Extracts: A New Source of Bioactive Agents. *Molecules* **2022**, *27*, 5886. [[CrossRef](#)]
11. Chen, X.Y.; Liu, T.; Hu, Y.Z.; Qiao, T.T.; Wu, X.J.; Sun, P.H.; Qian, C.W.; Ren, Z.; Zheng, J.X.; Wang, Y.F. Sesquiterpene lactones from *Artemisia vulgaris* L. as potential NO inhibitors in LPS-induced RAW264.7 macrophage cells. *Front. Chem.* **2022**, *10*, 948714. [[CrossRef](#)]

12. Chen, J.; Chen, F.; Peng, S.; Ou, Y.; He, B.; Li, Y.; Lin, Q. Effects of *Artemisia argyi* Powder on Egg Quality, Antioxidant Capacity, and Intestinal Development of Roman Laying Hens. *Front. Physiol.* **2022**, *13*, 902568. [[CrossRef](#)]
13. Su, S.H.; Sundhar, N.; Kuo, W.W.; Lai, S.C.; Kuo, C.H.; Ho, T.J.; Lin, P.Y.; Lin, S.Z.; Shih, C.Y.; Lin, Y.J.; et al. *Artemisia argyi* extract induces apoptosis in human gemcitabine-resistant lung cancer cells via the PI3K/MAPK signaling pathway. *J. Ethnopharmacol.* **2022**, *299*, 115658. [[CrossRef](#)]
14. Agrawal, P.K.; Agrawal, C.; Blunden, G. *Artemisia* Extracts and Artemisinin-Based Antimalarials for COVID-19 Management: Could These Be Effective Antivirals for COVID-19 Treatment? *Molecules* **2022**, *27*, 3828. [[CrossRef](#)]
15. Zhu, Z.; Turak, A.; Aisa, H.A. Sesquiterpene lactones from *Artemisia mongolica*. *Phytochemistry* **2022**, *199*, 113158. [[CrossRef](#)]
16. Zhang, J.J.; Qu, L.B.; Bi, Y.F.; Pan, C.X.; Yang, R.; Zeng, H.J. Antibacterial activity and mechanism of chloroform fraction from aqueous extract of mugwort leaves (*Artemisia argyi* L.) against *Staphylococcus aureus*. *Lett. Appl. Microbiol.* **2022**, *74*, 893–900. [[CrossRef](#)]
17. Wang, Y.; Li, J.; Chen, Q.; Zhou, J.; Xu, J.; Zhao, T.; Huang, B.; Miao, Y.; Liu, D. The role of antifungal activity of ethyl acetate extract from *Artemisia argyi* on *Verticillium dahliae*. *J. Appl. Microbiol.* **2022**, *132*, 1343–1356. [[CrossRef](#)]
18. Tao, A.; Feng, X.; Sheng, Y.; Song, Z. Optimization of the *Artemisia* Polysaccharide Fermentation Process by *Aspergillus niger*. *Front. Nutr.* **2022**, *9*, 842766. [[CrossRef](#)]
19. Suvaithenamudhan, S.; Ananth, S.; Mariappan, V.; Dhayabaran, V.V.; Parthasarathy, S.; Ganesh, P.S.; Shankar, E.M. In Silico Evaluation of Bioactive Compounds of *Artemisia pallens* Targeting the Efflux Protein of Multidrug-Resistant *Acinetobacter baumannii* (LAC-4 Strain). *Molecules* **2022**, *27*, 5188. [[CrossRef](#)]
20. Son, S.R.; Ju, I.G.; Kim, J.; Park, K.T.; Oh, M.S.; Jang, D.S. Chemical Constituents from the Aerial Parts of *Artemisia iwayomogi* and Their Anti-Neuroinflammatory Activities. *Plants* **2022**, *11*, 1954. [[CrossRef](#)]
21. Kolesar, J.M.; Seeberger, P.H. Editorial: Anticancer Potential of *Artemisia annua*. *Front. Oncol.* **2022**, *12*, 853406. [[CrossRef](#)]
22. Jung, E.J.; Paramanatham, A.; Kim, H.J.; Shin, S.C.; Kim, G.S.; Jung, J.M.; Hong, S.C.; Chung, K.H.; Kim, C.W.; Lee, W.S. Identification of Growth Factors, Cytokines and Mediators Regulated by *Artemisia annua* L. Polyphenols (pKAL) in HCT116 Colorectal Cancer Cells: TGF-beta1 and NGF-beta Attenuate pKAL-Induced Anticancer Effects via NF-kappaB p65 Upregulation. *Int. J. Mol. Sci.* **2022**, *23*, 1598. [[CrossRef](#)]
23. Bordoni, V.; Sanna, L.; Lyu, W.; Avitabile, E.; Zoroddu, S.; Medici, S.; Kelvin, D.J.; Bagella, L. Silver Nanoparticles Derived by *Artemisia arborescens* Reveal Anticancer and Apoptosis-Inducing Effects. *Int. J. Mol. Sci.* **2021**, *22*, 8621. [[CrossRef](#)]
24. Sahu, N.; Meena, S.; Shukla, V.; Chaturvedi, P.; Kumar, B.; Datta, D.; Arya, K.R. Extraction, fractionation and re-fractionation of *Artemisia nilagirica* for anticancer activity and HPLC-ESI-QTOF-MS/MS determination. *J. Ethnopharmacol.* **2018**, *213*, 72–80. [[CrossRef](#)]
25. Ahameethunisa, A.R.; Hopper, W. Antibacterial activity of *Artemisia nilagirica* leaf extracts against clinical and phytopathogenic bacteria. *BMC Complement. Altern. Med.* **2010**, *10*, 6. [[CrossRef](#)]
26. Naik, S.K.; Mohanty, S.; Padhi, A.; Pati, R.; Sonawane, A. Evaluation of antibacterial and cytotoxic activity of *Artemisia nilagirica* and *Murraya koenigii* leaf extracts against mycobacteria and macrophages. *BMC Complement. Altern. Med.* **2014**, *14*, 87. [[CrossRef](#)]
27. Panda, S.; Rout, J.R.; Pati, P.; Ranjit, M.; Sahoo, S.L. Antimalarial activity of *Artemisia nilagirica* against *Plasmodium falciparum*. *J. Parasit. Dis.* **2018**, *42*, 22–27. [[CrossRef](#)]
28. Gul, M.Z.; Chandrasekaran, S.; Manjulatha, K.; Bhat, M.Y.; Maurya, R.; Qureshi, I.A.; Ghazi, I.A. Bioassay-Guided Fractionation and In Vitro Antiproliferative Effects of Fractions of *Artemisia nilagirica* on THP-1 cell line. *Nutr. Cancer* **2016**, *68*, 1210–1224. [[CrossRef](#)]
29. Raju, S.R.; Balakrishnan, S.; Kollimada, S.; Chandrashekar, K.N.; Jampani, A. Anti-tumor effects of *Artemisia nilagirica* extract on MDA-MB-231 breast cancer cells: Deciphering the biochemical and biomechanical properties via TGF-beta upregulation. *Heliyon* **2020**, *6*, e05088. [[CrossRef](#)]
30. Parameswari, P.; Devika, R.; Vijayaraghavan, P. In vitro anti-inflammatory and antimicrobial potential of leaf extract from *Artemisia nilagirica* (Clarke) Pamp. *Saudi J. Biol. Sci.* **2019**, *26*, 460–463. [[CrossRef](#)]
31. Suseela, V.; Gopalakrishnan, V.K.; Varghese, S. In vitro Antioxidant Studies of Fruits of *Artemisia nilagirica* (Clarke) Pamp. *Indian J. Pharm. Sci.* **2010**, *72*, 644–649. [[CrossRef](#)] [[PubMed](#)]
32. Sati, S.C.; Sati, N.; Ahluwalia, V.; Walia, S.; Sati, O.P. Chemical composition and antifungal activity of *Artemisia nilagirica* essential oil growing in northern hilly areas of India. *Nat. Prod. Res.* **2013**, *27*, 45–48. [[CrossRef](#)] [[PubMed](#)]
33. Sonker, N.; Pandey, A.K.; Singh, P. Efficiency of *Artemisia nilagirica* (Clarke) Pamp. essential oil as a mycotoxicant against postharvest mycobiota of table grapes. *J. Sci. Food Agric.* **2015**, *95*, 1932–1939. [[CrossRef](#)] [[PubMed](#)]
34. Stappen, I.; Wanner, J.; Tabanca, N.; Wedge, D.E.; Ali, A.; Khan, I.A.; Kaul, V.K.; Lal, B.; Jaitak, V.; Gochev, V.; et al. Chemical composition and biological effects of *Artemisia maritima* and *Artemisia nilagirica* essential oils from wild plants of western Himalaya. *Planta Med.* **2014**, *80*, 1079–1087. [[CrossRef](#)]
35. Ekiert, H.; Świątkowska, J.; Klin, P.; Rzepiela, A.; Szopa, A. *Artemisia annua*—Importance in Traditional Medicine and Current State of Knowledge on the Chemistry, Biological Activity and Possible Applications. *Planta Med.* **2021**, *87*, 584–599. [[CrossRef](#)]
36. Matvieieva, N.; Drobot, K.; Duplij, V.; Ratushniak, Y.; Shakhovskiy, A.; Kyrpa-Nesmiian, T.; Mickevicius, S.; Brindza, J. Flavonoid content and antioxidant activity of *Artemisia vulgaris* L. “hairy” roots. *Prep. Biochem. Biotechnol.* **2019**, *49*, 82–87. [[CrossRef](#)]
37. Tavsan, Z.; Kayali, H.A. Flavonoids showed anticancer effects on the ovarian cancer cells: Involvement of reactive oxygen species, apoptosis, cell cycle and invasion. *Biomed. Pharmacother.* **2019**, *116*, 109004. [[CrossRef](#)]

38. Al-Ishaq, R.K.; Abotaleb, M.; Kubatka, P.; Kajo, K.; Büsselberg, D. Flavonoids and Their Anti-Diabetic Effects: Cellular Mechanisms and Effects to Improve Blood Sugar Levels. *Biomolecules* **2019**, *9*, 430. [[CrossRef](#)]
39. Tian, C.; Liu, X.; Chang, Y.; Wang, R.; Lv, T.; Cui, C.; Liu, M. Investigation of the anti-inflammatory and antioxidant activities of luteolin, kaempferol, apigenin and quercetin. *S. Afr. J. Bot.* **2021**, *137*, 257–264. [[CrossRef](#)]
40. Crasci, L.; Cardile, V.; Longhitano, G.; Nanfitò, F.; Panico, A. Anti-degenerative effect of Apigenin, Luteolin and Quercetin on human keratinocyte and chondrocyte cultures: SAR evaluation. *Drug Res.* **2018**, *68*, 132–138. [[CrossRef](#)]
41. Skowyra, M.; Gallego, M.G.; Segovia, F.; Almajano, M.P. Antioxidant Properties of *Artemisia annua* Extracts in Model Food Emulsions. *Antioxidants* **2014**, *3*, 116–128. [[CrossRef](#)]
42. Perillo, B.; Di Donato, M.; Pezone, A.; Di Zazzo, E.; Giovannelli, P.; Galasso, G.; Castoria, G.; Migliaccio, A. ROS in cancer therapy: The bright side of the moon. *Exp. Mol. Med.* **2020**, *52*, 192–203. [[CrossRef](#)]
43. Hayes, J.D.; Dinkova-Kostova, A.T.; Tew, K.D. Oxidative Stress in Cancer. *Cancer Cell* **2020**, *38*, 167–197. [[CrossRef](#)]
44. Alqahtani, A.S.; Hidayathulla, S.; Rehman, M.T.; ElGamal, A.A.; Al-Massarani, S.; Razmovski-Naumovski, V.; Alqahtani, M.S.; El Dib, R.A.; AlAjmi, M.F. Alpha-Amylase and Alpha-Glucosidase Enzyme Inhibition and Antioxidant Potential of 3-Oxolupenol and Katonic Acid Isolated from *Nuxia oppositifolia*. *Biomolecules* **2020**, *10*, 61. [[CrossRef](#)]
45. Jannapureddy, S.; Sharma, M.; Yepuri, G.; Schmidt, A.M.; Ramasamy, R. Aldose Reductase: An Emerging Target for Development of Interventions for Diabetic Cardiovascular Complications. *Front. Endocrinol.* **2021**, *12*, 636267. [[CrossRef](#)]
46. Kazeem, M.I.; Adeyemi, A.A.; Adenowo, A.F.; Akinsanya, M.A. Carica papaya Linn. fruit extract inhibited the activities of aldose reductase and sorbitol dehydrogenase: Possible mechanism for amelioration of diabetic complications. *Future J. Pharm. Sci.* **2020**, *6*, 96. [[CrossRef](#)]
47. Ali, A.N.M.; Saeed, N.; Omeir, H.A. The Anticancer Properties of *Artemisia aucheri* Boiss Extract on HT29 Colon Cancer Cells. *J. Gastrointest. Cancer* **2021**, *52*, 113–119. [[CrossRef](#)]
48. Mashati, P.; Esmaeili, S.; Dehghan-Nayeri, N.; Bashash, D.; Darvishi, M.; Gharehbaghian, A. Methanolic Extract from Aerial Parts of *Artemisia annua* L. Induces Cytotoxicity and Enhances Vincristine-Induced Anticancer Effect in Pre-B Acute Lymphoblastic Leukemia Cells. *Int. J. Hematol. Oncol. Stem Cell Res.* **2019**, *13*, 132–139. [[CrossRef](#)]
49. Choi, E.J.; Kim, G.H. Antioxidant and anticancer activity of *Artemisia princeps* var. *orientalis* extract in HepG2 and Hep3B hepatocellular carcinoma cells. *Chin. J. Cancer Res. Chung-Kuo Yen Cheng Yen Chiu* **2013**, *25*, 536–543. [[CrossRef](#)]
50. Choi, E.; Park, H.; Lee, J.; Kim, G. Anticancer, antiobesity, and anti-inflammatory activity of *Artemisia* species in vitro. *J. Tradit. Chin. Med. Chung I Tsa Chih Ying Wen Pan* **2013**, *33*, 92–97. [[CrossRef](#)]
51. Ghanbar, F.; Mirzaie, A.; Ashrafi, F.; Noorbazargan, H.; Dalirsaber Jalali, M.; Salehi, S.; Sadat Shandiz, S.A. Antioxidant, antibacterial and anticancer properties of phyto-synthesised *Artemisia quttensis* Podlech extract mediated AgNPs. *IET Nanobiotechnol.* **2017**, *11*, 485–492. [[CrossRef](#)] [[PubMed](#)]
52. Salehi, S.; Shandiz, S.A.; Ghanbar, F.; Darvish, M.R.; Ardestani, M.S.; Mirzaie, A.; Jafari, M. Phytosynthesis of silver nanoparticles using *Artemisia marschalliana* Sprengel aerial part extract and assessment of their antioxidant, anticancer, and antibacterial properties. *Int. J. Nanomed.* **2016**, *11*, 1835–1846. [[CrossRef](#)]
53. Briguglio, G.; Costa, C.; Pollicino, M.; Giambò, F.; Catania, S.; Fenga, C. Polyphenols in cancer prevention: New insights (Review). *Int. J. Funct. Nutr.* **2020**, *1*, 9. [[CrossRef](#)]
54. Kopustinskiene, D.M.; Jakstas, V.; Savickas, A.; Bernatoniene, J. Flavonoids as Anticancer Agents. *Nutrients* **2020**, *12*, 457. [[CrossRef](#)]
55. Tong, W.; Chen, X.; Song, X.; Chen, Y.; Jia, R.; Zou, Y.; Li, L.; Yin, L.; He, C.; Liang, X.; et al. Resveratrol inhibits LPS-induced inflammation through suppressing the signaling cascades of TLR4-NF- $\kappa$ B/MAPKs/IRF3. *Exp. Ther. Med.* **2020**, *19*, 1824–1834. [[CrossRef](#)]
56. Yücel, G.; Zhao, Z.; El-Battrawy, I.; Lan, H.; Lang, S.; Li, X.; Buljubasic, F.; Zimmermann, W.-H.; Cyganek, L.; Utikal, J.; et al. Lipopolysaccharides induced inflammatory responses and electrophysiological dysfunctions in human-induced pluripotent stem cell derived cardiomyocytes. *Sci. Rep.* **2017**, *7*, 2935. [[CrossRef](#)]
57. Tang, J.; Diao, P.; Shu, X.; Li, L.; Xiong, L. Quercetin and Quercitrin Attenuates the Inflammatory Response and Oxidative Stress in LPS-Induced RAW264.7 Cells: In Vitro Assessment and a Theoretical Model. *Biomed. Res. Int.* **2019**, *2019*, 7039802. [[CrossRef](#)]
58. Kim, Y.J.; Park, W. Anti-Inflammatory Effect of Quercetin on RAW 264.7 Mouse Macrophages Induced with Polyinosinic-Polycytidylic Acid. *Molecules* **2016**, *21*, 450. [[CrossRef](#)]
59. Lee, A.J.; Cho, K.-J.; Kim, J.-H. MyD88–BLT2-dependent cascade contributes to LPS-induced interleukin-6 production in mouse macrophage. *Exp. Mol. Med.* **2015**, *47*, e156. [[CrossRef](#)]
60. Sakai, J.; Cammarota, E.; Wright, J.A.; Cicuta, P.; Gottschalk, R.A.; Li, N.; Fraser, I.D.C.; Bryant, C.E. Lipopolysaccharide-induced NF- $\kappa$ B nuclear translocation is primarily dependent on MyD88, but TNF $\alpha$  expression requires TRIF and MyD88. *Sci. Rep.* **2017**, *7*, 1428. [[CrossRef](#)]
61. Shukla, A.; Vats, S.; Shukla, R.K. Phytochemical Screening, Proximate Analysis and Antioxidant Activity of *Dracaena reflexa* Lam. Leaves. *Indian J. Pharm. Sci.* **2015**, *77*, 640–644. [[CrossRef](#)]
62. Agidew, M.G. Phytochemical analysis of some selected traditional medicinal plants in Ethiopia. *Bull. Natl. Res. Cent.* **2022**, *46*, 87. [[CrossRef](#)]
63. Owolabi, O.O.; James, D.B.; Sani, I.; Andongma, B.T.; Fasanya, O.O.; Kure, B. Phytochemical analysis, antioxidant and anti-inflammatory potential of FERETIA APODANTHERA root bark extracts. *BMC Complement. Altern. Med.* **2018**, *18*, 12. [[CrossRef](#)]

64. House, N.C.; Puthenparampil, D.; Malayil, D.; Narayanankutty, A. Variation in the polyphenol composition, antioxidant, and anticancer activity among different *Amaranthus* species. *S. Afr. J. Bot.* **2020**, *135*, 408–412. [[CrossRef](#)]
65. Ortiz-Cruz, R.A.; Ramírez-Wong, B.; Ledesma-Osuna, A.I.; Torres-Chávez, P.I.; Sánchez-Machado, D.I.; Montañó-Leyva, B.; López-Cervantes, J.; Gutiérrez-Dorado, R. Effect of Extrusion Processing Conditions on the Phenolic Compound Content and Antioxidant Capacity of Sorghum (*Sorghum bicolor* (L.) Moench) Bran. *Plant Foods Hum. Nutr.* **2020**, *75*, 252–257. [[CrossRef](#)]
66. Wang, B.; Liu, L.; Huang, Q.; Luo, Y. Quantitative Assessment of Phenolic Acids, Flavonoids and Antioxidant Activities of Sixteen Jujube Cultivars from China. *Plant Foods Hum. Nutr.* **2020**, *75*, 154–160. [[CrossRef](#)]
67. Liu, D.; Guo, Y.; Wu, P.; Wang, Y.; Kwaku Golly, M.; Ma, H. The necessity of walnut proteolysis based on evaluation after in vitro simulated digestion: ACE inhibition and DPPH radical-scavenging activities. *Food Chem.* **2020**, *311*, 125960. [[CrossRef](#)]
68. Bi, X.; Zhang, J.; Chen, C.; Zhang, D.; Li, P.; Ma, F. Anthocyanin contributes more to hydrogen peroxide scavenging than other phenolics in apple peel. *Food Chem.* **2014**, *152*, 205–209. [[CrossRef](#)]
69. Dutta, S.; Ray, S. Comparative assessment of total phenolic content and in vitro antioxidant activities of bark and leaf methanolic extracts of *Manilkara hexandra* (Roxb.) Dubard. *J. King Saud Univ. Sci.* **2020**, *32*, 643–647. [[CrossRef](#)]
70. Tonisi, S.; Okaiyeto, K.; Mabinya, L.V.; Okoh, A.I. Evaluation of bioactive compounds, free radical scavenging and anticancer activities of bulb extracts of *Boophone disticha* from Eastern Cape Province, South Africa. *Saudi J. Biol. Sci.* **2020**, *27*, 3559–3569. [[CrossRef](#)]
71. Mechchate, H.; Es-safi, I.; Louba, A.; Alqahtani, A.S.; Nasr, F.A.; Noman, O.M.; Farooq, M.; Alharbi, M.S.; Alqahtani, A.; Bari, A.; et al. In Vitro Alpha-Amylase and Alpha-Glucosidase Inhibitory Activity and In Vivo Antidiabetic Activity of *Withania frutescens* L. Foliar Extract. *Molecules* **2021**, *26*, 293. [[CrossRef](#)]
72. Karakaya, S.; Gözcü, S.; Güvenalp, Z.; Özbek, H.; Yuca, H.; Dursunoğlu, B.; Kazaz, C.; Kılıç, C.S. The  $\alpha$ -amylase and  $\alpha$ -glucosidase inhibitory activities of the dichloromethane extracts and constituents of *Ferulago bracteata* roots. *Pharm. Biol.* **2018**, *56*, 18–24. [[CrossRef](#)]
73. Kim, T.H.; Kim, J.K.; Kang, Y.-H.; Lee, J.-Y.; Kang, I.J.; Lim, S.S. Aldose Reductase Inhibitory Activity of Compounds from *Zea mays* L. *Biomed. Res. Int.* **2013**, *2013*, 727143. [[CrossRef](#)]
74. Al-Yousef, H.M.; Fantoukh, O.I.; El-Sayed, M.A.; Amina, M.; Adel, R.; Hassan, W.H.B.; Abdelaziz, S. Metabolic profiling and biological activities of the aerial parts of *Micromeria imbricata* Forssk. growing in Saudi Arabia. *Saudi J. Biol. Sci.* **2021**, *28*, 5609–5616. [[CrossRef](#)]