

*Supporting information*

# Olive pomace phenolic compounds stability and safety evaluation: from raw material to future ophthalmic applications

Nikolaos Katsinas <sup>1,2,3</sup>, Amalia Enríquez-de-Salamanca <sup>2,3</sup>, Andreia Bento da Silva <sup>4,5,6</sup>, Maria Rosário Bronze <sup>4,7,8</sup> and Soraya Rodríguez-Rojo <sup>1,\*</sup>

<sup>1</sup> Research Institute on Bioeconomy (BioEcoUVa), High Pressure Processes Group, School of Engineering, University of Valladolid (UVa), Dr. Mergelina str., 47011, Valladolid, Spain; nkatsinas@ioba.med.uva.es

<sup>2</sup> Institute of Applied Ophthalmobiology (IOBA), Campus Miguel Delibes, University of Valladolid (UVa), Paseo de Belén 17, 47011 Valladolid, Spain; amalia@ioba.med.uva.es

<sup>3</sup> Biomedical Research Networking Center in Bioengineering, Biomaterials and Nanomedicine (CIBER-BBN), Av. Monforte de Lemos, 3-5, 28029 Madrid, Spain

<sup>4</sup> Instituto de Investigação do Medicamento (iMed.ULisboa), Faculdade de Farmácia, Universidade de Lisboa (FFULisboa), Av. Prof. Gama Pinto, 1649-019, Lisbon, Portugal; mrbronze@ff.ulisboa.pt; abentosilva@ff.ulisboa.pt

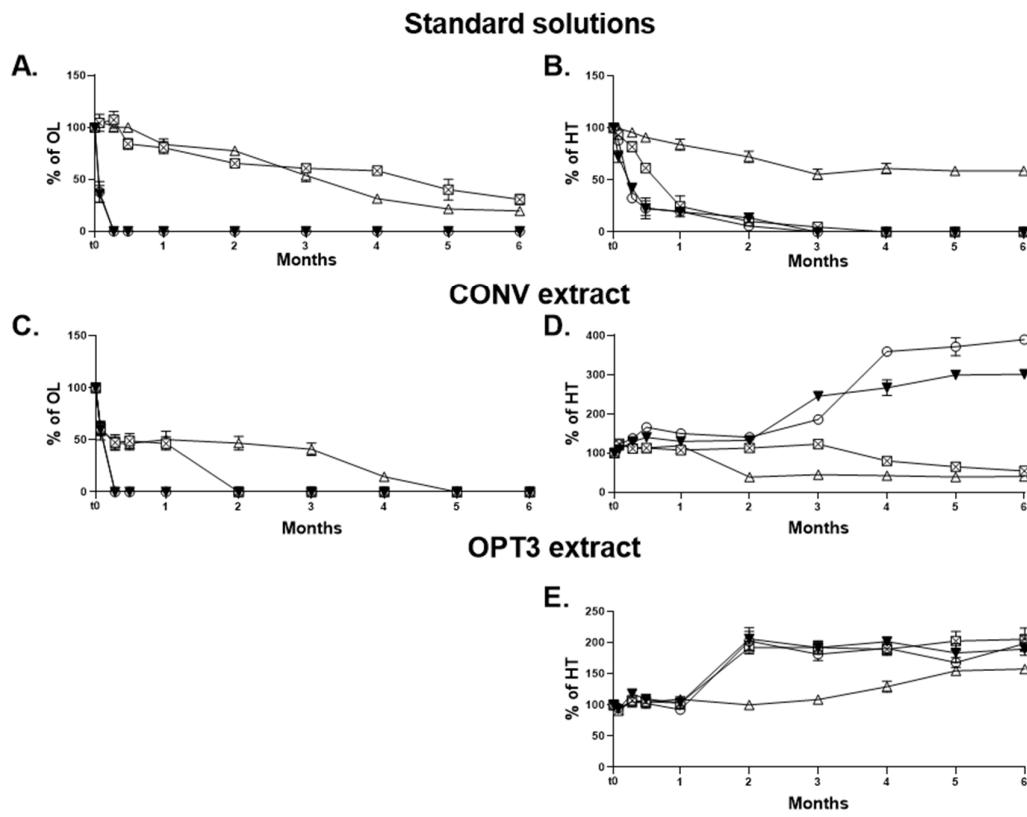
<sup>5</sup> Departamento de Ciências Farmacêuticas e do Medicamento (DCFM), Faculdade de Farmácia da Universidade de Lisboa (FFUL), Av. das Forças Armadas, 1649-003 Lisboa, Portugal

<sup>6</sup> Faculdade de Ciências e Tecnologia da Universidade Nova de Lisboa (FCT NOVA), Largo da Torre, 2829-516, Caparica, Portugal

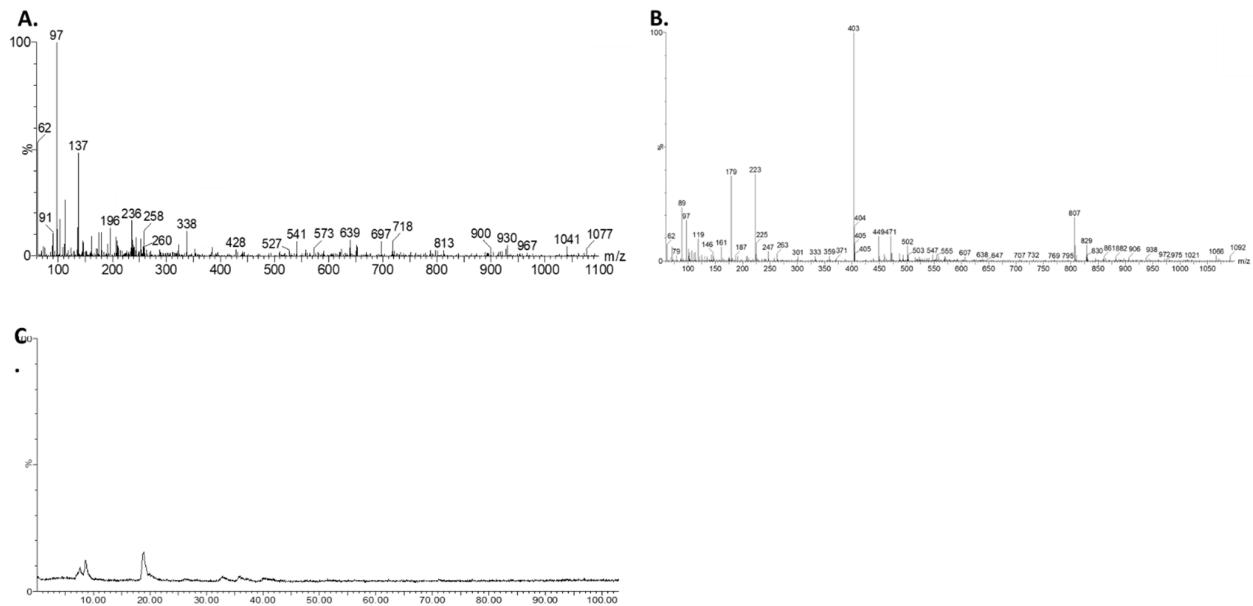
<sup>7</sup> Instituto de Biologia Experimental e Tecnológica (iBET), Apartado 12, 2780-901, Oeiras, Portugal

<sup>8</sup> Instituto de Tecnologia Química e Biológica da Universidade Nova de Lisboa (ITQB NOVA), Av. da República, 2780-157, Oeiras, Portugal

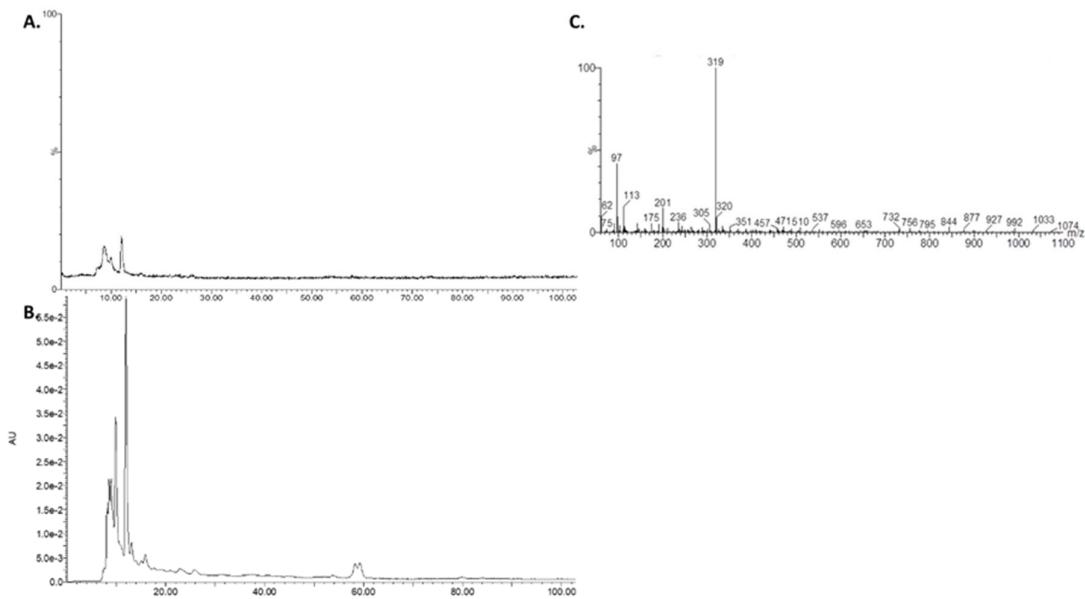
\* Correspondence: soraya.rodriguez@uva.es; Tel.: +34-983-184-077



**Figure S1.** Stability studies from 1 to 6 months of aqueous solution of oleuropein (OL) standard (A), hydroxytyrosol (HT) standard (B), OL (C) and HT (D) in conventional (CONV) extract, and HT (E) in optimized (OPT3) extract at 4 different conditions of temperature (T) and relative humidity (RH). Results are presented as average of percentage of each compound (HT or OL) with respect to the initial quantity of  $t_0 \pm$  standard deviation (SD). Lines added to guide the eye.



**Figure S2.** Full scan mass spectra acquired in electrospray ionization source in negative mode (ESI-) from peaks at 15.91 min (**A**) and 18.83 min (**B**) detected in the analysis of the oleuropein (OL) aqueous standard solution after exposure for 2 days at  $40 \pm 2$  °C and  $75 \pm 5\%$  relative humidity (RH). The scan chromatogram in is also presented (**C**).



**Figure S3.** UV Chromatogram at 280nm (**A**), scan chromatogram in electrospray ionization source in negative mode (ESI-) (**B**) and full scan mass spectra from peak at 10.14 min (**C**) detected in the analysis of the hydroxytyrosol (HT) aqueous standard solution after 30 days exposure at  $40 \pm 2$  °C and  $75 \pm 5\%$  relative humidity (RH).

**Table S1.** Analysis of variances (ANOVA) of all responses measured for extracts generated by conventional extraction conditions, using olive pomace (OP) stored at different conditions. Significance (*p*-value<0.05) is presented in bold.

Comparison	<i>p</i> -values							
	AA	TPC	TFC	OL	OLC	HT	TY	EY
Freeze-dried vs Fresh	<b>0.0057</b>	0.6992	<b>0.0042</b>	<b>0.0175</b>	<b>0.0015</b>	<b>0.0410</b>	0.1597	<b>0.0024</b>
Freeze-dried vs De-frozen	<b>0.0231</b>	0.9998	0.9622	<0.0001	<b>0.0052</b>	0.3166	<b>0.0302</b>	0.0623
Freeze-dried vs Dried	<b>0.0002</b>	0.2495	<0.0001	0.5729	<b>0.0006</b>	<b>0.0009</b>	<0.0001	0.1359
De-frozen vs Fresh	0.7141	0.7422	<b>0.0076</b>	<b>0.0011</b>	0.7101	0.4874	0.6505	0.1359
De-frozen vs Dried	<b>0.0161</b>	0.2762	<0.0001	<0.0001	0.2582	<b>0.0079</b>	<b>0.0014</b>	0.9429
Dried vs Fresh	0.0712	0.7836	<b>0.0062</b>	<b>0.0033</b>	0.7870	0.0592	<b>0.0004</b>	0.0623
Freeze-dried vs Fresh	<b>0.0057</b>	0.6992	<b>0.0042</b>	<b>0.0175</b>	<b>0.0015</b>	<b>0.0410</b>	0.1597	<b>0.0024</b>

\*Responses measured: Antioxidant activity (AA), total phenolic content (TPC), total flavonoid content (TFC), extract richness in oleuropein (OL), oleacein (OLC), hydroxytyrosol (HT) and tyrosol (TY), as well as extraction yield (EY)

**Table S2.** Analysis of variances (ANOVA) of the effect of 2 different defatting methods on the responses measured for extracts generated by conventional extraction conditions, using freeze-dried olive pomace (OP). An extract produced by non-defatted OP was used as the reference. Significance (*p*-value<0.05) is presented in bold.

Comparison	<i>p</i> -values							
	AA	TPC	TFC	OL	OLC	HT	TY	EY
Reference vs <i>n</i> -hexane	0.2550	0.4175	0.4327	0.2604	0.9984	>0.9999	0.7684	0.9968
Reference vs scCO <sub>2</sub>	0.4896	<b>0.0441</b>	>0.9999	0.9738	0.9935	0.8440	0.6892	0.1816
<i>n</i> -hexane vs scCO <sub>2</sub>	0.8428	0.2442	0.4327	0.3393	0.9854	0.8440	0.9891	0.1649

\*Responses measured: Antioxidant activity (AA), total phenolic content (TPC), total flavonoid content (TFC), extract richness in oleuropein (OL), oleacein (OLC), hydroxytyrosol (HT) and tyrosol (TY), as well as extraction yield (EY)

**Table S3.** Analysis of variances (ANOVA) of the effect of the extract drying on the stability of the responses measured compared to a freshly obtained liquid extract (reference). Significance (*p*-value<0.05) is presented in bold.

Comparison	<i>p</i> -values						
	AA	TPC	TFC	OL	OLC	HT	TY
Reference vs Step 1	0.6205	0.9791	0.9929	0.9053	0.9665	0.3077	0.9982
Reference vs Step 2	<0.0001	<b>0.0170</b>	0.6559	<b>0.0122</b>	0.1711	<b>0.0027</b>	<b>0.0094</b>
Step 1 vs Step 2	<0.0001	<b>0.0212</b>	0.5920	<b>0.0078</b>	0.2342	<b>0.0132</b>	<b>0.0089</b>

\*Responses measured: Antioxidant activity (AA), total phenolic content (TPC), total flavonoid content (TFC), and extract richness in oleuropein (OL), oleacein (OLC), hydroxytyrosol (HT) and tyrosol (TY).

**Table S4.** Analysis of variances (ANOVA) of the genotoxic effect (alkaline comet assay) of olive pomace (OP) extracts (0.8 mg/mL conventional – CONV and 0.4 mg/mL optimized – OPT3), together with 300 µM (162.2 mg/L) oleuropein (OL), 100 µM (15.4 mg/L) hydroxytyrosol (HT) and their mixture (5 µM/2.7 mg/L + 50 µM/7.7 mg/L OL+HT) on human corneal (HCE) and conjunctival (IM-ConjEpi) epithelial cells treated for 24 hours. Treatments are compared to control cells (treated with cell culture medium) and *p*-values lower than 0.05 were considered statistically significant.

Compared to control	<i>p</i> -values	
	HCE	IM-ConjEpi
OL	0.984	0.095
HT	1.000	0.982
OL+HT	0.450	0.603
CONV	1.000	0.359
OPT3	0.932	1.000