



Review

# The Impact of Electronic Nicotine Delivery System (ENDS) Flavors on Nicotinic Acetylcholine Receptors and Nicotine Addiction-Related Behaviors

Skylar Y. Cooper and Brandon J. Henderson \*

Department of Biomedical Sciences, Joan C. Edwards School of Medicine, Marshall University, Huntington, WV 25703, USA; cooper394@live.marshall.edu

\* Correspondence: hendersonbr@marshall.edu; Tel.: +1-304-696-7316

Academic Editors: Domenico Montesano and Riccardo Petrelli

Received: 19 August 2020; Accepted: 13 September 2020; Published: 15 September 2020



Abstract: Over the past two decades, combustible cigarette smoking has slowly declined by nearly 11% in America; however, the use of electronic cigarettes has increased tremendously, including among adolescents. While nicotine is the main addictive component of tobacco products and a primary concern in electronic cigarettes, this is not the only constituent of concern. There is a growing market of flavored products and a growing use of zero-nicotine e-liquids among electronic cigarette users. Accordingly, there are few studies that examine the impact of flavors on health and behavior. Menthol has been studied most extensively due to its lone exception in combustible cigarettes. Thus, there is a broad understanding of the neurobiological effects that menthol plus nicotine has on the brain including enhancing nicotine reward, altering nicotinic acetylcholine receptor number and function, and altering midbrain neuron excitability. Although flavors other than menthol were banned from combustible cigarettes, over 15,000 flavorants are available for use in electronic cigarettes. This review seeks to summarize the current knowledge on nicotine addiction and the various brain regions and nicotinic acetylcholine receptor subtypes involved, as well as describe the most recent findings regarding menthol and green apple flavorants, and their roles in nicotine addiction and vaping-related behaviors.

**Keywords:** nicotinic acetylcholine receptors; e-cigarettes; vaping; electronic nicotine delivery systems; flavorants; chemical flavors; terpene; menthol; green apple

## 1. Introduction

Cigarette smoking remains the leading cause of preventable disease and death worldwide, with nearly half a million deaths per year in the United States alone [1]. Additionally, more than 16 million Americans are suffering from a smoking-related disease including diabetes, stroke, cardiovascular disease, chronic obstructive pulmonary disease, or cancer [1]. Nearly 70% of adult smokers in the United States have a desire to quit, however, only 7% are successful [2], with an average of 10 cessation attempts needed for success [3].

In 2009, the Family Smoking Prevention and Tobacco Control Act was put in place to combat adolescent tobacco use and limit cigarette sales by banning all flavor additives other than menthol from being added to combustible cigarettes, yet this refrained from addressing other tobacco products including cigars, chewing tobacco, hookah, and more. As of 2018, mentholated cigarettes made up 36% of all cigarette sales in the United States [4], however, it has been established that regardless of labeling, even non-mentholated cigarettes contain traces of menthol [5].

Over the past two decades in the United States, combustible cigarette use has declined by nearly 11%, however, one form of nicotine administration has been replaced by another—electronic nicotine

Molecules **2020**, 25, 4223 2 of 28

delivery systems (ENDS) [6]. ENDS, or electronic cigarettes (e-cigarettes), are handheld devices that vaporize an e-liquid solution, commonly containing varying ratios of propylene glycol and vegetable glycerin, flavoring chemicals, nicotine, and in some cases additional sweeteners [7–9]. ENDS were initially intended to be a smoking cessation aid; however, ENDS companies have begun to target a new market of nicotine users among the adolescent population. According to the National Youth Tobacco Survey, usage rates continue to rise with over four million high school students and one million middle school students currently using ENDS products [10,11]. Nicotine dependence is thought to be intensified among adolescents when flavorants are present, as they mask the aversive sensations associated with nicotine and may promote pleasure on their own. This occurs primarily through masking the initial harshness of nicotine/tobacco that is aversive to new and beginning smokers and therefore increases smoking initiation [12,13]. Although non-menthol flavors are banned in combustible cigarettes, >15,000 flavor options are available for ENDS products with a 67% increase in flavor production from 2013 to 2014 [14,15]. In 2019, more than 50% and 60% of high school users used menthol and fruit flavored ENDS, respectively, and more than half of ENDS users prefer flavored products [10,11]. This has become a cause for concern with the number of adolescent ENDS users continuing to rise and the growing popularity of zero-nicotine flavored e-liquids. Yet, little is known regarding the effects of flavors on nicotine dependence and vaping-related behaviors.

Despite this gap in knowledge, there are numerous reports of menthol's effect on nicotine addiction, including menthol's ability to enhance nicotine reward and reinforcement [16–19]. These effects are due to menthol-induced nicotinic acetylcholine receptor (nAChR) upregulation [18,20,21], enhanced dopamine neuron excitability and dopamine release [22,23], and TrpM8-dependent mechanisms [24]. Based on a recent study reporting green apple and other fruity flavors to be the most popular of ENDS flavorants [9,25], additional reports have identified popular green apple flavorants, farnesol and farnesene, to not only enhance nicotine reward in a mouse model, but also display rewarding properties in the absence of nicotine [19,26,27]. These behavioral effects were found to be caused by changes in nAChR upregulation or stoichiometry, and ventral tegmental area dopamine neuron firing. Based on these findings, it is critical we further understand how ENDS flavoring chemicals may alter the addictive properties of nicotine in an attempt to combat the growing ENDS-use epidemic.

This review summarizes the current knowledge base of nicotine addiction and the major neurobiological and neurophysiological adaptations that contribute to dependence. Additionally, we summarize the effects of menthol on abuse liability and vaping-related behaviors and include the major impacts that another popular ENDS flavor, green apple, has on addiction-related behavior. This includes their effects on nicotine's actions in the brain and the major neurocircuitry involved in the induction of addiction.

# 2. Background of Nicotine Addiction

#### 2.1. Neuronal nAChRs: Structure and Function

nAChRs are ligand-gated ion channels in the Cys-loop superfamily, alongside N1-type acetylcholine, GABA<sub>A</sub> (ionotropic form), glycine, and 5-HT<sub>3</sub> receptors [28,29]. nAChRs are responsible for mediating fast synaptic transmission of nerve impulses [30–33]. Human nAChRs are assembled from various combinations of subunits, including  $\alpha 2-\alpha 7$  and  $\beta 2-\beta 4$ . Assembled subtypes consist of homomeric pentamers ( $\alpha 7$ ) or heteromeric pentamers ( $\alpha 2-\alpha 6$  with  $\beta 2-\beta 4$ ; Figure 1A). Each subunit comprises of a large *N*-terminal extracellular domain important for ligand binding, a short *C*-terminal extracellular domain, four hydrophobic transmembrane domains (M1–M4), with M2 lining the channel's central pore, and a large cytoplasmic loop between M3 and M4 that varies among different subunit complexes [34] (Figure 1B,C). The cytoplasmic loop is important for receptor trafficking in both the anterograde and retrograde directions between the endoplasmic reticulum to the plasma membrane [34–37]. These sites are also critical for protein interactions and serve as a site for phosphorylation.

Molecules **2020**, 25, 4223 3 of 28

nAChR agonists include the endogenous neurotransmitter, acetylcholine (ACh), as well as various exogenous molecules. Homomeric receptors have five identical orthosteric ACh-binding sites, whereas heteromeric receptors contain two or more orthosteric sites at the interfaces between an  $\alpha$  and  $\beta$ subunit (Figure 1A). The binding of two ACh molecules to the orthosteric sites on the receptors induces a conformational change allowing the channel to open. nAChRs are permeable to monovalent Na<sup>+</sup> and K<sup>+</sup> ions, as well as divalent Ca<sup>2+</sup> ions. Permeability to calcium acts on intracellular cascades that can play a vital role in neuronal signaling and plasticity [38]. Depending on the subtype assembly or stoichiometry of nAChRs,  $Ca^{2+}$  permeability varies. For instance, among  $\alpha 4\beta 2$  nAChRs, the high-sensitivity ( $\alpha 4_{(2)}\beta 2_{(3)}$ ) and low-sensitivity ( $\alpha 4_{(3)}\beta 2_{(2)}$ ) nAChR stoichiometries exhibit different  $Ca^{2+}$  permeabilities [39]; but homomeric  $\alpha 7$  nAChRs exhibit the highest  $Ca^{2+}$  permeability of all nAChR subtypes [40,41]. The net flow of positive ions inward depolarizes the cell membrane causing an excitatory postsynaptic potential [30]. nAChRs have a very widespread distribution throughout the brain and are found on presynaptic and postsynaptic neuronal membranes, as well as non-neuronal cells, such as glial cells [42,43]. nAChRs can control excitatory and inhibitory neurotransmission, which further excites and/or inhibits target cells [30,44]. Depending on the brain region and cell type, nAChR subunits form differing complexes that are associated with a variety of pathophysiological conditions. The most widely expressed neuronal subtypes in the brain are heteromeric  $\alpha 4\beta 2^*$  (\* = nAChR may contain other subunits) and homomeric  $\alpha$ 7 nAChRs [45–48]. Due to the variability in nAChR formation, each complex differs functionally with respect to channel opening, closing, and desensitization [47].

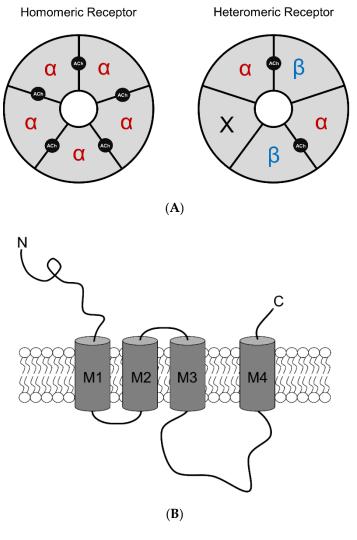
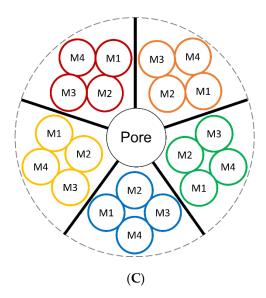


Figure 1. Cont.

Molecules **2020**, 25, 4223 4 of 28



**Figure 1.** Human nAChR structure and assembly. (**A**) Homomeric and heteromeric nAChR complexes assemble as solely  $\alpha$ -subunits or  $\alpha/\beta$ -subunits, respectively. Homomeric nAChRs possess five agonist binding sites at each  $\alpha$ - $\alpha$  interface while heteromeric nAChRs possess two agonist binding sites at the  $\alpha$ - $\beta$  interface but can still be weakly activated by 'non-canonical' sites at the  $\alpha$ - $\alpha$  interface (if present). X indicates other subunits may be present. (**B**) Single nAChR subunit topology consists of an extracellular domain, four transmembrane domains, and an intracellular loop that varies in length depending on subtype. (**C**) Pentameric nAChR complex from a top-side view identifying the formation of the transmembrane domains of individual nAChR subunits with respect to the nAChR central pore.

# 2.2. Nicotine's Actions in the Brain

Nicotine crosses the blood brain barrier and binds with high affinity to nAChRs widely distributed throughout the nervous system [32,38,48–50]. This interaction promotes a variety of neurophysiological changes including chaperone-mediated nAChR upregulation [30,35,37,51–54], activation of the mesocorticolimbic reward and reinforcement pathways [55,56], enhanced synaptic plasticity [57–60], and enhanced neuronal firing [53,60–63], ultimately leading to the development of nicotine addiction [33,64] (Table 1). This will be expounded in detail in the following paragraphs.

Nicotine binding promotes a conformational transition of nAChRs from a resting, closed state to an open state, allowing signal transduction to occur [65,66]. Initially, high affinity and high-sensitivity  $\alpha 4\beta 2^*$  and  $\alpha 4\alpha 6\beta 2^*$  nAChRs found on ventral tegmental area (VTA) dopamine (DA) neurons are quickly activated by low nicotine concentrations upon arrival into the midbrain region [67–70]. Further, nicotine acts on α7 nAChRs on glutamate neurons of local and distal regions resulting in a net excitatory effect on VTA DA neurons and synaptic strengthening between the two neuronal populations [47,60,71]. In addition to this excitatory transmission through DA and glutamate neurons, activation of the α4β2\* nAChRs found on GABA neurons of the midbrain elicits an inhibitory tone to VTA DA neurons, reducing the transmission of DA and decreasing reward through the mesolimbic pathway (dopaminergic tracts from the VTA to the nucleus accumbens (NAc)) [72]. However, the activation of α4β2\* and α7 nAChRs on midbrain DA and glutamate neurons, respectively, promotes a net excitatory effect on DA neurotransmission from the VTA to the NAc and prefrontal cortex (PFC), leading to the rewarding and reinforcing aspects of nicotine use [71]. More recently, Yan et al. discovered functional β2\* nAChRs on medial VTA (mVTA) glutamate neurons that further contribute to this net effect [63,73]. In general, nicotine exerts strong effects on the local microcircuits of the VTA, which have differing acute and chronic effects. This will be discussed further in a later section.

Molecules **2020**, 25, 4223 5 of 28

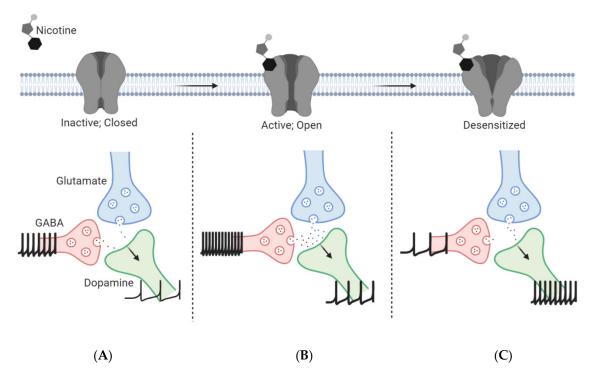
**Table 1.** Nicotine's actions in the brain.

Steps of Nicotine Addiction	nAChR Subunits	Brain Regions	Nicotine Concentration	Duration of Administration
Nicotine binding promotes a conformational transition of nAChRs from a resting, closed state to an open state, allowing signal transduction to occur [65,66].	α4β2*, α4α6β2*, α6β2β3, α7	Midbrain	Small (<1 μM)	Acute
When an individual becomes a long-term nicotine user, nAChR upregulation occurs.	α4, α6, β2, and β3	VTA		
Upregulated $\alpha 4\beta 2^*$ nAChRs found on GABA neurons of the midbrain elicit an inhibitory tone to VTA DA neurons, reducing the transmission of DA and contribute to nicotine-seeking behaviors [72].	α4β2*	Midbrain GABA	20–500 nM	Chronic
After long-term exposure to nicotine, $\alpha 4\beta 2^*$ nAChRs desensitize quickly, which decreases the GABAergic transmission onto DA neurons resulting in the disinhibition of DA neurons.	α4β2*	Midbrain GABA		
The activation of $\alpha 4^*$ , $\alpha 6^*$ , and $\alpha 7$ nAChRs on midbrain DA and glutamate neurons, respectively, promotes a net excitatory effect on DA neurotransmission from the VTA to the NAc and PFC, leading to the rewarding and reinforcing aspects of nicotine use [71].	α4β2*, α6β2β3, α4α6β2*, α7	VTA	20–500 nM	Chronic
These effects are reinforced by the enhanced glutamate neurotransmission from distal regions and local mVTA glutamate neurons to lateral VTA (latVTA) DA neurons [63,73], leading to long-term potentiation (LTP).	α7	mVTA, PFC		

Summarized details of the various steps (sensitization, upregulation, and desensitization) that take place during the development of nicotine dependence. The major findings for each step are indicated in the leftmost column. The specific nAChR subunits and brain region associated with these changes are in the second and third column. The nicotine concentration and duration of administration that induce these changes are located in the two rightmost columns. \* = nAChR may contain other subunits.

Persistent nicotine exposure further transitions the receptor to a desensitized state, where it is less responsive to agonist stimulation [34,44,45]. nAChRs are activated and desensitized to a degree depending on the subtype, brain region, and concentration of nicotine. According to Brody and colleagues, nAChRs will activate and desensitize at brain nicotine concentrations of 20–100 nM following cigarette smoking [74]. Following activation, most  $\alpha4\beta2^*$  nAChRs desensitize, which decreases the GABAergic transmission onto DA neurons resulting in the disinhibition of DA neurons (Figure 2C). These effects are reinforced by the enhanced glutamate neurotransmission from distal regions and local mVTA glutamate neurons to lateral VTA (latVTA) DA neurons [63,73]. This depolarization and enhanced action potential firing of glutamatergic neurons leads to long-term potentiation (LTP) and synaptic strength onto the midbrain DA neurons, which is a key role in the formation of nicotine dependence [53] (Figure 2). Additionally, although the  $\alpha4\beta2^*$  nAChRs present on VTA DA neurons desensitize similar to VTA GABA neurons, these DA neurons also express  $\alpha6\beta2\beta3^*$  and  $\alpha4\alpha6\beta2^*$  receptors, which do not desensitize as quickly as the high affinity  $\alpha4\beta2^*$  nAChRs [55]. Overall, there continues to be a net excitatory effect on DA release in the presence of nicotine molecules.

Molecules **2020**, 25, 4223 6 of 28



**Figure 2.** Involvement of ventral tegmental area (VTA) neuron types in nicotine reward and reinforcement. (**A**) In the absence of nicotine, glutamate and GABA inputs to VTA dopamine neurons modulate activity of the mesolimbic reward pathway. (**B**) Acute nicotine on  $\alpha$ 7 and  $\alpha$ 4 $\beta$ 2 nAChRs elicits enhanced glutamatergic and GABAergic firing, respectively, onto VTA dopamine neurons, resulting in a net enhancement of dopaminergic neuron firing and subsequent dopamine release. (**C**) Following long-term nicotine use,  $\alpha$ 4 $\beta$ 2 nAChRs desensitize rapidly with acute exposure to nicotine and result in reduced GABAergic firing while glutamatergic firing is enhanced to stimulate burst firing of dopaminergic neurons (see also Table 1 for additional details).

When an individual becomes a long-term nicotine user, further neuroadaptations occur. A major hallmark of nicotine dependence has been the occurrence of nAChR upregulation. This phenomenon has been understood by many to be a post-translational occurrence, based on no observable changes in mRNA levels [35,75,76]. Similar to desensitization, upregulation differs in response to nicotine concentration and time course [77], and is brain-region, cell-type, and nAChR subtype-specific, given that no upregulation has been noted in the thalamus, high-affinity nAChRs are favored for plasma upregulation, and only  $\alpha 4$ ,  $\alpha 6$ ,  $\beta 2$ , and  $\beta 3$  subunits upregulate. Upregulation occurs through a process termed pharmacological chaperoning that involves intracellular actions of nicotine that promote nAChR subunit assembly and enhanced trafficking of nAChRs through the secretory pathway [35,37,52,76]. This process is a physiological response following nAChR desensitization [45,78] that takes approximately ten days of long-term use among human and rodents but can also occur in cultured neuron and cell line preps as well [76,79]. Through the use of fluorescently tagged nAChRs, changes in nAChR number have been measured without the use of radioligand binding assays [18,20,35,37,53]. As previously stated, high-affinity  $\alpha 4\beta 2$  nAChRs are stabilized during this process [37,52,80]. With higher nicotine concentrations, further nAChRs are subjected to upregulation as well.

Given these effects are based on freebase nicotine that is present in combustible cigarettes, it is critical to point out the distinct differences between the combustible cigarette and the electronic cigarette that may affect the timespan of these steps towards addiction. With JUUL (pod-based e-cigarette) being one of the most popular ENDS devices among the adolescent population, studies have begun to surface identifying what makes JUUL so popular. The major difference between these products is the use of nicotine salt in JUULs [81]. Nicotine salt differs from nicotine freebase in that it contains benzoic acid in

Molecules **2020**, 25, 4223 7 of 28

order to sufficiently protonate most of the nicotine. The use of nicotine salt results in faster absorption, increased nicotine strength, and can be palatable at high concentrations, as opposed to nicotine freebase. Based on the different pharmacokinetic properties of these two nicotine forms, JUUL's average nicotine concentration per e-liquid pod is ~60 mg/mL, or 1.5 times the nicotine concentration in a pack of cigarettes. Although combustible cigarettes range in nicotine concentration, the majority contain 2 mg of nicotine per cigarette. Additionally, various other ENDS products (tank-based e-cigarette) also exhibit higher nicotine concentrations than the average cigarette (3–18 mg/mL), however, they are still much lower than the prefilled JUUL pods. Based on these differences, it is likely that ENDS enhance the steps of nicotine addiction and are further detrimental to the adolescent brain.

#### 2.3. nAChR Subtypes

#### 2.3.1. α4β2\* nAChRs

To date,  $\alpha 4\beta 2^*$  nAChRs have been the most-studied subtype in relation to nicotine addiction [33,37,45,55,61,82], as demonstrated through genetically modified mouse models [55,56,82,83]. Using knockout mice, Picciotto et al. investigated  $\alpha 4$  and  $\beta 2$  subunits and found attenuated nicotine self-administration and conditioned place preference in the absence of these subunits [83]. However, when  $\alpha 4$  or  $\beta 2$  subunits were re-expressed in the VTA, a prominent brain region associated with drug reward, it led to recovery of nicotine reinforcement and reward, demonstrating the importance of  $\alpha 4\beta 2^*$  nAChRs in the formation of nicotine dependence, specifically in the VTA. Alternatively, Tapper et al. genetically altered mice to express hypersensitive  $\alpha 4^*$  nAChRs [55]. This approach looked beyond the necessary presence of  $\alpha 4\beta 2^*$  nAChRs in addiction, and instead investigated the specific role(s) that  $\alpha 4^*$  may play in addiction. The presence of hypersensitive  $\alpha 4^*$  not only enhanced the nicotine reward at low concentrations, it also mediated nicotine-induced locomotion through  $\alpha 4^*$  nAChR sensitization, and facilitated tolerance to nicotine, deeming its importance in the induction and maintenance of nicotine dependence. Further, the administration of the  $\beta 2^*$  receptor-specific antagonist, dihydro- $\beta$ -erythroidine (Dh $\beta E$ ) hydrobromide, into the VTA decreases nicotine reward-related behavior as well as blocks the reinforcing effects of the drug [84,85].

 $\alpha4\beta2^*$  nAChRs exist in a high- and low-sensitivity stoichiometry,  $\alpha4_{(2)}\beta2_{(3)}$  and  $\alpha4_{(3)}\beta2_{(2)}$ , respectively [39,86]. Due to its high sensitivity stoichiometry,  $\alpha4\beta2^*$  receptors have a high affinity for low concentrations of nicotine commonly present following cigarette smoke inhalation [33,67,87]. Nicotine-induced upregulation of  $\alpha4\beta2^*$  nAChRs has been shown to depend on the specific subunit composition, with the high-sensitivity stoichiometry to be favored [35,37,80,88]. Further, it is important to note the more recent identification of genetic etiologies of CHRNA4, CHRNB2, and CHRNB4. Liu et al. positively correlated these genes with smoking phenotypes including 'age of smoking initiation', 'cigarettes per day', and 'smoking cessation' [89].

#### 2.3.2. $\alpha 6\beta 2\beta 3^*$ nAChRs

Given the knowledge that  $\alpha6^*$  nAChRs are expressed in dopaminergic neurons [90], initial studies aimed to identify the specific brain regions expressing these subtypes. The expression of  $\alpha6\beta2\beta3^*$  nAChRs is mostly limited to reward-related brain regions, including the DA neurons of the mesolimbic tract, making them critical factors in the induction of addiction [91–94]. However, investigations into the role of  $\alpha6^*$  nAChRs in addiction have been very limited, with one of the first critical findings being reported by Pons et al. [56]. Utilizing an intravenous nicotine self-administration paradigm and  $\alpha6$  knock-out mice, Pons et al. discovered the essential role of  $\alpha6^*$  nAChRs in nicotine reinforcement by demonstrating the lack of self-administration behaviors in  $\alpha6$  knock-out mice compared to their wildtype counterparts [56]. Furthermore, re-expression of  $\alpha6$  through a lentiviral vector into the VTA significantly increased self-administration behaviors to a similar extent as the wildtype control mice. Nicotine-induced activation of the mesolimbic system is dependent upon the presence of  $\alpha4\beta2^*$  and  $\alpha6\beta2\beta3^*$  nAChRs, as seen through conditioned place preference and self-administration

Molecules **2020**, 25, 4223 8 of 28

assays [56,83,95]. Similar to  $\alpha4\beta2^*$  nAChRs,  $\alpha6^*$  nAChRs consist of two stoichiometric forms,  $\alpha6\beta2$  or  $\alpha6_{(2)}\beta2_{(2)}\beta3$  [20,35]. The inclusion of the  $\beta3$  subunit results in a much higher sensitivity to nicotine. Further,  $\alpha6\beta2^*$  nAChRs desensitize to a similar extent as  $\alpha4\beta2^*$  nAChRs, however  $\alpha6\beta2^*$  nAChRs recover more quickly [96] and are thus more amenable to nicotine's long-term effects.

## 2.3.3. $\alpha 4\alpha 6\beta 2^*$ nAChRs

Despite the wide presence of nAChRs throughout the central nervous system, recent studies point to  $\alpha 4\alpha 6\beta 2^*$  nAChRs as a principle nAChR subtype that mediates nicotine reward. As discussed earlier, deletions of  $\alpha 4$  have shown the importance of this subunit; but additional studies with mouse models tell us that  $\alpha 4\alpha 6\beta 2^*$  nAChRs mediate the rewarding effects of nicotine. Deletion of  $\alpha 4$ ,  $\alpha 6$ , or  $\beta 2$  nAChR subunits is sufficient to block the self-administration of nicotine in mice (Pons et al., 2008). The selective re-expression of these deleted subunits in the VTA was sufficient to reinstate self-administration of nicotine (Pons et al., 2008). Re-expression of these subunits in neighboring midbrain regions, the substantia nigra pars reticulata (SNr) and substantia nigra pars compacta (SNc), did not 'rescue' nicotine reinforcement. The nicotine-induced enhancement of DA neuron excitability that is required for reward is dependent upon  $\alpha 4\alpha 6\beta 2^*$  nAChRs [69,70]. Here, both Liu et al. and Engle et al. revealed that the presence of both  $\alpha 4$  and  $\alpha 6$  were necessary for smoking-relevant concentrations of nicotine (300 nM) to depolarize nAChRs on VTA DA neurons. This suggests that while studies revealing the importance of  $\alpha 4$  and  $\alpha 6$  nAChR subunits independently, the physiological response to smoking-relevant concentrations of nicotine ( $\leq 300$  nM) depend on  $\alpha 4\alpha 6$ -containing nAChRs.

#### 2.3.4. α7 nAChRs

Alongside  $\alpha 4\beta 2^*$ ,  $\alpha 6\beta 2^*$ , and  $\alpha 4\alpha 6\beta 2^*$  nAChRs, the  $\alpha 7$  nAChR subtype also plays a prominent role in the formation of nicotine dependence. This homomeric nAChR subtype is largely present on glutamatergic neurons in the prefrontal cortex (PFC), hippocampus, and mVTA. Expression of  $\alpha 7$  nAChRs in these brain regions play multiple roles in nicotine addiction; they aid in the formation of functional synapses [49], promote the net excitatory effect on midbrain DA neurons of the reward pathway [60], and enhance postsynaptic excitation through NMDA receptors in the hippocampus and prefrontal cortex [49,60]. More recently, a population of functional heteromeric nAChRs has been discovered on mVTA glutamatergic neurons [63]. This novel finding supports the previous work demonstrating the intricate connections between VTA glutamate and DA neurons in reward and reinforcement processing [60,71,97], but further expands this knowledge by observing that nicotine mediates the excitatory transmission at this connection and thus, amplifies the mesocorticolimbic reward transmission [63].

The  $\alpha 7$  nAChR exhibits different receptor kinetics compared to  $\alpha 4\beta 2^*$  nAChRs, exemplified by fast activation and desensitization [44]. Due to their desensitization properties,  $\alpha 7$  nAChRs recover very rapidly, demonstrating the net excitation discussed earlier. The net excitation through rapid recovery following desensitization, and high permeability to calcium, makes  $\alpha 7$  nAChRs critical factors in the induction of long-term potentiation (LTP) [46,98]. Nicotine-induced synaptic plasticity, or LTP, of glutamatergic neurons in the VTA, PFC, and hippocampus are important in the tolerance and associative learning of nicotine addiction. Regardless of the fast depolarization of VTA DA neurons through nicotine's effect on  $\alpha 4\beta 2^*$ ,  $\alpha 6\beta 2^*$ , and  $\alpha 4\alpha 6\beta 2^*$  nAChRs, these receptors desensitize fairly quickly and recover slowly, rendering them inactive to the remaining nicotine molecules circulating in the brain. Thus, it is the presence of  $\alpha 7$  nAChRs and their rapid recovery that contribute to the long-standing effects of long-term nicotine. Additionally, administration of methyllycaconitine (MLA), an  $\alpha 7$ -selective antagonist, did not alter nicotine CPP, but decreased self-administration, confirming  $\alpha 7$  nAChRs' role in reinforcement but not reward [95]. These findings were further verified by the lack of change in nicotine self-administration behavior between  $\alpha 7$ -KO and  $\alpha 7$ -WT mice [56].

It is the reinforcing properties of nicotine that promote dependence, despite the many health risks. VTA DA neurons receive local and distal (mainly PFC) excitatory glutamatergic inputs that

Molecules **2020**, 25, 4223 9 of 28

further prompts DA release into the NAc, amygdala, and PFC, which makes up the mesocorticolimbic system [99]. However, when nicotine is persistently present in this system, it causes modification of neural circuits that promotes drug-induced synaptic plasticity [100]. This phenomenon outlasts the effects of the presence of nicotine molecules and contributes to the formation of tolerance and memory consolidation to nicotine. It is the repetitive use and reinforcing properties of nicotine that causes the brain to form an association between nicotine use and the physiological response to nicotine use. This association is what drives nicotine dependence by mediating the behavioral effects of nicotine addiction, including cue-induced craving and reinstatement (in rodents) or relapse (in humans) [101].

## 2.3.5. $\alpha$ 3\* and $\alpha$ 5\* nAChRs

Genetic polymorphisms in the  $\alpha 3/\alpha 5/\beta 4$  nAChR subunit gene cluster has been linked to increased risk to nicotine addiction [102,103]. Variation in the amino acid sequence of CHRNA5 results in reduced  $\alpha 5^*$  nAChR activity and increases the risk for dependence [102,104,105]. Additionally, Fowler et al. identified a null mutation in the CHRNA5 gene that results in significantly more intravenous self-administration nicotine consumption, compared to their control mice [106]. Interestingly, they found that  $\alpha 5^*$  nAChRs are involved in mediating aversive stimuli, titrating nicotine intake, and mediating somatic signs of withdrawal.

The  $\alpha5^*$  and  $\alpha3\beta4^*$  nAChRs are mostly expressed in the habenulo-IPN pathway, a group of cholinergic and glutamatergic neurons that project from the medial habenula (MHb) to the interpeduncular nucleus (IPN) through a bundle of axons termed the fasciculus retroflexus [33,106,107]. Due to their low affinity for nicotine, these subtypes require high concentrations of nicotine to sensitize them (2 mg/kg or more) [47,106,107]. The opening of  $\alpha5^*$  or  $\alpha3\beta4^*$  nAChRs activates the aversive pathways involved in the mediation of nicotine intake. The activation of either the habenulo-IPN pathway or the lateral habenula (LHb) projections to the rostromedial tegmental nucleus (RMTg) and VTA is a response to the aversive aspects of nicotine, including high doses of nicotine or nicotine withdrawal (discussed later).

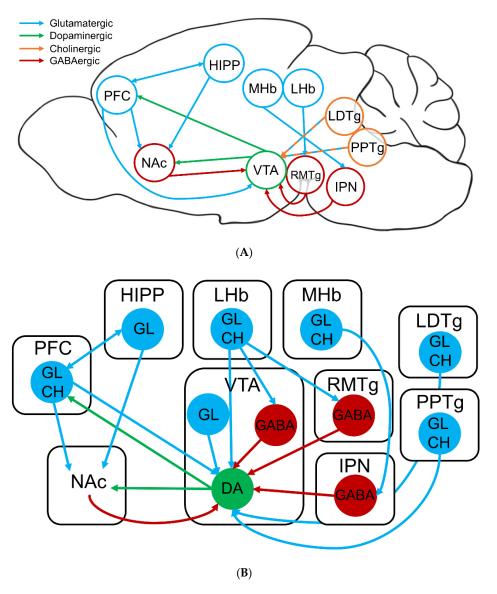
#### 3. Neurocircuitry Involved in Nicotine Addiction

## 3.1. Ventral Tegmental Area (VTA)

The ventral tegmental area is among the most vital brain regions involved in addiction. The VTA is a heterogeneous collection of neurons that in part makes up the midbrain. Although primarily studied for its DA neurons, the VTA also consists of  $\gamma$ -amino butyric acid (GABA) and glutamatergic neurons [108]. VTA DA neurons have been widely studied in various drug-related experiments for nearly all drugs of abuse acts on VTA DA neurons to stimulate DA neurotransmission into the ventral striatum, or nucleus accumbens (NAc)—commonly known as the mesolimbic pathway [68,109–111]. Further, nicotine-induced activation of this pathway stimulates reward, as seen in conditioned place preference (CPP) assays [26,55,82,112]. Whereas, lesioning of this pathway completely abolishes nicotine reward-related behavior, nicotine self-administration, and nicotine-induced locomotion [85,113,114]. Additionally, VTA DA neurons project to the prefrontal cortex (PFC) through the mesocortical pathway, and synapse on cortical pyramidal neurons and GABAergic interneurons. This dopaminergic pathway is largely associated with drug reinforcement and is commonly activated during nicotine self-administration studies [115,116]. Together, these paths make up the mesocorticolimbic DA pathway and are often linked in studies on drugs of abuse, although there are additional projections to various other brain regions, including the hippocampus and amygdala. Each of these neuronal paths circle back to the VTA and mediate the activity of the dopaminergic neurons (discussed further in their corresponding sections).

The VTA also receives afferent projections from numerous parts of the brain that mediate reward or aversion of certain stimuli. The cholinergic and glutamatergic projections of the laterodorsal tegmental nucleus (LDTg) and pedunculopontine tegmental nucleus (PPTg) excite the DA neurons of the VTA

leading to burst firing, enhanced dopamine release, and reward-related behavior (Figure 3) [117–119]. During long-term nicotine exposure, these glutamatergic projections can initiate long-term potentiation (LTP) in the VTA, promoting a continuous excitatory effect [120]. On the contrary, the medial (MHb) and lateral habenula (LHb) neurons innervate the interpeduncular nucleus (IPN) and rostromedial tegmental nucleus (RMTg) GABAergic neurons, respectively, which employ an inhibitory tone onto VTA DA neurons of the mesolimbic pathway (Figure 3) [121,122]. This net inhibition is stimulated during aversive stimuli, such as high nicotine concentrations, or during withdrawal [106,107,123]. Additionally, the LHb sends direct projections to the VTA in response to aversive stimuli, including the absence of an expected reward [124]. These different afferent projections mediate the firing patterns of VTA DA neurons and influence behavioral outputs.



**Figure 3.** Neurocircuitry involved in nicotine addiction. **(A)** Sagittal mouse brain schematic of the major brain regions and connections involved in nicotine addiction. **(B)** Intricate schematic of the above neurocircuitry. DA: dopaminergic, GABA: GABAergic, GL: glutamatergic, CH: cholinergic neurons. Blue arrows indicate excitatory projections; red arrows indicate inhibitory projections; and green arrows indicate modulatory projections.

Although many drugs of abuse studies have focused on DA neurotransmission, GABAergic and glutamatergic neurons of the VTA are prominent factors in the development of nicotine dependence.

Opioids, cannabinoids, and benzodiazepines primarily target the VTA GABAergic interneurons, inhibiting their activity and causing disinhibition and burst firing of the neighboring DA neurons [125]. Both GABAergic and glutamatergic interneurons of the VTA send projections to their neighboring DA neurons in order to maintain a homeostatic balance of neuronal firing. Following a cigarette or ENDS puff, nicotine activates the nAChRs found on each of these neuronal populations and increases neuronal firing [126]. However, in a long-term user, these neuronal firing patterns become altered following nAChR desensitization (see Figure 2). The  $\alpha$ 7 nAChRs expressed on glutamatergic neurons desensitize (and recover) the fastest while  $\beta 2^*$  nAChRs ( $\alpha 4\beta 2^*$ ,  $\alpha 6\beta 2^*$ , and  $\alpha 4\alpha 6\beta 2^*$ ) desensitize (and recover) much slower. As stated previously, a long-term nicotine user succumbs to various neuroadaptations, including nAChR upregulation and altered neuronal firing. Throughout abstinent periods, the GABAergic neuron firing patterns are enhanced, resulting in a net inhibitory effect on the DA neurons. This not only reduces baseline DA neurotransmission but also causes a depressive state that often triggers craving and relapse. In order to relieve this, the user will consume more nicotine, activate the nAChRs on the various neuronal populations, and increase firing and DA release. With repeated exposure and stimulation, nicotine renders many of the nAChRs inactive. Based on their desensitization properties, the GABA neurons (containing  $\alpha 4\beta 2^*$ ) have reduced output, resulting in disinhibition of the VTA DA neurons [71]. Additionally, the fast recovery of  $\alpha$ 7 nAChRs promotes a net excitatory effect on these DA neurons. It has long been considered that nicotine exerts its effects on α7 and β2\* nAChRs on VTA glutamate and GABA neurons, respectively, resulting in a mediatory action on VTA DA neurons. However, recent studies have shown that these glutamate neurons, found primarily in the mVTA, express  $\alpha 4$ ,  $\alpha 6$ , and  $\beta 2$  nAChR subunits in a somatodendritic manner [73]. Further, Yan et al. has discovered that of the three major neuronal populations in the mVTA (VGluT2<sup>+</sup> (glutamatergic neurons), Gad2<sup>+</sup> (GABAergic neurons), and VGluT2<sup>+</sup>/Gad2<sup>+</sup> (glutamatergic/GABAergic co-releasing neurons)), nicotine enhanced glutamate release into the latVTA via VGluT2<sup>+</sup> neurons but decreased glutamate (and GABA) release via Gad2<sup>+</sup> and VGluT2<sup>+</sup>/Gad2<sup>+</sup> neurons [73]. This further demonstrates the complexity of the VTA, and the various microcircuits involved in reward processing. It is due to these changes that a person dependent on nicotine requires more nicotine over time to achieve the rewarding feeling they once felt. Additionally, more studies are identifying the role of glycine in this microcircuit. Although little work has been performed on glycine's impact on nicotine addiction, it has been shown that glycinergic neurotransmission significantly alters ethanol intake most likely by impacting glutamate release via glycine receptors on glutamatergic terminals of the VTA [127], however these receptors are also present on GABAergic terminals and have resulted in reduced GABA release and enhanced dopamine release [128,129]. Accordingly, glycine receptors may play a role in nicotine's actions; but this has yet to be determined.

# 3.2. Substantia Nigra (SN)

The other contributing cluster of neurons that makes up the midbrain is the substantia nigra. The SN is composed of two subgroups: pars compacta (SNc) and pars reticulata (SNr). The SNc is composed of dopaminergic, glutamatergic, and GABAergic neurons while the SNr is largely made up of GABA neurons. The third most extensively studied dopaminergic pathway in the brain aside from the mesocorticolimbic paths is the nigrostriatal pathway, a dopaminergic tract from the SNc to the dorsal striatum [130]. Activation of this pathway is involved in the motor loop of the basal ganglia and has been shown to be acted on by drugs of abuse [131]. The loss of dopaminergic neurons in this pathway is a pathological classification of Parkinson's disease, characterized by tremors and motor deficits.

#### 3.3. Nucleus Accumbens (NAc)

A major output target of VTA DA neurons is the ventral striatum, or the nucleus accumbens (NAc), which makes up the mesolimbic pathway. This pathway is highly involved in reward-related effects of drugs of abuse [68,109]. The NAc contains two subregions: the core and the shell, and is

made up of specialized GABAergic neurons, termed medium spiny neurons (MSNs). MSNs consist of two types: D1 (direct pathway) and D2 (indirect pathway) based on the DA receptors present, further explained by Cooper et al. [132]. These neurons receive excitatory projections from the PFC, hippocampus (HIPP), and the VTA [100] to mediate the reinforcing and drug-seeking behaviors of nicotine addiction [133,134]. Repeated activation of the afferent glutamatergic projections can lead to LTP in the NAc, further driving drug-taking behaviors, as demonstrated through reduced AMPAR and AMPAR/NMDAR ratios on MSNs following drug consumption [100]. The D1 MSNs of the lateral NAc shell project in a cyclical manner back to the VTA and synapse on VTA GABA neurons [100,135]. These projections further mediate and disinhibit the VTA DA neuron activity, resulting in enhanced DA release [136,137].

# 3.4. Prefrontal Cortex (PFC)

Dopaminergic projections from the VTA to the PFC make up the mesocortical pathway involved in reinforcement and motivational salience [115,132,133]. This pathway has enhanced neurotransmission during nicotine self-administration, demonstrated by an increase in postsynaptic ionotropic glutamate receptors in the PFC [138]. PFC lesions significantly reduced nicotine self-administration in rats, however, to date, this has only been performed in neonates [139]. Additionally, glutamatergic pyramidal neurons of the PFC project back to the VTA, amygdala, hippocampus, and NAc. The connection between the PFC and NAc is stimulated during drug seeking and reinstatement [140,141]. Due et al. has demonstrated PFC activation of smoker's brains via functional magnetic resonance imaging in the presence of a smoking-related cue (an image of a person smoking) [142]. Activation of the PFC to VTA pathway results in enhanced firing of the VTA DA neurons and increases DA release through the mesolimbic pathway [143], in part due to upregulation of NMDA and AMPA receptor activation [60].

Based on the growing number of adolescent ENDS users, it is important to discuss the effects of nicotine exposure on the developing brain. More specifically, the PFC is one of the last brain regions to fully reach maturation in the developing brain, which indicates the executive control functions, including attention and working memory are not fully developed upon initial nicotine exposure. Nicotine use or exposure during this critical development period is known to negatively impact the developing process, resulting in impaired cognition and psychiatric disorders, including depression [144,145], and also often results in an increased risk for drug abuse behaviors [146].

### 3.5. Hippocampus (HIPP)

The hippocampus (HIPP) is highly involved in learning and memory, which is of importance to the tolerance-related aspect of nicotine addiction. The glutamatergic neurons of the HIPP are simultaneously excited by VTA DA and PFC pyramidal neurons, which induces LTP in the hippocampus. The strengthening of hippocampal synapses is a key feature to nicotine addiction because of its underlying mechanism involved in cue-induced drug-seeking behavior often leading to relapse [147]. The changes in synaptic machinery are due to the initial formation of immature glutamatergic synapses from the HIPP onto NAc MSNs, leading to silent synapses, which promotes no electrical changes on the inhibitory neurons of the NAc. These immature synapses are due to a large NMDA to AMPA ratio, however, during nicotine withdrawal, stimulation of active synapses occur in part due to the insertion of AMPA receptors on the presynaptic glutamate neurons. This alteration leads to the propagation of inhibitory signals through the reward pathway, ultimately driving drug craving behaviors [148].

#### 3.6. Habenula (Hb)

Although most drugs of abuse studies focus on reward pathways, aversive pathways play an important role in addiction as well. The habenula consists of a medial (MHb) and lateral (LHb) portion that has been known to be implemented during nicotine aversion. The most widely studied aversive pathway is the habenulo-IPN tract, which runs from the MHb to the interpeduncular nucleus. The excitatory glutamatergic and cholinergic projections of the MHb activate the GABAergic neurons

of the IPN, which then inhibits the DA neurons of the VTA [149]. This, in turn, leads to decreased activation of the mesolimbic pathway and thus, reduced reward.

Although many studies have shown the rewarding and reinforcing aspects of nicotine that drive the drug-seeking and taking behaviors, nicotine is known to have an inverted U-shaped dose response curve in rodents as well as humans [64]. Due to this dose-dependent effect, nicotine tends to be aversive at high concentrations. Humans have been shown to titrate the amount of nicotine they consume when smoking or vaping because of the aversive properties of too much nicotine. The process of titration occurs through  $\alpha 3/\alpha 5/\beta 4^*$  nAChRs, which are only activated in the presence of high nicotine concentrations (2 mg/kg in a rodent) [47,106,107]. These nAChR subunits are highly populated in the habenulo-IPN pathway that mediates the aversive properties of nicotine. In the absence of these subunits, individuals no longer titrate their nicotine consumption and lose the mediatory effects of this pathway on the negative effects of high doses of nicotine. These subunits are also important in the manifestation of withdrawal, including the somatic and physical symptoms. Knockout mice lacking these subunits exhibited fewer somatic withdrawal symptoms [150,151] in chronic nicotine administered mice.

Further, the less studied but still relevant aversive LHb pathway is a collection of glutamatergic and cholinergic excitatory neurons that project to the tail of the VTA, the rostromedial tegmental nucleus (RMTg). The GABAergic neurons of the RMTg then inhibit DA cell firing of the VTA [152]. This pathway is acted on by aversive stimuli, such as the aversive signals associated with nicotine withdrawal and the absence of an expected reward, in an attempt to suppress the mesocorticolimbic DA system [153]. Activation of the Hb pathways is important in the development of addiction, for their influence on drug-seeking and taking behaviors.

## 4. Flavoring Chemicals in Nicotine Addiction

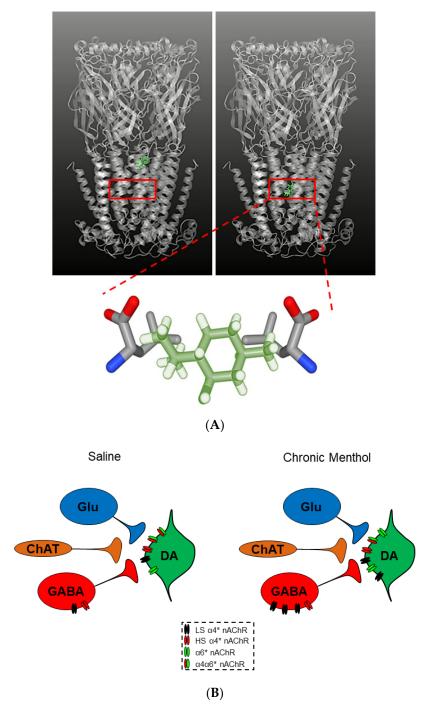
In the late 1900s, cigarette smoking had little to no restrictions and was acceptable in restaurants, hospitals, public transportation, and more, making them not only freely accessible but a new trend among the United States, with 50% of young women and 60–70% of young men being long-term smokers [154]. This popularity was in part attributed to flavoring additives. Mentholated cigarettes became a huge success, especially among the adolescent population after the discovery that menthol provided a cooling sensation in the cigarette smoke. Adolescent cigarette use continued to rise over the years due to the sweet aroma and taste that "characterizing" flavors provided. It was not until 2009 that the Family Smoking Prevention and Tobacco Control Act (FSPTCA) gave the FDA the authority to ban "characterizing" flavor additives such as strawberry and vanilla (not including tobacco or menthol) in combustible cigarettes in an attempt to mitigate adolescent cigarette use [155].

Since the FSPTCA, menthol has been the most widely studied flavor additive, for its popularity and sole acceptance as a flavorant in combustible cigarettes. Aside from mentholated cigarette popularity among adolescents, menthol has also long been popular among African Americans, with more than 80% of non-Hispanic black adults using menthol cigarettes [156]. It was recently determined that African Americans exhibit unique variants that promote a significant increase in the odds of menthol cigarette smoking [157]. Now that ENDS products are commonly used among those that were life-long smokers, menthol is still a very prominent flavor in e-liquids. Menthol is known to intensify various smoking-related behaviors and facilitates first-time nicotine use by masking the aversive properties of nicotine [158], which may explain the popularity of flavored ENDS and signifies the concerns with adolescent ENDS use [24,159].

Initially, menthol was considered to be an inert flavor additive, with the focus of tobacco-related research being on nicotine dependence. It was not until the early 2000s that studies on menthol began to surface. These initial studies suggested menthol smokers to be more nicotine dependent based on (1) how quickly they smoke upon waking in the morning compared to non-menthol smokers [160,161] and (2) the higher nicotine/cotinine levels among menthol smokers versus non-menthol smokers [161,162]. Further, Ahijevych and Garrett identified menthol to be a conditioned stimulus that enhances the

rewarding and reinforcing properties of nicotine through its positive sensory effects [162], resulting in drug-craving and drug-taking behaviors. These behaviors were mimicked in female rats that earned more intravenous nicotine infusions in the presence of an oral menthol cue during self-administration assays [16]. Conditioned stimuli paired with nicotine enhance the acquisition and maintenance of nicotine use and are known to be a driving force in nicotine dependence [163]. Menthol has the unique minty taste profile and a cooling sensory effect that, when combined with nicotine can contribute to craving and relapse when a person is going through an abstinent period [164]. These sensory cues are common among other sweet oral flavorants as well, including sucrose and saccharin. Interestingly, Wickham et al. utilized an intraoral delivery method that resulted in sucrose- and saccharin-induced phasic DA release, and significantly increased nicotine self-administration behavior [165]. It is these conditioned sensory cues that often make cessation so difficult.

Based on these initial sensory effects, menthol was studied for its pharmacological impact in the brain. Menthol was determined to act as a negative allosteric modulator (NAM) of nAChRs following the observation of reduced nicotine-induced (but not ACh-induced) inward currents through α4β2\* nAChRs expressed in human embryonic kidney (HEK) cells [166]. This finding was supported by additional electrophysiological characterization utilizing single-channel recordings that revealed menthol shifted  $\alpha 4\beta 2$  nAChRs towards the desensitized conformation state. In separate investigations, menthol was also determined to act as a noncompetitive antagonist on both  $\alpha 7$  and α3β4\* nAChRs [167,168]. A combination of computational docking, site-directed mutagenesis, and electrophysiology revealed that menthol may exert its effect on nAChRs by binding to the 9' position in the transmembrane 2 (M2) helix of nAChRs [22] (Figure 4A). Here, a series of mutations at the 9' leucine residues (L9') of the high-sensitivity α4β2 nAChRs were made and characterized with electrophysiology to investigate the connection between menthol's pharmacology and the L9' site [22]. It was concluded that menthol's inhibitory actions rely on the presence of the L9' site of the M2 helix and only one menthol molecule is sufficient for  $\alpha 4\beta 2$  nAChR inhibition. Another recent study identified additional menthol binding sites, including the M3-M4 extracellular interface on  $\alpha$ 4 subunits, various sites at the M1–M4 interface on β2 subunits, and M2 residues positioned extracellularly [169]. However, these binding sites have not been validated using functional assays. Here, it is important to distinguish the fact that the concentrations of menthol used to examine its inhibitory activity on nAChRs is several orders of magnitude higher (>30 µM) than what is considered smoking/vaping-relevant ( $<2 \mu M$ ). The studies that have used low concentrations of menthol ( $0.5 \mu M$ ) have determined that menthol combined with nicotine does enhance nAChR upregulation on VTA dopamine neurons and also enhances the excitability of these same neurons (discussed further below) [18,22]. A previously published review has extensively disseminated the recent investigations that have elucidated the mechanism of menthol's actions on nicotine reward and reinforcement [158]. To avoid repeating what has been eloquently explained and summarized previously, we direct interest into the specifics to the review by Wickham et al. [158].



**Figure 4.** Menthol binding patterns and mechanisms of action. **(A)** Top panel: Menthol molecule (in green) binding in the transmembrane 2 (M2) helix lining the central pore of the nAChR. Bottom panel: A closer look at menthol binding within the 9' leucine residues of the M2 helix of the nAChR. **(B)** Compared to saline, menthol induces upregulation of low-sensitivity (LS)  $\alpha 4^*$  nAChRs on VTA GABA neurons (in red) and  $\alpha 4^*$  (LS) and  $\alpha 6^*$  nAChRs on VTA dopamine neurons (in green). This results in reduced neuronal firing based on the desensitization properties of the nAChRs, with a net excitatory effect of VTA dopamine release. \* = nAChR may contain other subunits.

Much like nicotine's prominent ability to promote nAChR upregulation, menthol was determined to induce nAChR upregulation in the brainstem, cerebellum, corpus callosum, PFC [21], hippocampus, striatum [170], and VTA [18,20]. More specifically, Henderson and colleagues observed increased levels

of  $\alpha 4$  and  $\alpha 6$  nAChR subunit expression in VTA neurons following 10-day chronic menthol exposure [20], as well as increased levels of  $\alpha 4$  and  $\alpha 4\alpha 6^*$  nAChRs in VTA neurons following 10-day chronic menthol + nicotine exposure [18] through osmotic minipumps and daily intraperitoneal injections in mice. It is important to once again distinguish the concentrations used in these studies [18,20]. However, unlike nicotine, chronic menthol treatment caused a shift from  $\alpha 4_{(2)}\beta 2_{(3)}$  (high-sensitivity) to  $\alpha 4_{(3)}\beta 2_{(2)}$  (low-sensitivity) nAChRs in cultured cells (Figure 4B) [20]. This stoichiometry shift was accompanied by reduced VTA DA neuron firing frequency and an attenuation of reward-related behavior, dissimilar to nicotine. Interestingly, menthol + nicotine not only resulted in  $\alpha 4\alpha 6^*$  nAChR upregulation, but also enhanced VTA DA neuron excitability, and enhanced nicotine reward-related behavior [18].

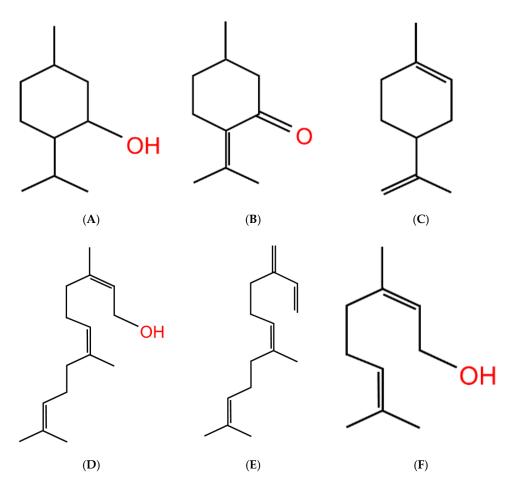
These neurobiological and neurophysiological alterations via menthol contribute to the behavioral effects demonstrated in rodent studies. As stated previously, menthol lacks the rewarding properties that nicotine presents, but menthol has the ability to enhance nicotine reward-related behavior with rodents in a conditioned place preference assay [18]. Further, to determine menthol's effect on smoking initiation, self-administration paradigms have been used to determine menthol's role in nicotine acquisition. In numerous studies, menthol facilitated intravenous nicotine self-administration and increased the rate of nicotine intake compared to control groups [16,17]. This behavior was followed by enhanced withdrawal-like behaviors, demonstrated through somatic signs and anxiety-related assays [170], and enhanced menthol-induced reinstatement [16]. Menthol presents a minty and cooling sensation through a transient receptor potential M8 (TRPM8)-mediated mechanism, which in turn, masks the aversive and harsh taste of nicotine by shifting nicotine's inverted U-shaped dose-response curve to the left and contributing to the enhanced nicotine acquisition behavior. Aside from menthol's ability to enhance smoking initiation, chronic menthol enhances the reinforcing properties of nicotine, leading to enhanced nicotine self-administration and dependence over time [17]. This may partially be due to menthol's effect on nicotine metabolism [171]. More specifically, menthol smokers exhibit reduced nicotine metabolism compared to non-menthol smokers [170,172], likely due to a competitive effect of menthol on the availability of CYP2A6, the major cytochrome enzyme involved in metabolizing nicotine [171].

With the common knowledge that smokers of menthol cigarettes exhibit lower cessation rates [173] and the growing use of flavored ENDS products (see Table 2), studies are beginning to arise on the role of other flavor additives. Recently these studies have included the popular green apple ENDS flavorants, farnesol [26] and farnesene [27], for they are structurally similar to menthol in the terpene class (see Figure 5). According to Espino-Diaz et al., apple biochemistry varies during the maturation process, with aldehydes primarily making up the apple flavor profile during the beginning, and following maturity, the flavor profiles transition to primarily alcohols and esters [25]. Farnesol, like menthol, is a terpene alcohol, which makes them more water soluble than a typical hydrocarbon, given their hydroxyl functional group. Both farnesene and farnesol differ from menthol given they are acyclical sesquiterpenes (natural 15-carbon organic compounds consisting of three isoprene units). Sesquiterpenes, alongside monoterpenes, are the main aroma components that make up apple flavorants. Additional to these two flavorants, green apple flavor includes numerous other flavorant compounds including hexyl acetate, ethyl acetate, and methylbutyl acetate [174]. However, ENDS-related studies on these flavorants have not yet been performed.

Flavor Chemical	Chemical Class	Flavor Profile	
Vanillin	Aldehyde	Vanilla, Chocolate, Cotton Candy, Mint, Coffee, Tobacco	
Ethyl Vanillin	Aldehyde	Vanilla, Chocolate, Cotton Candy, Coffee, Tobacco	
Ethyl Maltol	Alcohol	Vanilla, Chocolate, Cotton Candy, Mint, Coffee, Tobacco, Grape, Cherry	
Maltol	Alcohol	Vanilla, Chocolate, Mint, Coffee, Tobacco, Grape	
Benzaldehyde	Aldehyde	Cherry, Bubble Gum	
Benzyl Alcohol	Alcohol	Cherry, Vanilla, Coffee, Tobacco	
Ethyl Butyrate	Ester	Vanilla, Cherry, Bubble Gum, Apple, Tobacco, Grape	
Menthol	Alcohol	Mint	
Hexyl Acetate	Ester	Apple	
Ethyl Acetate	Ester	Bubble Gum, Apple, Grape, Tobacco	
Methylbutyl Acetate	Ester	Bubble Gum, Apple	
Farnesol	Sesquiterpene	Apple	
Farnesene	Sesquiterpene	Apple	

**Table 2.** Popular flavoring chemicals in e-liquids.

Comprehensive list of popular flavoring chemicals including their chemical class association and various flavor profiles that varying concentrations of these flavoring chemicals can be found in.



**Figure 5.** Flavorant chemical profiles. Similarity in chemical structure formation of **(A)** menthol, **(B)** pulegone (peppermint), **(C)** limonene (citrus), **(D)** farnesol, **(E)** farnesene, and **(F)** geraniol (fruity), despite the wide array of flavor profiles.

Similar to menthol, both farnesol and farnesene exhibited a significant enhancement of nicotine reward-related behavior in mice [26,27]. In the case of farnesol, there was an observed sex-dependent effect as only male mice (at the doses tested) exhibited changes in reward-related behavior. Interestingly, unlike menthol, both green apple flavorants exhibited significant reward-related behavior in the absence of nicotine compared to saline-control. The finding that ENDS flavors may produce reward-related

behavior on their own is a significant contribution to the field of nicotine addiction and may explain the continued use by adolescents and their strong preference for fruity flavors. This also reveals some insight into why there may be a rise in vaping of zero-nicotine flavored e-liquids.

Much like the previous menthol findings, Avelar et al., identified farnesol-induced upregulation of  $\alpha6^*$  and  $\alpha4\alpha6^*$  nAChRs on VTA DA neurons and a complementary increase in the firing frequency of these neurons [26]. Here, it was reported that farnesene, by itself, produced a greater change in nAChR upregulation and enhancement in firing frequency when compared to farnesol plus nicotine. They speculated that the differences in the flavorant-induced reward is likely due to the fact that menthol-alone only upregulates low-sensitivity  $\alpha6^*$  (not  $\alpha4\alpha6^*$ ) nAChRs and decreases VTA DA neuron firing [20], whereas farnesol not only increased VTA DA neuron firing but also upregulated both  $\alpha6^*$  and  $\alpha4\alpha6^*$  nAChRs [26]. Desensitization of the  $\alpha4\beta2^*$  nAChRs on SNr GABA neurons reduces the inhibitory outputs, and results in disinhibiting the VTA DA neurons [175]. Based on these previous findings, farnesol treatment was observed on SNr GABA neurons for their potential downstream effects on VTA DA neurons [26]. Here, in male mice only, they found a significant downregulation of  $\alpha4$  subunits on SNr GABA neurons and this contributed to a decrease in the firing frequency of these neurons.

Similar to the farnesol study, Cooper et al. performed conditioned place preference assays with mice and observed significant farnesene-induced reward-related behavior [27]. Unlike farnesol, farnesene exerted its effects through changes in nAChR stoichiometry, not nAChR upregulation. Through the use of mouse brain slices and cultured neuroblastoma 2a cells, it was determined that farnesene triggered a stoichiometric shift toward high-sensitivity  $\alpha4\beta2$  nAChRs on VTA DA neurons. This was accompanied by a leftward shift in the concentration-response of nicotine-induced inward currents on VTA DA neurons in a brain slice preparation. Finally, VTA DA neurons in farnesene-treated mice exhibited an increase in excitatory postsynaptic current frequency and amplitude. Considering that this finding was the result of a coronal brain-slice preparation, this suggests farnesene alters local VTA GABAergic or glutamatergic transmission and results in a net increase of excitatory input on to VTA DA neurons. The above flavorant studies demonstrate the potential risk for the abuse potential of ENDS flavorants and highlight the need to understand zero-nicotine flavored products.

# 5. Future Directions

Although flavorant studies are becoming more prevalent, there is still much to be learned. Currently, menthol and green apple are the only flavors studied for their effects on nicotine addiction-related behaviors. Given the distinct differences between menthol and green apple, and more specifically between green apple flavorants: farnesol and farnesene, it is safe to assume that the various flavor profiles on the market will have differing effects in future studies as well. This is especially true given that prominent chemical flavorants used in ENDS e-liquids have similar chemical scaffolds as menthol, farnesene, and farnesol (see Figure 5). It is likely these effects will also differ among specific classes of flavor chemicals being studied. For example, majority of the flavorants used in these devices are alcohols or aldehydes, and some flavorants are present in numerous flavor subcategories outside of their major flavor category (i.e., vanillin (vanilla) flavorant in chocolate, mint, and cotton candy flavored products; [174]). It will be interesting to determine how heavily used flavorants (like vanillin) differ from farnesol or farnesene, which are primarily found in apple flavored products. As well as general differences in chemical classes.

Another important gap in this field of research is the gap in knowledge regarding the difference between adult, adolescent, and in utero exposure. This is a critical gap to fill considering the constant rise in adolescent ENDS use. Despite not knowing the age-dependent effects on flavored ENDS use, it is common knowledge that adolescent nicotine use has a variety of detrimental impacts on the developing brain including greater tolerance for high doses of nicotine [176], enhanced nicotine reward sensitivity [177], reduced PFC activation and thus impaired cognitive function [178,179], and enhanced risk for other drug abuse later in life through a process known as "priming" [146,180]. With the current

flavor studies resulting in an enhancing effect on nicotine pharmacology, it is likely flavors may also enhance the above neurological impacts as well. Yet, more information must be gathered.

Currently, menthol and green apple have been studied for their effects on midbrain dopamine and GABA neurons, however glutamatergic and cholinergic afferents are important contributors to the activity of the mesocorticolimbic pathway. It would be interesting to determine the role that flavors may play in glutamate firing, including synaptic plasticity. It is also critical to utilize fast-scan cyclic voltammetry in the determining of flavorant-induced DA release in the NAc. Although we have observed flavorant-induced alterations in firing on presynaptic midbrain neurons, it is important to determine the postsynaptic effects as well.

With crucial evidence pointing to the aversive pathways as a mediator for nicotine addiction, it is also important to determine if flavors also elicit effects in these specific regions. Flavorants may be acting on these pathways by reducing the inhibitory transmission from the RMTg and IPN to the VTA, resulting in net excitation and dopamine release. Furthermore, the habenula consists of more diverse nAChR subtypes than the midbrain. Flavorants may have a more (or less) pronounced effect on these subtypes. Regardless, there is plenty of research that still needs to be done in order to fully understand how these devices and their additives affect a chronic user.

Lastly, although combustible cigarette plus electronic cigarette use (termed 'dual-use') has been explored. These studies have focused more on lung-related and cardiovascular-related impacts, as opposed to the impacts on the brain. It will be interesting to identify the effect that dual use has on a chronic user's brain and whether ENDS use exacerbates the already-addicted brain, or merely maintains the combustible cigarette-induced neuroalterations.

#### 6. Conclusions

Here, we discussed the major flavorant-induced changes in neurobiology and neurophysiology that include changes in nAChR function, nAChR subtype assembly, and nAChR upregulation. Together, these changes cause an impact on reward- and reinforcement-related behavior. Thus far, basic science investigations into smoking- and vaping-related behavior have only focused on menthol and green apple flavorants. These studies suggest that there are complex effects that ENDS flavorants exert on nicotine reward and reinforcement. Based on these findings, and the current knowledge regarding nicotine, it is important to continue investigating ENDS flavorants to determine their impact on the neurobiology and neurochemistry of addiction. In parallel, there is a need to understand the toxicology of these ENDS flavorants as well.

**Author Contributions:** Conceptualization, S.Y.C. and B.J.H.; writing—original draft preparation, S.Y.C.; writing—review and editing, S.Y.C. and B.J.H.; funding acquisition, S.Y.C. and B.J.H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was supported by the National Institutes on Drug Abuse (NIDA) at the National Institutes of Health Grant DA040047 (to B.J.H.). Research reported in this publication was supported by NIDA and Food and Drug Administration Center for Tobacco Products Grant DA046335 (to B.J.H.). Funding was also provided by the PhRMA Foundation (Predoctoral Fellowship in Pharmacology/Toxicology to S.Y.C.) and startup funds to B.J.H. by the Marshall University Research Corporation.

Conflicts of Interest: The authors declare no conflict of interest.

# References

- SGR. Reports of the Surgeon General. In The Health Consequences of Smoking-50 Years of Progress: A Report of the Surgeon General; Centers for Disease Control and Prevention (US): Atlanta, GA, USA, 2014.
- 2. Babb, S.; Malarcher, A.; Schauer, G.; Asman, K.; Jamal, A. Quitting Smoking among Adults—United States, 2000–2015. *Mmwr. Morb. Mortal. Wkly. Rep.* 2017, 65, 1457–1464. [CrossRef] [PubMed]
- 3. Office on Smoking and Health. Publications and Reports of the Surgeon General. In *Women and Smoking: A Report of the Surgeon General;* Centers for Disease Control and Prevention (US): Atlanta, GA, USA, 2001. Available online: https://www.hhs.gov/surgeongeneral/reports-and-publications/tobacco/index.html (accessed on 12 May 2020).

Molecules **2020**, 25, 4223 20 of 28

 FTC Cigarette Report for 2018. Available online: https://www.ftc.gov/system/files/documents/reports/federaltrade-commission-cigarette-report-2018-smokeless-tobacco-report-2018/p114508cigarettereport2018.pdf (accessed on 27 March 2020).

- 5. Ai, J.; Taylor, K.M.; Lisko, J.G.; Tran, H.; Watson, C.H.; Holman, M.R. Menthol Content in US Marketed Cigarettes. *Nicotine Tob. Res.* **2015**, *18*, 1575–1580. [CrossRef] [PubMed]
- 6. Glasser, A.M.; Collins, L.; Pearson, J.L.; Abudayyeh, H.; Niaura, R.S.; Abrams, D.B.; Villanti, A.C.; Katz, L. Overview of Electronic Nicotine Delivery Systems: A Systematic Review. *Am. J. Prev. Med.* **2017**, 52, e33–e66. [CrossRef]
- 7. SGR. Reports of the Surgeon General. In *Facing Addiction in America: The Surgeon General's Report on Alcohol, Drugs, and Health;* US Department of Health and Human Services: Washington, DC, USA, 2016.
- 8. SGR. Publications and Reports of the Surgeon General. In *E-Cigarette Use Among Youth and Young Adults: A Report of the Surgeon General;* Centers for Disease Control and Prevention (US): Atlanta, GA, USA, 2016.
- 9. Omaiye, E.E.; McWhirter, K.J.; Luo, W.; Tierney, P.A.; Pankow, J.F.; Talbot, P. High concentrations of flavor chemicals are present in electronic cigarette refill fluids. *Sci. Rep.* **2019**, *9*, 2468. [CrossRef] [PubMed]
- 10. Office on Smoking and Health. 2019 National Youth Tobacco Survey: Methodology Report; Department of Health and Human Services, Centers for Disease Control and Prevention National Center for Chronic Disease Prevention and Health Promotion, Office on Smoking and Health: Atlanta, GA, USA, 2019. Available online: <a href="https://www.cdc.gov/tobacco/data\_statistics/surveys/nyts/index.htm">https://www.cdc.gov/tobacco/data\_statistics/surveys/nyts/index.htm</a> (accessed on 8 September 2020).
- 11. Cullen, K.A.; Gentzke, A.S.; Sawdey, M.D.; Chang, J.T.; Anic, G.M.; Wang, T.W.; Creamer, M.R.; Jamal, A.; Ambrose, B.K.; King, B.A. E-cigarette Use Among Youth in the United States, 2019. *JAMA* 2019, 322, 2095. [CrossRef]
- 12. Mead, E.; Duffy, V.B.; Oncken, C.; Litt, M.D. E-cigarette palatability in smokers as a function of flavorings, nicotine content and propylthiouracil (PROP) taster phenotype. *Addict. Behav.* **2019**, *91*, 37–44. [CrossRef] [PubMed]
- 13. Willis, D.N.; Liu, B.; Ha, M.A.; Jordt, S.-E.; Morris, J.B. Menthol attenuates respiratory irritation responses to multiple cigarette smoke irritants. *FASEB J.* **2011**, *25*, 4434–4444. [CrossRef]
- 14. Zhu, S.-H.; Sun, J.Y.; Bonnevie, E.E.; Cummins, S.; Gamst, A.; Yin, L.; Lee, M. Four hundred and sixty brands of e-cigarettes and counting: Implications for product regulation. *Tob. Control.* **2014**, 23, iii3–iii9. [CrossRef]
- 15. Hsu, G.; Sun, J.Y.; Zhu, S.-H.; Allem, J.-P.; Chen, J.C.; Gundel, L. Evolution of Electronic Cigarette Brands From 2013–2014 to 2016–2017: Analysis of Brand Websites. *J. Med Internet Res.* **2018**, *20*, e80. [CrossRef]
- 16. Wang, T.; Wang, B.; Chen, H. Menthol facilitates the intravenous self-administration of nicotine in rats. *Front. Behav. Neurosci.* **2014**, *8*, 437. [CrossRef]
- 17. Biswas, L.; Harrison, E.; Gong, Y.; Avusula, R.; Lee, J.; Zhang, M.; Rousselle, T.; Lage, J.; Liu, X. Enhancing effect of menthol on nicotine self-administration in rats. *Psychopharmacology* **2016**, 233, 3417–3427. [CrossRef] [PubMed]
- Henderson, B.J.; Wall, T.R.; Henley, B.M.; Kim, C.H.; McKinney, S.; Lester, H.A. Menthol Enhances Nicotine Reward-Related Behavior by Potentiating Nicotine-Induced Changes in nAChR Function, nAChR Upregulation, and DA Neuron Excitability. *Neuropsychopharmacology* 2017, 42, 2285–2291. [CrossRef] [PubMed]
- 19. Cooper, S.Y.; Akers, A.T.; Henderson, B.J. Flavors enhance nicotine vapor self-administration in male mice. *Nicotine Tob. Res.* **2020**. [CrossRef]
- 20. Henderson, B.J.; Wall, T.R.; Henley, B.M.; Kim, C.H.; Nichols, W.A.; Moaddel, R.; Xiao, C.; Lester, H.A. Menthol Alone Upregulates Midbrain nAChRs, Alters nAChR Subtype Stoichiometry, Alters Dopamine Neuron Firing Frequency, and Prevents Nicotine Reward. *J. Neurosci.* 2016, *36*, 2957–2974. [CrossRef]
- 21. Brody, A.L.; Mukhin, A.G.; La Charite, J.; Ta, K.; Farahi, J.; Sugar, C.A.; Mamoun, M.S.; Vellios, E.; Archie, M.; Kozman, M.; et al. Up-regulation of nicotinic acetylcholine receptors in menthol cigarette smokers. *Int. J. Neuropsychopharmacol.* **2013**, *16*, 957–966. [CrossRef] [PubMed]
- 22. Henderson, B.J.; Grant, S.; Chu, B.W.; Shahoei, R.; Huard, S.M.; Saladi, S.S.M.; Tajkhorshid, E.; Dougherty, D.A.; Lester, H.A. Menthol stereoisomers exhibit different effects on a4b2 nAChR upregulation and dopamine neuron spontaneous firing. *eNeuro* **2018**. [CrossRef]
- 23. Zhang, M.; Harrison, E.; Biswas, L.; Tran, T.; Liu, X. Menthol facilitates dopamine-releasing effect of nicotine in rat nucleus accumbens. *Pharmacol. Biochem. Behav.* **2018**, 175, 47–52. [CrossRef] [PubMed]

Molecules **2020**, 25, 4223 21 of 28

24. Fan, L.; Balakrishna, S.; Jabba, S.V.; Bonner, P.E.; Taylor, S.R.; Picciotto, M.R.; Jordt, S.-E. Menthol decreases oral nicotine aversion in C57BL/6 mice through a TRPM8-dependent mechanism. *Tob. Control.* **2016**, 25, ii50–ii54. [CrossRef]

- 25. Espino-Díaz, M.; Sepúlveda, D.R.; González-Aguilar, G.; Olivas, G.I. Biochemistry of Apple Aroma: A Review. *Food Technol. Biotechnol.* **2016**, *54*, 375–397. [CrossRef]
- 26. Avelar, A.J.; Akers, A.T.; Baumgard, Z.J.; Cooper, S.Y.; Casinelli, G.P.; Henderson, B.J. Why flavored vape products may be attractive: Green apple tobacco flavor elicits reward-related behavior, upregulates nAChRs on VTA dopamine neurons, and alters midbrain dopamine and GABA neuron function. *Neuropharmacology* **2019**, *158*, 107729. [CrossRef]
- 27. Cooper, S.Y.; Akers, A.T.; Henderson, B.J. Green apple e-cigarette flavorant farnesene triggers reward-related behavior by promoting high-sensitivity nAChRs in the ventral tegmental area. *Eneuro* **2020**. [CrossRef] [PubMed]
- 28. Sine, S.M.; Engel, A.G. Recent advances in Cys-loop receptor structure and function. *Nature* **2006**, 440, 448–455. [CrossRef]
- 29. Le Novère, N.; Changeux, J.-P. Molecular evolution of the nicotinic acetylcholine receptor: An example of multigene family in excitable cells. *J. Mol. Evol.* **1995**, *40*, 155–172. [CrossRef]
- 30. Albuquerque, E.X.; Pereira, E.F.R.; Alkondon, M.; Rogers, S.W. Mammalian Nicotinic Acetylcholine Receptors: From Structure to Function. *Physiol. Rev.* **2009**, *89*, 73–120. [CrossRef]
- 31. Dang, N.; Meng, X.; Song, H. Nicotinic acetylcholine receptors and cancer. *Biomed. Rep.* **2016**, *4*, 515–518. [CrossRef] [PubMed]
- 32. Zoli, M.; Pistillo, F.; Gotti, C. Diversity of native nicotinic receptor subtypes in mammalian brain. *Neuropharmacology* **2015**, *96*, 302–311. [CrossRef] [PubMed]
- 33. Penton, R.E.; Lester, R.A. Cellular events in nicotine addiction. *Semin. Cell Dev. Boil.* **2009**, 20, 418–431. [CrossRef]
- 34. Gregg, B.W.; Wells, G.B. Structural answers and persistent questions about how nicotinic receptors work. *Front. Biosci.* **2008**, *13*, 5479–5510. [CrossRef]
- 35. Henderson, B.J.; Srinivasan, R.; Nichols, W.A.; Dilworth, C.N.; Gutierrez, D.F.; Mackey, E.D.; McKinney, S.; Drenan, R.M.; Richards, C.I.; Lester, H.A. Nicotine exploits a COPI-mediated process for chaperone-mediated up-regulation of its receptors. *J. Gen. Physiol.* **2013**, *143*, 51–66. [CrossRef]
- Wiesner, A.; Führer, C. Regulation of nicotinic acetylcholine receptors by tyrosine kinases in the peripheral and central nervous system: Same players, different roles. Cell. Mol. Life Sci. 2006, 63, 2818–2828. [CrossRef]
- 37. Srinivasan, R.; Pantoja, R.; Moss, F.J.; Mackey, E.D.; Son, C.D.; Miwa, J.M.; Lester, H.A. Nicotine up-regulates  $\alpha 4\beta 2$  nicotinic receptors and ER exit sites via stoichiometry-dependent chaperoning. *J. Gen. Physiol.* **2010**, 137, 59–79. [CrossRef] [PubMed]
- 38. Gotti, C.; Clementi, F.; Fornari, A.; Gaimarri, A.; Guiducci, S.; Manfredi, I.; Moretti, M.; Pedrazzi, P.; Pucci, L.; Zoli, M. Structural and functional diversity of native brain neuronal nicotinic receptors. *Biochem. Pharmacol.* **2009**, *78*, 703–711. [CrossRef] [PubMed]
- 39. Tapia, L.; Kuryatov, A.; Lindström, J. Ca<sup>2+</sup> Permeability of the  $(\alpha 4)_3(\beta 2)_2$  Stoichiometry Greatly Exceeds That of  $(\alpha 4)_2(\beta 2)_3$  Human Acetylcholine Receptors. *Mol. Pharmacol.* **2006**, *71*, 769–776. [CrossRef]
- 40. Zhao, L.; Kuo, Y.-P.; George, A.A.; Peng, J.-H.; Purandare, M.S.; Schroeder, K.M.; Lukas, R.J.; Wu, J. Functional Properties of Homomeric, Human α7-Nicotinic Acetylcholine Receptors Heterologously Expressed in the SH-EP1 Human Epithelial Cell Line. *J. Pharmacol. Exp. Ther.* **2003**, *305*, 1132–1141. [CrossRef] [PubMed]
- 41. Conroy, W.G.; Ogden, L.F.; Berg, D.K. Cluster formation of α7-containing nicotinic receptors at interneuronal interfaces in cell culture. *Neuropharmacology* **2000**, *39*, 2699–2705. [CrossRef]
- 42. Zoli, M.; Pucci, S.; Vilella, A.; Gotti, C. Neuronal and Extraneuronal Nicotinic Acetylcholine Receptors. *Curr. Neuropharmacol.* **2018**, *16*, 338–349. [CrossRef]
- 43. McGehee, D.; Heath, M.; Gelber, S.; DeVay, P.; Role, L. Nicotine enhancement of fast excitatory synaptic transmission in CNS by presynaptic receptors. *Science* **1995**, *269*, 1692–1696. [CrossRef]
- 44. Jensen, A.A.; Froelund, B.; Liljefors, T.; Krogsgaard-Larsen, P. Neuronal Nicotinic Acetylcholine Receptors: Structural Revelations, Target Identifications, and Therapeutic Inspirations. *J. Med. Chem.* **2005**, *48*, 4705–4745. [CrossRef]

Molecules **2020**, 25, 4223 22 of 28

45. Picciotto, M.R.; Addy, N.A.; Mineur, Y.S.; Brunzell, D.H. It is not "either/or": Activation and desensitization of nicotinic acetylcholine receptors both contribute to behaviors related to nicotine addiction and mood. *Prog. Neurobiol.* **2008**, *84*, 329–342. [CrossRef]

- 46. Alkondon, M.; Albuquerque, E.X. Nicotinic acetylcholine receptor alpha7 and alpha4beta2 subtypes differentially control GABAergic input to CA1 neurons in rat hippocampus. *J. Neurophysiol.* **2001**, *86*, 3043–3055. [CrossRef]
- 47. Dani, J.A. Neuronal Nicotinic Acetylcholine Receptor Structure and Function and Response to Nicotine. *Int. Rev. Neurobiol.* **2015**, *124*, 3–19. [CrossRef] [PubMed]
- 48. Picciotto, M.R.; Kenny, P.J. Molecular Mechanisms Underlying Behaviors Related to Nicotine Addiction. *Cold Spring Harb. Perspect. Med.* **2012**, *3*, a012112. [CrossRef] [PubMed]
- 49. Molas, S.; Dierssen, M. The role of nicotinic receptors in shaping and functioning of the glutamatergic system: A window into cognitive pathology. *Neurosci. Biobehav. Rev.* **2014**, *46*, 315–325. [CrossRef] [PubMed]
- 50. Shivange, A.V.; Borden, P.M.; Muthusamy, A.K.; Nichols, A.L.; Bera, K.; Bao, H.; Bishara, I.; Jeon, J.; Mulcahy, M.J.; Cohen, B.; et al. Determining the pharmacokinetics of nicotinic drugs in the endoplasmic reticulum using biosensors. *J. Gen. Physiol.* **2019**, *151*, 738–757. [CrossRef] [PubMed]
- 51. Lester, H.A.; Xiao, C.; Srinivasan, R.; Son, C.D.; Miwa, J.M.; Pantoja, R.; Banghart, M.R.; Dougherty, D.A.; Goate, A.; Wang, J.C. Nicotine is a Selective Pharmacological Chaperone of Acetylcholine Receptor Number and Stoichiometry. Implications for Drug Discovery. *AAPS J.* 2009, *11*, 167–177. [CrossRef]
- 52. Kuryatov, A.; Luo, J.; Cooper, J.; Lindstrom, J. Nicotine acts as a pharmacological chaperone to up-regulate human α4β2 acetylcholine receptors. *Mol. Pharmacol.* **2005**, *68*, 1839–1851. [CrossRef]
- 53. Nashmi, R.; Xiao, C.; Deshpande, P.; McKinney, S.; Grady, S.R.; Whiteaker, P.; Huang, Q.; McClure-Begley, T.; Lindstrom, J.M.; Labarca, C.; et al. Chronic Nicotine Cell Specifically Upregulates Functional α4\* Nicotinic Receptors: Basis for Both Tolerance in Midbrain and Enhanced Long-Term Potentiation in Perforant Path. *J. Neurosci.* 2007, 27, 8202–8218. [CrossRef]
- 54. Sallette, J.; Pons, S.; Devillers-Thiéry, A.; Soudant, M.; De Carvalho, L.P.; Changeux, J.-P.; Corringer, P.J. Nicotine Upregulates Its Own Receptors through Enhanced Intracellular Maturation. *Neuron* **2005**, *46*, 595–607. [CrossRef]
- 55. Tapper, A.R.; McKinney, S.L.; Nashmi, R.; Schwarz, J.; Deshpande, P.; Labarca, C.; Whiteaker, P.; Marks, M.J.; Collins, A.C.; Lester, H.A. Nicotine Activation of α4\* Receptors: Sufficient for Reward, Tolerance, and Sensitization. *Science* **2004**, *306*, 1029–1032. [CrossRef]
- 56. Pons, S.; Fattore, L.; Cossu, G.; Tolu, S.; Porcu, E.; McIntosh, J.M.; Changeux, J.P.; Maskos, U.; Fratta, W. Crucial role of α4 and α6 nicotinic acetylcholine receptor subunits from ventral tegmental area in systemic nicotine self-administration. *J. Neurosci.* **2008**, *28*, 12318–12327. [CrossRef]
- 57. Dani, J.A.; Ji, D.; Zhou, F.-M. Synaptic plasticity and nicotine addiction. *Neuron* **2001**, *31*, 349–352. [CrossRef]
- 58. McKay, B.E.; Placzek, A.N.; Dani, J.A. Regulation of synaptic transmission and plasticity by neuronal nicotinic acetylcholine receptors. *Biochem. Pharmacol.* **2007**, *74*, 1120–1133. [CrossRef]
- 59. Mansvelder, H.D.; Mertz, M.; Role, L. Nicotinic modulation of synaptic transmission and plasticity in cortico-limbic circuits. *Semin. Cell Dev. Boil.* **2009**, *20*, 432–440. [CrossRef]
- 60. Mansvelder, H.D.; McGehee, D.S. Long-term potentiation of excitatory inputs to brain reward areas by nicotine. *Neuron* **2000**, *27*, 349–357. [CrossRef]
- 61. Xiao, C.; Nashmi, R.; McKinney, S.; Cai, H.; McIntosh, J.M.; Lester, H.A. Chronic nicotine selectively enhances α4β2\* nicotinic acetylcholine receptors in the nigrostriatal dopamine pathway. *J. Neurosci.* **2009**, 29, 12428–12439. [CrossRef]
- 62. Mansvelder, H.D.; Keath, J.R.; McGehee, D.S. Synaptic mechanisms underlie nicotine-induced excitability of brain reward areas. *Neuron* **2002**, *33*, 905–919. [CrossRef]
- 63. Yan, Y.; Peng, C.; Arvin, M.C.; Jin, X.-T.; Kim, V.; Ramsey, M.D.; Wang, Y.; Banala, S.; Wokosin, D.L.; McIntosh, J.M.; et al. Nicotinic Cholinergic Receptors in VTA Glutamate Neurons Modulate Excitatory Transmission. *Cell Rep.* **2018**, 23, 2236–2244. [CrossRef] [PubMed]
- 64. Corrigall, W.A.; Coen, K.M. Nicotine maintains robust self-administration in rats on a limited-access schedule. *Psychopharmacology* **1989**, *99*, 473–478. [CrossRef] [PubMed]
- 65. Miyazawa, A.; Fujiyoshia, Y.; Unwin, N. Structure and gating mechanism of the acetylcholine receptor pore. *Nature* **2003**, 423, 949–955. [CrossRef]

Molecules **2020**, 25, 4223 23 of 28

66. Unwin, N.; Miyazawa, A.; Li, J.; Fujiyoshia, Y. Activation of the Nicotinic Acetylcholine Receptor Involves a Switch in Conformation of the α Subunits. *J. Mol. Boil.* **2002**, *319*, 1165–1176. [CrossRef]

- 67. Picciotto, M.R.; Zoli, M.; Rimondini, R.; Lena, C.; Marubio, L.M.; Pich, E.M.; Fuxe, K.; Changeux, J.P. Acetylcholine receptors containing the beta2 subunit are involved in the reinforcing properties of nicotine. *Nature* **1998**, *391*, 173–177. [CrossRef]
- 68. Pidoplichko, V.I.; DeBiasi, M.; Williams, J.T.; Dani, J.A. Nicotine activates and desensitizes midbrain dopamine neurons. *Nature* **1997**, 390, 401–404. [CrossRef] [PubMed]
- 69. Engle, S.E.; Shih, P.Y.; McIntosh, J.M.; Drenan, R.M.  $\alpha 4\alpha 6\beta 2^*$  nicotinic acetylcholine receptor activation on ventral tegmental area dopamine neurons is sufficient to stimulate a depolarizing conductance and enhance surface AMPA receptor function. *Mol. Pharmacol.* **2013**, *84*, 393–406. [CrossRef] [PubMed]
- 70. Liu, L.; Zhao-Shea, R.; McIntosh, J.M.; Gardner, P.D.; Tapper, A.R. Nicotine persistently activates ventral tegmental area dopaminergic neurons via nicotinic acetylcholine receptors containing α4 and α6 subunits. *Mol. Pharmacol.* **2012**, *81*, 541–548. [CrossRef] [PubMed]
- 71. Mansvelder, H.D.; McGehee, D.S. Cellular and synaptic mechanisms of nicotine addiction. *J. Neurobiol.* **2002**, 53, 606–617. [CrossRef] [PubMed]
- 72. Johnson, S.W.A.; North, R. Two types of neurone in the rat ventral tegmental area and their synaptic inputs. *J. Physiol.* **1992**, 450, 455–468. [CrossRef] [PubMed]
- 73. Yan, Y.; Beckley, N.A.; Kim, V.; Drenan, R.M. Differential Nicotinic Modulation of Glutamatergic and GABAergic VTA Microcircuits. *Eneuro* **2019**, *6*. [CrossRef]
- 74. Brody, A.L.; Mandelkern, M.A.; London, E.D.; Olmstead, R.E.; Farahi, J.; Scheibal, D.; Jou, J.; Allen, V.; Tiongson, E.; Chefer, S.I.; et al. Cigarette smoking saturates brain α4β2 nicotinic acetylcholine receptors. *Arch Gen Psychiatry* 2006, 63, 907–915. [CrossRef] [PubMed]
- 75. Marks, M.; Pauly, J.; Gross, S.D.; Deneris, E.; Hermans-Borgmeyer, I.; Heinemann, S.; Collins, A. Nicotine binding and nicotinic receptor subunit RNA after chronic nicotine treatment. *J. Neurosci.* **1992**, *12*, 2765–2784. [CrossRef]
- 76. Henderson, B.J.; Lester, H.A. Inside-out neuropharmacology of nicotinic drugs. *Neuropharmacology* **2015**, *96*, 178–193. [CrossRef]
- 77. Hurst, R.; Rollema, H.; Bertrand, D. Nicotinic acetylcholine receptors: From basic science to therapeutics. *Pharmacol. Ther.* **2013**, 137, 22–54. [CrossRef]
- 78. Dani, J.A.; Heinemann, S. Molecular and Cellular Aspects of Nicotine Abuse. *Neuron* **1996**, *16*, 905–908. [CrossRef]
- 79. Mukhin, A.G.; Kimes, A.S.; Chefer, S.I.; Matochik, J.A.; Contoreggi, C.S.; Horti, A.G.; Vaupel, D.B.; Pavlova, O.; Stein, E.A. Greater nicotinic acetylcholine receptor density in smokers than in nonsmokers: A PET study with 2-18F-FA-85380. *J. Nucl. Med.* **2008**, *49*, 1628–1635. [CrossRef] [PubMed]
- 80. Srinivasan, R.; Richards, C.I.; Xiao, C.; Rhee, D.; Pantoja, R.; Dougherty, D.A.; Miwa, J.M.; Lester, H.A. Pharmacological chaperoning of nicotinic acetylcholine receptors reduces the endoplasmic reticulum stress response. *Mol. Pharmacol.* **2012**, *81*, 759–769. [CrossRef] [PubMed]
- 81. Omaiye, E.E.; McWhirter, K.J.; Luo, W.; Pankow, J.F.; Talbot, P. High-Nicotine Electronic Cigarette Products: Toxicity of JUUL Fluids and Aerosols Correlates Strongly with Nicotine and Some Flavor Chemical Concentrations. *Chem. Res. Toxicol.* **2019**, *32*, 1058–1069. [CrossRef] [PubMed]
- 82. Grieder, T.E.; Besson, M.; Maal-Bared, G.; Pons, S.; Maskos, U.; van der Kooy, D. beta2\* nAChRs on VTA dopamine and GABA neurons separately mediate nicotine aversion and reward. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 25968–25973. [CrossRef]
- 83. Picciotto, M.R.; Caldarone, B.J.; Brunzell, D.H.; Zachariou, V.; Stevens, T.R.; King, S.L. Neuronal nicotinic acetylcholine receptor subunit knockout mice: Physiological and behavioral phenotypes and possible clinical implications. *Pharmacol. Ther.* **2001**, *92*, 89–108. [CrossRef]
- 84. Watkins, S.S.; Epping-Jordan, M.P.; Koob, G.F.; Markou, A. Blockade of Nicotine Self-Administration with Nicotinic Antagonists in Rats. *Pharmacol. Biochem. Behav.* **1999**, *62*, 743–751. [CrossRef]
- 85. Corrigall, W.A.; Coen, K.M.; Adamson, K.L. Self-administered nicotine activates the mesolimbic dopamine system through the ventral tegmental area. *Brain Res.* **1994**, *653*, 278–284. [CrossRef]
- 86. Nelson, M.E.; Kuryatov, A.; Choi, C.H.; Zhou, Y.; Lindstrom, J. Alternate stoichiometries of α4β2 nicotinic acetylcholine receptors. *Mol. Pharmacol.* **2003**, *63*, 332–341. [CrossRef]

Molecules **2020**, 25, 4223 24 of 28

87. Kuryatov, A.; Onksen, J.; Lindstrom, J. Roles of Accessory Subunits in α4β2\* Nicotinic Receptors. *Mol. Pharmacol.* **2008**, 74, 132–143. [CrossRef]

- 88. Schwartz, R.; Kellar, K. Nicotinic cholinergic receptor binding sites in the brain: Regulation In Vivo. *Science* **1983**, 220, 214–216. [CrossRef]
- 89. Liu, M.; Jiang, Y.; Wedow, R.; Li, Y.; Brazel, D.M.; Chen, F.; Datta, G.; Davila-Velderrain, J.; McGuire, D.; Tian, C.; et al. Association studies of up to 1.2 million individuals yield new insights into the genetic etiology of tobacco and alcohol use. *Nat. Genet.* **2019**, *51*, 237–244. [CrossRef] [PubMed]
- 90. Champtiaux, N.; Han, Z.-Y.; Bessis, A.; Rossi, F.M.; Zoli, M.; Marubio, L.; McIntosh, J.M.; Changeux, J.-P. Distribution and Pharmacology of α6-Containing Nicotinic Acetylcholine Receptors Analyzed with Mutant Mice. *J. Neurosci.* **2002**, 22, 1208–1217. [CrossRef] [PubMed]
- 91. Champtiaux, N.; Gotti, C.; Cordero-Erausquin, M.; David, D.J.; Przybylski, C.; Léna, C.; Clementi, F.; Moretti, M.; Rossi, F.M.; Le Novère, N.; et al. Subunit Composition of Functional Nicotinic Receptors in Dopaminergic Neurons Investigated with Knock-Out Mice. *J. Neurosci.* 2003, 23, 7820–7829. [CrossRef] [PubMed]
- 92. Salminen, O.; Murphy, K.L.; McIntosh, J.M.; Drago, J.; Marks, M.J.; Collins, A.C.; Grady, S.R. Subunit Composition and Pharmacology of Two Classes of Striatal Presynaptic Nicotinic Acetylcholine Receptors Mediating Dopamine Release in Mice. *Mol. Pharmacol.* 2004, 65, 1526–1535. [CrossRef]
- 93. Wonnacott, S.; Sidhpura, N.; Balfour, D.J. Nicotine: From molecular mechanisms to behaviour. *Curr. Opin. Pharmacol.* **2005**, *5*, 53–59. [CrossRef] [PubMed]
- 94. Mackey, E.D.; Engle, S.E.; Kim, M.R.; O'Neill, H.C.; Wageman, C.R.; Patzlaff, N.E.; Wang, Y.; Grady, S.R.; McIntosh, J.M.; Marks, M.J.; et al. α6\* nicotinic acetylcholine receptor expression and function in a visual salience circuit. *J. Neurosci.* **2012**, 32, 10226–10237. [CrossRef] [PubMed]
- 95. Walters, C.L.; Brown, S.; Changeux, J.-P.; Martin, B.; Damaj, M.I. The β2 but not α7 subunit of the nicotinic acetylcholine receptor is required for nicotine-conditioned place preference in mice. *Psychopharmacology* **2006**, *184*, 339–344. [CrossRef]
- 96. Xiao, C.; Srinivasan, R.; Drenan, R.M.; Mackey, E.D.W.; McIntosh, J.M.; Lester, H.A. Characterizing functional α6β2 nicotinic acetylcholine receptors in vitro: Mutant β2 subunits improve membrane expression, and fluorescent proteins reveal responsive cells. *Biochem. Pharmacol.* **2011**, *82*, 852–861. [CrossRef]
- 97. Wang, J.; Kuryatov, A.; Sriram, A.; Jin, Z.; Kamenecka, T.M.; Kenny, P.J.; Lindstrom, J. An accessory agonist binding site promotes activation of α4β2\* nicotinic acetylcholine receptors. *J. Biol. Chem.* **2015**, 290, 13907–13918. [CrossRef] [PubMed]
- 98. Alkondon, M.; Albuquerque, E.X. Initial Characterization of the Nicotinic Acetylcholine Receptors in Rat Hippocampal Neurons. *J. Recept. Res.* **1991**, *11*, 1001–1021. [CrossRef] [PubMed]
- 99. Swanson, L.W. The projections of the ventral tegmental area and adjacent regions: A combined fluorescent retrograde tracer and immunofluorescence study in the rat. *Brain Res. Bull.* **1982**, *9*, 321–353. [CrossRef]
- 100. Lüscher, C.; Malenka, R.C. Drug-Evoked Synaptic Plasticity in Addiction: From Molecular Changes to Circuit Remodeling. *Neuron* **2011**, *69*, *650–663*. [CrossRef]
- 101. Van Huijstee, A.N.; Mansvelder, H.D. Glutamatergic synaptic plasticity in the mesocorticolimbic system in addiction. *Front. Cell. Neurosci.* **2015**, *8*, 466. [CrossRef] [PubMed]
- 102. Berrettini, W.; Yuan, X.; Tozzi, F.; Song, K.; Francks, C.; Chilcoat, H.; Waterworth, D.; Muglia, P.; Mooser, V. α5/α3 nicotinic receptor subunit alleles increase risk for heavy smoking. *Mol. Psychiatry* **2008**, *13*, 368–373. [CrossRef]
- 103. Saccone, S.F.; Hinrichs, A.L.; Saccone, N.L.; Chase, G.A.; Konvicka, K.; Madden, P.A.; Breslau, N.; Johnson, E.O.; Hatsukami, D.; Pomerleau, O.; et al. Cholinergic nicotinic receptor genes implicated in a nicotine dependence association study targeting 348 candidate genes with 3713 SNPs. *Hum. Mol. Genet.* 2006, 16, 36–49. [CrossRef]
- 104. Bierut, L.J.; Stitzel, J.A.; Wang, J.C.; Hinrichs, A.L.; Grucza, R.A.; Xuei, X.; Saccone, N.L.; Saccone, S.F.; Bertelsen, S.; Fox, L.; et al. Variants in nicotinic receptors and risk for nicotine dependence. *Am. J. Psychiatry* **2008**, *165*, 1163–1171. [CrossRef]
- 105. Wang, J.C.; Grucza, R.A.; Cruchaga, C.; Hinrichs, A.L.; Bertelsen, S.; Budde, J.; Fox, L.; Goldstein, E.; Reyes, O.; Saccone, N.; et al. Genetic variation in the CHRNA5 gene affects mRNA levels and is associated with risk for alcohol dependence. *Mol. Psychiatry* **2008**, *14*, 501–510. [CrossRef]
- 106. Fowler, C.D.; Lu, Q.; Johnson, P.M.; Marks, M.J.; Kenny, P.J. Habenular α5 nicotinic receptor subunit signalling controls nicotine intake. *Nature* **2011**, *471*, 597–601. [CrossRef]

Molecules **2020**, 25, 4223 25 of 28

107. Fowler, C.D.; Kenny, P.J. Nicotine aversion: Neurobiological mechanisms and relevance to tobacco dependence vulnerability. *Neuropharmacology* **2013**, *76*, 533–544. [CrossRef] [PubMed]

- 108. Nair-Roberts, R.; Chatelain-Badie, S.; Benson, E.; White-Cooper, H.; Bolam, J.P.; Ungless, M.A. Stereological estimates of dopaminergic, GABAergic and glutamatergic neurons in the ventral tegmental area, substantia nigra and retrorubral field in the rat. *Neuroscience* **2008**, *152*, 1024–1031. [CrossRef] [PubMed]
- 109. Di Chiara, G.; Imperato, A. Drugs abused by humans preferentially increase synaptic dopamine concentrations in the mesolimbic system of freely moving rats. *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 5274–5278. [CrossRef] [PubMed]
- 110. Koob, G.F.; Le Moal MD, M. Drug Addiction, Dysregulation of Reward, and Allostasis. *Neuropsychopharmacology* **2001**, 24, 97–129. [CrossRef]
- 111. Wise, R.A.; Rompre, P.P. Brain dopamine and reward. Annu. Rev. Psychol. 1989, 40, 191–225. [CrossRef]
- 112. Wise, R.A. Dopamine and reward: The anhedonia hypothesis 30 years on. *Neurotox. Res.* **2008**, *14*, 169–183. [CrossRef]
- 113. Corrigall, W.A.; Franklin, K.B.J.; Coen, K.M.; Clarke, P.B.S. The mesolimbic dopaminergic system is implicated in the reinforcing effects of nicotine. *Psychopharmacology* **1992**, *107*, 285–289. [CrossRef]
- 114. Louis, M.; Clarke, P.B. Effect of ventral tegmental 6-hydroxydopamine lesions on the locomotor stimulant action of nicotine in rats. *Neuropharmacology* **1998**, *37*, 1503–1513. [CrossRef]
- 115. Capriles, N.; Rodaros, D.; Sorge, R.E.; Stewart, J. A role for the prefrontal cortex in stress- and cocaine-induced reinstatement of cocaine seeking in rats. *Psychopharmacology* **2002**, *168*, 66–74. [CrossRef]
- 116. Counotte, D.S.; Goriounova, N.A.; Moretti, M.; Smoluch, M.T.; Irth, H.; Clementi, F.; Schoffelmeer, A.N.M.; Mansvelder, H.D.; Smit, A.B.; Gotti, C.; et al. Adolescent nicotine exposure transiently increases high-affinity nicotinic receptors and modulates inhibitory synaptic transmission in rat medial prefrontal cortex. FASEB J. 2012, 26, 1810–1820. [CrossRef]
- 117. Lodge, D.J.; Grace, A.A. The laterodorsal tegmentum is essential for burst firing of ventral tegmental area dopamine neurons. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 5167–5172. [CrossRef] [PubMed]
- 118. Omelchenko, N.; Sesack, S.R. Laterodorsal tegmental projections to identified cell populations in the rat ventral tegmental area. *J. Comp. Neurol.* **2005**, *483*, 217–235. [CrossRef] [PubMed]
- 119. Paladini, C.; Roeper, J. Generating bursts (and pauses) in the dopamine midbrain neurons. *Neuroscience* **2014**, 282, 109–121. [CrossRef] [PubMed]
- 120. Wolf, M.E. Addiction: Making the Connection Between Behavioral Changes and Neuronal Plasticity in Specific Pathways. *Mol. Interv.* **2002**, *2*, 146–157. [CrossRef] [PubMed]
- 121. Lammel, S.; Lim, B.; Ran, C.; Huang, K.W.; Betley, M.J.; Tye, K.M.; Deisseroth, K.; Malenka, R.C. Input-specific control of reward and aversion in the ventral tegmental area. *Nature* **2012**, 491, 212–217. [CrossRef]
- 122. Matsui, A.; Williams, J.T. Opioid-sensitive GABA inputs from rostromedial tegmental nucleus synapse onto midbrain dopamine neurons. *J. Neurosci.* **2011**, *31*, 17729–17735. [CrossRef]
- 123. Fowler, C.D.; Tuesta, L.; Kenny, P.J. Role of α5\* nicotinic acetylcholine receptors in the effects of acute and chronic nicotine treatment on brain reward function in mice. *Psychopharmacology* **2013**, 229, 503–513. [CrossRef]
- 124. Hikosaka, O. The habenula: From stress evasion to value-based decision-making. *Nat. Rev. Neurosci.* **2010**, 11, 503–513. [CrossRef]
- 125. Sulzer, D. How Addictive Drugs Disrupt Presynaptic Dopamine Neurotransmission. *Neuron* **2011**, *69*, 628–649. [CrossRef]
- 126. Maskos, U.; Molles, B.E.; Pons, S.; Besson, M.; Guiard, B.P.; Guilloux, J.-P.; Evrard, A.; Cazala, P.; Cormier, A.; Mameli-Engvall, M.; et al. Nicotine reinforcement and cognition restored by targeted expression of nicotinic receptors. *Nature* 2005, 436, 103–107. [CrossRef]
- 127. Guan, Y.-Z.; Ye, J.-H. Glycine blocks long-term potentiation of GABAergic synapses in the ventral tegmental area. *Neuroscience* **2016**, *318*, 134–142. [CrossRef] [PubMed]
- 128. Ye, J.H.; Wang, F.; Krnjevic, K.; Wang, W.; Xiong, Z.-G.; Zhang, J. Presynaptic Glycine Receptors on GABAergic Terminals Facilitate Discharge of Dopaminergic Neurons in Ventral Tegmental Area. *J. Neurosci.* **2004**, 24, 8961–8974. [CrossRef] [PubMed]
- 129. Li, J.; Nie, H.; Bian, W.; Dave, V.; Janak, P.H.; Ye, J.H. Microinjection of glycine into the ventral tegmental area selectively decreases ethanol consumption. *J. Pharmacol. Exp. Ther.* **2012**, *341*, 196–204. [CrossRef] [PubMed]

Molecules **2020**, 25, 4223 26 of 28

130. Tritsch, N.X.; Sabatini, B.L. Dopaminergic Modulation of Synaptic Transmission in Cortex and Striatum. *Neuron* **2012**, *76*, 33–50. [CrossRef] [PubMed]

- 131. Drenan, R.M.; Grady, S.R.; Steele, A.D.; McKinney, S.; Patzlaff, N.E.; McIntosh, J.M.; Marks, M.J.; Miwa, J.M.; Lester, H.A. Cholinergic Modulation of Locomotion and Striatal Dopamine Release Is Mediated by α6α4\* Nicotinic Acetylcholine Receptors. *J. Neurosci.* **2010**, *30*, 9877–9889. [CrossRef]
- 132. Cooper, S.; Robison, A.J.; Mazei-Robison, M.S. Reward Circuitry in Addiction. *Neurotherapeutics* **2017**, *14*, 687–697. [CrossRef]
- 133. McFarland, K.; Kalivas, P.W. The Circuitry Mediating Cocaine-Induced Reinstatement of Drug-Seeking Behavior. *J. Neurosci.* **2001**, *21*, 8655–8663. [CrossRef]
- 134. Knackstedt, L.A.; Kalivas, P.W. Glutamate and reinstatement. *Curr. Opin. Pharmacol.* **2009**, *9*, 59–64. [CrossRef]
- 135. Yang, H.; De Jong, J.W.; Tak, Y.; Peck, J.; Bateup, H.S.; Lammel, S. Nucleus Accumbens Subnuclei Regulate Motivated Behavior via Direct Inhibition and Disinhibition of VTA Dopamine Subpopulations. *Neuron* **2018**, 97. [CrossRef]
- 136. Bocklisch, C.; Pascoli, V.; Wong, J.; House, D.R.C.; Yvon, C.; De Roo, M.; Tan, K.R.; Lüscher, C. Cocaine Disinhibits Dopamine Neurons by Potentiation of GABA Transmission in the Ventral Tegmental Area. *Science* **2013**, *341*, 1521–1525. [CrossRef]
- 137. Xia, Y.; Driscoll, J.R.; Wilbrecht, L.; Margolis, E.B.; Fields, H.L.; Hjelmstad, G.O. Nucleus Accumbens Medium Spiny Neurons Target Non-Dopaminergic Neurons in the Ventral Tegmental Area. *J. Neurosci.* **2011**, *31*, 7811–7816. [CrossRef] [PubMed]
- 138. Wang, F.; Chen, H.; Steketee, J.D.; Sharp, B.M. Upregulation of ionotropic glutamate receptor subunits within specific mesocorticolimbic regions during chronic nicotine self-administration. *Neuropsychopharmacology* **2006**, 32, 103–109. [CrossRef] [PubMed]
- 139. Rezvani, A.H.; Eddins, N.; Slade, S.; Hampton, D.S.; Christopher, N.C.; Petro, A.; Horton, K.; Johnson, M.; Levin, E.D. Neonatal 6-hydroxydopamine lesions of the frontal cortex in rats: Persisting effects on locomotor activity, learning and nicotine self-administration. *Neuroscience* **2008**, *154*, 885–897. [CrossRef] [PubMed]
- 140. Kalivas, P.W.; Volkow, N.D. The Neural Basis of Addiction: A Pathology of Motivation and Choice. *Am. J. Psychiatry* **2005**, *162*, 1403–1413. [CrossRef]
- 141. Cornish, J.L.; Kalivas, P.W. Glutamate Transmission in the Nucleus Accumbens Mediates Relapse in Cocaine Addiction. *J. Neurosci.* **2000**, 20, RC89. [CrossRef]
- 142. Due, D.L.; Huettel, S.A.; Hall, W.G.; Rubin, D.C. Activation in Mesolimbic and Visuospatial Neural Circuits Elicited by Smoking Cues: Evidence From Functional Magnetic Resonance Imaging. *Am. J. Psychiatry* **2002**, 159, 954–960. [CrossRef]
- 143. Taber, M.T.; Das, S.; Fibiger, H.C. Cortical Regulation of Subcortical Dopamine Release: Mediation via the Ventral Tegmental Area. *J. Neurochem.* **2002**, *65*, 1407–1410. [CrossRef]
- 144. Deas, D.; Brown, E.S. Adolescent substance abuse and psychiatric comorbidities. *J. Clin. Psychiatry* **2006**, *67*, 2. [CrossRef]
- 145. Brown, R.A.; Lewinsohn, P.M.; Seeley, J.R.; Wagner, E.F. Cigarette Smoking, Major Depression, and Other Psychiatric Disorders among Adolescents. *J. Am. Acad. Child. Adolesc. Psychiatry* **1996**, *35*, 1602–1610. [CrossRef]
- 146. Yuan, M.; Cross, S.J.; Loughlin, S.E.; Leslie, F.M. Nicotine and the adolescent brain. *J. Physiol.* **2015**, *593*, 3397–3412. [CrossRef]
- 147. Pascoli, V.; Terrier, J.; Espallergues, J.; Valjent, E.; O'Connor, E.C.; Lüscher, C. Contrasting forms of cocaine-evoked plasticity control components of relapse. *Nature* **2014**, *509*, 459–464. [CrossRef] [PubMed]
- 148. Huang, Y.H.; Schlüter, O.M.; Dong, Y. Silent Synapses Speak Up. *Neuroscience* **2015**, *21*, 451–459. [CrossRef] [PubMed]
- 149. McLaughlin, I.; Dani, J.A.; De Biasi, M. The medial habenula and interpeduncular nucleus circuitry is critical in addiction, anxiety, and mood regulation. *J. Neurochem.* **2017**, *142*, 130–143. [CrossRef] [PubMed]
- 150. Salas, R.; Sturm, R.M.; Boulter, J.; De Biasi, M. Nicotinic receptors in the habenulo-interpeduncular system are necessary for nicotine withdrawal in mice. *J. Neurosci.* **2009**, *29*, 3014–3018. [CrossRef]
- 151. Salas, R.; Pieri, F.; De Biasi, M. Decreased signs of nicotine withdrawal in mice null for the β4 nicotinic acetylcholine receptor subunit. *J. Neurosci.* **2004**, *24*, 10035–10039. [CrossRef]

Molecules **2020**, 25, 4223 27 of 28

152. Ji, H.; Shepard, P.D. Lateral habenula stimulation inhibits rat midbrain dopamine neurons through a GABA(A) receptor-mediated mechanism. *J. Neurosci.* **2007**, 27, 6923–6930. [CrossRef]

- 153. Matsumoto, M.; Hikosaka, O. Lateral habenula as a source of negative reward signals in dopamine neurons. *Nature* **2007**, *447*, 1111–1115. [CrossRef]
- 154. Wellmann, K.F. Smoking and health. On the report of the advisory committee to the surgeon general of the public health service. *DMW—Dtsch. Med. Wochenschr.* **1964**, *89*, 45–46.
- 155. Lundeen, R.E. Tobacco under the FDA: A summary of the Family Smoking Prevention and Tobacco Control Act. *Health Care Law Mon.* **2009**, *9*, 2–9.
- 156. Odani, S.; Armour, B.S.; Graffunder, C.M.; Willis, G.; Hartman, A.M.; Agaku, I.T. The 2014–2015 Tobacco Use Supplement to the Current Population Survey. *MMWR Morb Mortal Wkly Rep.* **2018**, *67*, 97–102. [CrossRef]
- 157. Kozlitina, J.; Risso, D.; Lansu, K.; Olsen, R.H.J.; Sainz, E.; Luiselli, D.; Barik, A.; Domingues, C.E.F.; Pagani, L.; Wooding, S.; et al. An African-specific haplotype in MRGPRX4 is associated with menthol cigarette smoking. *PLoS Genet.* **2019**, *15*, e1007916. [CrossRef] [PubMed]
- 158. Wickham, R. How Menthol Alters Tobacco-Smoking Behavior: A Biological Perspective. *Yale J. Boil. Med.* **2015**, *88*, 279–287.
- 159. DeVito, E.E.; Krishnan-Sarin, S. E-cigarettes: Impact of E-Liquid Components and Device Characteristics on Nicotine Exposure. *Curr. Neuropharmacol.* **2018**, *16*, 438–459. [CrossRef]
- 160. Baker, T.B.; Piper, M.E.; McCarthy, D.E.; Bolt, D.M.; Smith, S.S.; Kim, S.-Y.; Colby, S.; Conti, D.; Giovino, G.A.; Hatsukami, D.; et al. Time to first cigarette in the morning as an index of ability to quit smoking: Implications for nicotine dependence. *Nicotine Tob. Res.* **2007**, *9*, 555–570. [CrossRef]
- 161. Ahijevych, K.; Parsley, L.A. Smoke constituent exposure and stage of change in black and white women cigarette smokers. *Addict. Behav.* **1999**, 24, 115–120. [CrossRef]
- 162. Ahijevych, K.; Garrett, B.E. Menthol pharmacology and its potential impact on cigarette smoking behavior. *Nicotine Tob. Res.* **2004**, *6*, 17–28. [CrossRef]
- 163. Caggiula, A.R.; Donny, E.C.; White, A.R.; Chaudhri, N.; Booth, S.A.; Gharib, M.; Hoffman, A.; Perkins, K.A.; Sved, A.F. Cue dependency of nicotine self-administration and smoking. *Pharmacol. Biochem. Behav.* **2001**, *70*, 515–530. [CrossRef]
- 164. Harrison, E.; Biswas, L.; Avusula, R.; Zhang, M.; Gong, Y.; Liu, X. Effects of menthol and its interaction with nicotine-conditioned cue on nicotine-seeking behavior in rats. *Psychopharmacology* **2017**, 234, 3443–3453. [CrossRef]
- 165. Wickham, R.J.; Nunes, E.J.; Hughley, S.; Silva, P.; Walton, S.N.; Park, J.; Addy, N.A. Evaluating oral flavorant effects on nicotine self-administration behavior and phasic dopamine signaling. *Neuropharmacology* **2017**, 128, 33–42. [CrossRef]
- 166. Hans, M.; Wilhelm, M.; Swandulla, D. Menthol Suppresses Nicotinic Acetylcholine Receptor Functioning in Sensory Neurons via Allosteric Modulation. *Chem. Senses* **2012**, *37*, 463–469. [CrossRef]
- 167. Ashoor, A.; Nordman, J.C.; Veltri, D.; Yang, K.-H.S.; Al Kury, L.; Shuba, Y.; Mahgoub, M.; Howarth, F.C.; Sadek, B.; Shehu, A.; et al. Menthol Binding and Inhibition of α7-Nicotinic Acetylcholine Receptors. *PLoS ONE* **2013**, *8*, e67674. [CrossRef]
- 168. Ton, H.T.; Smart, A.E.; Aguilar, B.L.; Olson, T.T.; Kellar, K.J.; Ahern, G.P. Menthol Enhances the Desensitization of Human alpha3beta4 Nicotinic Acetylcholine Receptors. *Mol. Pharmacol.* **2015**, *88*, 256–264. [CrossRef]
- 169. Shahoei, R.; Tajkhorshid, E. Menthol Binding to the Human α4β2 Nicotinic Acetylcholine Receptor Facilitated by Its Strong Partitioning in the Membrane. *J. Phys. Chem. B* **2020**, *124*, 1866–1880. [CrossRef] [PubMed]
- 170. AlSharari, S.D.; King, J.R.; Nordman, J.C.; Muldoon, P.P.; Jackson, A.; Zhu, A.Z.X.; Tyndale, R.F.; Kabbani, N.; Damaj, M.I. Effects of Menthol on Nicotine Pharmacokinetic, Pharmacology and Dependence in Mice. *PLoS ONE* **2015**, *10*, e0137070. [CrossRef] [PubMed]
- 171. Wickham, R.J. The Biological Impact of Menthol on Tobacco Dependence. Nicotine Tob. Res. 2019. [CrossRef]
- 172. Valentine, G.E.; DeVito, E.; Jatlow, P.; Gueorguieva, R.V.; Sofuoglu, M. Acute effects of inhaled menthol on the rewarding effects of intravenous nicotine in smokers. *J. Psychopharmacol.* **2018**, 32, 986–994. [CrossRef]
- 173. Villanti, A.C.; Collins, L.K.; Niaura, R.; Gagosian, S.Y.; Abrams, D.B. Menthol cigarettes and the public health standard: A systematic review. *BMC Public Health* **2017**, *17*, 1–13. [CrossRef] [PubMed]
- 174. Tierney, P.A.; Karpinski, C.D.; Brown, J.E.; Luo, W.; Pankow, J.F. Flavour chemicals in electronic cigarette fluids. *Tob. Control.* **2015**, 25, e10–e15. [CrossRef] [PubMed]

Molecules **2020**, 25, 4223 28 of 28

175. Keath, J.R.; Iacoviello, M.P.; Barrett, L.E.; Mansvelder, H.D.; McGehee, D.S. Differential Modulation by Nicotine of Substantia Nigra Versus Ventral Tegmental Area Dopamine Neurons. *J. Neurophysiol.* **2007**, *98*, 3388–3396. [CrossRef]

- 176. Chen, H.; Matta, S.G.; Sharp, B.M. Acquisition of Nicotine Self-Administration in Adolescent Rats Given Prolonged Access to the Drug. *Neuropsychopharmacology* **2006**, *32*, 700–709. [CrossRef]
- 177. Kota, D.; Martin, B.R.; Robinson, S.E.; Damaj, M.I. Nicotine Dependence and Reward Differ between Adolescent and Adult Male Mice. *J. Pharmacol. Exp. Ther.* **2007**, 322, 399–407. [CrossRef] [PubMed]
- 178. Goriounova, N.A.; Mansvelder, H.D. Short- and Long-Term Consequences of Nicotine Exposure during Adolescence for Prefrontal Cortex Neuronal Network Function. *Cold Spring Harb. Perspect. Med.* **2012**, 2, a012120. [CrossRef] [PubMed]
- 179. Goriounova, N.A.; Mansvelder, H.D. Nicotine exposure during adolescence alters the rules for prefrontal cortical synaptic plasticity during adulthood. *Front. Synaptic Neurosci.* **2012**, *4*, 3. [CrossRef] [PubMed]
- 180. Romoli, B.; Lozada, A.F.; Sandoval, I.M.; Manfredsson, F.P.; Hnasko, T.S.; Berg, D.K.; Dulcis, D. Neonatal Nicotine Exposure Primes Midbrain Neurons to a Dopaminergic Phenotype and Increases Adult Drug Consumption. *Boil. Psychiatry* **2019**, *86*, 344–355. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).