

## SUPPLEMENTARY MATERIALS

### Dual miRNases for triple incision of miRNA target: design concept and catalytic performance

Olga A. Patutina <sup>1</sup>, Daria A. Chiglintseva <sup>1</sup>, Elena V. Bichenkova <sup>2</sup>, Svetlana K. Gaponova <sup>1</sup>, Nadezhda L. Mironova <sup>1</sup>, Valentin V. Vlassov <sup>1</sup> and Marina A. Zenkova <sup>1,\*</sup>

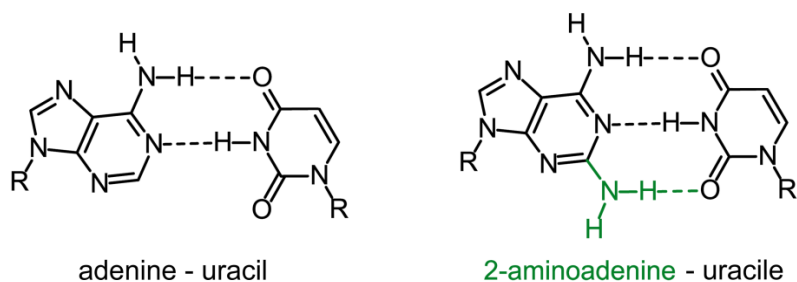
<sup>1</sup> Laboratory of Nucleic Acids Biochemistry, Institute of Chemical Biology and Fundamental Medicine SB RAS, Lavrentiev's ave. 8, 630090, Novosibirsk, Russia

<sup>2</sup> School of Health Sciences, Faculty of Biology, Medicine and Health, University of Manchester, Oxford Rd, Manchester, M13 9PT, UK

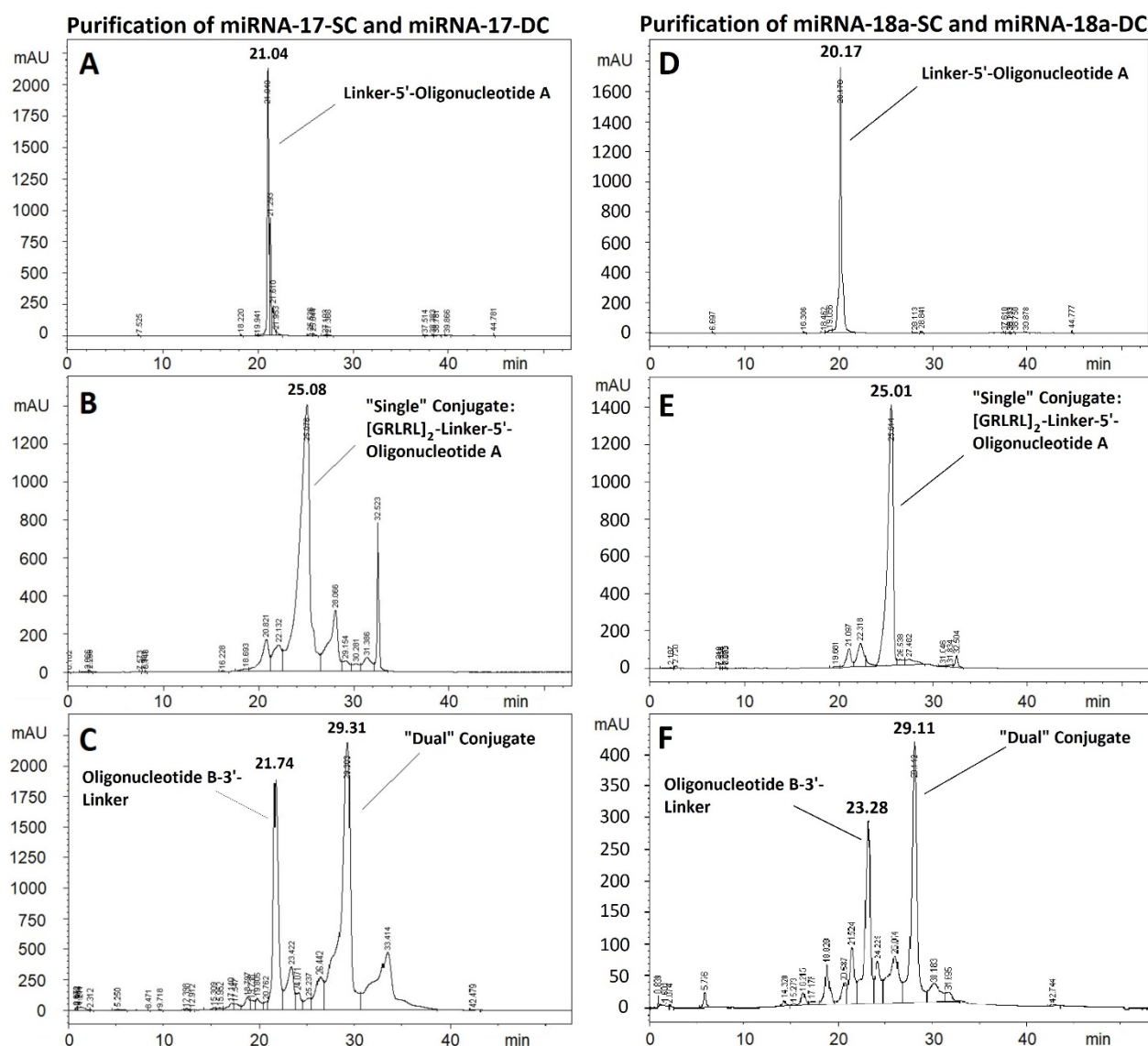
\* Correspondence: marzen@niboch.nsc.ru; Tel.: +7-383-363-51-60

**Table S1.** The role of miR-21, miR-155, miR-17 and miR-18a in oncological diseases.

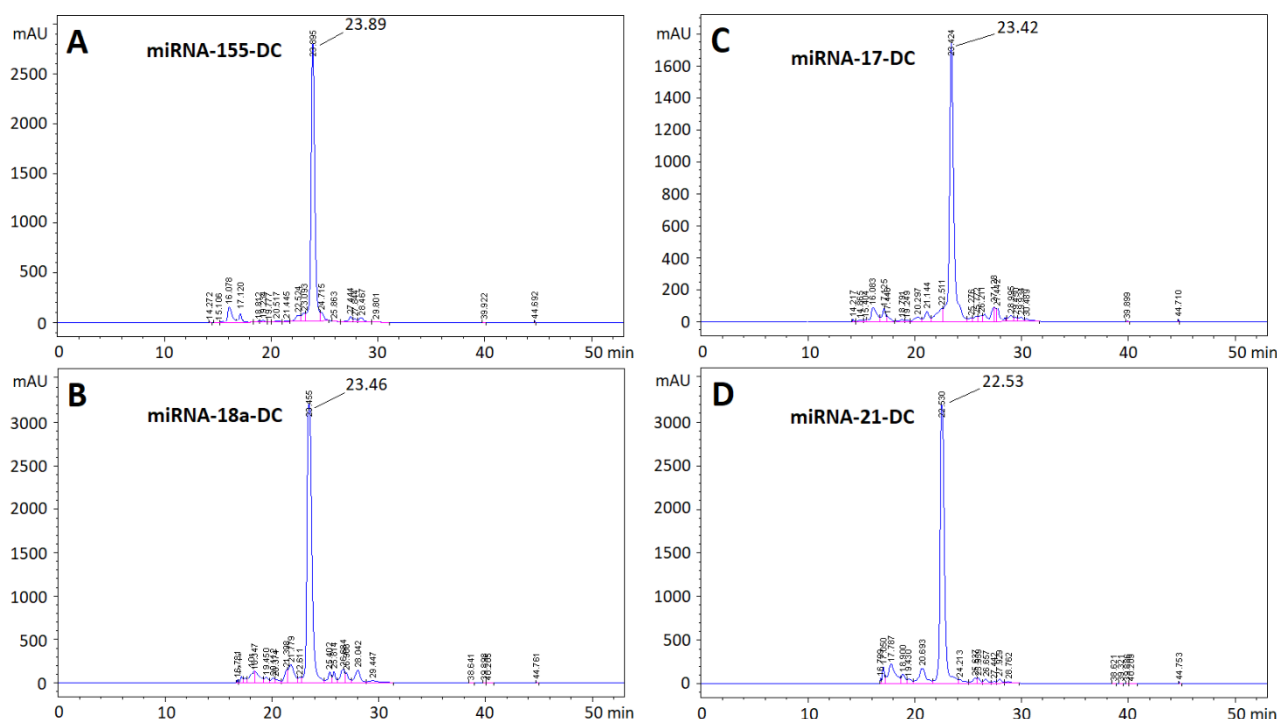
miRNA	mRNA targets	Cellular function under control	Malignancy	Ref.
miR-21	PDCD4, TPM1, PTEN, SPRY1, SPRY2, MARCKS	Proliferation, differentiation, migration, invasion, angiogenesis, apoptosis	Breast, non-small cellular lung cancer, lymphosarcoma, melanoma, gastric and colorectal cancer	[4,14,15]
miR-155	GABRA1, FOXO3a, BACH1, VHL	Proliferation, migration, invasion, angiogenesis	B-cell lymphoma, hepatocellular carcinoma, lung and breast cancer	[4,16,20]
miR-17	Transcription factors, matrix metalloproteinases	Proliferation, migration, apoptosis	Colorectal, gastric and breast cancer	[4,17]
miR-18a	Proteins of phosphatidylinositol 3-kinase-protein kinase B (PI3K/AKT), MEK/extracellular signal-regulated kinase (ERK), mTOR and Wnt/ $\beta$ -catenin pathways	Proliferation, migration, invasion	Nasopharyngeal carcinoma, clear cell renal cell carcinoma, cervical cancer	[4,18,19]



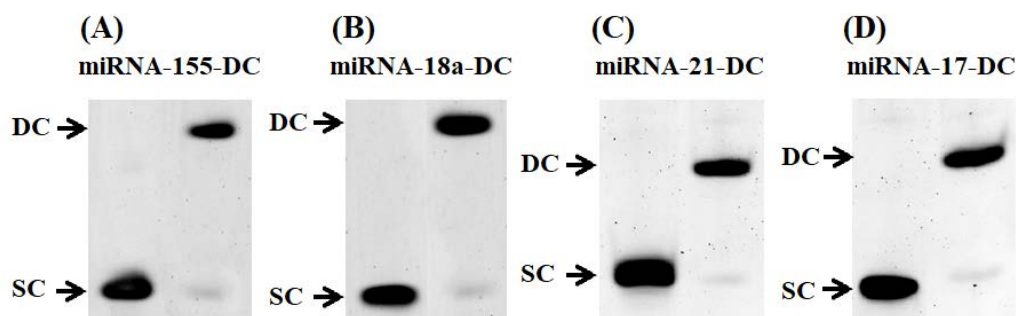
**Figure S1.** The structure of the pair adenine-uracile and 2-aminoadenine-uracil.



**Figure S2.** Representative examples of IEX HPLC showing purification of the Single and Dual conjugates after the first and second coupling reactions, respectively, as a part of the synthetic route of miRNA-17-DC (left; **A – C**) and miRNA-18a-DC (right; **D – F**). (**A** and **D**) Starting Oligonucleotide A bearing 5'-thiohexyl linker with the retention time of 21.04 or 20.17 min, respectively. (**B** and **E**) Single conjugate (miRNA-17-SC or miRNA-18a-SC, respectively) after the first coupling step, showing the shift in the retention time by 4 or 5 min as compared to unmodified Oligonucleotide A. (**C** and **F**) Dual conjugate (miRNA-17-DC or miRNA-18a-DC, respectively) after the second coupling step, showing the additional 4-minute shift in the retention time as compared to that of the Single conjugate. IEX was performed using a Clarity Oligo-WAX column. The mobile phase included 10 mM Tris-Cl (pH 7.25) and 10% AcCN in water as eluent A, and 2 M NaCl, 10 mM Tris-Cl (pH 7.25) and 10% AcCN in water as eluent B. The following gradient was applied: 100% A for 5 min, followed by 0% B to 100% B in 32 min. The flow rate was maintained at 2.0 mL/min, and the UV absorbance was monitored at 260 nm. Similar shifts in the retention times were witnessed during IEX HPLC purification of the other conjugates.

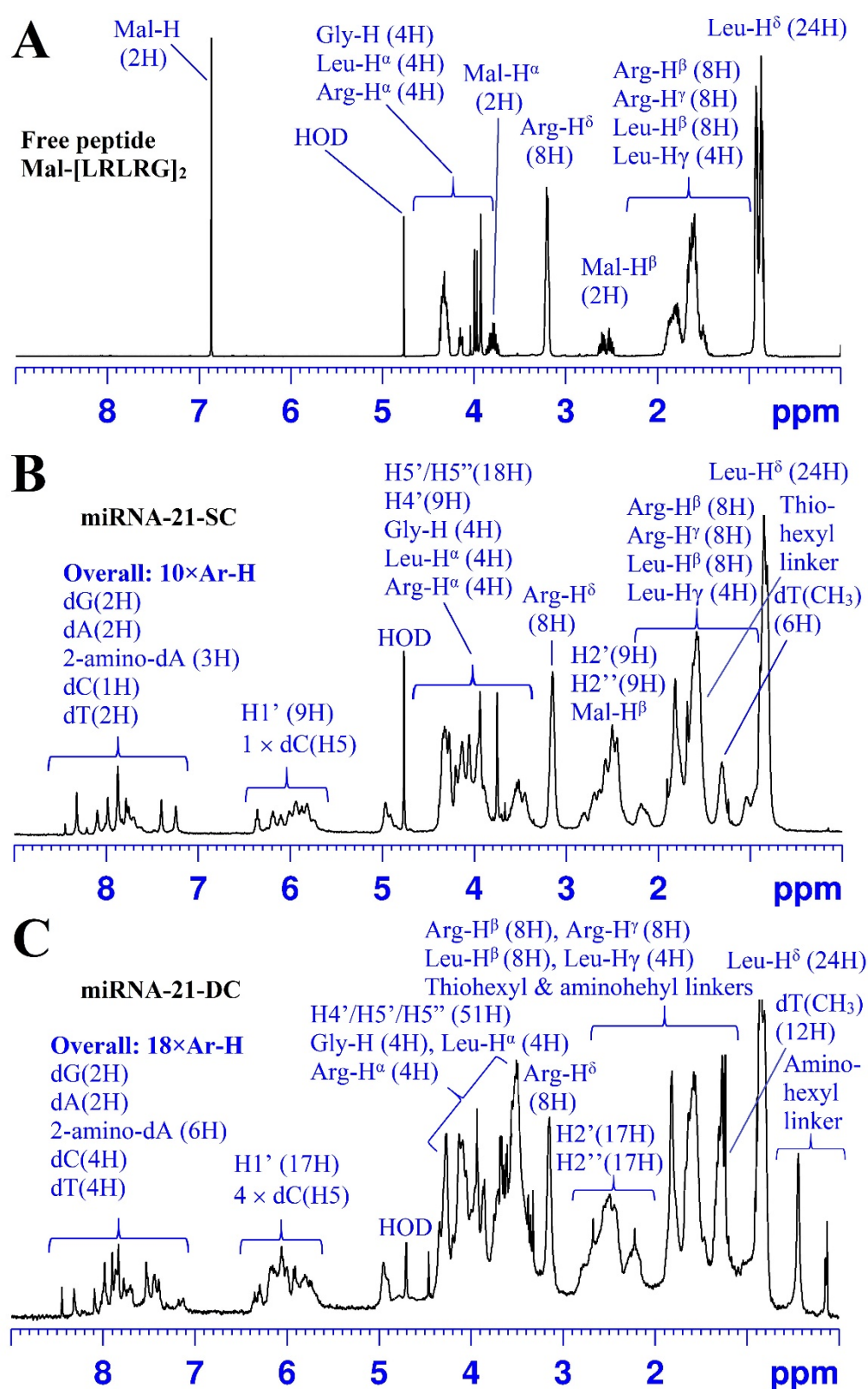


**Figure S3.** Reversed phase HPLC purification of the Dual conjugates miRNA-155-DC (A), miRNA-18a-DC (B), miRNA-17-DC (C) and miRNA-21-DC (D). RP-HPLC was carried out using a semi-preparative column Luna C18 (Phenomenex; CA, USA). The mobile phase included 0.05 M LiClO<sub>4</sub> in water as eluent A and 0.05 M LiClO<sub>4</sub> in AcCN as eluent B. The absorbance was monitored at 260 nm, and the following gradient was applied: 100% A for 3 min, 0% B to 100% B in 30 min, followed by washing with 100% B over 10 min and with 100% A over the next 20 min. The flow rate was maintained at 2.0 mL/min. The Single conjugates miRNA-155-SC, miRNA-18a-SC, miRNA-17-SC and miRNA-21-SC were purified by RP-HPLC in a similar way.

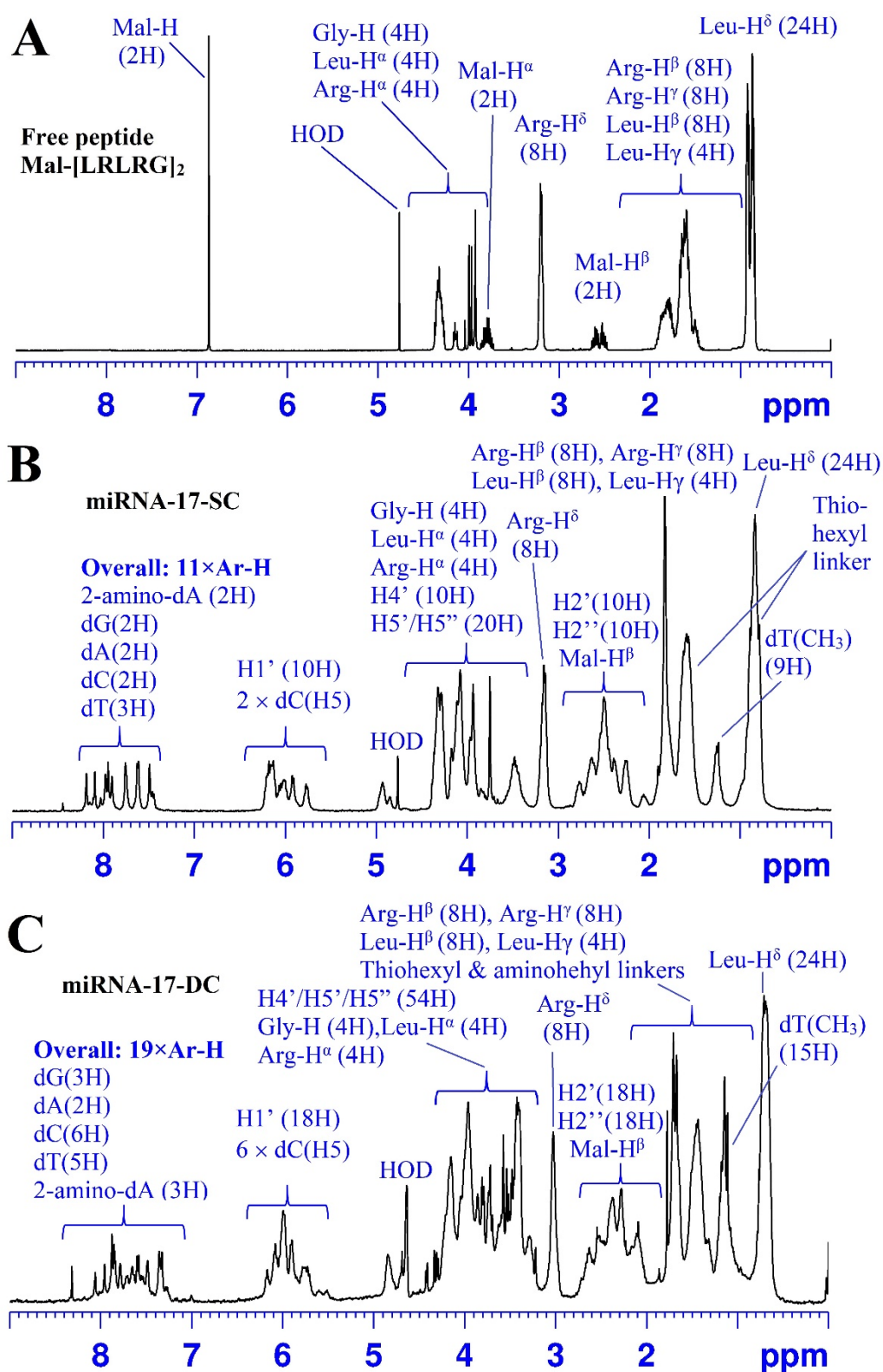


PAGE	Dual Conjugate	Purity (%)
(A)	miRNA-155-DC	95.2%
(B)	miRNA-18a-DC	95.6%
(C)	miRNA-21-DC	96.5%
(D)	miRNA-17-DC	95.6%

**Figure S4:** Urea-PAGE (20% PAA/8 M Urea) analysis showing homogeneity of the Dual conjugates (DC) miRNA-155-DC (A), miRNA-18a-DC (B), miRNA-21-DC (C) and miRNA-17-DC (D) isolated as the main fractions between 22 and 25 min of the reversed phase HPLC purification (see **Figure S3**). Urea-PAGE analysis shows the reduced electrophoretic mobility of the Dual conjugates as compared to that for the corresponding Single conjugates (SC) miRNA-155-SC, miRNA-18a-SC, miRNA-21-SC and miRNA-17-SC, respectively.

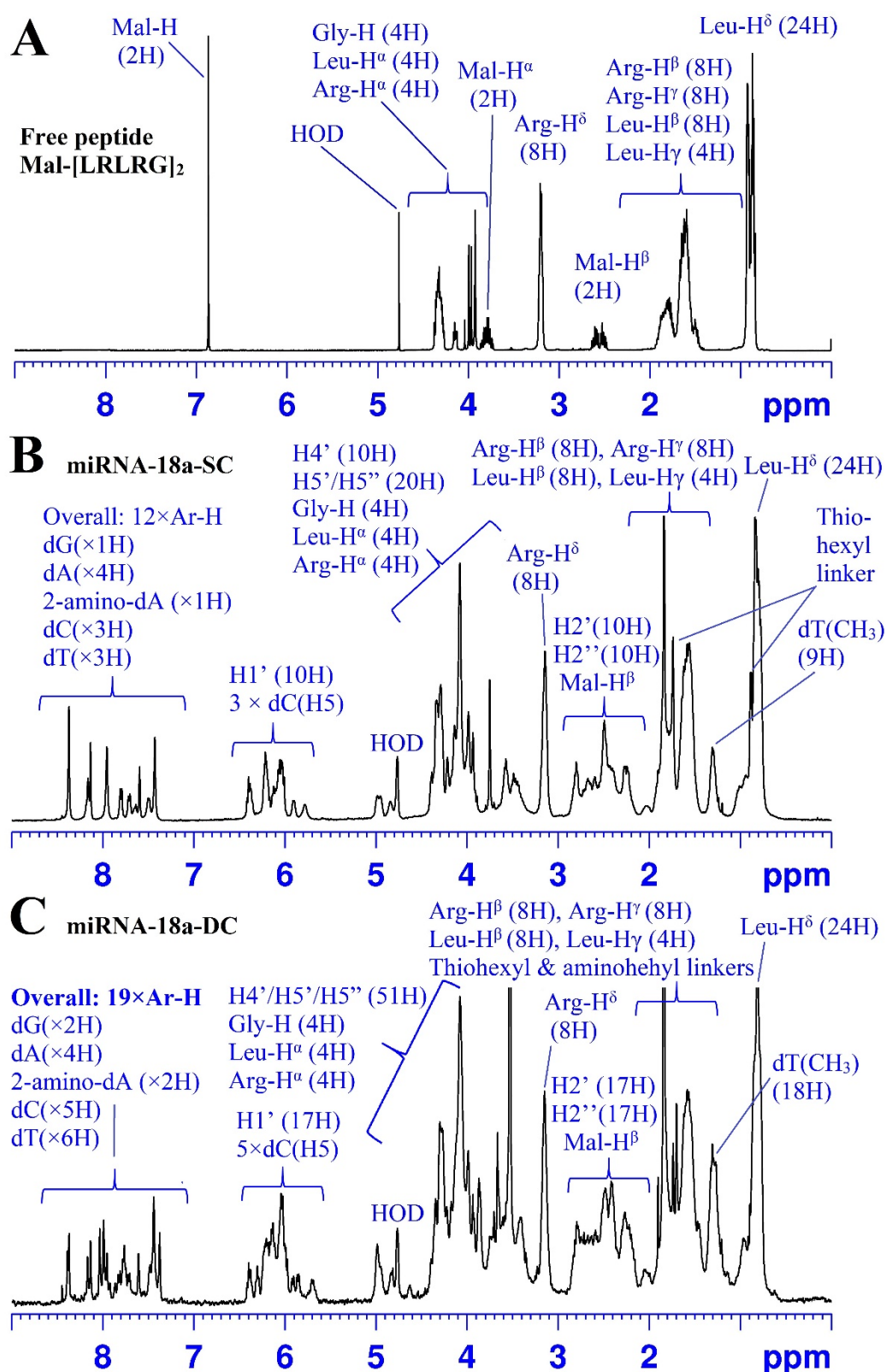


**Figure S5.** Step-by-step characterization of the Single and Dual peptidyl-oligonucleotide conjugates produced for targeting miR-21 sequence using <sup>1</sup>H-NMR spectroscopy. NMR spectra of the unconjugated **Mal-[LRLRG]<sub>2</sub>** peptide (**A**), intermediate Single conjugate **miRNA-21-SC** (**B**) and Dual conjugate **miRNA-21-DC** (**C**). Spectra were recorded in D<sub>2</sub>O at 25 °C using 400 MHz, Bruker Avance II+ 400 NMR spectrometer.

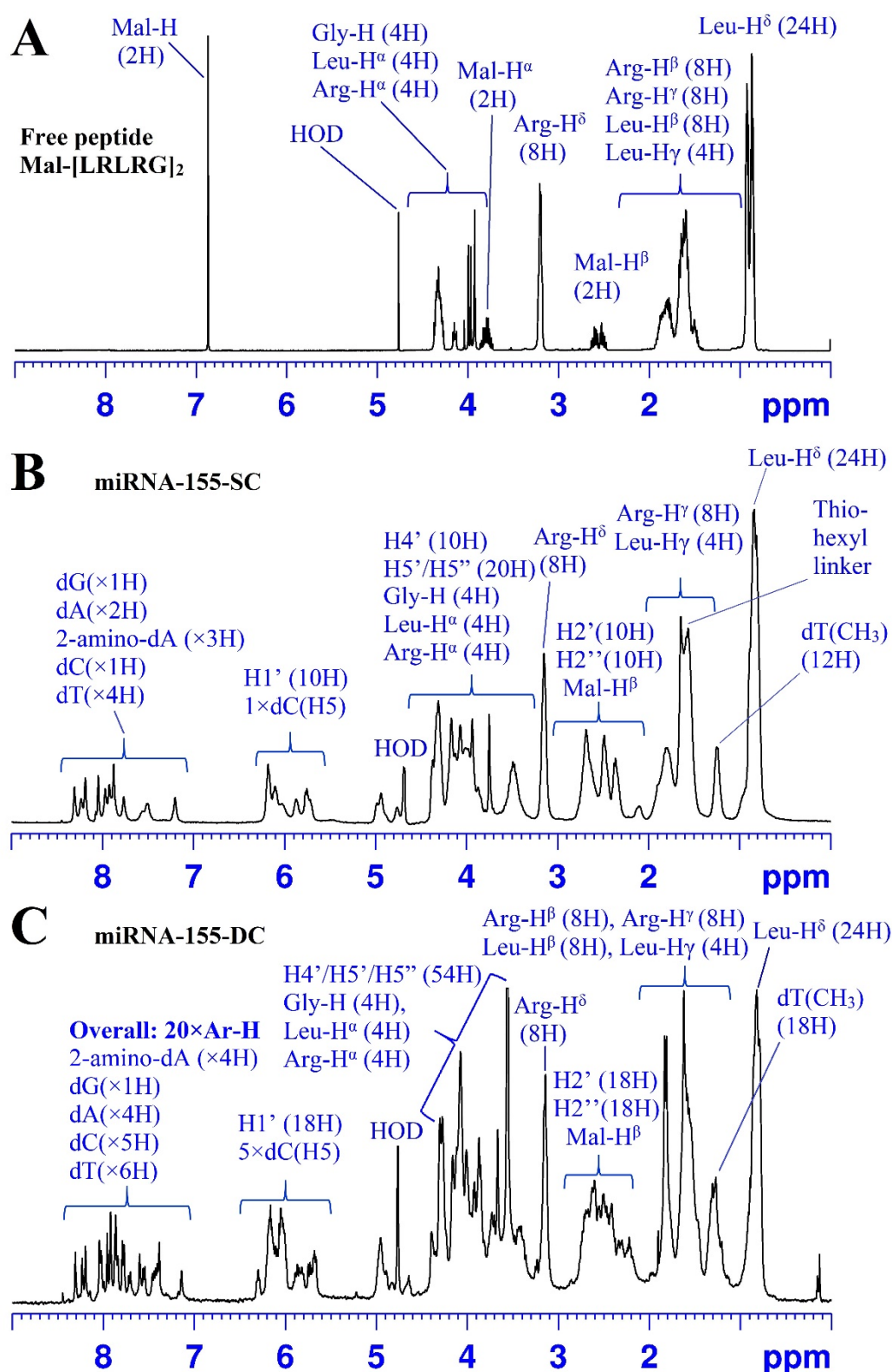


**Figure S6.** Step-by-step characterization of the Single and Dual peptidyl-oligonucleotide conjugates produced for targeting miR-17 sequence using <sup>1</sup>H-NMR spectroscopy. NMR spectra of the unconjugated **Mal-[LRLRG]<sub>2</sub>** peptide (A), intermediate Single conjugate **miRNA-17-SC** (B) and Dual conjugate **miRNA-17-DC** (C). Spectra were recorded in D<sub>2</sub>O at 25 °C using 400 MHz, Bruker Avance II+ 400 NMR spectrometer.

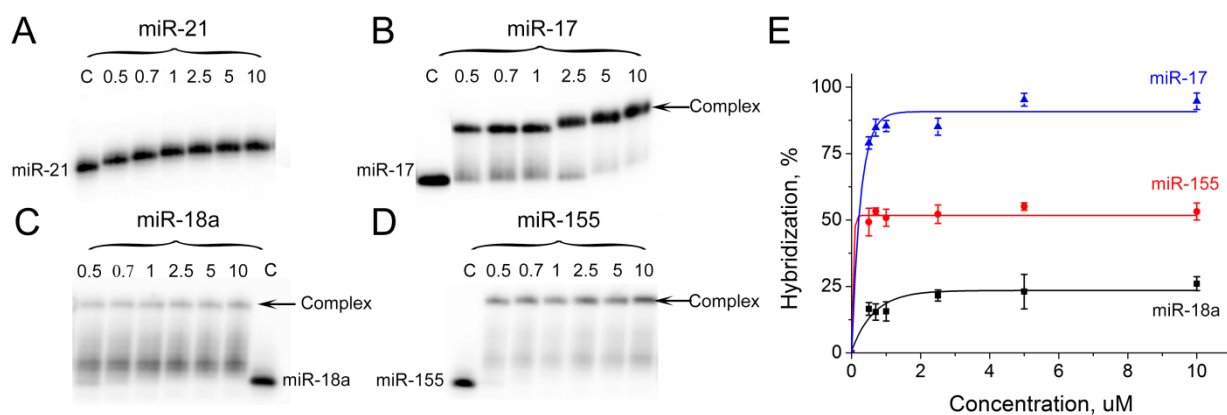




**Figure S7.** Step-by-step characterization of the Single and Dual peptidyl-oligonucleotide conjugates produced for targeting miR-18a sequence using <sup>1</sup>H-NMR spectroscopy. NMR spectra of the unconjugated **Mal-[LRLRG]<sub>2</sub>** peptide (**A**), intermediate Single conjugate **miRNA-18a-SC** (**B**) and Dual conjugate **miRNA-18a-DC** (**C**). Spectra were recorded in D<sub>2</sub>O at 25 °C using 400 MHz, Bruker Avance II+ 400 NMR spectrometer.



**Figure S8.** Step-by-step characterization of the Single and Dual peptidyl-oligonucleotide conjugates produced for targeting miR-155 sequence using <sup>1</sup>H-NMR spectroscopy. NMR spectra of the unconjugated **Mal-[LRLRG]<sub>2</sub>** peptide (**A**), intermediate Single conjugate **miRNA-155-SC** (**B**) and **miRNA-155-DC** (**C**). Spectra were recorded in D<sub>2</sub>O at 25 °C using 400 MHz, Dual conjugate Bruker Avance II+ 400 NMR spectrometer.



**Figure S9.** Hybridization of 5'-[<sup>32</sup>P]-labeled miRNAs with the Dual oligonucleotides (ONs) miRNA-21-ON, miRNA-17-ON, miRNA-18a-ON and miRNA-155-ON. **A, B, C** and **D.** Radioautographs of 15% native PAAG, showing hybridization of the corresponding ONs with miR-21, miR-17, miR-18a and miR-155, respectively. miRNAs (1 μM) were incubated with ONs (0.1 – 10 μM) in buffer (1), containing 50 mM Tris-HCl, pH 7.0, 200 mM KCl and 1 mM EDTA at 37 °C for 45 min. The samples were loaded onto the running gel immediately after quenching the reaction with 1 min intervals. The concentration (μM) of ONs is indicated on the top of electrophoregrams. **E.** Concentration dependencies of ONs hybridization with miR-17, miR-18a and miR-155. Data are presented as mean ± s.e.



**Table S2.** Efficiency of miRNA cleavage and observed rate constants ( $K_{obs}$ ) of miRNA-targeted DCs in buffer (1) and (2).

	Total Cleavage, %								
	miR-17		miR-18a		miR-21	miR-21_1		miR-155	
Time, h	<i>buff 1</i>	<i>buff 2</i>	<i>buff 1</i>	<i>buff 2</i>	<i>buff 2</i>	<i>buff 1</i>	<i>buff 2</i>	<i>buff 1</i>	<i>buff 2</i>
2	0	1,5	0	1,4	4	0,7	-	0	15
6	0	4	1	5	13	-	14	0	41
24	1	18	3	19	30	10	48	0,5	56
48	2	32	7	23	30	18	63	16	57
72	5	40	8	37	34	-	-	21	72
$K_{obs}$ , $\times 10^{-6}, s^{-1}$	18.0± 1.6	190.0± 4.5	34.2± 2.4	222.0± 6.1	328.0± 32.9	107.0± 4.3	373.0± 55.9	85.2± 17.0	543.0± 173.0

**Table S3.** Cleavage of 5'-[ $^{32}$ P]-labeled miR-18a by DC in buffer, containing different concentration of  $MgCl_2$ .

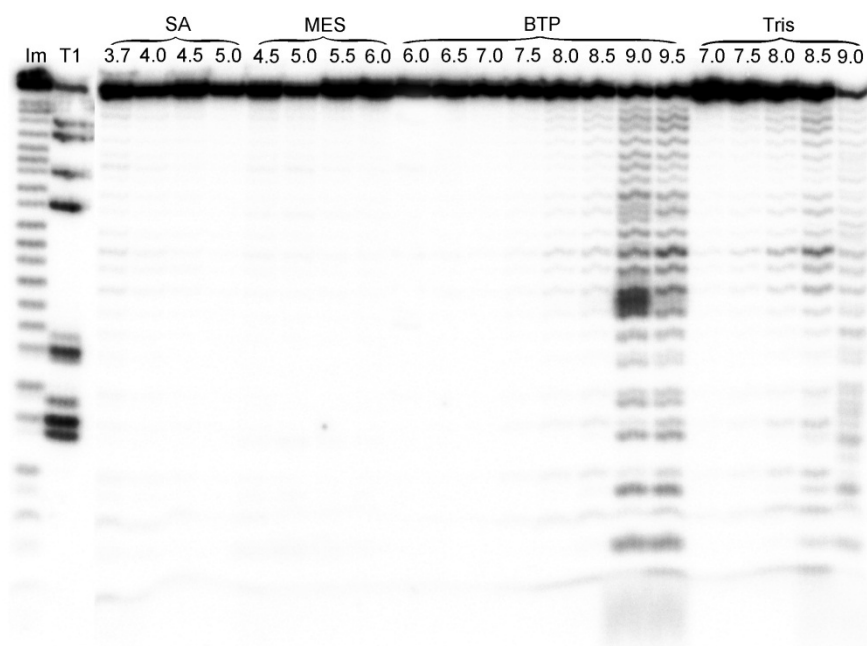
$Mg^{2+}$ , mM	0	2	4	8	15
Cleavage, %	26.4 $\pm$ 4.0	28.8 $\pm$ 3.9	24.3 $\pm$ 2.7	23.1 $\pm$ 3.1	27.2 $\pm$ 4.3

miRNA [1  $\mu$ M] and DC [20  $\mu$ M] were incubated at 37°C for 48 h. Buffer: 50 mM Tris-HCl, pH 7.0, 100 mM KCl, 1 mM DTT, 0.02 mg/mL BSA and  $MgCl_2$  at a concentration range from 0 to 15 mM.

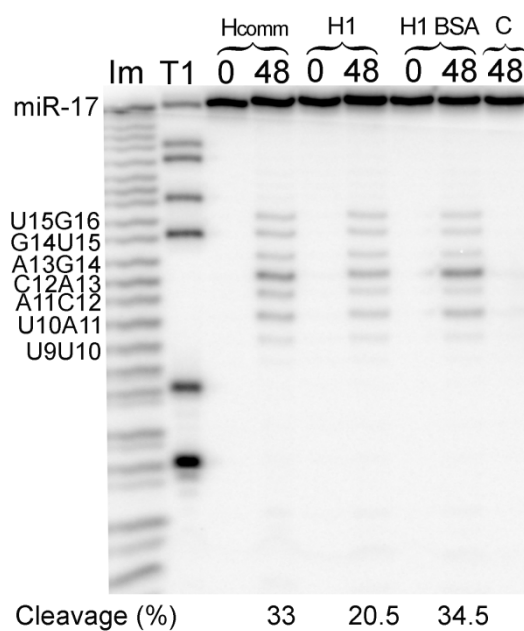
**Table S4.** Cleavage of 5'-[ $^{32}$ P]-labeled miR-17 by DC in buffer, containing different concentration of KCl.

$K^+$ , mM	0	40	100	200	400
Cleavage, %	13.2 $\pm$ 5.0	22.5 $\pm$ 4.3	26.4 $\pm$ 2.0	13.7 $\pm$ 3.5	14.1 $\pm$ 3.8

miRNA [1  $\mu$ M] and DC [20  $\mu$ M] were incubated at 37°C for 48 h. Buffer: Buffer: 20 mM Tris-HCl, pH 7.8, 8 mM  $MgCl_2$ , 1 mM DTT, 0.02 mg/mL BSA and KCl at a concentration range from 0 to 400 mM.



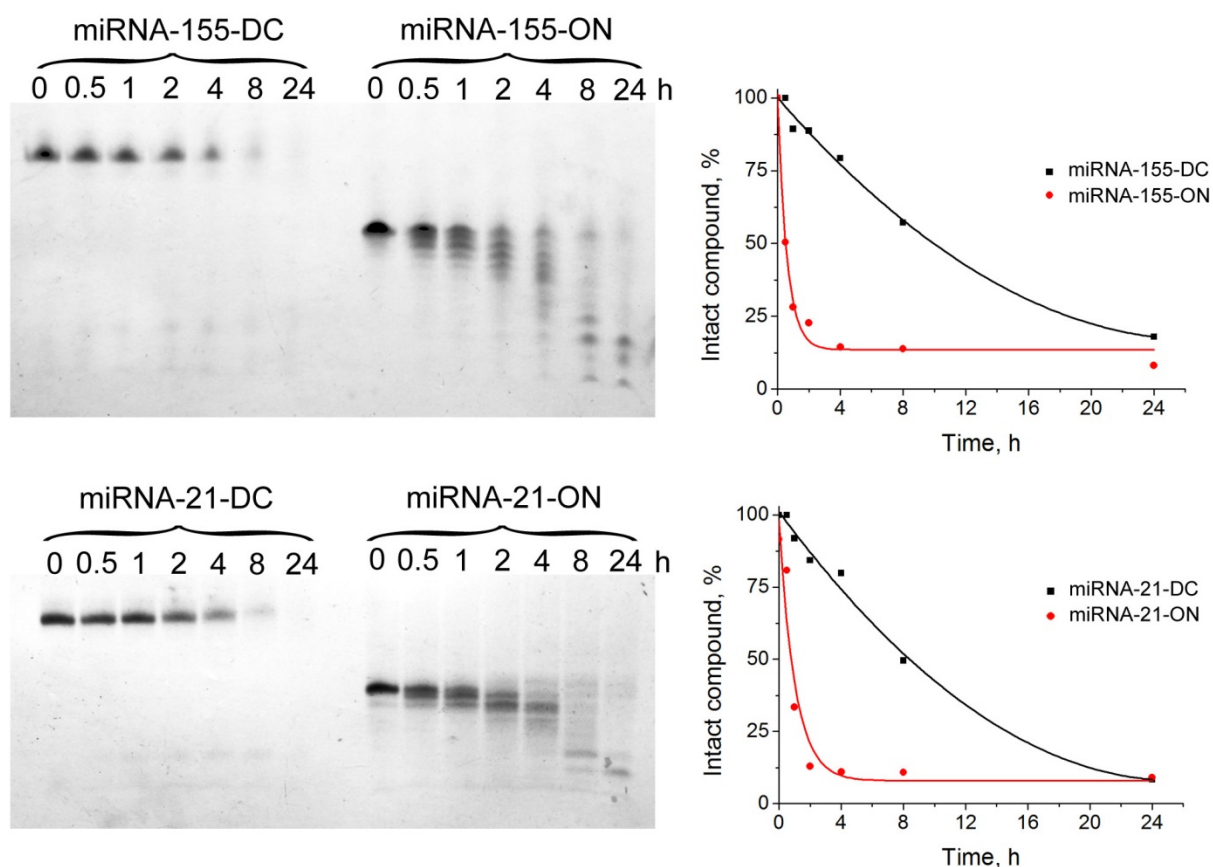
**Figure S10.** Self-cleavage of 5'-[<sup>32</sup>P]-labeled miR-17 at different pH. Buffers are listed in the “Materials and methods” section. Radioautograph of 18% denaturing PAAG showing the cleavage products of miR-17. miRNA-17 at 1 μM concentration was incubated at 37°C for 48 h. Lanes Im and T1 – imidazole ladder and partial RNA digestion with RNase T1, respectively.



**Figure S11.** Cleavage of 5'-[<sup>32</sup>P]-labeled miR-17 by DC in three buffers: Hcomm – commercial RNase H buffer; H1 – buffer, containing 20 mM Tris-HCl, pH 7.8, 40 mM KCl, 8 mM MgCl<sub>2</sub>, 1 mM DTT, and H1 BSA – buffer H1 with addition of 0.02 mg/mL BSA. Radioautograph of 18% denaturing PAAG, showing the cleavage products of miR-17. miRNA (1 μM) and DC (20 μM) were incubated at 37°C for 48 h. Lanes Im and T1 – imidazole ladder and partial RNA digestion with RNase T1, respectively; C – miRNA was incubated in the absence of the conjugate in buffer H1. The incubation time (in hours) is indicated at the top.

**Table S5.** Sequences of miRNAs used in the study.

miRNA	Sequence 5'-3'
miRNA-21	UAGCUUAUCAGACUGAUGUUGA
miR-21_1	UAGCUUAUCAUACAGAUGUUGA
miRNA-17	CAAAGUGCUUACAGUGCAGGUAG
miRNA-18a	UAAGGUGCAUCUAGUGCAGUA
miRNA-155	UUAAUGCUGAAUUGUGAUAGGGGU



**Figure S12.** Stability of Dual conjugates (DCs) and corresponding Dual oligonucleotides (ONs) targeted to miR-155 and miR-21 in cell medium DMEM supplemented with 10% FBS. Images of the 12% PAAG/8M urea, stained with Stains-All and kinetics of degradation of DCs and ONs. Conjugates and oligonucleotides at a concentration of 0.1 mg/mL were incubated in DMEM supplemented with 10% FBS at 37 °C for 24 h.