

# A deeper investigation of drug degradation mixtures using a combination of MS and NMR data: application to indapamide

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## SUPPLEMENTARY MATERIALS

**Table S1**

Methods for the determination of detection and quantification limits of API for NMR and UV/MS experiments and their respective calculated values.

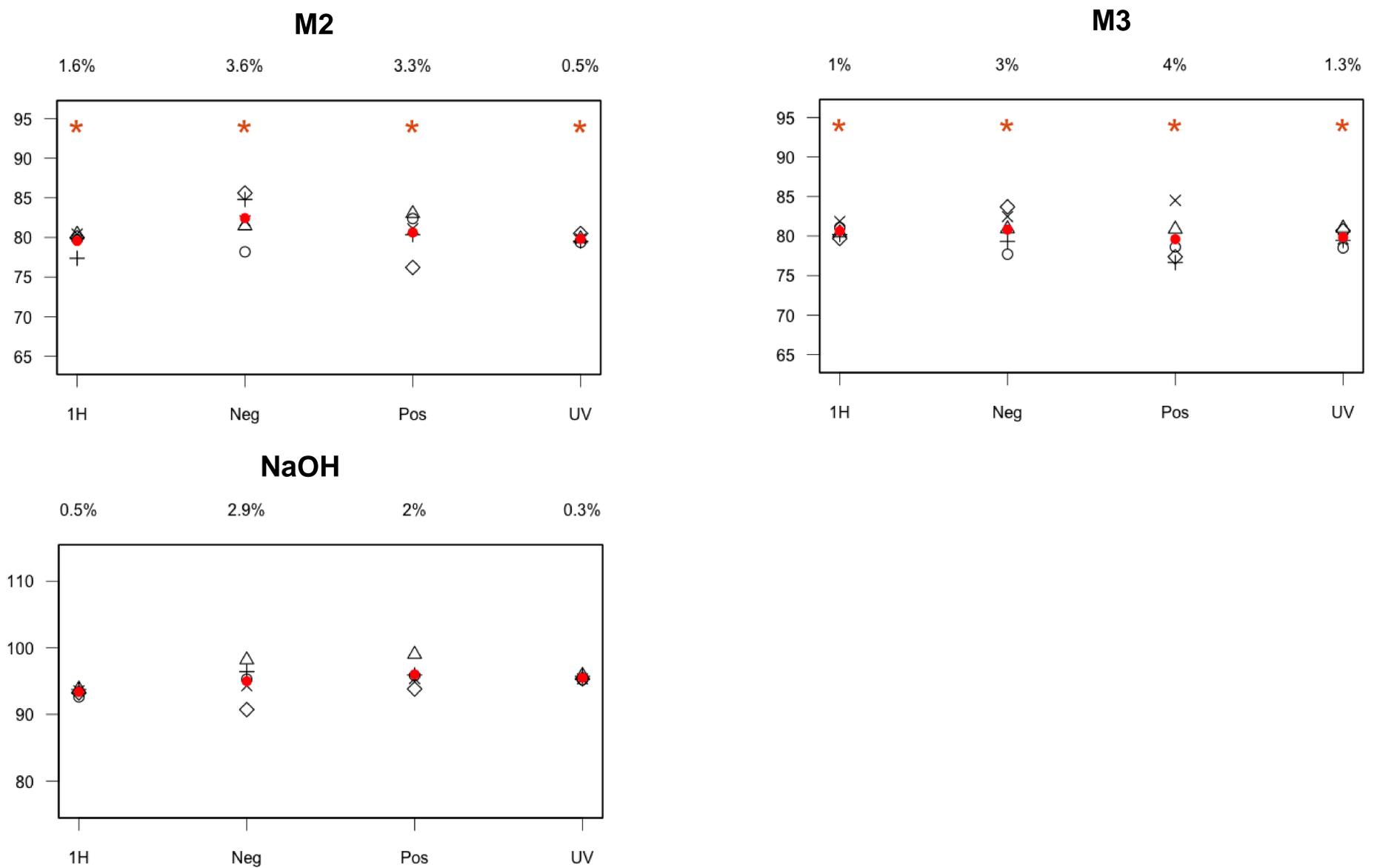
Calibration curves provided linear equations:  $y = a_0 + a_1x$ , where  $y$  is the response,  $x$  the API concentration,  $a_1$  the slope coefficient and  $a_0$  the intercept coefficient corresponding to noise.

$$\text{LOD} = 3.3 \times \frac{\text{Standard deviation of the intercept } (S_{a_0})}{\text{Slope of linear equation } (a_1)}$$

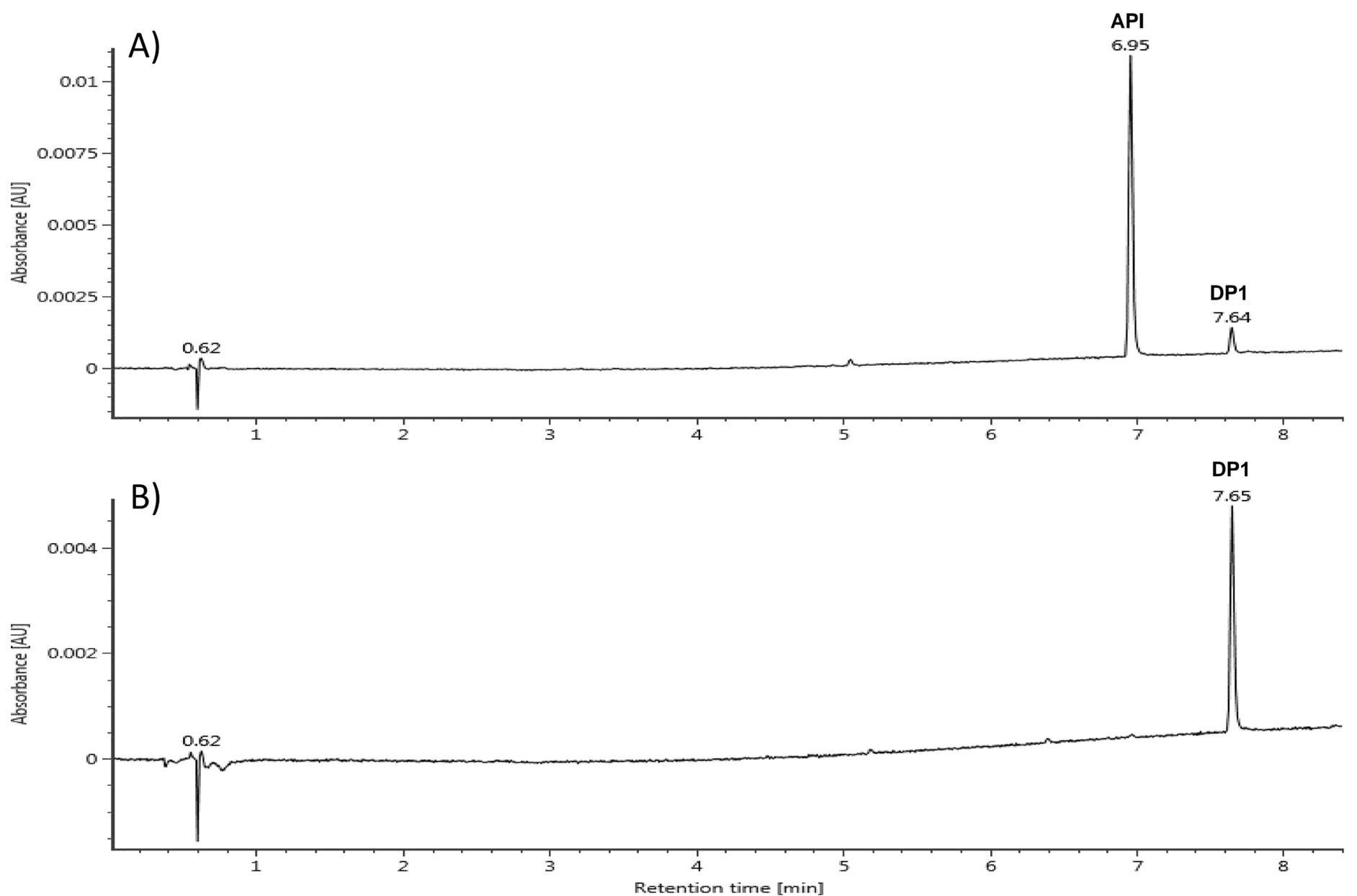
$$\text{LOQ} = 10 \times \frac{\text{Standard deviation of the intercept } (S_{a_0})}{\text{Slope of linear equation } (a_1)}$$

We calculated the lack of fit of the linear model using the R software and more particularly thanks to the `pure.error.anova` function of the `alr3` package. This test is based on an F-test (with  $\alpha = 0.05$ ): under the  $H_0$  hypothesis: there is no lack of fit in the linear regression model while under the  $H_1$  hypothesis: there is a lack of fit in the linear regression model.

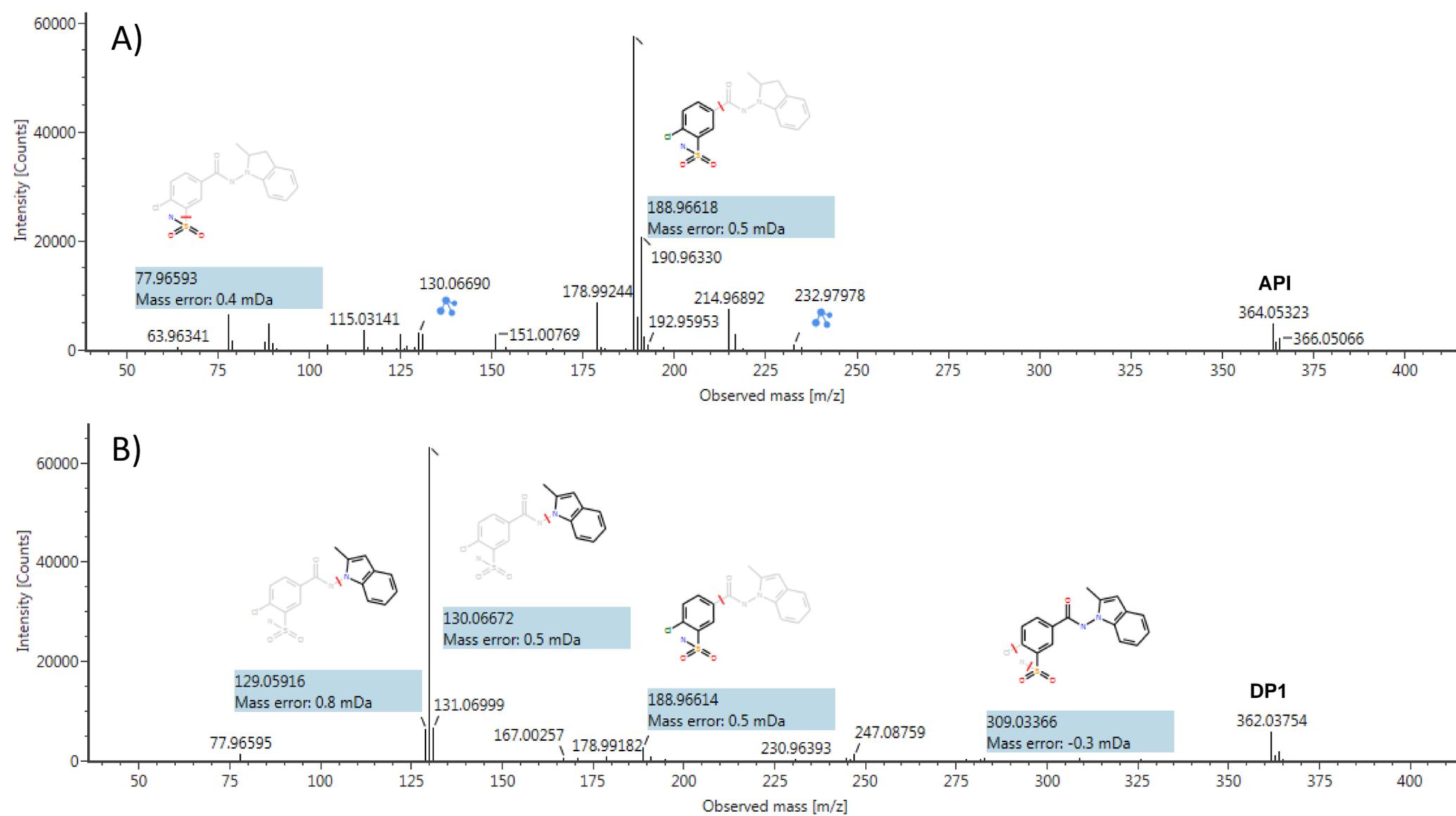
	Slope ( $\pm$ SD)	Intercept ( $\pm$ SD)	R <sup>2</sup>	p-value (F-test)	LOD (mM)	LOQ (mM)
UV	3.43E6 $\pm$ 3.66 E4	-3.08E3 $\pm$ 1.12E3	0.9961	0.4135	0.001	0.003
ESI+ MS	2.83E6 $\pm$ 3.32E4	-1.94 $\pm$ 1.02E3	0.9961	0.8608	0.001	0.004
ESI- MS	7.39E6 $\pm$ 7.82E4	-1.75E3 $\pm$ 7.39E4	0.9969	0.5840	0.001	0.003
<sup>1</sup> H NMR	1.84E-2 $\pm$ 1.15E-4	-2.21E-3 $\pm$ 6.99E-4	0.9989	0.2481	0.125	0.380



**Figure S1.** Quantification of API (%; Y axis) for M2, M3 and NaOH degradation with different methods ( $^1\text{H}$  NMR, ESI: negative and positive ion mode, UV at 275 nm; X axis) and for the 5 replicates ( $\times$ ,  $\diamond$ ,  $\circ$ ,  $+$ ,  $\Delta$  with  $\bullet$  represents the mean value). The coefficient of variation (CV) for each technique is indicated (top) and the significant of result is described by  $*$  for M2, M3 where theoretical values are available. NB: API degradation with Cu(II) is total.



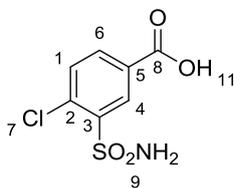
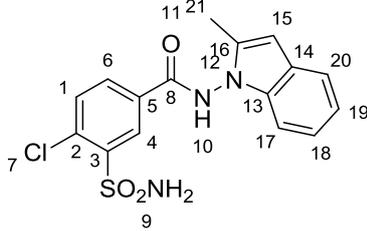
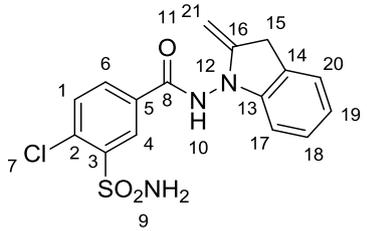
**Figure S2.** UV chromatograms of NaOH (A) and Cu(II) (B) degradations.



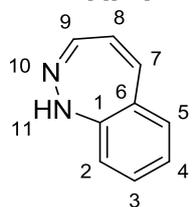
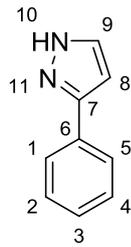
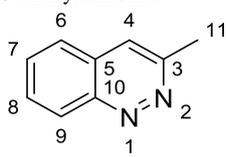
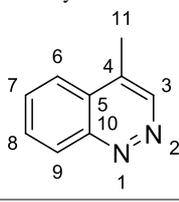
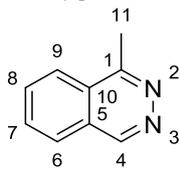
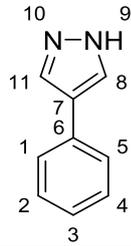
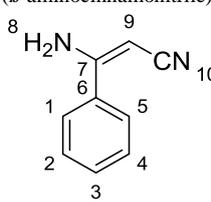
**Figure S3.** Negative HDMS<sup>E</sup> spectra of API (A) and DP1 (B).

**Table S2**

Structure hypotheses for DP1 and DP3 and predicted  $^1\text{H}/^{13}\text{C}$  NMR data (MNova).

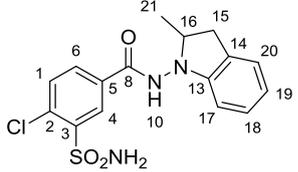
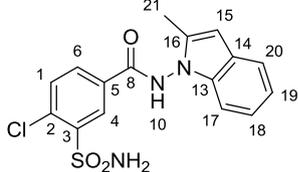
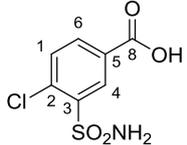
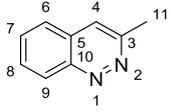
Compounds	Positions	$\delta\text{C}$ (ppm)	$\delta\text{H}$ (ppm)	Multiplicity, $J$ (Hz), nH	
DP3					
Tr (min) = 3.22					
	1	128.76	7.95	d, $J = 8.4$ Hz, 1H	
	2	129.77	-	-	
	3	136.57	-	-	
	4	129.73	8.47	d, $J = 1.9$ Hz, 1H	
	5	128.07	-	-	
	6	131.28	8.39	dd, $J = 8.3, 1.9$ Hz, 1H	
	8	166.27	-	-	
	11	-	-	-	
DP1					
Tr (min) = 7.67					
	1	131.09	7.92	d, $J = 8.2$ Hz, 1H	
	2	130.60	-	-	
	3	137.29	-	-	
	4	126.11	8.62	d, $J = 1.9$ Hz, 1H	
	5	131.91	-	-	
	6	130.71	8.26	dd, $J = 8.2, 1.9$ Hz, 1H	
	8	163.36	-	-	
	13	140.32	-	-	
	14	128.43	-	-	
	15	102.53	6.34	dh, $J = 2.0, 0.5$ Hz, 1H	
	16	142.36	-	-	
	17	111.61	7.50	ddt, $J = 6.2, 1.7, 0.5$ Hz, 1H	
	18	124.35	7.05	ddd, $J = 6.9, 6.2, 1.2$ Hz, 1H	
	19	120.63	7.11	dddd, $J = 7.4, 6.9, 1.6, 0.5$ Hz, 1H	
	20	121.21	7.22	dddd, $J = 7.4, 1.9, 1.2, 0.5$ Hz, 1H	
	21	11.61	2.29	d, $J = 0.5$ Hz, 3H	
		1	131.09	7.92	d, $J = 8.2$ Hz, 1H
		2	130.60	-	-
		3	137.29	-	-
		4	126.11	8.62	d, $J = 1.9$ Hz, 1H
		5	131.91	-	-
6		130.71	8.25	dd, $J = 8.2, 1.9$ Hz, 1H	
8		163.36	-	-	
13		141.72	-	-	
14		128.07	-	-	
15		29.12	3.22	q, $J = 1.0$ Hz, 2H	
16		150.33	-	-	
17		112.35	6.87	dd, $J = 7.8, 1.5$ Hz, 1H	
18		126.04	7.26	td, $J = 7.7, 2.4$ Hz, 1H	
19		125.02	6.59	td, $J = 7.7, 1.5$ Hz, 1H	
20		123.93	7.02	ddt, $J = 7.7, 2.4, 1.0$ Hz, 1H	
21		86.20	4.19	t, $J = 1.0$ Hz, 2H	

**Table S3**Structure hypotheses for DP5 and predicted  $^1\text{H}/^{13}\text{C}$  NMR data (MNova).

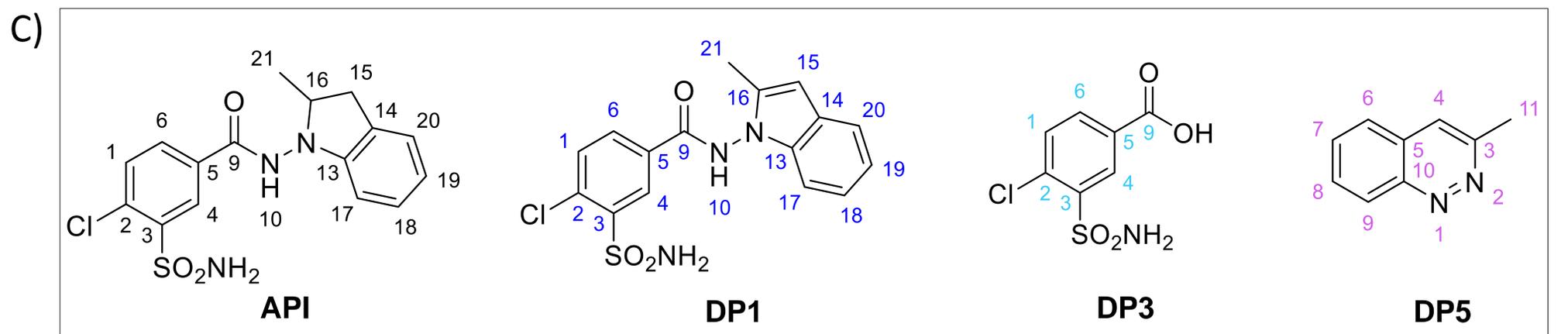
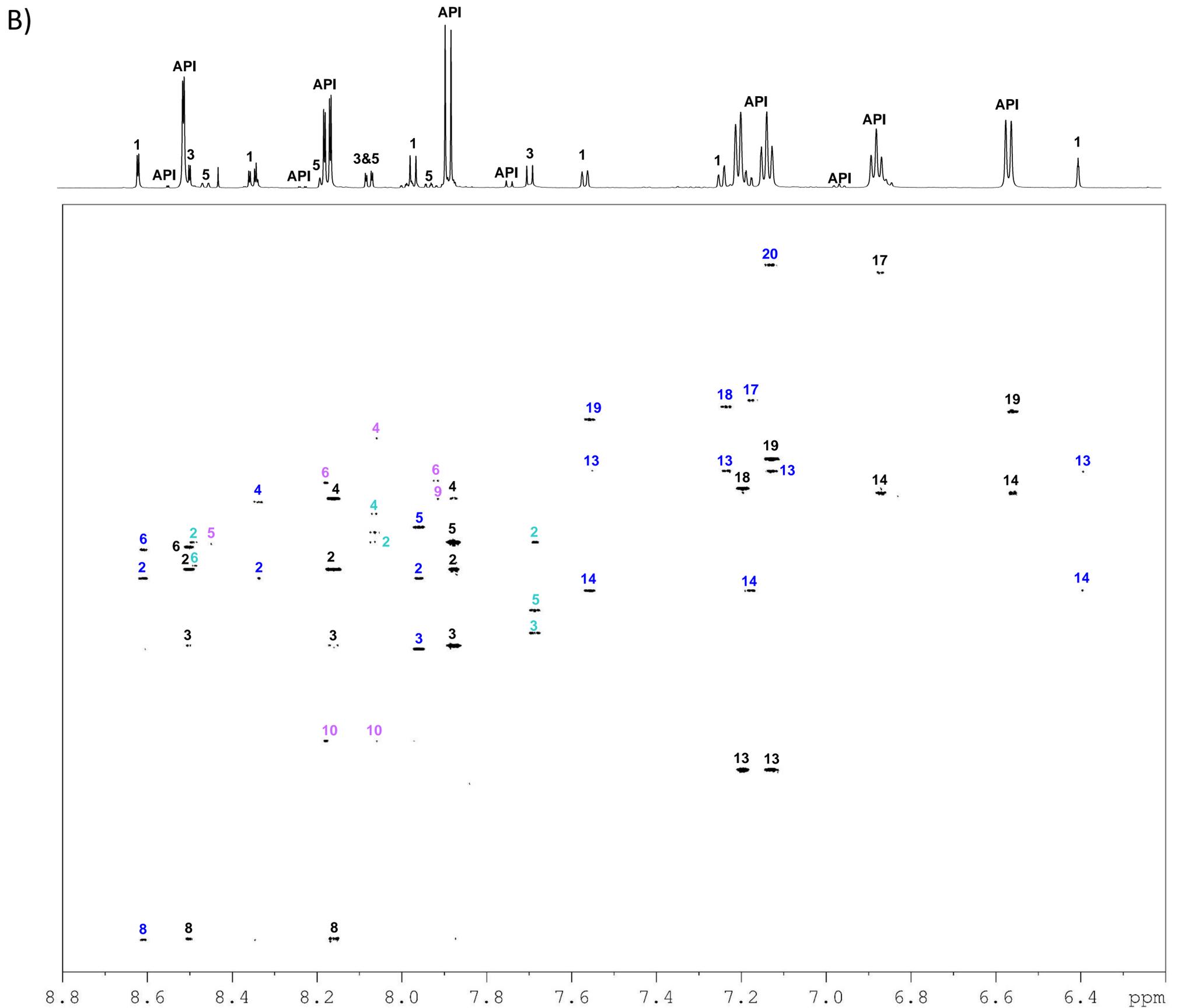
Compounds	Positions	$\delta\text{C}$ (ppm)	$\delta\text{H}$ (ppm)	Multiplicity, $J$ (Hz), nH
DP5	Tr (min) = 3.91			
1H-benzo[c][1,2]diazepine (benzodiazepine)	1	140.09	-	-
	2	118.51	7.30	dd, $J = 7.5, 1.5$ Hz, 1H
	3	126.31	7.19-7.13	m, 1H
	4	123.88	7.42	ddd, $J = 7.8, 7.1, 1.5$ Hz, 1H
	5	127.33	7.48	ddd, $J = 7.8, 1.6, 0.6$ Hz, 1H
	6	127.52	-	-
	7	130.63	6.90	ddd, $J = 10.7, 1.3, 0.6$ Hz, 1H
	8	127.72	5.30	dd, $J = 10.7, 8.9$ Hz, 1H
	9	144.17	7.94	dd, $J = 8.9, 1.3$ Hz, 1H
5-phenyl-1H-pyrazole	1	128.88	7.85-7.81	m, 1H
	2	127.83	7.47-7.39	m, 1H
	3	129.85	7.47-7.39	m, 1H
	4	127.83	7.47-7.39	m, 1H
	5	128.88	7.85-7.81	m, 1H
	6	131.80	-	-
	7	146.72	-	-
	8	102.45	6.53	d, $J = 2.5$ Hz, 1H
	9	131.76	7.51	d, $J = 2.5$ Hz, 1H
3-methylcinnoline	3	155.24	-	-
	4	121.95	7.49	dh, $J = 2.1, 0.5$ Hz, 1H
	5	130.07	-	-
	6	128.34	7.22	dddd, $J = 8.5, 2.2, 1.2, 0.5$ Hz, 1H
	7	129.55	7.72	dddd, $J = 8.4, 7.8, 1.2, 0.5$ Hz, 1H
	8	129.39	7.37	ddd, $J = 8.3, 7.8, 1.1$ Hz, 1H
	9	128.81	8.05	ddt, $J = 8.4, 1.1, 0.5$ Hz, 1H
	10	148.30	-	-
	11	21.80	2.08	d, $J = 0.5$ Hz, 3H
4-methylcinnoline	3	145.49	8.90	p, $J = 0.6$ Hz, 1H
	4	140.13	-	-
	5	128.54	-	-
	6	125.97	7.43-7.37	m, 1H
	7	134.41	7.81	ddd, $J = 8.1, 7.0, 1.2$ Hz, 1H
	8	129.10	7.43-7.37	m, 2H
	9	129.10	8.13	ddd, $J = 8.4, 1.2, 0.6$ Hz, 1H
	10	148.92	-	-
	11	18.90	2.39	d, $J = 0.6$ Hz, 3H
1-methylphthalazine	1	156.21	-	-
	4	149.30	9.64-9.60	m, 1H
	5	124.41	-	-
	6	123.94	8.25-8.19	m, 1H
	7	126.22	8.11-8.06	m, 1H
	8	127.49	8.11-8.06	m, 1H
	9	126.72	8.25-8.19	m, 1H
	10	126.96	-	-
	11	20.07	1.93	s, 3H
4-phenyl-1-H-pyrazole	1	126.61	7.60-7.54	m, 1H
	2	128.91	7.48-7.42	m, 1H
	3	127.69	7.42-7.36	m, 1H
	4	128.91	7.48-7.42	m, 1H
	5	126.61	7.60-7.54	m, 1H
	6	133.38	-	-
	7	124.95	-	-
	8	127.79	8.04	s, 1H
	11	132.51	8.04	s, 1H
3-amino-3-phenylacrylonitrile ( $\beta$ -aminocinnamionitrile)	1	127.47	7.72-7.66	m, 1H
	2	128.07	7.58-7.50	m, 1H
	3	128.29	7.58-7.50	m, 1H
	4	128.07	7.58-7.50	m, 1H
	5	127.47	7.72-7.66	m, 1H
	6	132.62	-	-
	7	163.72	-	-
	9	58.52	4.19	s, 1H
	10	117.19	-	-

**Table S4**

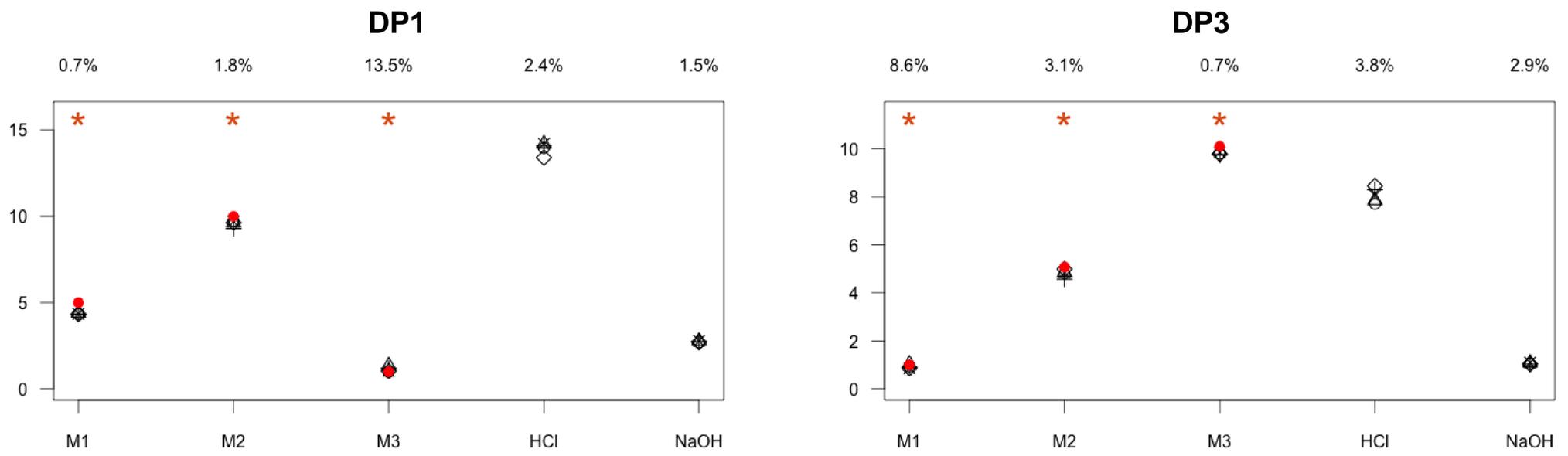
Experimental NMR assignment of API and its degradation products (DPs) in HCl mixture. The selected proton for qNMR is described by \*.

	Positions	$\delta C$ (ppm)	$\delta H$ (ppm)	Multiplicity, $J$ (Hz), nH	
<b>API (S1520)</b> 	1	134.02	7.88	d, $J = 8.3$ Hz, 1H	
	2	135.87	-	-	
	3	142.37	-	-	
	4	129.94	8.50	d, $J = 2.2$ Hz, 1H	
	5	133.63	-	-	
	6	134.03	8.16	dd, $J = 8.3, 2.2$ Hz, 1H	
	8	167.19	-	-	
	13	152.85	-	-	
	14	129.43	-	-	
	15	37.09	2.66	dd, $J = 11.2, 15.6$ Hz, 1H	
	15*	37.09	3.21	dd, $J = 8.1, 15.6$ Hz, 1H	
	16	-	3.95	s, 1H	
	17*	110.80	6.56	d, $J = 7.7$ Hz, 1H	
	18	129.06	7.12	t, $J = 7.7$ Hz, 1H	
	19	122.58	6.87	t, $J = 7.4$ Hz, 1H	
	20	126.57	7.20	d, $J = 7.4$ Hz, 1H	
	21	19.93	1.35	d, $J = 6.2$ Hz, 3H	
	<b>DP1 (Y38)</b> 	1	134.33	7.96	d, $J = 8.3$ Hz, 1H
		2	136.78	-	-
		3	142.64	-	-
		4*	129.04	8.61	d, $J = 2.2$ Hz, 1H
5		132.38	-	-	
6		134.37	8.34	dd, $J = 8.3, 2.2$ Hz, 1H	
8		167.29	-	-	
13		127.60	-	-	
14		137.39	-	-	
15		100.59	6.40	t, $J = 0.9$ Hz, 1H	
16		138.95	-	-	
17		121.64	7.56	d, $J = 7.7$ Hz, 1H	
18		122.14	7.13	td, $J = 7.1, 0.9$ Hz, 1H	
19	123.25	7.18	td, $J = 7.1, 0.9$ Hz, 1H		
20	110.08	7.24	d, $J = 8.0$ Hz, 1H		
21	12.86	2.33	d, $J = 0.9$ Hz, 3H		
<b>DP3 (Y36)</b> 	1*	133.06	7.69	d, $J = 8.2$ Hz, 1H	
	2	133.54	-	-	
	3	141.27	-	-	
	4	131.22	8.49	d, $J = 2.0$ Hz, 1H	
	5	139.36	-	-	
	6	135.61	8.07	dd, $J = 8.2, 2.0$ Hz, 1H	
	8	170.88	-	-	
	<b>DP5</b> 	3	156.00	-	-
4		124.85	8.18	s, 1H	
5		133.67	-	-	
6		128.60	8.06	d, $J = 0.9$ Hz, 1H	
7		133.81	7.92	d, $J = 0.4$ Hz, 1H	
8		132.81	7.98	td, $J = 6.7, 1.2$ Hz, 1H	
9*		129.92	8.45	dd, $J = 8.5, 0.7$ Hz, 1H	
10		150.47	-	-	
11		23.02	2.92	d, $J = 0.4$ Hz, 3H	

A) NMR spectrum was recorded on Bruker Avance NEO 900 MHz spectrometer with a 5-mm TCI cryoprobe. The 2D HMBC (Heteronuclear Multiple-Bond Correlation spectroscopy) spectrum was acquired with the Bruker library *hmbcetgpl3nd*, with a 2 s relaxation delay using 64 scans per 8 K increments, which were collected into 4 K data points, using spectral widths of 9090 Hz in F2 and 45276 Hz in F1. The number of NUS sampling points was 256 complex points (3.125 % sampling density of 8 K points).



**Figure S4.** Complete NMR assignment of indapamide, DP1, DP3 and DP5 (labeled API, 1, 3 and 5) for HCl hydrolysis: A) Acquisition parameters; B) 2D HMBC spectrum with correlations  $^1\text{H}$ - $^{13}\text{C}$  of each compounds; C) Structures and numbering of each compounds.



**Figure S5.** Quantification of DP1 and DP3 (in Y axis) in reconstitute mixtures and acid/alkaline degradations with qNMR methods for the pools ( $\times$ ,  $\diamond$ ,  $\circ$ ,  $+$ ,  $\Delta$  with  $\bullet$  represents the mean value). In the top of each figure, the coefficient of variation were calculated and the significant of result was described by  $*$ .